# The genes associated with early-onset Alzheimer's disease

### Meng-Hui Dai<sup>1,\*</sup>, Hui Zheng<sup>2,\*</sup>, Ling-Dan Zeng<sup>1</sup> and Yan Zhang<sup>1</sup>

Department of Neurology, Union Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan, 430022, China

<sup>2</sup>Department of Radiology, Union Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan, 430022, China

<sup>\*</sup>These authors contributed equally to this work

Correspondence to: Ling-Dan Zeng, email: 34187746@qq.com Yan Zhang, email: 799896551@qq.com

Keywords: Alzheimer's disease; early-onset AD; APP; PSEN1; PSEN2

Received: June 06, 2017 Accepted: October 14, 2017 Epub: December 15, 2017 Published: March 13, 2018

**Copyright:** Dai et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License 3.0 (CC BY 3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

#### ABSTRACT

Alzheimer's disease (AD) is a progressive neurodegenerative disorder that accounts for the most cases of dementia, which is characterized by the deposition of dense plaques of amyloid beta (A $\beta$ ) plaques and neurofibrillary tangles consisting of hyperphosphorylated tau. The two main types of AD can be classified as early-onset AD (EOAD, onset < 65 years) and late-onset AD (LOAD, onset  $\geq$  65 years). Evidence from family and twin studies indicate that genetic factors are estimated to play a role in at least 80% of AD cases. The first milestone with linkage analysis revealed the mutations in *APP*, *PSEN1*, and *PSEN2* genes that cause EOAD. But pathogenic mutations in these three genes can only explain a small fraction of EOAD families. The additional disease-causing genes have not yet been identified. This review provides an overview of the genetic basis of EOAD and the relationship between the functions of these risk genes and the neuropathologic features of AD. A better understanding of genetic mechanisms underlying EOAD pathogenesis and the potentially molecular mechanisms of neurodegeneration will lead to the development of effective diagnosis and treatment strategies for this devastating disease.

### **INTRODUCTION**

Alzheimer's disease (AD) is a neurodegenerative disease and the most common form of dementia in elderly people, leading to progressive and widespread damage to the brain and, ultimately, death [1]. Worldwide, 47.5 million people are living with dementia, and nearly 7.7 million new diagnoses are made every year [2]. The total number of people with dementia is projected to 75.6 million by 2030 and almost tripling to 135.5 million by 2050 [2]. The alarming rise of AD will impose a mounting financial and social burden worldwide. We urgently need to development of effective treatment strategies to prevent, delay the onset, slow the progression, or improve the symptoms of AD [3].

The key pathological changes observed in AD brain tissue are amyloid beta  $(A\beta)$  peptide deposited and neuritic plaques, hyperphosphorylated tau protein

and neurofibrillary tangles [4, 5]. Additional changes include microgliosis and loss of neurons, white matter and synapses. At present, the exact etiology of AD is unclear. AD is considered as a complex disease, resulted from the complicated interactions between the multiple factors, such as the age, education, genetic, and environmental factors. The familial aggregation of AD shows that genetic factors may play a vital essential role in the development of the disease [6-8]. Based on its age of onset, AD is classified into early-onset AD (EOAD, onset < 65 years) and late-onset AD (LOAD, onset  $\ge 65$  years). The proportion of EOAD in all AD cases is between 5% and 10%. LOAD is a highly polygenic disease. By contrast, EOAD or autosomal dominantly inherited AD is substantially or even entirely genetically determined frequently associated with genetic causes [7]. EOAD is generally associated with a more rapid rate of progression, so better understanding of genetic mechanisms underlying

EOAD pathogenesis will lead to the development of effective diagnosis and treatment strategies.

Three causal genes, which encode proteins involved in amyloid precursor protein (APP) breakdown and A $\beta$  generation, have been identified so far for EOAD, including the APP gene on chromosome 21 [9], the presenilin 1 (PSENI) gene on chromosome 14 [10], and presenilin 2 (PSEN2) on chromosome 1 [11]. AD-linked mutations in these three genes exhibit high penetrance (>85%), are mostly autosomal dominantly inherited, and lead with certainty to early-onset disease. Consequently, they are considered 'diagnostic biomarkers' of the disease [5]. However, mutations in these three genes can only explain a small fraction (5 to 10%) of EOAD cases [7, 12]; over 50% of Mendelian cases and most of sporadic EOAD cases remain genetically unexplained [7, 13–15]. In this review, we provide a summary of the genetic basis of EOAD, the relationship between these risk genes and the pathogenesis of EOAD.

### The genetic architecture of EOAD

### APP

### The gene and protein structure of APP

The gene encoding APP is located on chromosome 21q21.3. It is a highly conserved gene containing 18 exons and spanning 290 kilobases [16]. APP protein is a ubiquitously expressed single-pass type I transmembrane protein, with a large extra-membranous N-terminal region, a single transmembrane domain and a small cytoplasmic C-terminal tail [17, 18]. APP proteins range from 365 to 770 amino acids due to different splicing isoforms [9, 19].

Alternative splicing of transcripts from the single APP gene results in several isoforms of the gene product. APP695, APP751 or APP770 are the most common APP isoforms [20, 21]. APP695 is preferentially expressed in the central nervous system. APP751 and APP770 expressed both in the peripheral and central nervous systems. The ratios of APP770 mRNA and APP770plus-APP751 mRNAs were increased significantly in AD brain [21]. Both APP751 and APP770 contain the Kunitz protease inhibitor (KPI) domain, and APP770 also contains an OX-2 domain [22]. APP695 on the other hand, lacks both of these domains. The up-regulation of the KPIcontaining APP isoforms has been reported in the brains of AD patients and could be associated with the disease's progression [23]. Except for the protease-inhibitor role of the KPI domain, no other definite functional differences have been found in the different APP isoforms [20, 24, 25]. Other isoforms are referred to as leukocyte-derived APP (L-APP). L-APP mRNA is either seldom or never expressed in the central nervous system tissues [26].

As we known, extracellular deposition of the  $A\beta$ , the major constituent of senile plaques, is derived from the APP by proteolysis. There are at least two major pathways (the non-amyloidogenic and amyloidogenic pathways) leading to proteolytic cleavage of APP by  $\alpha$ -,  $\beta$ -and  $\gamma$ -secretases [27]. The non-amyloidogenic pathway, a proteolysis process including  $\alpha$ - and  $\gamma$ -secretases, results in nonpathogenic soluble  $\alpha$ -Cleaved soluble APP (sAPP $\alpha$ ) and a membrane-bound  $\alpha$ -C-terminal fragment ( $\alpha$ CTF). In this pathway, the APP protein is cleaved by  $\alpha$ -secretase in the middle of the A $\beta$  sequence, resulting in the release of sAPP $\alpha$ . Subsequently, the remaining  $\alpha$ CTF is cleaved by  $\gamma$ -secretase, resulting in the release of the P3 peptide and the amyloid intracellular domain (AICD) [27-29]. In the amyloidogenic pathway, which is common in neurons, the APP protein is cleaved by  $\beta$ -secretase at the 1st residue or at the 11th residue of A $\beta$  sequence, resulting in the release of soluble  $\beta$ -Cleaved soluble APP (sAPP $\beta$ ). Next, the remaining  $\beta$ CTF in the membrane is cleaved by  $\gamma$ -secretase, leading to a mixture of A $\beta$  peptides of different lengths (38 to 42 amino acids). AB1-40 (around 90%) and A $\beta$ 1-42 (around 10%) are the two major A $\beta$ peptides [30, 31].

### The function of APP in the central nervous system

APP is essential for physiological processes such as neural proliferation, migration, differentiation, plasticity and synaptogenesis [32–36]. Acutely knock down *APP* in the developing cortex of Sprague Dawley rats revealed that neuronal precursor cells in the embryonic cortex require APP to migrate correctly into the nascent cortical plate [32]. APP was also shown to play an important role in the cell cycle progression of neural stem cells through its interaction with APP binding protein-1 [33]. APP regulates netrin-1-mediated commissural axon outgrowth [34]. The functions of reelin and integrins on neurite development, neuronal migration and synapse functions may also be modulated by APP [35].

Genetically modified mice have shown that mice lacking APP are viable, fertile and exhibit a relatively mild phenotype, including a reduced body and brain weight, as well as several neurological symptoms such as reduced grip strength, deficits in spatial memory and increased susceptibility to seizures [37-40]. These symptoms are probably related to changes at the cellular and network level, including decreased dendritic spines, reduced hippocampal long-term potentiation and altered short-term neural plasticity [37, 41-43]. The development of a neuronal circuit requires the maintenance of synaptic homeostasis demands on a coordinated proteomic response at both pre- and postsynaptic sites. A very recent study showed that APP proteins are one of the unique sets of proteins regulating proper presynaptic physiology [44, 45]. Synaptic homeostasis entails a stable but plastic network and neuronal circuit. The combination of bioinformatics tools and biochemical approaches has shown that APP is a structural and functional regulator in a context-sensitive manner within the hippocampal active zone network [46].

During neuronal differentiation, *APP* expression remained stable, whereas the proteolytic processing of APP changed at various stages of differentiation [47]. sAPP $\alpha$  was secreted in the early stages of differentiation (neuronal progenitors), and sAPP $\beta$  was usually secreted after the production of deep-layer neurons. A recent study has shown that sAPP $\alpha$ , a neurotrophic fragment released from the metabolites of APP, is also important to synaptogenesis [36, 48].

### The mutation spectrum of APP

To date, about 50 pathogenic mutations of APP have been reported (Alzheimer Disease and Frontotemporal Dementia Mutation Database. http://www.molgen.ua.ac. be/admutations.), most of which affect the proteolysis of APP in such a way that A<sub>β1</sub>-42 levels are changed relative to other A $\beta$  isoforms [49–54]. Most pathogenic mutations of APP occur near the  $\beta$ -secretase cleavage site (amino acids 670aa-682aa), near the  $\gamma$ -secretase cleavage site (amino acids 713aa-724aa) or in the A $\beta$  sequence (amino acids 692aa-705aa) of the APP protein [55]. The mutations (D7H, E682K and K16N) that affect the  $\beta$ -secretase cleavage site cause a significant increase in the total levels of A $\beta$  and A $\beta$ 1-42/40 [56–58]. The mutations (T714I, V715M, V715A, I716V, V717I, V717L, L723P and K724N) that affect the  $\gamma$ -secretase cleavage site cause an increased relative ratio of A $\beta$ 42 to A $\beta$ 40 [59–61]. The mutations clustered around the central hydrophobic  $A\beta$ core near the  $\alpha$ -secretase cleavage site (E693G Arctic mutation, E693K Italian mutation, D694N Iowa mutation and 692G Flemish mutation) could result in a variety of polymorphic aggregates in a mutation-dependent manner, and could disrupt the integrity of the bilayer [62, 63].

The missense mutation V717I, the first described and best characterized of all *APP* mutations, is located in the transmembrane domain near the  $\gamma$ -secretase cleavage site, and affects both the  $\beta$ - and  $\gamma$ -secretase cleavage of APP protein [64]. This mutation changes the initial cleavage site of  $\gamma$ -secretase, which results in the increase of both A $\beta$ 42 and A $\beta$ 38 peptides. The initial clinical phenotype for a Chinese AD patient carrying the V717I mutation is characterized by prominent early affective symptoms, executive dysfunction and disorientation in comparison with Western patients [65]. This finding shows that the phenotypes of AD patients with identical *APP* mutations are affected by ethnic differences, environment or other unknown factors [65, 66].

Other than these dominant mutations, a recessive amino acid deletion mutation (E693Delta), which is more resistant to proteolytic degradation, and a recessive missense mutation (A673V) with a dominant negative effect on amyloidogenesis haves been reported [50, 67]. The E693Delta mutation has been suggested as a cause of dementia because of enhanced formation of synaptotoxic A $\beta$  oligomers [67]. The A673V mutation shows highly amyloidogenic effect in the homozygous state and antiamyloidogenic effect in the heterozygous state [50]. In the same amino acid position, another mutation (A673T) enriched in the Icelandic population is reported to be a protective variant [68]. This variant is adjacent to the  $\beta$ -secretase cleavage site of APP, and results in a reduction of approximately 40% in the formation of amyloidogenic peptides *in vitro*. This variant also protects against cognitive decline in elderly people without AD.

In addition, genomic duplications of variable size containing *APP* have also been identified in autosomal dominant EOAD [69]. In contrast to missense mutations, which show a near-complete disease penetrance, genomic duplications of *APP* are rare and display a higher variability in the age of onset. Moreover, the dementia of patients with *APP* duplication show a virtually complete penetrance by the age of 65 [69, 70]. Their phenotype is not associated with the size of the duplication. Patients with a duplicated *APP* gene are more often affected by seizures compared with patients suffering from other missense variants [71].

## Presenilin 1 (PSEN1)

### The gene and protein structure of PSEN1

After the discovery of the APP mutations that cause AD, PSEN1 was identified as the most common cause of autosomal dominant EOAD. PSEN1 mutations account for 70% to 80% of autosomal dominant EOAD cases [54, 72]. PSEN1 is located on chromosome 14q24.2. It consists of 12 exons encoding a 467-amino-acid protein, which has 9 C-terminal transmembrane domains locating to the lumen/ extracellular space [73]. Full-length PSEN1 is an inactive precursor and could be transformed into a heterodimer composed of a 30 kDa N-terminal fragment (NTF) and a 20 kDa C-terminal fragment (CTF) by an endoproteolytic cleavage within hydrophobic domain 7 in a large cytosolic loop [74]. The NTF/CTF heterodimers are the active forms of PSEN1 as well as the predominant forms detected in cells, whereas the full-length proteins not targeted for the cleavage pathway are rapidly degraded. PSEN1 endoproteolysis may be an important step in the process of activating the  $\gamma$ -secretase complex. Furthermore, the NTF/CTF heterodimers may form the catalytic core of the  $\gamma$ -secretase complex [75, 76]. The two catalytic aspartate residues (Asp257 at NTF and Asp385 at CTF) in PSEN1 are important for the activity of  $\gamma$ -secretase, and each mutation of the two conserved aspartates could abolish the  $\gamma$ -secretase activity [75, 77, 78].

# The function of PSEN1 in the central nervous system

PSEN1 is one of the four core proteins (others include nicastrin, anterior pharynx-defective 1 and presenilin enhancer 2) in the  $\gamma$ -secretase complex, which

is considered to play an important role in the generation of A $\beta$  from APP [79]. Some studies have shown that y-secretase activity in hippocampal neurons lacking PSEN1 is inhibited obviously than wild-type neurons [80]. The PSEN1 knockin studies showed that PSEN1 plays an important role in synaptic plasticity [81-83]. The knockout mice exhibit perinatal lethality with developmental defects and impaired neurogenesis. Conditional PSEN1 knockout studies have shown that the loss of PSEN1 activity causes synaptic dysfunction, memory impairment and agedependent neurodegeneration in the excitatory neurons of the postnatal forebrains of mice and a pronounced deficiency in enrichment-induced neurogenesis in the dentate gyrus [84-86]. These studies have suggested that PSEN1 may play an important role in promoting and maintaining memory and neuronal survival [87, 88]. At the cellular level of the knockout model, this inactivity has no effect on evoked neurotransmitter release, shortterm plasticity or the apparent calcium dependence of the evoked release [89]. This physiological function maybe associated with the interaction with the pre-synaptic protein synaptotagmin 1 (Syt1), a calcium sensor in synaptic vesicle exocytosis [90]. The binding between PSEN1 and Syt1 could regulate synaptic vesicle trafficking along neuronal processes, promoting exocytosis and neurotransmitter release and leading to neurodegeneration in the affected circuits in neurodegenerative diseases [90].

### The mutation spectrum of PSEN1

Patients with *PSEN1* mutations typically have an earlier age of onset, with symptoms starting an average of 8.4 years earlier than in *APP* mutation carriers (an average of 42.9 years of age vs. 51.3 years) and 14.2 years earlier than in *PSEN2* mutation carriers (an average of 57.1 years of age) [54]. Other studies have shown that very early-onset AD (VEOAD), which starts before the age of 35 years, is almost entirely caused by *PSEN1* mutations [91, 92]. Like *APP*, mutations in the promoter region of *PSEN1* also were found to be associated with increased risks of EOAD, perhaps due to an alteration of *PSEN1* gene expression with a subsequent influence on A $\beta$  load [93, 94]. Seizures and myoclonus is common feature of autosomal dominant EOAD, which was associated with *PSEN1* mutation [95, 96].

To date, more than 200 pathogenetic mutations have been described in *PSEN1* throughout the world, of which 70% mutations occur in exons 5, 6, 7 and 8 (Alzheimer Disease and Frontotemporal Dementia Mutation Database. http://www.molgen.ua.ac.be/admutations.). The majority of *PSEN1* mutations in EOAD are missense mutations, which cause amino acid substitutions. Mutations in the transmembrane domains 2 and 4 are associated with an earlier age of onset and death than those in the transmembrane domains 6 and 8 [97]. *PSEN1* mutations affect A $\beta$  production through a relative increase in the ratio of A\u03c642/A\u03c640 [98, 99]. A\u03c642 is more prone to forming amyloidogenic aggregates in brain than Aβ40 [100]. Investigations into the effects of PSEN1 mutations on  $\gamma$ -secretase have shown that around 90% of those reported mutations lead to the reduced production of A $\beta$ 42 and A $\beta$ 40, while the majority of mutations lead to increased Aβ42/Aβ40 ratios in vitro. There is no statistically significant correlation between the  $A\beta 42/A\beta 40$ ratio affected by PSEN1 mutations and the mean onset age of patients carrying the mutation [98]. These patients with PSEN1 mutations usually have higher amounts of total A $\beta$  deposits in the brain than sporadic AD patients. Research into PSEN1 mutation mechanisms that affect APP processing by  $\gamma$ -secretase has had conflicting results in different experimental systems, which may be affected by endogenous PSEN1, PSEN2 or other components of γ-secretas [101, 102].

## Presenilin-2 (PSEN2)

### The gene and protein structure of PSEN2

Less than a year after mapping *PSEN1*, another gene encoding the transmembrane protein PSEN2 showed a significant association with AD. *PSEN2*, located on the long arm of chromosome 1 (1q42.13), has a nearly 60% homology to *PSEN1* [103, 104]. It consists of 12 exons encoding a 448-amino-acid protein that is predicted to consist of 9 transmembrane domains and a large cytoplasmic loop domain between the 6th and 7th domains [104, 105]. *PSEN2* has two isoforms. Isoform 1 is found in the placenta, skeletal muscle and heart, while isoform 2, which lacks amino acids 263–296, is found in the brain, heart, placenta, liver, skeletal muscle, and kidney [103, 104, 106–108].

PSEN2 is also one of the four core proteins in the  $\gamma$ -secretase complex and provides the catalytic activity of the complex. *PSEN2* has been poorly studied and is considered to be a compensatory partner of *PSEN1* [109]. In contrast to the broadly distributed PSEN1, PSEN2 is known to be mainly restricted to a specific subcellular compartment, such as late endosomes and lysosomes [110]. The more restricted localization of the PSEN2 contributes to the intracellular pool of A $\beta$  peptide previously associated with an early event in AD [111].

### The function of PSEN2 in the central nervous system

Some controversial results have suggested that PSEN2 has a role in apoptosis [112–115]. Research into PSEN2-overexpressing Neuro2a cells has shown that PSEN2 could result in reduced viability and condensed chromatin. It could also affect the expression of Bax associated with apoptosis, but not the expression of p53 and PSEN1 [116]. Lots of studies suggest that PSEN2 proteins in the immune system have a variety of biologically important roles. The loss of presenilin function is associated with neuroinflammation and neurodegeneration [84, 85, 117]. PSEN2 could increase A $\beta$ -induced classical proinflammatory cytokines such as IL-1 $\beta$ , IL-1 $\alpha$  and TNF- $\alpha$  in knockout microglial cells [118]. PSEN2 could play a potential role in modulating lipopolysaccharide-mediated immune responses [119]. PSEN2 also modulates endoplasmic reticulummitochondria coupling in the presence of mitofusin-2, which is crucial for the regulation of various physiological and pathophysiological processes [120].

### The mutation spectrum of PSEN2

Unlike the PSEN1, mutations in the PSEN2 gene are extremely rare. Less than 40 mutations in PSEN2 have been identified (Alzheimer Disease and Frontotemporal Dementia Mutation Database. http://www.molgen. ua.ac.be/admutations.). PSEN2 mutation might increase  $\gamma$ -secretase activity. The known pathogenic mutations of PSEN2 lead to a significant decrease in extracellular A $\beta$ 40, an increase A $\beta$ 42 and a dramatic rise in the A $\beta$ 42/40 ratios. This change is more pronounced in the intracellular pool of AB. Except for two frame shift mutations, Glu126fs and Lys306fs, the others are nonsynonymous substitutions [54]. Familial AD with PSEN2 mutations have a later age of onset, longer disease duration compared with families with PSEN1 mutations [121]. Except in the case of familial AD, some PSEN2 mutations are associated with other disorders, such as dementia with Lewy bodies, frontotemporal dementia, breast cancer, dilated cardiomyopathy and Parkinson's disease with dementia (PDD) [105]. The penetrance of the disease in AD patients with PSEN2 mutations is variable and the age of onset ranges widely, from 40 to 80 years of age [105, 122, 123]. Only 17 of the 38 are predicted to be disease-causing mutations. Ten of the mutations are not pathogenic and the others are still unclear. The mutations T122P, N141I, M239I and M239V cause the increase of A<sub>β</sub> amount [124]. The mutations T122R, S130L and M239I were found to alter calcium signaling [125, 126].

### Genetically unexplained EOAD

Research into the genetics of EOAD has made great progress, but pathogenic mutations in *APP*, *PSEN1* and *PSEN2* can only explain a small fraction of EOAD families. The large number of genetically unexplained EOAD patients suggests that the additional diseasecausing genes have not yet been identified. In the last few years the next generation sequencing technologies, such as whole genome sequencing and whole exome sequencing, offer new insights into the genetic etiology of EOAD.

The homozygosity for the *APOE*  $\varepsilon$ 4 allele, as a major genetic risk factor for LOAD, was shown to be an independent genetic factor that significantly increases the risk of EOAD [127]. But, unlike mutations in *APP*, *PSEN1* and *PSEN2*, the *APOE*  $\varepsilon$ 4 allele was not considered a

significant cause of EOAD and was only a risk factor [128]. Some research into EOAD families using nextgeneration sequencing technology has identified some genes that could be candidates for causing EOAD, such as TYROBP, NOTCH3 and SORL1 [129-132]. The use of exome sequencing in EOAD patients identified some TYROBP variants might contribute to the risk of EOAD [129]. TYROBP might be involved in AB turnover. The partial loss of the TYROBP signaling pathway has been suggested to be responsible for the neurological phenotypes of cognitive decline. The whole exome sequencing technology identified a mutation in NOTCH3 is associated with AD [130]. NOTCH3 has been associated with cerebral autosomal dominant arteriopathy with subcortical infarcts and leukoencephalopathy (CADASIL), a dementia disorder which clinical phenotype overlaps with EOAD. Dysfunctional Notch signaling may also induce or inhibit genes that are important in the pathogenesis of AD or in PSEN function. Several mutations in SORL1, a sorting protein-related receptor gene involved in the trafficking of APP and A $\beta$ , was found in autosomal dominant EOAD patients without mutation on the known genes (APP, PSEN1 and PSEN2) by exome sequencing and whole genome sequencing studies [131, 132]. The screening of NOTCH3, SORL1 and TYROBP in larger patient/control groups would help to define the contribution of rare genetic variants in the three candidate genes to the etiology of EOAD.

### Summary

AD is one of the most challenging disorders and is characterized by dementia that typically begins with the progressive recession of memory and slowly becomes more severe and incapacitating. Understanding the genetics of AD may lead to its early detection, prevention and treatment. Compared with LOAD, learning more about the genetics of the rare, early-onset familial form of AD could result in a better understanding of the pathophysiology of the disease. In this review, we have summarized the main genetic aspects of EOAD and their role in the physiological function of the nervous system and the pathological function of disease mechanisms in EOAD patients. We have identified 3 high-penetrant EOAD genes (APP, PSEN1 and PSEN2) by genetic linkage studies and the gene cloning method in exceptionally large and informative monogenic pedigrees. The APOE ε4 allele also increased the risk of EOAD in carriers of at least one ɛ4 allele, and was a significant risk factor for EOAD independent of other genetic factors. Thanks to new high-throughput sequencing technologies, the systematic screening of the causal genes for dementia in both familial and sporadic patients has uncovered some other candidate genes. Investigations into the missing genetic etiology in unexplained EOAD patients still has a vast potential to discover new and crucial genetics aspects

that will lead to a better understanding of the pathological mechanisms leading to AD. Moreover, these findings will accelerate the development of new therapeutic strategies for preventing, stopping and even reversing AD.

# **CONFLICTS OF INTEREST**

None.

# REFERENCES

- Burns A, Iliffe S. Alzheimer's disease. Bmj. 2009; 338:b158. https://doi.org/10.1136/bmj.b158.
- Gallaway PJ, Miyake H, Buchowski MS, Shimada M, Yoshitake Y, Kim AS, Hongu N. Physical Activity: A Viable Way to Reduce the Risks of Mild Cognitive Impairment, Alzheimer's Disease, and Vascular Dementia in Older Adults. Brain Sci. 2017; 7. https://doi.org/10.3390/ brainsci7020022.
- Cummings JL, Morstorf T, Zhong K. Alzheimer's disease drug-development pipeline: few candidates, frequent failures. Alzheimers Res Ther. 2014; 6:37. https://doi. org/10.1186/alzrt269.
- Castellani RJ, Rolston RK, Smith MA. Alzheimer disease. Dis Mon. 2010; 56:484–546. https://doi.org/10.1016/j. disamonth.2010.06.001.
- Reitz C, Mayeux R. Alzheimer disease: epidemiology, diagnostic criteria, risk factors and biomarkers. Biochem Pharmacol. 2014; 88:640–51. https://doi.org/10.1016/j. bcp.2013.12.024.
- Dosunmu R, Wu J, Basha MR, Zawia NH. Environmental and dietary risk factors in Alzheimer's disease. Expert Rev Neurother. 2007; 7:887–900. https://doi. org/10.1586/14737175.7.7.887.
- Wingo TS, Lah JJ, Levey AI, Cutler DJ. Autosomal recessive causes likely in early-onset Alzheimer disease. Arch Neurol. 2012; 69:59–64. https://doi.org/10.1001/ archneurol.2011.221.
- Gatz M, Reynolds CA, Fratiglioni L, Johansson B, Mortimer JA, Berg S, Fiske A, Pedersen NL. Role of genes and environments for explaining Alzheimer disease. Arch Gen Psychiatry. 2006; 63:168–74. https://doi.org/10.1001/ archpsyc.63.2.168.
- Kang J, Lemaire HG, Unterbeck A, Salbaum JM, Masters CL, Grzeschik KH, Multhaup G, Beyreuther K, Muller-Hill B. The precursor of Alzheimer's disease amyloid A4 protein resembles a cell-surface receptor. Nature. 1987; 325:733–6. https://doi.org/10.1038/325733a0.
- Sherrington R, Rogaev EI, Liang Y, Rogaeva EA, Levesque G, Ikeda M, Chi H, Lin C, Li G, Holman K, Tsuda T, Mar L, Foncin JF, et al. Cloning of a gene bearing missense mutations in early-onset familial Alzheimer's disease. Nature. 1995; 375:754–60. https://doi. org/10.1038/375754a0.

- Levy-Lahad E, Wasco W, Poorkaj P, Romano DM, Oshima J, Pettingell WH, Yu CE, Jondro PD, Schmidt SD, Wang K, Crowley AC, Fu YH, Guenette SY, et al. Candidate gene for the chromosome 1 familial Alzheimer's disease locus. Science. 1995; 269:973–7.
- Brouwers N, Sleegers K, Van Broeckhoven C. Molecular genetics of Alzheimer's disease: an update. Ann Med. 2008; 40:562–83. https://doi.org/10.1080/07853890802186905.
- Jarmolowicz AI, Chen HY, Panegyres PK. The patterns of inheritance in early-onset dementia: Alzheimer's disease and frontotemporal dementia. Am J Alzheimers Dis Other Demen. 2015; 30:299–306. https://doi. org/10.1177/1533317514545825.
- Wallon D, Rousseau S, Rovelet-Lecrux A, Quillard-Muraine M, Guyant-Marechal L, Martinaud O, Pariente J, Puel M, Rollin-Sillaire A, Pasquier F, Le Ber I, Sarazin M, Croisile B, et al. The French series of autosomal dominant early onset Alzheimer's disease cases: mutation spectrum and cerebrospinal fluid biomarkers. J Alzheimers Dis. 2012; 30:847–56. https://doi.org/10.3233/jad-2012-120172.
- Janssen JC, Beck JA, Campbell TA, Dickinson A, Fox NC, Harvey RJ, Houlden H, Rossor MN, Collinge J. Early onset familial Alzheimer's disease: Mutation frequency in 31 families. Neurology. 2003; 60:235–9.
- Yoshikai S, Sasaki H, Doh-ura K, Furuya H, Sakaki Y. Genomic organization of the human amyloid beta-protein precursor gene. Gene. 1990; 87:257–63.
- Matsui T, Ingelsson M, Fukumoto H, Ramasamy K, Kowa H, Frosch MP, Irizarry MC, Hyman BT. Expression of APP pathway mRNAs and proteins in Alzheimer's disease. Brain Res. 2007; 1161:116–23. https://doi.org/10.1016/j. brainres.2007.05.050.
- Kong GK, Adams JJ, Harris HH, Boas JF, Curtain CC, Galatis D, Masters CL, Barnham KJ, McKinstry WJ, Cappai R, Parker MW. Structural studies of the Alzheimer's amyloid precursor protein copper-binding domain reveal how it binds copper ions. J Mol Biol. 2007; 367:148–61. https://doi.org/10.1016/j.jmb.2006.12.041.
- Lamb BT, Sisodia SS, Lawler AM, Slunt HH, Kitt CA, Kearns WG, Pearson PL, Price DL, Gearhart JD. Introduction and expression of the 400 kilobase amyloid precursor protein gene in transgenic mice [corrected]. Nat Genet. 1993; 5:22–30. https://doi.org/10.1038/ng0993-22.
- Golde TE, Estus S, Usiak M, Younkin LH, Younkin SG. Expression of beta amyloid protein precursor mRNAs: recognition of a novel alternatively spliced form and quantitation in Alzheimer's disease using PCR. Neuron. 1990; 4:253–67.
- Tanaka S, Shiojiri S, Takahashi Y, Kitaguchi N, Ito H, Kameyama M, Kimura J, Nakamura S, Ueda K. Tissuespecific expression of three types of beta-protein precursor mRNA: enhancement of protease inhibitor-harboring types in Alzheimer's disease brain. Biochem Biophys Res Commun. 1989; 165:1406–14.

- 22. Caporaso GL, Gandy SE, Buxbaum JD, Ramabhadran TV, Greengard P. Protein phosphorylation regulates secretion of Alzheimer beta/A4 amyloid precursor protein. Proc Natl Acad Sci U S A. 1992; 89:3055–9.
- 23. Tanaka S, Nakamura S, Ueda K. [Expression of amyloid beta-protein gene in Alzheimer's disease]. [Article in Japanese]. Rinsho Byori. 1990; 38:489–93.
- 24. Tanzi RE, McClatchey AI, Lamperti ED, Villa-Komaroff L, Gusella JF, Neve RL. Protease inhibitor domain encoded by an amyloid protein precursor mRNA associated with Alzheimer's disease. Nature. 1988; 331:528–30. https://doi.org/10.1038/331528a0.
- Kitaguchi N, Takahashi Y, Tokushima Y, Shiojiri S, Ito H. Novel precursor of Alzheimer's disease amyloid protein shows protease inhibitory activity. Nature. 1988; 331:530– 2. https://doi.org/10.1038/331530a0.
- Ohgami T, Kitamoto T, Tateishi J. Alzheimer's amyloid precursor protein mRNA without exon 15 is ubiquitously expressed except in the rat central nervous system. Brain Res Mol Brain Res. 1993; 20:240–4.
- Selkoe DJ. Amyloid beta-protein precursor: new clues to the genesis of Alzheimer's disease. Curr Opin Neurobiol. 1994; 4:708–16.
- Sinha S, Lieberburg I. Cellular mechanisms of beta-amyloid production and secretion. Proc Natl Acad Sci U S A. 1999; 96:11049–53.
- 29. Nunan J, Small DH. Regulation of APP cleavage by alpha-, beta- and gamma-secretases. FEBS Lett. 2000; 483:6–10.
- Zhang H, Ma Q, Zhang YW, Xu H. Proteolytic processing of Alzheimer's beta-amyloid precursor protein. J Neurochem. 2012; 120:9–21. https://doi.org/10.1111/ j.1471-4159.2011.07519.x.
- O'Brien RJ, Wong PC. Amyloid precursor protein processing and Alzheimer's disease. Annu Rev Neurosci. 2011; 34:185– 204. https://doi.org/10.1146/annurev-neuro-061010-113613.
- 32. Young-Pearse TL, Bai J, Chang R, Zheng JB, LoTurco JJ, Selkoe DJ. A critical function for beta-amyloid precursor protein in neuronal migration revealed by in utero RNA interference. J Neurosci. 2007; 27:14459–69. https://doi. org/10.1523/JNEUROSCI.4701-07.2007.
- 33. Joo Y, Ha S, Hong BH, Kim J, Chang KA, Liew H, Kim S, Sun W, Kim JH, Chong YH, Suh YH, Kim HS. Amyloid precursor protein binding protein-1 modulates cell cycle progression in fetal neural stem cells. PLoS One. 2010; 5:e14203. https://doi.org/10.1371/journal.pone.0014203.
- Rama N, Goldschneider D, Corset V, Lambert J, Pays L, Mehlen P. Amyloid precursor protein regulates netrin-1mediated commissural axon outgrowth. J Biol Chem. 2012; 287:30014–23. https://doi.org/10.1074/jbc.M111.324780.
- 35. Hoe HS, Lee KJ, Carney RS, Lee J, Markova A, Lee JY, Howell BW, Hyman BT, Pak DT, Bu G, Rebeck GW. Interaction of reelin with amyloid precursor protein promotes neurite outgrowth. J Neurosci. 2009; 29:7459–73. https://doi.org/10.1523/JNEUROSCI.4872-08.2009.

- Vasques JF, Heringer PVB, Goncalves RGJ, Campello-Costa P, Serfaty CA, Faria-Melibeu ADC. Monocular denervation of visual nuclei modulates APP processing and sAPPalpha production: A possible role on neural plasticity. Int J Dev Neurosci. 2017; 60:16–25. https://doi. org/10.1016/j.ijdevneu.2017.03.003.
- 37. Weyer SW, Klevanski M, Delekate A, Voikar V, Aydin D, Hick M, Filippov M, Drost N, Schaller KL, Saar M, Vogt MA, Gass P, Samanta A, et al. APP and APLP2 are essential at PNS and CNS synapses for transmission, spatial learning and LTP. Embo j. 2011; 30:2266–80. https://doi.org/10.1038/emboj.2011.119.
- Puzzo D, Privitera L, Fa M, Staniszewski A, Hashimoto G, Aziz F, Sakurai M, Ribe EM, Troy CM, Mercken M, Jung SS, Palmeri A, Arancio O. Endogenous amyloid-beta is necessary for hippocampal synaptic plasticity and memory. Ann Neurol. 2011; 69:819–30. https://doi.org/10.1002/ ana.22313.
- Steinbach JP, Muller U, Leist M, Li ZW, Nicotera P, Aguzzi A. Hypersensitivity to seizures in beta-amyloid precursor protein deficient mice. Cell Death Differ. 1998; 5:858–66. https://doi.org/10.1038/sj.cdd.4400391.
- Caldwell JH, Klevanski M, Saar M, Muller UC. Roles of the amyloid precursor protein family in the peripheral nervous system. Mech Dev. 2013; 130:433–46. https://doi. org/10.1016/j.mod.2012.11.001.
- Jedlicka P, Owen M, Vnencak M, Tschape JA, Hick M, Muller UC, Deller T. Functional consequences of the lack of amyloid precursor protein in the mouse dentate gyrus *in vivo*. Exp Brain Res. 2012; 217:441–7. https://doi. org/10.1007/s00221-011-2911-9.
- Korte M, Herrmann U, Zhang X, Draguhn A. The role of APP and APLP for synaptic transmission, plasticity, and network function: lessons from genetic mouse models. Exp Brain Res. 2012; 217:435–40. https://doi.org/10.1007/ s00221-011-2894-6.
- 43. Seabrook GR, Smith DW, Bowery BJ, Easter A, Reynolds T, Fitzjohn SM, Morton RA, Zheng H, Dawson GR, Sirinathsinghji DJ, Davies CH, Collingridge GL, Hill RG. Mechanisms contributing to the deficits in hippocampal synaptic plasticity in mice lacking amyloid precursor protein. Neuropharmacology. 1999; 38:349–59.
- Weingarten J, Weingarten M, Wegner M, Volknandt W. APP-A Novel Player within the Presynaptic Active Zone Proteome. Front Mol Neurosci. 2017; 10:43. https://doi. org/10.3389/fnmol.2017.00043.
- Schanzenbacher CT, Sambandan S, Langer JD, Schuman EM. Nascent Proteome Remodeling following Homeostatic Scaling at Hippocampal Synapses. Neuron. 2016; 92:358– 71. https://doi.org/10.1016/j.neuron.2016.09.058.
- 46. Lassek M, Weingarten J, Wegner M, Mueller BF, Rohmer M, Baeumlisberger D, Arrey TN, Hick M, Ackermann J, Acker-Palmer A, Koch I, Muller U, Karas M, et al. APP Is a Context-Sensitive Regulator of the Hippocampal

Presynaptic Active Zone. PLoS Comput Biol. 2016; 12:e1004832. https://doi.org/10.1371/journal.pcbi.1004832.

- 47. Bergstrom P, Agholme L, Nazir FH, Satir TM, Toombs J, Wellington H, Strandberg J, Bontell TO, Kvartsberg H, Holmstrom M, Borestrom C, Simonsson S, Kunath T, et al. Amyloid precursor protein expression and processing are differentially regulated during cortical neuron differentiation. Sci Rep. 2016; 6:29200. https://doi.org/10.1038/srep29200.
- Baumkotter F, Schmidt N, Vargas C, Schilling S, Weber R, Wagner K, Fiedler S, Klug W, Radzimanowski J, Nickolaus S, Keller S, Eggert S, Wild K, et al. Amyloid precursor protein dimerization and synaptogenic function depend on copper binding to the growth factor-like domain. J Neurosci. 2014; 34:11159–72. https://doi.org/10.1523/ jneurosci.0180-14.2014.
- 49. De Jonghe C, Esselens C, Kumar-Singh S, Craessaerts K, Serneels S, Checler F, Annaert W, Van Broeckhoven C, De Strooper B. Pathogenic APP mutations near the gammasecretase cleavage site differentially affect Abeta secretion and APP C-terminal fragment stability. Hum Mol Genet. 2001; 10:1665–71.
- Di Fede G, Catania M, Morbin M, Rossi G, Suardi S, Mazzoleni G, Merlin M, Giovagnoli AR, Prioni S, Erbetta A, Falcone C, Gobbi M, Colombo L, et al. A recessive mutation in the APP gene with dominant-negative effect on amyloidogenesis. Science. 2009; 323:1473–7. https://doi. org/10.1126/science.1168979.
- Kwok JB, Li QX, Hallupp M, Whyte S, Ames D, Beyreuther K, Masters CL, Schofield PR. Novel Leu723Pro amyloid precursor protein mutation increases amyloid beta42(43) peptide levels and induces apoptosis. Ann Neurol. 2000; 47:249–53.
- 52. Scheuner D, Eckman C, Jensen M, Song X, Citron M, Suzuki N, Bird TD, Hardy J, Hutton M, Kukull W, Larson E, Levy-Lahad E, Viitanen M, et al. Secreted amyloid betaprotein similar to that in the senile plaques of Alzheimer's disease is increased *in vivo* by the presenilin 1 and 2 and APP mutations linked to familial Alzheimer's disease. Nat Med. 1996; 2:864–70.
- Suzuki N, Cheung TT, Cai XD, Odaka A, Otvos L Jr, Eckman C, Golde TE, Younkin SG. An increased percentage of long amyloid beta protein secreted by familial amyloid beta protein precursor (beta APP717) mutants. Science. 1994; 264:1336–40.
- Cruts M, Theuns J, Van Broeckhoven C. Locus-specific mutation databases for neurodegenerative brain diseases. Hum Mutat. 2012; 33:1340–4. https://doi.org/10.1002/ humu.22117.
- 55. Ringman JM, Goate A, Masters CL, Cairns NJ, Danek A, Graff-Radford N, Ghetti B, Morris JC. Genetic heterogeneity in Alzheimer disease and implications for treatment strategies. Curr Neurol Neurosci Rep. 2014; 14:499. https://doi.org/10.1007/s11910-014-0499-8.
- 56. Chen WT, Hong CJ, Lin YT, Chang WH, Huang HT, Liao JY, Chang YJ, Hsieh YF, Cheng CY, Liu HC, Chen YR,

Cheng IH. Amyloid-beta (Abeta) D7H mutation increases oligomeric Abeta42 and alters properties of Abeta-zinc/ copper assemblies. PLoS One. 2012; 7:e35807. https://doi. org/10.1371/journal.pone.0035807.

- 57. Zhou L, Brouwers N, Benilova I, Vandersteen A, Mercken M, Van Laere K, Van Damme P, Demedts D, Van Leuven F, Sleegers K, Broersen K, Van Broeckhoven C, Vandenberghe R, et al. Amyloid precursor protein mutation E682K at the alternative beta-secretase cleavage beta'-site increases Abeta generation. EMBO Mol Med. 2011; 3:291–302. https://doi.org/10.1002/emmm.201100138.
- Kaden D, Harmeier A, Weise C, Munter LM, Althoff V, Rost BR, Hildebrand PW, Schmitz D, Schaefer M, Lurz R, Skodda S, Yamamoto R, Arlt S, et al. Novel APP/Abeta mutation K16N produces highly toxic heteromeric Abeta oligomers. EMBO Mol Med. 2012; 4:647–59. https://doi. org/10.1002/emmm.201200239.
- Bergman A, Religa D, Karlstrom H, Laudon H, Winblad B, Lannfelt L, Lundkvist J, Naslund J. APP intracellular domain formation and unaltered signaling in the presence of familial Alzheimer's disease mutations. Exp Cell Res. 2003; 287:1–9.
- Hecimovic S, Wang J, Dolios G, Martinez M, Wang R, Goate AM. Mutations in APP have independent effects on Abeta and CTFgamma generation. Neurobiol Dis. 2004; 17:205–18. https://doi.org/10.1016/j.nbd.2004.04.018.
- 61. Pasalar P, Najmabadi H, Noorian AR, Moghimi B, Jannati A, Soltanzadeh A, Krefft T, Crook R, Hardy J. An Iranian family with Alzheimer's disease caused by a novel APP mutation (Thr714Ala). Neurology. 2002; 58:1574–5.
- Nilsberth C, Westlind-Danielsson A, Eckman CB, Condron MM, Axelman K, Forsell C, Stenh C, Luthman J, Teplow DB, Younkin SG, Naslund J, Lannfelt L. The 'Arctic' APP mutation (E693G) causes Alzheimer's disease by enhanced Abeta protofibril formation. Nat Neurosci. 2001; 4:887–93. https://doi.org/10.1038/nn0901-887.
- 63. De Jonghe C, Zehr C, Yager D, Prada CM, Younkin S, Hendriks L, Van Broeckhoven C, Eckman CB. Flemish and Dutch mutations in amyloid beta precursor protein have different effects on amyloid beta secretion. Neurobiol Dis. 1998; 5:281–6. https://doi.org/10.1006/nbdi.1998.0202.
- 64. Goate A, Chartier-Harlin MC, Mullan M, Brown J, Crawford F, Fidani L, Giuffra L, Haynes A, Irving N, James L, Mant R, Newton P, Rooke K, et al. Segregation of a missense mutation in the amyloid precursor protein gene with familial Alzheimer's disease. Nature. 1991; 349:704–6. https://doi.org/10.1038/349704a0.
- Zhang G, Xie Y, Wang W, Feng X, Jia J. Clinical characterization of an APP mutation (V717I) in five Han Chinese families with early-onset Alzheimer's disease. J Neurol Sci. 2017; 372:379–86. https://doi.org/10.1016/j. jns.2016.10.039.
- 66. Jiao B, Tang B, Liu X, Xu J, Wang Y, Zhou L, Zhang F, Yan X, Zhou Y, Shen L. Mutational analysis in early-onset familial Alzheimer's disease in Mainland China. Neurobiol

Aging. 2014; 35:1957.e1–6. https://doi.org/10.1016/j. neurobiolaging.2014.02.014.

- 67. Tomiyama T, Nagata T, Shimada H, Teraoka R, Fukushima A, Kanemitsu H, Takuma H, Kuwano R, Imagawa M, Ataka S, Wada Y, Yoshioka E, Nishizaki T, et al. A new amyloid beta variant favoring oligomerization in Alzheimer's-type dementia. Ann Neurol. 2008; 63:377–87. https://doi.org/10.1002/ana.21321.
- Jonsson T, Atwal JK, Steinberg S, Snaedal J, Jonsson PV, Bjornsson S, Stefansson H, Sulem P, Gudbjartsson D, Maloney J, Hoyte K, Gustafson A, Liu Y, et al. A mutation in APP protects against Alzheimer's disease and age-related cognitive decline. Nature. 2012; 488:96–9. https://doi. org/10.1038/nature11283.
- Sleegers K, Brouwers N, Gijselinck I, Theuns J, Goossens D, Wauters J, Del-Favero J, Cruts M, van Duijn CM, Van Broeckhoven C. APP duplication is sufficient to cause early onset Alzheimer's dementia with cerebral amyloid angiopathy. Brain. 2006; 129:2977–83. https://doi. org/10.1093/brain/awl203.
- Rovelet-Lecrux A, Hannequin D, Raux G, Le Meur N, Laquerriere A, Vital A, Dumanchin C, Feuillette S, Brice A, Vercelletto M, Dubas F, Frebourg T, Campion D. APP locus duplication causes autosomal dominant early-onset Alzheimer disease with cerebral amyloid angiopathy. Nat Genet. 2006; 38:24–6. https://doi.org/10.1038/ng1718.
- 71. Wiseman FK, Al-Janabi T, Hardy J, Karmiloff-Smith A, Nizetic D, Tybulewicz VL, Fisher EM, Strydom A. A genetic cause of Alzheimer disease: mechanistic insights from Down syndrome. Nat Rev Neurosci. 2015; 16:564–74. https://doi.org/10.1038/nrn3983.
- 72. Theuns J, Del-Favero J, Dermaut B, van Duijn CM, Backhovens H, Van den Broeck MV, Serneels S, Corsmit E, Van Broeckhoven CV, Cruts M. Genetic variability in the regulatory region of presenilin 1 associated with risk for Alzheimer's disease and variable expression. Hum Mol Genet. 2000; 9:325–31.
- 73. Hutton M, Hardy J. The presenilins and Alzheimer's disease. Hum Mol Genet. 1997; 6:1639–46.
- 74. Van Gassen G, De Jonghe C, Pype S, Van Criekinge W, Julliams A, Vanderhoeven I, Woodrow S, Beyaert R, Huylebroeck D, Van Broeckhoven C. Alzheimer's disease associated presenilin 1 interacts with HC5 and ZETA, subunits of the catalytic 20S proteasome. Neurobiol Dis. 1999; 6:376–91. https://doi.org/10.1006/nbdi.1999.0265.
- 75. Laudon H, Mathews PM, Karlstrom H, Bergman A, Farmery MR, Nixon RA, Winblad B, Gandy SE, Lendahl U, Lundkvist J, Naslund J. Co-expressed presenilin 1 NTF and CTF form functional gamma-secretase complexes in cells devoid of full-length protein. J Neurochem. 2004; 89:44–53. https://doi.org/10.1046/j.1471-4159.2003.02298.x.
- Vetrivel KS, Zhang YW, Xu H, Thinakaran G. Pathological and physiological functions of presenilins. Mol Neurodegener. 2006; 1:4. https://doi.org/10.1186/1750-1326-1-4.

- Veugelen S, Saito T, Saido TC, Chavez-Gutierrez L, De Strooper B. Familial Alzheimer's Disease Mutations in Presenilin Generate Amyloidogenic Abeta Peptide Seeds. Neuron. 2016; 90:410–6. https://doi.org/10.1016/j. neuron.2016.03.010.
- Levitan D, Lee J, Song L, Manning R, Wong G, Parker E, Zhang L. PS1 N- and C-terminal fragments form a complex that functions in APP processing and Notch signaling. Proc Natl Acad Sci U S A. 2001; 98:12186–90. https://doi. org/10.1073/pnas.211321898.
- 79. Schroeter EH, Ilagan MX, Brunkan AL, Hecimovic S, Li YM, Xu M, Lewis HD, Saxena MT, De Strooper B, Coonrod A, Tomita T, Iwatsubo T, Moore CL, et al. A presenilin dimer at the core of the gamma-secretase enzyme: insights from parallel analysis of Notch 1 and APP proteolysis. Proc Natl Acad Sci USA. 2003; 100:13075–80. https://doi.org/10.1073/pnas.1735338100.
- De Strooper B, Saftig P, Craessaerts K, Vanderstichele H, Guhde G, Annaert W, Von Figura K, Van Leuven F. Deficiency of presenilin-1 inhibits the normal cleavage of amyloid precursor protein. Nature. 1998; 391:387–90. https://doi.org/10.1038/34910.
- Wang Y, Greig NH, Yu QS, Mattson MP. Presenilin-1 mutation impairs cholinergic modulation of synaptic plasticity and suppresses NMDA currents in hippocampus slices. Neurobiol Aging. 2009; 30:1061–8. https://doi. org/10.1016/j.neurobiolaging.2007.10.009.
- Xia D, Watanabe H, Wu B, Lee SH, Li Y, Tsvetkov E, Bolshakov VY, Shen J, Kelleher RJ 3rd. Presenilin-1 knockin mice reveal loss-of-function mechanism for familial Alzheimer's disease. Neuron. 2015; 85:967–81. https://doi.org/10.1016/j.neuron.2015.02.010.
- Xia D, Kelleher RJ 3rd, Shen J. Loss of Abeta43 Production Caused by Presenilin-1 Mutations in the Knockin Mouse Brain. Neuron. 2016; 90:417–22. https://doi.org/10.1016/j. neuron.2016.03.009.
- 84. Saura CA, Choi SY, Beglopoulos V, Malkani S, Zhang D, Shankaranarayana Rao BS, Chattarji S, Kelleher RJ 3rd, Kandel ER, Duff K, Kirkwood A, Shen J. Loss of presenilin function causes impairments of memory and synaptic plasticity followed by age-dependent neurodegeneration. Neuron. 2004; 42:23–36.
- 85. Feng R, Wang H, Wang J, Shrom D, Zeng X, Tsien JZ. Forebrain degeneration and ventricle enlargement caused by double knockout of Alzheimer's presenilin-1 and presenilin-2. Proc Natl Acad Sci U S A. 2004; 101:8162–7. https://doi.org/10.1073/pnas.0402733101.
- 86. Feng R, Rampon C, Tang YP, Shrom D, Jin J, Kyin M, Sopher B, Miller MW, Ware CB, Martin GM, Kim SH, Langdon RB, Sisodia SS, et al. Deficient neurogenesis in forebrain-specific presenilin-1 knockout mice is associated with reduced clearance of hippocampal memory traces. Neuron. 2001; 32:911–26.
- 87. Lee SH, Sharma M, Sudhof TC, Shen J. Synaptic function of nicastrin in hippocampal neurons. Proc Natl Acad

Sci USA. 2014; 111:8973-8. https://doi.org/10.1073/ pnas.1408554111.

- Tabuchi K, Chen G, Sudhof TC, Shen J. Conditional forebrain inactivation of nicastrin causes progressive memory impairment and age-related neurodegeneration. J Neurosci. 2009; 29:7290–301. https://doi.org/10.1523/ jneurosci.1320-09.2009.
- Pratt KG, Zhu P, Watari H, Cook DG, Sullivan JM. A novel role for {gamma}-secretase: selective regulation of spontaneous neurotransmitter release from hippocampal neurons. J Neurosci. 2011; 31:899–906. https://doi. org/10.1523/jneurosci.4625-10.2011.
- 90. Kuzuya A, Zoltowska KM, Post KL, Arimon M, Li X, Svirsky S, Maesako M, Muzikansky A, Gautam V, Kovacs D, Hyman BT, Berezovska O. Identification of the novel activity-driven interaction between synaptotagmin 1 and presenilin 1 links calcium, synapse, and amyloid beta. BMC Biol. 2016; 14:25. https://doi.org/10.1186/s12915-016-0248-3.
- 91. Holmes C. Genotype and phenotype in Alzheimer's disease. Br J Psychiatry. 2002; 180:131–4.
- 92. Campion D, Brice A, Dumanchin C, Puel M, Baulac M, De La Sayette V, Hannequin D, Duyckaerts C, Michon A, Martin C, Moreau V, Penet C, Martinez M, et al. A novel presenilin 1 mutation resulting in familial Alzheimer's disease with an onset age of 29 years. Neuroreport. 1996; 7:1582–4.
- 93. van Duijn CM, Cruts M, Theuns J, Van Gassen G, Backhovens H, van den Broeck M, Wehnert A, Serneels S, Hofman A, Van Broeckhoven C. Genetic association of the presenilin-1 regulatory region with early-onset Alzheimer's disease in a population-based sample. Eur J Hum Genet. 1999; 7:801–6. https://doi.org/10.1038/sj.ejhg.5200373.
- 94. Lambert JC, Mann DM, Harris JM, Chartier-Harlin MC, Cumming A, Coates J, Lemmon H, StClair D, Iwatsubo T, Lendon C. The -48 C/T polymorphism in the presenilin 1 promoter is associated with an increased risk of developing Alzheimer's disease and an increased Abeta load in brain. J Med Genet. 2001; 38:353–5.
- 95. Rudzinski LA, Fletcher RM, Dickson DW, Crook R, Hutton ML, Adamson J, Graff-Radford NR. Early onset familial Alzheimer Disease with spastic paraparesis, dysarthria, and seizures and N135S mutation in PSEN1. Alzheimer Dis Assoc Disord. 2008; 22:299–307. https://doi.org/10.1097/ WAD.0b013e3181732399.
- Jacquemont ML, Campion D, Hahn V, Tallaksen C, Frebourg T, Brice A, Durr A. Spastic paraparesis and atypical dementia caused by PSEN1 mutation (P264L), responsible for Alzheimer's disease. J Med Genet. 2002; 39:E2.
- Lippa CF, Swearer JM, Kane KJ, Nochlin D, Bird TD, Ghetti B, Nee LE, St George-Hyslop P, Pollen DA, Drachman DA. Familial Alzheimer's disease: site of mutation influences clinical phenotype. Ann Neurol. 2000; 48:376–9.

- 98. Sun L, Zhou R, Yang G, Shi Y. Analysis of 138 pathogenic mutations in presenilin-1 on the *in vitro* production of Abeta42 and Abeta40 peptides by gamma-secretase. Proc Natl Acad Sci USA. 2017; 114:E476-e85. https://doi. org/10.1073/pnas.1618657114.
- 99. Borchelt DR, Thinakaran G, Eckman CB, Lee MK, Davenport F, Ratovitsky T, Prada CM, Kim G, Seekins S, Yager D, Slunt HH, Wang R, Seeger M, et al. Familial Alzheimer's disease-linked presenilin 1 variants elevate Abeta1-42/1-40 ratio *in vitro* and *in vivo*. Neuron. 1996; 17:1005–13.
- 100. Jan A, Gokce O, Luthi-Carter R, Lashuel HA. The ratio of monomeric to aggregated forms of Abeta40 and Abeta42 is an important determinant of amyloid-beta aggregation, fibrillogenesis, and toxicity. J Biol Chem. 2008; 283:28176– 89. https://doi.org/10.1074/jbc.M803159200.
- 101. Cacquevel M, Aeschbach L, Houacine J, Fraering PC. Alzheimer's disease-linked mutations in presenilin-1 result in a drastic loss of activity in purified gamma-secretase complexes. PLoS One. 2012; 7:e35133. https://doi. org/10.1371/journal.pone.0035133.
- 102. Bentahir M, Nyabi O, Verhamme J, Tolia A, Horre K, Wiltfang J, Esselmann H, De Strooper B. Presenilin clinical mutations can affect gamma-secretase activity by different mechanisms. J Neurochem. 2006; 96:732–42. https://doi. org/10.1111/j.1471-4159.2005.03578.x.
- 103. Rogaev EI, Sherrington R, Rogaeva EA, Levesque G, Ikeda M, Liang Y, Chi H, Lin C, Holman K, Tsuda T, Mar L, Sorbi S, Nacmias B, et al. Familial Alzheimer's disease in kindreds with missense mutations in a gene on chromosome 1 related to the Alzheimer's disease type 3 gene. Nature. 1995; 376:775–8. https://doi.org/10.1038/376775a0.
- 104. Levy-Lahad E, Wijsman EM, Nemens E, Anderson L, Goddard KA, Weber JL, Bird TD, Schellenberg GD. A familial Alzheimer's disease locus on chromosome 1. Science. 1995; 269:970–3.
- 105. Cai Y, An SS, Kim S. Mutations in presenilin 2 and its implications in Alzheimer's disease and other dementiaassociated disorders. Clin Interv Aging. 2015; 10:1163–72. https://doi.org/10.2147/cia.s85808.
- 106. Hutton M, Busfield F, Wragg M, Crook R, Perez-Tur J, Clark RF, Prihar G, Talbot C, Phillips H, Wright K, Baker M, Lendon C, Duff K, et al. Complete analysis of the presenilin 1 gene in early onset Alzheimer's disease. Neuroreport. 1996; 7:801–5.
- 107. Anwar R, Moynihan TP, Ardley H, Brindle N, Coletta PL, Cairns N, Markham AF, Robinson PA. Molecular analysis of the presenilin 1 (S182) gene in "sporadic" cases of Alzheimer's disease: identification and characterisation of unusual splice variants. J Neurochem. 1996; 66:1774–7.
- 108. Prihar G, Fuldner RA, Perez-Tur J, Lincoln S, Duff K, Crook R, Hardy J, Philips CA, Venter C, Talbot C, Clark RF, Goate A, Li J, et al. Structure and alternative splicing of the presenilin-2 gene. Neuroreport. 1996; 7:1680–4.

- 109. Jayadev S, Case A, Eastman AJ, Nguyen H, Pollak J, Wiley JC, Moller T, Morrison RS, Garden GA. Presenilin 2 is the predominant gamma-secretase in microglia and modulates cytokine release. PLoS One. 2010; 5:e15743. https://doi.org/10.1371/journal.pone.0015743.
- 110. Sannerud R, Esselens C, Ejsmont P, Mattera R, Rochin L, Tharkeshwar AK, De Baets G, De Wever V, Habets R, Baert V, Vermeire W, Michiels C, Groot AJ, et al. Restricted Location of PSEN2/gamma-Secretase Determines Substrate Specificity and Generates an Intracellular Abeta Pool. Cell. 2016; 166:193–208. https://doi.org/10.1016/j. cell.2016.05.020.
- 111. Pensalfini A, Albay R 3rd, Rasool S, Wu JW, Hatami A, Arai H, Margol L, Milton S, Poon WW, Corrada MM, Kawas CH, Glabe CG. Intracellular amyloid and the neuronal origin of Alzheimer neuritic plaques. Neurobiol Dis. 2014; 71:53–61. https://doi.org/10.1016/j.nbd.2014.07.011.
- 112. Araki W, Yuasa K, Takeda S, Takeda K, Shirotani K, Takahashi K, Tabira T. Pro-apoptotic effect of presenilin 2 (PS2) overexpression is associated with down-regulation of Bcl-2 in cultured neurons. J Neurochem. 2001; 79:1161–8.
- 113. Nguyen HN, Lee MS, Hwang DY, Kim YK, Yoon DY, Lee JW, Yun YP, Lee MK, Oh KW, Hong JT. Mutant presenilin 2 increased oxidative stress and p53 expression in neuronal cells. Biochem Biophys Res Commun. 2007; 357:174–80. https://doi.org/10.1016/j.bbrc.2007.03.119.
- 114. Ghidoni R, Paterlini A, Benussi L, Binetti G. Presenilin 2 is secreted in mouse primary neurons: a release enhanced by apoptosis. Mech Ageing Dev. 2007; 128:350–3. https://doi. org/10.1016/j.mad.2007.01.003.
- 115. Gamliel A, Teicher C, Hartmann T, Beyreuther K, Stein R. Overexpression of wild-type presenilin 2 or its familial Alzheimer's disease-associated mutant does not induce or increase susceptibility to apoptosis in different cell lines. Neuroscience. 2003; 117:19–28.
- 116. Kumar A, Sivanandam TM, Thakur MK. Presenilin 2 overexpression is associated with apoptosis in Neuro2a cells. Transl Neurosci. 2016; 7:71–5. https://doi. org/10.1515/tnsci-2016-0011.
- 117. Beglopoulos V, Sun X, Saura CA, Lemere CA, Kim RD, Shen J. Reduced beta-amyloid production and increased inflammatory responses in presenilin conditional knockout mice. J Biol Chem. 2004; 279:46907–14. https://doi. org/10.1074/jbc.M409544200.
- 118. Qin J, Zhang X, Wang Z, Li J, Zhang Z, Gao L, Ren H, Qian M, Du B. Presenilin 2 deficiency facilitates Abetainduced neuroinflammation and injury by upregulating P2X7 expression. Sci China Life Sci. 2017; 60:189–201. https://doi.org/10.1007/s11427-016-0347-4.
- 119. Agrawal V, Sawhney N, Hickey E, McCarthy JV. Loss of Presenilin 2 Function Is Associated with Defective LPS-Mediated Innate Immune Responsiveness. Mol Neurobiol. 2016; 53:3428–38. https://doi.org/10.1007/s12035-015-9285-0.

- 120. Filadi R, Greotti E, Turacchio G, Luini A, Pozzan T, Pizzo P. Presenilin 2 Modulates Endoplasmic Reticulum-Mitochondria Coupling by Tuning the Antagonistic Effect of Mitofusin 2. Cell Rep. 2016; 15:2226–38. https://doi. org/10.1016/j.celrep.2016.05.013.
- 121. Jayadev S, Leverenz JB, Steinbart E, Stahl J, Klunk W, Yu CE, Bird TD. Alzheimer's disease phenotypes and genotypes associated with mutations in presenilin 2. Brain. 2010; 133:1143–54. https://doi.org/10.1093/brain/awq033.
- 122. Youn YC, Bagyinszky E, Kim H, Choi BO, An SS, Kim S. Probable novel PSEN2 Val214Leu mutation in Alzheimer's disease supported by structural prediction. BMC Neurol. 2014; 14:105. https://doi.org/10.1186/1471-2377-14-105.
- 123. Ryan NS, Rossor MN. Correlating familial Alzheimer's disease gene mutations with clinical phenotype. Biomark Med. 2010; 4:99–112. https://doi.org/10.2217/bmm.09.92.
- 124. Walker ES, Martinez M, Brunkan AL, Goate A. Presenilin 2 familial Alzheimer's disease mutations result in partial loss of function and dramatic changes in Abeta 42/40 ratios. J Neurochem. 2005; 92:294–301. https://doi.org/10.1111/ j.1471-4159.2004.02858.x.
- 125. Zatti G, Ghidoni R, Barbiero L, Binetti G, Pozzan T, Fasolato C, Pizzo P. The presenilin 2 M239I mutation associated with familial Alzheimer's disease reduces Ca2+ release from intracellular stores. Neurobiol Dis. 2004; 15:269–78. https://doi.org/10.1016/j.nbd.2003.11.002.
- 126. Zatti G, Burgo A, Giacomello M, Barbiero L, Ghidoni R, Sinigaglia G, Florean C, Bagnoli S, Binetti G, Sorbi S, Pizzo P, Fasolato C. Presenilin mutations linked to familial Alzheimer's disease reduce endoplasmic reticulum and Golgi apparatus calcium levels. Cell Calcium. 2006; 39:539–50. https://doi.org/10.1016/j.ceca.2006.03.002.
- 127. Lopez-Riquelme N, Alom-Poveda J, Viciano-Morote N, Llinares-Ibor I, Tormo-Diaz C. Apolipoprotein E epsilon4 allele and malondialdehyde level are independent risk factors for Alzheimer's disease. SAGE Open Med. 2016; 4:2050312115626731. https://doi.org/10.1177/2050312115626731.
- 128. Genin E, Hannequin D, Wallon D, Sleegers K, Hiltunen M, Combarros O, Bullido MJ, Engelborghs S, De Deyn P, Berr C, Pasquier F, Dubois B, Tognoni G, et al. APOE and Alzheimer disease: a major gene with semi-dominant inheritance. Mol Psychiatry. 2011; 16:903–7. https://doi.org/10.1038/mp.2011.52.
- 129. Pottier C, Ravenscroft TA, Brown PH, Finch NA, Baker M, Parsons M, Asmann YW, Ren Y, Christopher E, Levitch D, van Blitterswijk M, Cruchaga C, Campion D, et al. TYROBP genetic variants in early-onset Alzheimer's disease. Neurobiol Aging. 2016; 48:222 e9-e15. https://doi. org/10.1016/j.neurobiolaging.2016.07.028.
- 130. Guerreiro RJ, Lohmann E, Kinsella E, Bras JM, Luu N, Gurunlian N, Dursun B, Bilgic B, Santana I, Hanagasi H, Gurvit H, Gibbs JR, Oliveira C, et al. Exome sequencing reveals an unexpected genetic cause of disease: NOTCH3 mutation in a Turkish family with Alzheimer's disease.

Neurobiol Aging. 2012; 33:1008 e17–23. https://doi. org/10.1016/j.neurobiolaging.2011.10.009.

- 131. Pottier C, Hannequin D, Coutant S, Rovelet-Lecrux A, Wallon D, Rousseau S, Legallic S, Paquet C, Bombois S, Pariente J, Thomas-Anterion C, Michon A, Croisile B, et al. High frequency of potentially pathogenic SORL1 mutations in autosomal dominant early-onset Alzheimer disease. Mol Psychiatry. 2012; 17:875–9. https://doi.org/10.1038/ mp.2012.15.
- 132. Nicolas G, Charbonnier C, Wallon D, Quenez O, Bellenguez C, Grenier-Boley B, Rousseau S, Richard AC, Rovelet-Lecrux A, Le Guennec K, Bacq D, Garnier JG, Olaso R, et al. SORL1 rare variants: a major risk factor for familial early-onset Alzheimer's disease. Mol Psychiatry. 2016; 21:831–6. https://doi.org/10.1038/mp.2015.121.