



## (1→3)- $\beta$ -D-Glucan: A Biomarker for **Microbial Translocation in Individuals** with Acute or Early HIV Infection?

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Background: The extent of gut microbial translocation, which plays roles in HIV disease progression and non-AIDS comorbidities, appears to vary with the composition of the gut microbiome, particularly the presence of Lactobacillales, which reduce mucosal injury. While low proportions of Lactobacillales in the distal gut microbiome are a very promising indicator of microbial translocation, measurement is expensive and complicated and not feasible for clinical routine.  $(1\rightarrow 3)$ - $\beta$ -D-Glucan (BDG) is a component of most fungal cell walls and might be a surrogate marker for Lactobacillales proportion in the gut and a useful indicator of HIV-associated gut injury. This study evaluated BDG as a biomarker of gut integrity in adults with acute or early HIV infection (AEH).

Methods: Study samples were collected longitudinally during study visits at weeks 0, 12, and 24 in a cohort of 11 HIV-infected men starting antiretroviral therapy during AEH. Blood plasma levels of BDG, soluble cluster of differentiation 14 (sCD14) and lipopolysaccharide (LPS) were measured and then correlated with the proportion of Lactobacillales in the distal gut microbiome, as measured by 16s rDNA sequencing by using mixed-effects models with random intercepts.

Results: Mean BDG and sCD14 levels across subjects were associated with Lactobacillales after controlling for time effects and within-subjects correlations (p-values < 0.05), while LPS levels were not. Specifically, each point increase in mean BDG and sCD14 levels across participants was associated with 0.31  $\pm$  0.14 and 0.03  $\pm$  0.01 percent decrease in mean Lactobacillales proportions, respectively.

Conclusion: BDG and sCD14 may be indicators of low Lactobacillales in the gut in adults with acute or early HIV infection, and serve as biomarkers of gut integrity and microbial translocation in HIV infection. Larger studies are needed to confirm our findings.

Keywords: beta-p-glucan, HIV, microbial translocation, Lactobacillales, microbiome, acute HIV infection

1

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#### INTRODUCTION

CD4+ T cells trigger many elements of the immune response, including regulation of CD8+ T cell activation (1). Early HIV infection is characterized by a dramatic depletion of CD4+ T cells and impaired polarization of Th17 cells in the gastrointestinal tract and a massive expansion of activated CD8+ T cells causing CD8+ T cell-mediated enteropathy (2-4), which is also characterized by microbial overgrowth and translocation of microbial products, including bacteria, fungi, and viruses, from the gut into the systemic circulation (5). Lactobacillales influence gut mucosal immunity by increasing the suppressive function of CD4+ regulatory T cells of colon lamina propria, which can alleviate HIV-associated colitis (1). High proportions of Lactobacillales in the distal gut microbiome appear to reduce mucosal injury, lower inflammatory responses, reduce barrier disruption (6, 7), and may result in less microbial translocation during HIV infection (4, 8, 9). Low proportions of Lactobacillales may, therefore, serve as an indicator of microbial translocation, which likely plays a role in HIV disease progression and non-AIDS comorbidities, even when antiretroviral treatment is initiated early in the course of infection (10-12). However, determination of gut Lactobacillales proportion is expensive and complicated. Surrogate markers for Lactobacillales proportion in the gut microbiome are, therefore, needed for the clinical routine.

The polysaccharide  $(1\rightarrow 3)$ - $\beta$ -D-glucan (BDG) is a cell wall component of most fungal species, and is used as a serum biomarker for early diagnosis of invasive fungal infections (13–15). BDG is not highly specific for fungal infections, however, and among individuals with HIV infection but without invasive fungal infection, elevated levels of BDG in serum correlate with HIV-associated immunosuppression, inflammation, and cardiopulmonary comorbidity (16, 17). Consistent with these links, a recent report indicates that BDG may be a promising biomarker for neurocognitive impairment in virally suppressed HIV-infected adults (18). The pathogenic mechanism behind this finding remains unclear.

We hypothesized that in the absence of invasive fungal infection, BDG may be a biomarker of gut mucosal barrier disruption (19, 20) and microbial translocation (21), potentially contributing to HIV-associated morbidity (22). The objective of this pilot study was to evaluate BDG as a marker of gut permeability by correlating blood BDG with proportions of *Lactobacillales* in the distal gut microbiome of individuals diagnosed with acute and early HIV infection.

#### MATERIALS AND METHODS

In this longitudinal observational analysis, we retrospectively measured levels of BDG in blood plasma samples in a cohort of adults with acute and early HIV infection, and compared BDG levels and established biomarkers of microbial translocation with 16s rDNA sequencing of the gut microbiome.

All 13 individuals participated in the San Diego Primary HIV Infection Research Consortium (SD PIRC), which is composed of individuals diagnosed with a cuteorearly HIV-infection (acute: HIV nucleic acid amplification testing+/antibody- consistent with

infection <30 days; early: HIV antibody+/detuned HIV Ab consistent with infection <70 days) followed longitudinally (23–25). The UCSD Human Research Protections Program approved the study protocol, consent document, and all study procedures. All participants provided voluntary, written informed consent. All participants started ART within 2 weeks of study enrollment with a combination of tenofovir, emtricitabine, and ritonavir-boosted atazanavir, with or without maraviroc according to a randomization schedule at entry (baseline). Paired anal swabs [anal swabs have been shown to produce highly reproducible microbiota profiles resembling the human gut microbiota (26)] and blood samples were collected sequentially during study visits at baseline and at weeks 12 (±2 weeks) and 24 (±6 weeks) between August 2010 and September 2011 at the University of California, San Diego and stored at -80°C on the day of collection (4).

 $(1\rightarrow 3)$ -β-D-glucan was measured by the Fungitell assay in June 2015 at the Associates of Cape Cod, Inc., research laboratories (Associates of Cape Cod, Inc, East Falmouth, MA, USA). Both comparator biomarkers were measured in 2012 and have been – in part – published before (4). Soluble cluster of differentiation 14 (sCD14) was measured by immunoassay (Quantikine, R&D Systems, Minneapolis, MN, USA). Lipopolysaccharide (LPS) was measured by the Limulus Amebocyte Lysate QCL-1000 assay.

DNA extractions from anal swabs were performed in 2012 using the QIAamp Stool DNA kit (Qiagen) (4). Amplification of bacterial DNA and pyrosequencing amplification of the V6 hypervariable region of the 16 S rDNA gene was carried out in a 50  $\mu l$  reaction using the highly purified Amplitaq Gold Low DNA polymerase (Applied Biosystems, Foster City, CA, USA) (4). For classification of bacteria, we kept bacterial sequences with at least 90 continuous base pairs with a quality score of at least 20 for further analyses as described before (4).

Statistical analyses were performed using SPSS 22 (SPSS Inc., Chicago, IL, USA). As study samples were collected longitudinally (i.e., repeated measures) we applied mixed-effects models with random intercepts to test whether Lactobacillales proportions were associated with between-subjects (Bw) as well as withinsubjects (Wi) BDG levels after adjustment for repeated measures. The same analysis was repeated for other biomarkers (sCD14, LPS, and CD4+ T cell counts). We used mixed-effects models to analyze our repeated-measures data because its maximum-likelihood estimation of the missing values allows for retaining the participants with missing values. In addition, the mixed-effects model approach increases power to detect effects and allows for a more accurate estimate of correlations. In addition, we also calculated cross-sectional correlations between BDG, Lactobacillales and other biomarkers using Pearson correlation coefficient for weeks 12 and 24 and Spearman correlation coefficient for baseline due to skewed distributions at week 0 (i.e., secondary analysis).

#### **RESULTS**

Thirteen men diagnosed with acute (8/13) or early (5/13) HIV infection enrolled. Median age was 29 years (range 21–55 years). About a third (4/13) reported Hispanic ethnicity, the rest reporting White (7/13) or Asian race (2/13). Eleven of the 13 men completed their 12- and 24-week visits. All 11 initiated ART

between baseline and week 12, 5/11 (45%) achieving viral suppression (i.e. <50 copies/mL) by week 12, and 8/11 (73%) by week 24. Participants who were not virally suppressed (i.e., HIV RNA >50 copies/mL) at week 12 (6/11) showed a trend for higher BDG levels in plasma (median 52 pg/mL, range 12–122 pg/mL) when compared to those who were virally suppressed (median 9 pg/mL, range 3–46 pg/mL; p = 0.052, Mann–Whitney U test), while there was no difference at week 24. In addition, no difference in the levels of BDG was observed between participants with and without maraviroc (n = 7; median 39, range 13–96 vs. n = 6; median 39, range 20–89; p > 0.20 at all timepoints). At baseline, BDG levels did not differ between participants with early HIV infection (median 63 pg/mL, range 13–89 pg/mL) and those with acute HIV infection (median 37 pg/mL, range 20–96 pg/mL).

Levels of clinical and immunological variables as well as the correlation analyses for each time point separately are depicted in **Table 1**. The median proportion of *Lactobacillales* in the distal gut was 41.5% [range 5.2–52.1%; n=11 (two individuals who received systemic antibacterial therapy were excluded)] at baseline, 26.4% (range 6.2–41.7%; n=11) at week 12, and 22.8% (range 3.0–47.9%) at week 24. While in cross-sectional analysis, there was no correlation between levels of BDG and *Lactobacillales* at baseline, there was a negative correlation at week 12 and a trend at week 24. Scatter plots showing correlations of blood BDG levels and proportions of *Lactobacillales* at weeks 12 and 24 are displayed in **Figure 1**.

In the mixed-effects model, higher proportions of *Lactobacillales* were associated with lower Bw BDG (p = 0.023, **Table 2**) and

TABLE 1 | Levels of plasma beta p-glucan (BDG) and cross-sectional correlations with other biomarkers, viral loads, and *Lactobacillales* proportions of distal gut bacterial flora (median and range are displayed) at baseline, week 12, and week 24.

Biomarkers	Baseline ( $n = 13$ )			Week 12 (n = 11)			Week 24 (n = 11)		
	Results [median (range)]	Spearman correlation with BDG	p-Value	Results [median (range)]	Pearson correlation with BDG	p-value	Results [median (range)]	Pearson correlation with BDG	•
BDG (pg/mL)	39 (13–96)	_	-	45 (3–122)	_		35 (3–89)		
LPS (pg/mL)	0.358 (0.278-0.409)	0.275	n.s.	0.306 (0.278-0.393)	0.263	n.s.	0.311 (0.183-0.411)	-0.241	n.s.
sCD14 (ng/mL)	1,084 (706-2,683)	0.363	n.s	1,290 (728-1,844)	0.352	n.s.	981 (858-1,493)	-0.192	n.s.
CD4+ cell count (cells/µL)	520 (91–971)	-0.126	n.s.	822 (245–1,408)	-0.363	n.s.	793 (262–1,150)	-0.456	n.s.
CD8+ cell count (cells/µL)	856 (558–5,216)	-0.357	n.s.	813 (436–2,196)	0.044	n.s.	774 (588–1,906)	-0.009	n.s.
CD4/CD8 ratio	0.576 (0.101-1.279)	-0.055	n.s.	1.077 (0.267-1.372)	-0.475	n.s.	0.888 (0.382-1.070)	-0.493	n.s.
Viral loads (log10 RNA)	5.25 (3.10-7.00)	0.055	n.s.	1.914 (1.623-4.153)	0.147	n.s.	1.681 (1.681–2.702)	0.113	n.s.
Lactobacillales (%)	41.49 (5.15-52.12; n = 11)	-0.109	n.s.	26.40 (6.16-41.68)	-0.639	0.034	22.85 (3.04-47.94)	-0.542	0.085

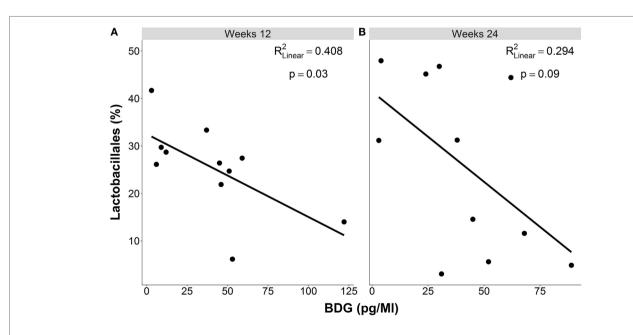


FIGURE 1 | Cross-sectional associations between levels of beta-p-glucan and proportions of *Lactobacillales* in the distal gut (A) at week 12 (B) at week 24 of follow up in individuals with acute or early HIV infection.

TABLE 2 | Summary of mixed-effects models for BDG and other biomarkers predicting Lactobacillales proportions.

	BDG		LPS		sCD14		CD4 <sup>+</sup> cell count	
	β	SMD	β	SMD	β	SMD	β	SMD
(Intercept)	30.47 (4.05)***	3.5	31.04 (4.57)***	3.3	31.91 (4.11)***	3.7	31.12 (4.48)***	3.3
Time	-0.19 (0.23)	0.4	-0.26 (0.25)	0.5	-0.29 (0.23)	0.6	-0.22 (0.27)	0.4
Bw	-0.31 (0.14)*	1.4	-29.03 (79.42)	0.2	-0.03 (0.01)*	1.3	0.02 (0.01)†	1.0
Wi	0.05 (0.13)	0.2	-5.61 (86.71)	0.0	-0.01 (0.01)	0.5	0.00 (0.02)	0.0

Bw, between-subjects effect; Wi, within-subjects effect, SMD, standardized mean difference effect-size measure (>0.8 = large effect). Bw and Wi are centered at the grand mean and subject means, respectively. The value inside the parentheses indicates the SE for  $\beta$ . †p < 0.10, \*p < 0.05, \*\*\*p < 0.001 based on likelihood-ratio tests.

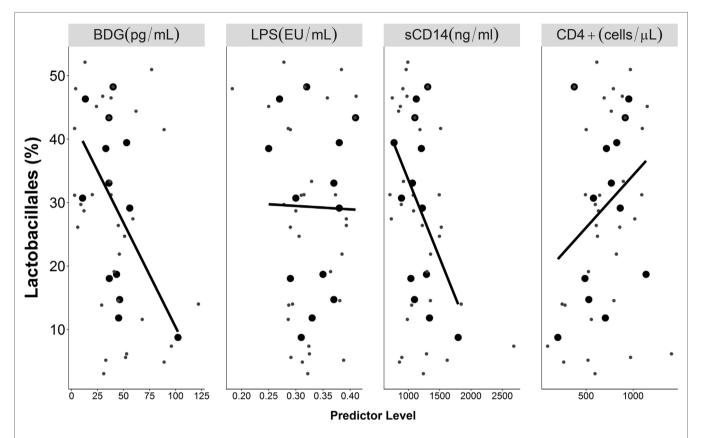


FIGURE 2 | Longitudinal associations of BDG, LPS, and sCD14 with proportions of *Lactobacillales* proportion in the distal gut. Big dots indicate mean levels for each subject, and small dots indicate individual observations. Black lines indicate linear regression lines fitted to the mean values, while gray lines indicate non-linear smooth lines.

lower Bw sCD14 (p=0.035). Specifically, each point increase in mean BDG and sCD14 levels across participants were associated with  $0.31\pm0.14$  and  $0.03\pm0.01$  percent decrease in mean Lactobacillales proportions, respectively. These two predictors (BDG and sCD 14) were highly correlated (r=0.69, p=0.01). Lactobacillales was not associated with Bw LPS (p>0.2) nor with Bw CD4+ cell count (p=0.095). None of the time or Wi effects showed significant associations (p-values >0.2). Scatter plots of mean Wi BDG, LPS, and sCD14 levels predicting Lactobacillales proportion in the distal gut are displayed in Figure 2. The big dots are means across longitudinal observations, representing more stable estimates of predictor levels and proportions of Lactobacillales. The regression lines fit to those means and, therefore, represent stable estimates of correlations. The small dots indicate individual observations.

### DISCUSSION

This is the first study to evaluate blood levels of the fungal polysaccharide BDG in individuals with acute or early HIV infection. We found that higher levels of blood BDG correlated with higher levels of sCD14 and lower proportions of gut *Lactobacillales*. High proportions of *Lactobacillales* in the distal gut microbiome have been shown to reduce barrier disruption (6, 7), and result in less microbial translocation during HIV infection (4, 8, 9), even when administered as a probiotic treatment (27). Correlation of BDG with high levels of sCD14 and low proportions of *Lactobacillales* may, therefore, suggest that BDG may be a biomarker of gut integrity and microbial translocation in individuals with acute or early HIV infection.

Our findings may have important clinical implications, as microbial translocation may be a major contributing factor to HIV-associated morbidity (22, 28). Two previous studies found that blood BDG levels correlated directly with non-AIDS comorbidities among individuals with chronic HIV infection. Morris and colleagues found that high blood BDG was associated with HIV-associated immunosuppression, inflammation (plasma interleukin-8 and tumor necrosis factor- $\alpha$ ), and cardiopulmonary comorbidity in 132 HIV-infected outpatients, the majority of whom had measurable HIV RNA in their blood (17). Our study team recently reported that higher blood BDG levels correlated with worse neurocognitive performance in 19 virologically suppressed HIV-infected adults (18). While both studies explained their findings with the hypothesis that elevated plasma BDG may primarily reflect translocation of products from endogenous fungal flora from the gastrointestinal tract into systemic circulation, this is the first study that actually evaluated BDG as a biomarker of the composition of the gut microbiome, namely Lactobacillales (4, 6-8, 22, 29, 30).

In this study, the mean levels of BDG and sCD14 across subjects correlated strongly and were significantly associated with *Lactobacillales* after controlling for time effects and withinsubjects correlations, while no significant correlations were found between LPS and *Lactobacillales*. The lack of associations of LPS with other biomarkers of microbial translocation may be explained by the fact that currently available LPS assays rely on limulus amebocyte lysate (LAL) reagents, which are not specific for endotoxin and may be subject to cross-reactivity (31, 32).

Our results indicate that BDG correlates with sCD14 and may be an indicator of low *Lactobacillales* in the gut and increased microbial translocation in adults with acute or early HIV infection. This theory is supported by a number of studies reporting positive BDG levels during hemodialysis (33, 34), most likely explained by transient reduced blood flow within the splanchnic region (35), potentially resulting in ischemia and a transient barrier damage in the gut. By contrast, another recent study did not find elevated serum BDG levels in hematological malignancy patients suffering for mild to moderate mucositis (36).

We also observed a trend toward higher BDG levels in those who were not virologically suppressed at week 12. This finding is in line with results from Morris and colleagues who found that high BDG levels were associated with high viral loads and also low CD4+ cell counts among chronically infected individuals (17). While elevated blood BDG levels may be associated with microbial translocation in all HIV-infected individuals (i.e., independent of CD4+ cell counts), interpretation of elevated blood BDG levels in individuals with CD4+ cell counts below 200–300 cells/µL may be more complicated. While it seems intuitive that deteriorating CD4+ counts are associated with worse mucosal barrier function (21), other reasons for elevated BDG may include potential colonization or subclinical infection with *Candida* spp. or *Pneumocystis* (15) that may occur more frequently in individuals with lower CD4 counts.

Major limitations of this small study include its small sample size, its cross-sectional, single-site design, and its focus on adults with acute or early HIV infection. While the mechanistic hypothesis of the correlation between *Lactobacillales* proportion

in the gut and microbial translocation has been evaluated in a number of studies, very few studies to date have actually reported that the proportion of Lactobacillales may be an indicator of microbial translocation, and the most important study showing this correlation used in part the same samples that were used in this study. Also BDG has been primarily used as a biomarker for fungal infection and BDG cut-off levels for microbial translocation have yet not been defined. However, our study cohort of individuals with AEH presented with markedly higher BDG levels when compared to previously published levels from healthy individuals undergoing elective plastic surgery procedures (37). In addition, cross-sectional analyses of this longitudinal data were limited by particularly small sample sizes. Another explanation for the absence of correlation between BDG levels and Lactobacillales in cross-sectional analysis at baseline may be the fact that Lactobacillales proportions were markedly lower at week 12 and week 24 of follow-up when compared to baseline (median 22-26% vs. median 41%). The clear negative correlations between BDG and Lactobacillales and positive correlations with sCD14 levels in the mixed-effects model suggest nevertheless that BDG may be an indicator of gut mucosal barrier interruption and microbial translocation. To further examine the role of BDG as a potential biomarker for translocation of gut luminal contents, more comprehensive studies will be necessary. BDG levels in the intestinal luminal contents are also likely to be highly variable on an individual basis, and a standardized oral BDG challenge approach may be more suitable for assessing gut integrity.

In conclusion, high BDG levels may be a useful indicator of low *Lactobacillales* in the gut and microbial translocation in individuals with acute and early HIV infection. These findings indicate that BDG may be a promising marker for gut integrity. Larger studies are needed to confirm our findings.

#### **AUTHOR CONTRIBUTIONS**

MH had the study idea. JP-S, MO, DS, and SG provided the samples for the study. All authors (MH, JP-S, MO, YZ, MF, SL, DS, and SG) contributed to the study design. YZ and MF tested the samples. MH, JS, and SG conducted the data analysis and interpretation. MH, JS, and SG prepared the manuscript. MO, YZ, MF, SL, and DS revised it critical for intellectual contact. All authors have given final approval of the version to be published.

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