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Sirtuin 1 Activity in Peripheral Blood Mononuclear Cells of Patients with Osteoporosis

Authors' Contribution:
Study Design A
Data Collection B
Statistical Analysis C
Data Interpretation D
Manuscript Preparation E
Literature Search F
Funds Collection G

B 1 **Marie Godfrin-Valnet**
C 2 **Kashif A. Khan**
B 1 **Xavier Guillot**
B 1 **Clément Prati**
C 2 **Lucile Baud**
C 2 **Wasim Abbas**
B 1,2 **Eric Toussirot**
ABCDEF 1,2 **Daniel Wendling***
ACDEF 2,3 **Georges Herbein***

1 Department of Rheumatology, Centre Hospitalier Régional Universitaire (CHRU) Besançon, Besançon, France
2 Pathogens & Inflammation Laboratory, University of Franche-Comté, Besançon, France
3 Department of Virology, Centre Hospitalier Régional Universitaire (CHRU) Besançon, Besançon, France

* GH and DW contributed equally to the work

Corresponding Authors: Georges Herbein, e-mail: georges.herbein@univ-fcomte.fr, Daniel Wendling, e-mail: dwendling@chu-besancon.fr
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Background: Sirtuin 1 (SIRT1) is a class III histone deacetylase that may play a critical role in several biological functions, including lifespan, stress, and inflammation. Our main objective was to evaluate SIRT1 activity in peripheral blood mononuclear cells (PBMCs) in patients with osteoporosis and to analyze the relationship between the SIRT 1 activity and markers of inflammation and bone remodelling.

Material/Methods: We performed a prospective monocentric study of patients with osteoporosis and measured the nuclear and cytoplasmic activities of SIRT1 in PBMCs. Levels of proinflammatory cytokines were assessed in culture supernatants of PBMCs isolated from the osteoporosis patients. The level of serum C-terminal cross-linking telopeptide of type I collagen (CTX), a marker of bone resorption, was measured in the serum of osteoporosis patients.

Results: Sixteen women with osteoporosis were included. A statistically significant correlation between the cytoplasmic and nuclear SIRT 1 activities was found in PBMCs of patients with osteoporosis. Although non-significant, we observed a negative trend between nuclear SIRT 1 activity and the rate of serum CTX and a positive trend between IL-6 and CTX levels in patients with osteoporosis.

Conclusions: This study shows that the cytoplasmic and nuclear SIRT 1 activities are measurable in circulating PBMCs of patients with osteoporosis and that these 2 activities are correlated. The potential role of inflammation in bone resorption in patients with osteoporosis was also studied.

MeSH Keywords: **Sirtuin 1 • SIRT1 • Osteoporosis • PBMCs**

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Background

Osteoporosis, which affects nearly 1 out of 2 women after 50 years of age, is a cause of morbidity and mortality in proven cases of fracture [1]. The sirtuin proteins (Silent Information Regulator proteins) belong to the class III histone deacetylases (HDAC) [2,3]. HDACs, of which 18 have been identified, are classified into 4 groups and are involved in epigenetic regulation, especially the deacetylation of histones [4,5]. These processes of acetylation and deacetylation are regulated by 2 enzyme systems, histone acetyl transferases (HATs) and HDACs, which include 7 sirtuins, especially sirtuin 1 (SIRT1) [6]. In mice, in the mesenchymal stem cells, SIRT1 activation promotes osteoblast differentiation via action on the p53 protein [7]. SIRT1 is regulated by a natural activator, resveratrol, which is a polyphenol abundant in red wine [8,9]. A study by Shakibaei demonstrated the role of resveratrol in inhibiting osteoclastogenesis via inhibition of RANK ligand [10]. Altogether studies suggested that SIRT1 dysregulation might play a role in the development of osteoporosis [3,5]. The presence of SIRT1 activity in animal mesenchymal cells has been highlighted, but so far no study has measured SIRT1 activity in human peripheral blood cells of osteoporotic patients [8]. We hypothesized that SIRT1 activity is measurable in PBMCs of patients with osteoporosis. The objective of our study was to evaluate the activity of SIRT 1 in PBMCs of osteoporotic patients and to analyze the relationship between SIRT1 activity and markers of inflammation IL-6, TNF alpha, and IL-8 and a marker of bone resorption, the serum C-terminal cross-linking telopeptide of type I collagen (CTX).

Material and Methods

Patients

A prospective monocentric study was conducted to measure the activity of SIRT1 in PBMCs of 16 osteoporotic patients. Sixteen control subjects were also enrolled. The criteria for inclusion of osteoporotic patients were the presence of a vertebral osteoporosis, defined by low bone mineral density (T score less than -2 SD) and the presence of at least 1 vertebral fracture without significant associated comorbidity. Exclusion criteria were the presence of significant comorbidity (e.g., diabetes and dyslipidemia) or treatment with corticosteroid or immunosuppressive therapy. The protocol was previously approved by the local Ethics Committee (CPP EST-II). All the patients and controls gave written informed consent.

PBMC isolation from whole blood

PBMCs were isolated by Ficoll gradient centrifugation. Blood from a patient was diluted with equal amounts of PBS, overlaid on Ficoll medium (Eurobio), and centrifuged at 900 × g for 30 min

at 25°C. The PBMC band was removed and washed twice with PBS. Cell count was determined by Malassez cytometer (Poly Labo), and cells were resuspended in serum-free RPMI 1640.

Isolation of nuclear and cytoplasmic extracts from PBMCs

After isolation, PBMCs were harvested and washed with wash buffer (10 mM HEPES (pH 7.6), 10 mM KCl, 2 mM MgCl₂, 1 mM EDTA). Cell pellets were then incubated on ice with cytoplasmic isolation buffer (10 mM HEPES (pH 7.6), 10 mM KCl, 2 mM MgCl₂, 1 mM EDTA, and 0.02% Nonidet P-40). Cytoplasmic extracts were collected by centrifugation, and the nuclear pellets were washed twice in wash buffer, spun, and incubated for 15 min on ice with nuclear isolation buffer (20 mM HEPES (pH 7.6), 420 mM NaCl, 1.5 mM MgCl₂, 0.2 mM EDTA, and 25% glycerol). Supernatants containing nuclear extracts were collected by centrifugation and stored at -80°C. Protease inhibitors (1 mM DTT, 1 mM PMSF, 1 µg/ml aprotinin, 1 µg/ml leupeptin, and 1 µg/ml pepstatin) were added to all solutions. Protein concentration in nuclear and cytoplasmic extracts was determined by the Bradford method using a Bio-Photometer (Eppendorf).

Measurement of SIRT1 activity in PBMC extracts

SIRT1 activity was evaluated from cytoplasmic and nuclear compartments using a fluorometric assay at the 15-min point (SIRT1 fluorometric kit, BML-AK-555, Enzo Life Sciences, Villeurbanne, France) [11].

Measurement of proinflammatory cytokines and serum CTX

The levels of TNF alpha, IL-6, and IL-8 were quantified in PBMC culture supernatants at 48 h using ELISA assays according to the manufacturer's recommendations (Quantikine Kits, R&D Systems, Minneapolis, MN). The biological marker of bone turnover sCTX-I was measured in the patient's serum.

Statistical analysis

The correlation coefficient R² was used to assess correlations between different data. This coefficient has been tested by a z-test at alpha=5% risk. Comparisons of continuous variables were made between groups using the t-test. The significance level used was p<0.05. Spearman's Rank correlation coefficient was used to identify and test the strength of a relationship between 2 sets of data.

Results

Sixteen osteoporotic women were included, with a mean age 68±9 years and an average of 2 vertebral fractures. We found

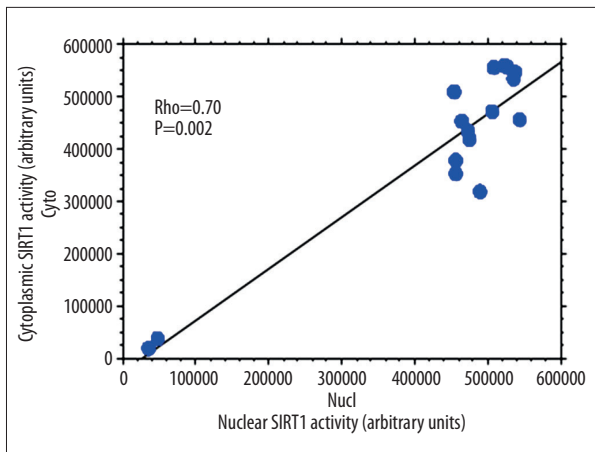


Figure 1. Positive correlation between nuclear and cytoplasmic SIRT1 activity in PBMCs isolated from patients with osteoporosis (rho=0.70; p=0.002).

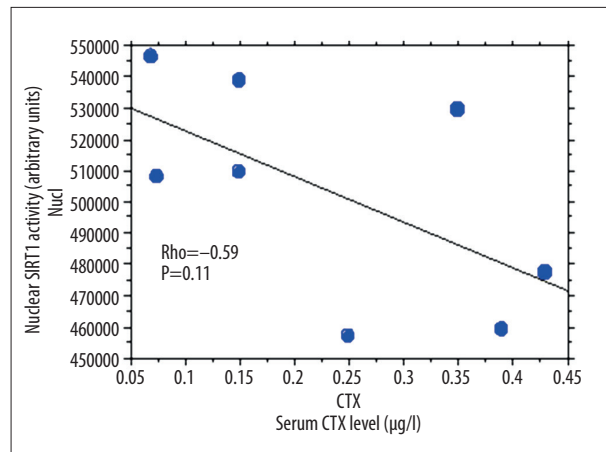


Figure 2. PBMC nuclear SIRT1 activity and serum CTX levels in patients with osteoporosis (rho=-0.59; p=0.11).

Table 1. Values of biological parameters in patients with osteoporosis and controls.

	SIRT 1 nuclear activity	SIRT 1 cytoplasmic activity	CRP mg/l	ESR mm/h	TNF α pg/ml	IL 6 pg/ml	IL 8 pg/ml
Osteoporosis patients (mean \pm SD) (n=16)	441,818 \pm 158,525	409,403 \pm 162,651	4.3 \pm 4.3	23.7 \pm 14.1	22.0 \pm 17.0	7.2 \pm 5.2	1,140.4 \pm 1,398.2
Controls (mean \pm SD) (n=16)	377,564 \pm 196,262	354,613 \pm 200,762	2.8 \pm 1.6	9 \pm 5.5	30.5 \pm 27.1	8.7 \pm 4.9	564.8 \pm 720.4
p	0.16	0.20	0.11	0.006	0.16	0.22	0.09

a statistically significant correlation between the cytoplasmic and nuclear activity of SIRT1 in PBMCs of osteoporotic patients ($R^2=0.866$) (Figure 1). By Spearman rank correlation analysis, cytoplasmic SIRT1 activity correlated positively with nuclear SIRT1 activity in PBMCs of osteoporotic patients (rho=0.70, P=0.002) (Figure 1). Nuclear and cytoplasmic SIRT1 activities were not significantly different in PBMCs of osteoporotic patients and healthy controls (p=0.16 and p=0.20, respectively). Erythrocyte sedimentation rate (ESR) was significantly higher in osteoporotic patients compared to controls (p=0.006). We then measured the levels of proinflammatory cytokines TNF alpha, IL-6, and IL-8 in the culture supernatants of PBMCs isolated from patients with osteoporosis and controls (Table 1). We did not find any statistically significant difference for TNF alpha (p=0.16), IL-6 (p=0.22), and IL-8 (p=0.09) levels in the culture supernatants of PBMCs isolated from osteoporotic patients compared to controls (Table 1). No significant correlation between the cytoplasmic and nuclear activity of SIRT 1 and the levels of the proinflammatory cytokines TNF alpha (rho=0.30, p=0.27), IL-6 (rho=0.05, p=0.82), and IL-8 (rho=0.40, p=0.12) was observed using Spearman rank correlation. Although not reaching significance, a negative trend was observed between the nuclear activity of SIRT1 and the levels of serum CTX using

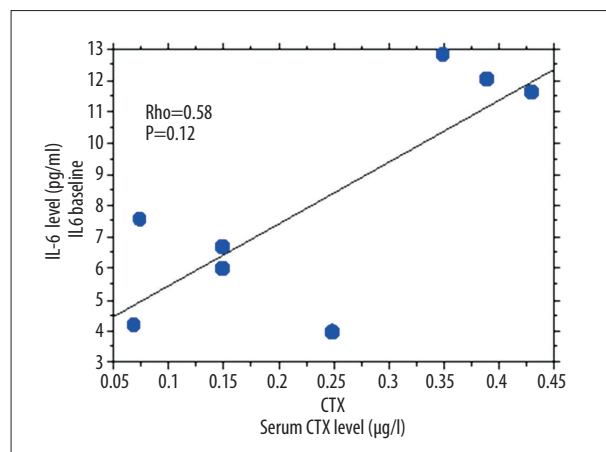


Figure 3. IL-6 levels in PBMC culture supernatants and serum CTX levels in patients with osteoporosis (rho=0.58; p=0.12).

Spearman rank correlation (rho=-0.59, p=0.11) (Figure 2). Similarly, using Spearman rank correlation, a non-significant positive trend was observed between the level of serum CTX and IL-6 levels (rho=0.58, p=0.12) (Figure 3).

Discussion

Our study assessed the detection of SIRT1 activity in PBMCs of patients with osteoporosis. We observed a statistically significant relationship between nuclear and cytoplasmic activity of SIRT1 in PBMCs of osteoporotic patients. Our results indicate that SIRT1 also has cytoplasmic activity in human PBMCs, although the primarily nuclear activity of SIRT1 was shown previously in the literature [4]. Additionally, nucleo-cytoplasmic shuttling of SIRT1 may participate in the differentiation and in the modulation of cell death [12]. We did not find any significant association between the cytoplasmic and nuclear activities of SIRT1 and the levels of inflammation markers (CRP and ESR), and between the rate of bone resorption marker (CTX) and inflammation markers. We did not observe a statistical correlation between nuclear and cytoplasmic activities of SIRT1 and the levels of proinflammatory cytokines in the culture supernatants of PBMCs isolated from osteoporotic patients. Our results can be explained by the fact that in addition to SIRT1, several other HDACs, including HDAC1 and HDAC2, regulate gene expression of proinflammatory cytokines such as TNF alpha [13]. Although not reaching significance, a negative trend was observed between the nuclear activity of SIRT1 and the levels of serum CTX, a bone resorption marker, using Spearman rank correlation ($\rho = -0.59$, $p = 0.11$)

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(Figure 2). Similarly, using Spearman rank correlation, a non-significant positive trend was observed between the level of serum CTX and IL-6 levels ($\rho = 0.58$, $p = 0.12$) (Figure 3). He et al. observed that resveratrol, a SIRT1 activator, inhibits osteoclastogenesis [14]. Further studies are needed to define the exact molecular mechanisms linking SIRT1 activity and bone homeostasis [15] and to support the potential role of inflammation in bone resorption.

Conclusions

We show here that the cytoplasmic and nuclear activities of SIRT1 are measurable in human blood PBMCs isolated from patients with osteoporosis, and that SIRT1 activity is correlated in these 2 cellular compartments. Our results also indicate a potential role of SIRT1 activity in the regulation of bone resorption. Further studies are needed to determine the exact role of SIRT1 and the molecular mechanisms involved in bone remodeling, with the goal to develop new therapeutic approaches to cure osteoporosis.

Conflict of interest

The authors declare no conflict of interest.