

Augmented Reduced-Intensity Regimen Does Not Improve Postallogeneic Transplant Outcomes in Acute Myeloid Leukemia

Charles Craddock, MD^{1,2}; Aimee Jackson, BSc²; Justin Loke, MD¹; Shamyia Siddique, BSc²; Andrea Hodgkinson, PhD²; John Mason, BSc²; Georgia Andrew, BSc³; Sandeep Nagra, MBChB¹; Ram Malladi, MD⁴; Andrew Peniket, MD⁵; Maria Gilleece, MD⁶; Rahuman Salim, MD⁷; Eleni Tholouli, MD⁸; Victoria Potter, MD⁹; Charles Crawley, MD⁴; Keith Wheatley, DPhil²; Rachel Protheroe, MD¹⁰; Paresh Vyas, MD⁵; Ann Hunter, MD¹¹; Anne Parker, MD¹²; Keith Wilson, MD¹³; Jiri Pavlu, MD¹⁴; Jenny Byrne, MD¹⁵; Richard Dillon, MD¹⁶; Naeem Khan, PhD³; Nicholas McCarthy, PhD³; and Sylvie D. Freeman, MD³

PURPOSE Reduced-intensity conditioning (RIC) regimens have extended the curative potential of allogeneic stem-cell transplantation to older adults with high-risk acute myeloid leukemia (AML) and myelodysplasia (MDS) but are associated with a high risk of disease relapse. Strategies to reduce recurrence are urgently required. Registry data have demonstrated improved outcomes using a sequential transplant regimen, fludarabine/amsacrine/cytarabine-busulphan (FLAMSA-Bu), but the impact of this intensified conditioning regimen has not been studied in randomized trials.

PATIENTS AND METHODS Two hundred forty-four patients (median age, 59 years) with high-risk AML (n = 164) or MDS (n = 80) were randomly assigned 1:1 to a fludarabine-based RIC regimen or FLAMSA-Bu. Pretransplant measurable residual disease (MRD) was monitored by flow cytometry (MFC-MRD) and correlated with outcome.

RESULTS There was no difference in 2-year overall survival (hazard ratio 1.05 [85% CI, 0.80 to 1.38] $P = .81$) or cumulative incidence of relapse (CIR) (hazard ratio 0.94 [95% CI, 0.60 to 1.46] $P = .81$) between the control and FLAMSA-Bu arms. Detectable pretransplant MFC-MRD was associated with an increased CIR (2-year CIR 41.0% v 20.0%, $P = .01$) in the overall trial cohort with a comparable prognostic impact when measured by an unsupervised analysis approach. There was no evidence of interaction between MRD status and conditioning regimen intensity for relapse or survival. Acquisition of full donor T-cell chimerism at 3 months abrogated the adverse impact of pretransplant MRD on CIR and overall survival.

CONCLUSION The intensified RIC conditioning regimen, FLAMSA-Bu, did not improve outcomes in adults transplanted for high-risk AML or MDS regardless of pretransplant MRD status. Our data instead support the exploration of interventions with the ability to accelerate acquisition of full donor T-cell chimerism as a tractable strategy to improve outcomes in patients allografted for AML.

J Clin Oncol 39:768-778. © 2020 by American Society of Clinical Oncology

Creative Commons Attribution Non-Commercial No Derivatives 4.0 License 

INTRODUCTION

Allogeneic stem-cell transplantation (allo-SCT) is an increasingly important treatment modality in adults with acute myeloid leukemia (AML) and myelodysplasia (MDS). The advent of reduced-intensity conditioning (RIC) regimens has permitted the extension of a potentially curative graft-versus-leukemia (GVL) effect to older patients in whom transplantation using myeloablative conditioning (MAC) is precluded by excess toxicity.¹ Indeed, the majority of allografts performed in the United States now use an RIC regimen.

In patients with AML and MDS, the use of an RIC regimen is associated with a higher rate of disease relapse than is observed with myeloablative transplants.² Despite the fact that relapse remains the dominant cause of transplant failure, no effective strategies have yet been identified to reduce the risk of disease recurrence after an RIC allograft. Indeed,

although a multiplicity of RIC regimens have been developed, most using a fludarabine backbone,^{3,4} there have been very few randomized studies to inform choice of regimen, and as a result, clinical practice worldwide is heterogeneous. Single-arm studies using a sequential fludarabine/amasacrine/cytarabine regimen, in which amsacrine-based cyto-reductive chemotherapy is delivered 7-14 days prior to a conventional RIC allograft incorporating either low dose total body irradiation or busulphan (Bu), have been reported to reduce the risk of relapse in high-risk AML.⁵⁻⁷ However, despite the widespread adoption of this regimen in the management of high-risk AML, its benefits have never been examined in a randomized trial.

The presence of measurable residual disease (MRD) measured by flow cytometry, quantitative polymerase chain reaction, or more recently next-generation

ASSOCIATED CONTENT

Data Supplement Protocol

Author affiliations and support information (if applicable) appear at the end of this article.

Accepted on December 4, 2020 and published at ascopubs.org/journal/jco on December 29, 2020; DOI <https://doi.org/10.1200/JCO.20.02308>

CONTEXT

Key Objective

The FIGARO study is the first prospective trial to examine the impact of an intensified conditioning regimen (FLAMSA-Bu) alongside the impact of pretransplant flow cytometric measurable residual disease (MRD) on transplant outcome in patients allografted for acute myeloid leukemia (AML) or myelodysplasia (MDS).

Knowledge Generated

The results of FIGARO demonstrate that pretransplant flow cytometric MRD is correlated with an increased risk of disease relapse after a reduced-intensity allograft by both conventional and unsupervised MRD analyses. Random assignment to an intensified sequential conditioning regimen failed to improve transplant outcome regardless of pretransplant MRD status.

Relevance

Our data do not support the use of an intensified sequential conditioning regimen as a strategy to improve transplant outcome, regardless of pretransplant MRD status. The results further demonstrate the importance of flow cytometry-determined MRD as a pretransplant risk characteristic in patients with AML or high-risk MDS.

sequencing (NGS) is an important determinant of disease relapse in adults with AML treated with intensive chemotherapy.^{8-15,16} Although retrospective studies have shown that the presence of pretransplant MRD is associated with an increased risk of relapse post-transplant,¹⁷⁻³⁰ the reported effect size varies widely and prospective studies addressing the prognostic value of pretransplant flow cytometric MRD, the most commonly used and widely applicable AML MRD technology, are lacking. Hourigan et al³¹ recently reported that the presence of pretransplant MRD in the peripheral blood, determined by an innovative but currently research-restricted NGS strategy, was associated with an increase in relapse in recipients of RIC but not MAC regimens, but this study did not examine the impact of flow-determined pretransplant MRD on outcome.

The FIGARO trial compared the outcomes of patients with high-risk AML and MDS transplanted using an intensified FLAMSA-Bu regimen with those receiving a conventional fludarabine-based RIC regimen. The impact of flow cytometric MRD on transplant outcomes was prospectively determined in trial patients.

PATIENTS AND METHODS

Study Design

FIGARO, an open label phase II randomized trial, was performed in 20 UK transplant centers and recruited patients from October 2013 to February 2017. The trial Protocol (online only; EudraCT 2012-005538-12) was approved by the UK research ethics service, National Research Ethics Service (NRES). An independent data monitoring committee oversaw the trial. Patients were randomly assigned in a one-to-one ratio via a minimization algorithm stratified by underlying disease, cytogenetic risk group, disease status at transplant, intended control transplant regimen, age, and donor type.

Patients

Patients were eligible for trial entry if they had a WHO-defined diagnosis of AML or high-risk MDS, were undergoing their first allo-SCT from a matched sibling or unrelated donor, and had been deemed ineligible for a MAC regimen on the grounds of age or comorbidity. Patients were of age 22 to 75, had a Hematopoietic Cell Transplant-Comorbidity Index (HCT-CI) score of 0-6, and were transplanted using peripheral blood- or bone marrow (BM)-derived stem cells from an HLA identical (HLA-A/-B/-C/-DRbeta1) matched sibling or $\geq 7/8$ HLA-A/-B/-C/-DRbeta1 adult-unrelated donor. All patients with AML were in complete remissions (CR1 and CR2) or had primary refractory AML (defined by failure to achieve a morphological CR after two courses of induction chemotherapy). High-risk MDS was defined as patients with an International Prognostic Scoring System score of intermediate-1 with $> 5\%$ blasts or intermediate-2 or high risk who had $< 5\%$ blasts at the time of random assignment. Cytogenetic risk group was classified as described previously.³²

Conditioning Regimens and GVHD Prophylaxis

Patients were randomly assigned 1:1 to a control arm determined by the investigator's choice of Flu/B₂/anti-thymocyte globulin (ATG), Flu/Mel/alemtuzumab (A), or Flu/Bu₂/A (details given in the Data Supplement, online only) versus an experimental arm of FLAMSA-Bu (Flu, cytarabine [araC] 2 g/m² once a day \times 4 days, amsacrine [AMSA] 100 mg/m² once a day \times 4 days, intravenous Bu total dose 11.2 mg/kg) and ATG 5 mg/kg over 3 days. Patients of age > 60 years received an adjusted FLAMSA-Bu regimen using a reduced dose of araC (1 g/m² once a day \times 4 days) and Bu (8 mg/kg total). However, after the first 31 patients had received treatment on the experimental arm, additional safety information was published with regard to the FLAMSA-Bu regimen in patients of age ≥ 60 years.

The experimental regimen in the subsequent 77 patients was modified to Flu, araC 1 g/m² once a day × 4 days, AMSA 100 mg/m² once a day × 4 days, and Bu 6.4 mg/kg for those patients who were > 60 years.

All patients received ciclosporin graft-versus-host-disease (GVHD) prophylaxis. Supportive care was according to institutional guidelines. All patients were formally reviewed at day + 100, 6, and 12 months post-transplant. BMs to determine remission status were reviewed at day + 42, and months 3, 6, 9, and 12 post-transplant. T-cell lineage chimerism was assessed at months 3, 6, 9, and 12 post-transplant.

MRD Quantitation

BMs for multiparameter flow cytometric (MFC) detection of MRD were obtained pretransplant (within 4 weeks of transplant) and at day + 42 post-transplant. Sample logistics, processing, and analysis strategy are provided in the Data Supplement. MFC-MRD analysis was performed centrally, using a standardized manual gating strategy that screened blasts for different-from-normal leukemia-associated immunophenotypes (LAIPs) and any previously identified baseline LAIPs when available. Samples were reported as MRD-negative if no baseline and/or different-from-normal LAIP cells could be quantitated above the limit of detection (approximately 0.02%-0.05%). The results were not reported to treating clinicians.

Recognizing the potential for variation in manual MFC-MRD analysis, an unsupervised approach was applied as an independent measurement of LAIPs. This incorporated (1) a multidimensional clustering algorithm to maximize information from the LAIP marker combinations and (2) statistical criteria to discriminate blast subpopulations that were immunophenotypically most aberrant (compared with reference ranges established from 40 control BMs) and above the limit of quantitation (Data Supplement). The analytic method, similar to standard different-from-normal MFC-MRD, did not require diagnostic samples. Unsupervised MFC-MRD percentages were higher than conventional MFC-MRD as the former summated all quantifiable nonoverlapping abnormal blast subpopulations from an antibody combination, whereas conventional MFC-MRD values are from a single LAIP. Concordance between methods was strongest at higher MRD levels (Data Supplement). The unsupervised MFC-MRD combined test criteria included results from a third antibody combination (stem and progenitor) in addition to standard LAIP markers; positivity required detection of aberrant blasts in at least two of the three antibody combinations (Data Supplement).

Outcomes

The primary outcome was overall survival (OS) defined on an intention-to-treat basis. A sensitivity analysis was conducted to assess OS in a per-protocol population. Secondary outcome measures included event-free survival

(EFS), cumulative incidence of relapse (CIR), incidence of GVHD, and transplant-related mortality (TRM). Acute and chronic GVHD were scored according to published criteria.^{33,34} Nonhematological grade 3-4 adverse events were classified according to Common Terminology Criteria for Adverse Events Version 4.0.

Statistical Analysis

The sample size was calculated on the basis of previously published data and clinical judgment. Assuming a 2-year OS in the control arm of 25%, to detect a 15% improvement in the experimental arm, a total of at least 214 patients (two-sided $\alpha = 0.15$ and $\beta = 0.16$) were required. To account for the likelihood that 10% of randomly assigned patients would not proceed to transplant, the trial aimed to recruit a minimum of 240 patients. Analysis was conducted in line with the predefined statistical analysis plan on the intention-to-treat population unless otherwise stated. Per-protocol population analysis was restricted to patients who had commenced the conditioning regimen. Standard analysis methods were employed as further outlined in the Data Supplement.

Additional analysis in the per-protocol populations was conducted to assess the effect on OS, CIR, and TRM of pretransplant MRD by the different MFC-MRD analysis methods and for various MRD thresholds. No adjustment for multiple testing was made within the MRD threshold analysis; however, the results are interpreted with caution and focused on identifying the highest level of discrimination from a range of significant results.

RESULTS

Enrollment

Of 255 patients screened for trial entry, 244 fulfilled eligibility criteria and were randomly assigned to receive trial therapy (Fig 1). Twenty-eight randomly assigned patients did not receive their allocated treatment (two deaths, 14 withdrawn because of clinical deterioration or patient or physician choice, and 12 relapses prior to transplant). Of the 108 patients who were transplanted on the control arm, 63 received Flu/B₂/ATG, 31 Flu/Mel/A, and 14 Flu/B₂/A. The median follow-up was 35 months. Patient and transplant characteristics of randomly assigned patients are summarized in Table 1. One hundred sixty-four patients had an initial diagnosis of AML of whom 154 were in CR1 or CR2 and nine had primary refractory AML at the time of random assignment. Eighty patients had high-risk MDS. The median age of the study population was 59 years (range, 22-75 years).

Survival

The 2-year OS was 58.8% in patients treated on the control arm and 60.9% in patients assigned to FLAMSA-Bu (hazard ratio [HR] 1.05 [85% CI, 0.80 to 1.38] log-rank *P* value = .81; Fig 2A). The EFS at 2 years was 48.7% in the

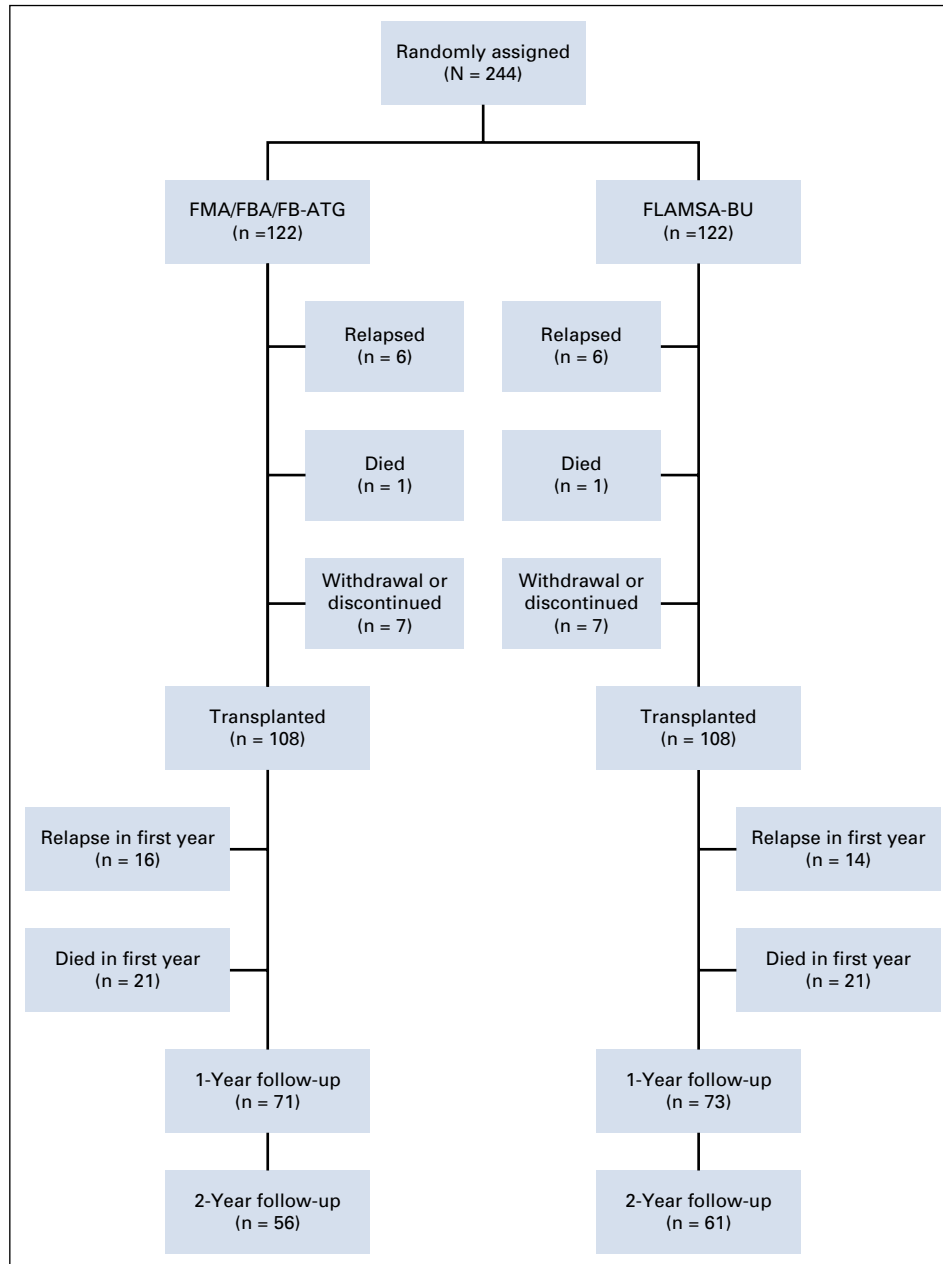


FIG 1. Trial CONSORT diagram. FBA, fludarabine/busulphan/alemtuzumab; FB-ATG, fludarabine/busulphan/antithymocyte globulin; FMA, fludarabine/melphalan/alemtuzumab; FLAMSA-Bu, fludarabine/amsacrine/cytarabine-busulphan.

control arm versus 54.2% for FLAMSA-Bu (HR 0.96 [95% CI, 0.68 to 1.35] log-rank P value = .82; Fig 2B). Two-year OS and EFS were similar between both arms in a per-protocol sensitivity analysis (Data Supplement). In the preplanned subgroup analysis, no survival benefit of the FLAMSA-Bu regimen was evident in patients diagnosed with either AML or MDS, in patients with AML according to cytogenetic risk category, or in patients under or over 60 years of age. No difference in outcome was evident when analysis was restricted to patients over 60 in the experimental arm after adoption of the Protocol amendment.

Transplant-Related Mortality, GVHD, and Disease Relapse

The 1-year TRM was 16.8% in the control arm and 20.5% in the experimental arm (HR 1.20 [95% CI, 0.68 to 2.13], Gray's test P value = .53). There were no statistically significant differences in the cumulative incidences of acute GVHD at day + 100 (with death and relapse as competing events) between the control and FLAMSA-Bu arms (grades 2-4, 10.1% v 8.3%, Gray's test P value = .93; grade 3-4, 1.7% v 5.8%, Gray's test P value = .23). The cumulative incidence of chronic GVHD at 1 year was

TABLE 1. Patient Characteristics by Random Assignment

Characteristic		Control (FMA/FBA/FB-ATG) n (%)	FLAMSA-BU n (%)	Overall n (%)
Age	≤ 60 years	71 (58)	69 (57)	140 (57)
	> 60 years	51 (42)	53 (43)	104 (43)
Sex	Female	51 (42)	48 (39)	99 (41)
	Male	71 (58)	74 (61)	145 (59)
HCT-comorbidity index	≤ 2	66 (57)	79(68)	145 (62)
	≥ 3	33 (28)	18 (15)	51 (22)
	Unknown	23(19)	25(20)	48 (20)
Diagnosis	AML	82 (67)	82 (67)	164 (67)
	MDS	40 (33)	40 (33)	80 (33)
AML cytogenetic risk	Adverse	24 (29)	26 (32)	50 (30)
	Intermediate	53 (65)	52 (63)	105 (64)
	Favourable	4 (5)	3 (4)	7 (4)
	Unknown	1 (1)	1 (1)	2 (1)
AML disease status	CR1 or CR2	77 (94)	77 (94)	154 (94)
	Primary refractory	5 (6)	4 (5)	9 (5)
	Unknown	0	1 (1)	1 (1)
AML <i>FLT3</i> -ITD	Yes	20 (24)	23 (28)	43 (26)
	No	49 (60)	52 (63)	101(62)
	Unknown	13 (16)	7 (9)	20 (12)
AML-mutated <i>NPM1</i>	Yes	17 (21)	23 (28)	40 (24)
	No	50 (61)	53 (65)	103 (63)
	Unknown	15 (18)	6 (7)	21 (13)
MDS IPSS	≤ 2	33 (83)	33 (83)	66 (83)
	> 2	0 (0)	2 (5)	2 (3)
	Unknown	7 (18)	5 (13)	12 (15)
Transplant				
Donor type	Sibling	25 (20)	24 (20)	49 (20)
	Unrelated	97 (80)	98 (80)	195 (80)
Graft type	PBSCs	101 (94)	107 (99)	208 (96)
	BM	7 (6)	1 (1)	8 (4)
Pretransplant MFC-MRD status	Positive	43 (35)	52 (43)	95(39)
	Negative	46 (38)	35 (29)	81 (33)
	Inadequate	14 (11)	13 (11)	27 (11)
	No sample	19 (16)	22 (18)	41 (17)

Abbreviations: AML, acute myeloid leukemia; BM, bone marrow; CR, complete remission; FBA, fludarabine/busulphan/alemtuzumab; FB-ATG, fludarabine/busulphan/antithymocyte globulin; FLAMSA-Bu, fludarabine/amsacrine/cytarabine-busulphan; FMA, fludarabine/melphalan/alemtuzumab; HCT-CI, hematopoietic cell transplant-comorbidity index; IPSS, International Prognostic Scoring System; ITD, internal tandem duplication; MDS, myelodysplasia; MFC, multiparameter flow cytometric; MRD, minimal residual disease; PBSC, peripheral blood stem cells.

25.2% and 19.2% in the control and FLAMSA-Bu arms, respectively (Gray's test P value = .53). Twenty-eight patients received DLI in the control arm (19 for mixed chimerism from day + 115, nine for relapse) and 14 in the experimental arm (10 for mixed chimerism from day + 104, four for relapse) (Data Supplement). There was no evidence of DLI impact on the incidence of GVHD with eight

and five episodes of chronic GVHD post-DLI in the control and FLAMSA-Bu arms, respectively.

The 2-year CIR was 29.5% in patients in the control arm and 26.7% in patients assigned FLAMSA-Bu (Fig 2C) (Gray's test P value = .81). There was no statistically significant effect of disease (AML v MDS), patient age, and

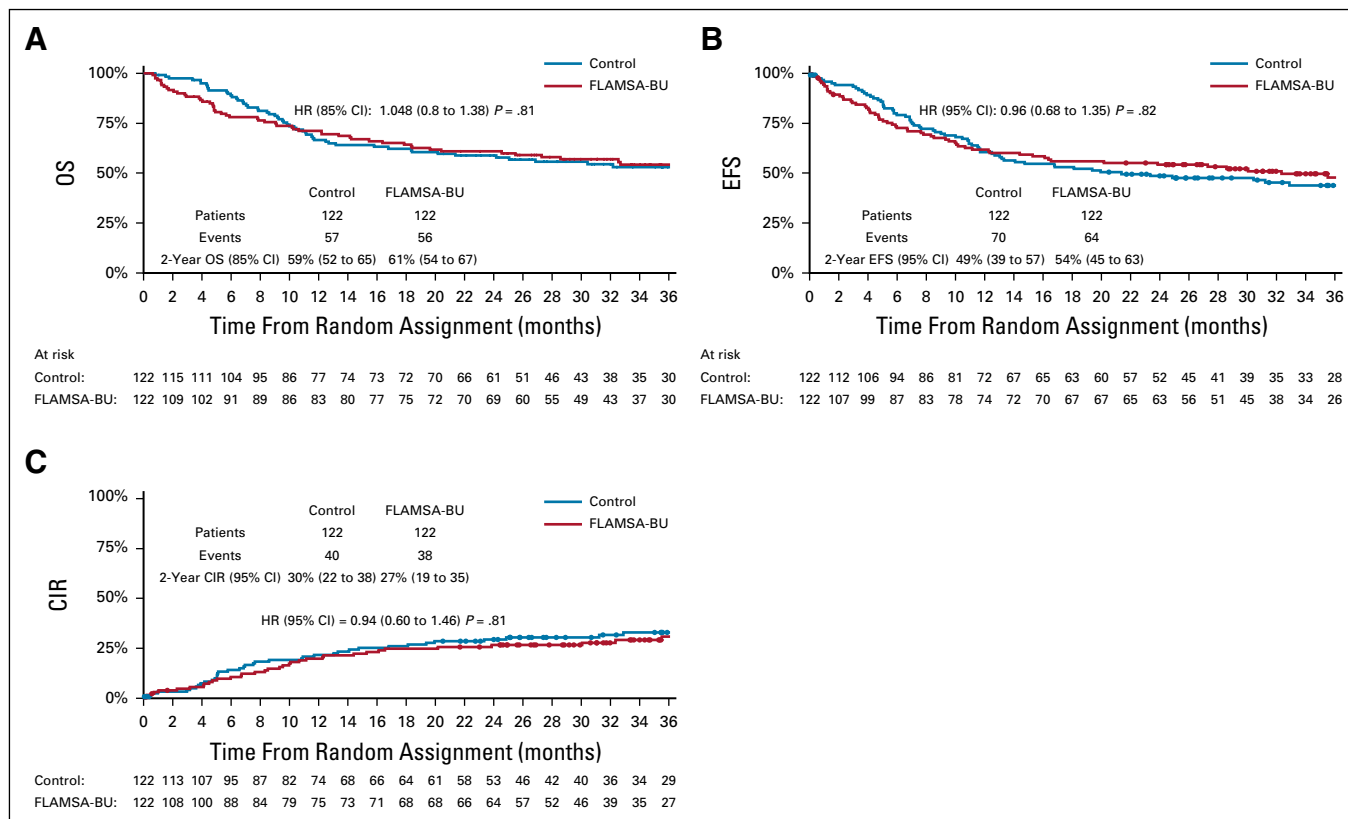


FIG 2. (A) OS, (B) EFS, and (C) CIR by conditioning regimen in the intention-to-treat population. 85% CIs are reported for overall survival to align with the type I error rate applied in the sample size calculation (described in the Data Supplement). CIR, cumulative incidence of relapse; EFS, event-free survival; FLAMSA-Bu, fludarabine/amsacrine/cytarabine-busulphan; HR, hazard ratio; OS, overall survival.

treatment arm (including by different control regimens) on relapse risk (Data Supplement).

Pretransplant MRD Status and Post-Transplant Outcome

Pretransplant MRD data, excluding inadequate BMs, were available in 176 randomly assigned patients of whom 156 proceeded to transplant (Data Supplement, distribution of clinical characteristics by MRD status in Data Supplement). MRD at any level was detected by flow cytometry in 43% of the 156 patients (38 of 79 receiving control regimens and 29 of 77 receiving FLAMSA-Bu) (median MRD level of 0.2%, range 0.02%-12.3%). In randomly assigned patients, pretransplant MRD positivity was associated with an increased relapse risk (2-year CIR 41.0% ν 20.0% (HR 1.97 [95% CI, 1.18 to 3.28], Gray's test P value = .01) and a borderline significant reduction in 2-year OS (70.1%-51.4% log-rank P value = .05) (Data Supplement). No statistically significant difference was observed in TRM (2-year TRM 12.1% MRD-positive ν 21.6% MRD-negative (HR 0.60 (95% CI, 0.29 to 1.27), Gray's test P value = .18). There was no interaction between MRD status and conditioning intensity in the preplanned subgroup survival analysis (heterogeneity test $P = .56$) or on relapse risk (treatment MRD interaction term $P = .92$). No difference in post-transplant MRD clearance was apparent between treatment arms from MRD results

at day + 42 (Data Supplement). Although flow cytometric methodology represents the most widely applicable MRD assay in AML, its reliance on operator analysis expertise is a recognized limitation that may potentially contribute to variation in its prognostic value.²⁷ We therefore used an unsupervised computational approach to analyze flow cytometric sample files to obtain independent evaluation of the impact of conventionally determined MFC-MRD (Data Supplement) on outcome in the transplanted cohort. Twenty patients with pretransplant conventional MFC-MRD results were excluded since their samples had fewer than the minimum requirement of 1,000 blast events. Outcomes (Table 2, Fig 3) and test accuracy for relapse prediction (Data Supplement) were comparable between both methods in transplanted patients, supporting reproducibility of the prognostic effect from immunophenotypic MRD. The prognostic significance of pretransplant MRD above the thresholds that provided the most discrimination in this RIC allo-SCT setting (0.2% by conventional analysis, 1% by unsupervised) (Figs 3B and 3C, Table 2) was retained for relapse in an analysis adjusted for additional factors with the potential to determine transplant outcome (Table 2). To further test the robustness of these MFC-MRD-predicted outcomes, we applied stringent criteria (quantifiable, unsupervised MFC-MRD in at least 2 different antibody

TABLE 2. Conventional and Unsupervised MRD Comparison: Outcomes by Pretransplant MRD Status

Pretransplant MRD status	2-Year CIR (95% CI)	Unadjusted P	Adjusted HR (95% CI) P	2-Year TRM (95% CI)	P	2-Year OS (95% CI)	Unadjusted P	Adjusted HR (95% CI) P
MRD-negative n = 73	20.7% (12.2 to 30.7)	.034	1.8 (0.94 to 3.42) .075	16.6% (9.1 to 26.1)	.63	72.1% (60.2 to 81)	.08	1.54 (0.88 to 2.7) .13
MRD-positive n = 63	38.3% (26.3 to 50.2)			12.9% (6 to 22.6)		53% (39.9 to 64.6)		
UnSup MRD-negative n = 86	22.1% (14 to 31.4)	.022	1.82 (1.00 to 3.34) .051	16.5% (9.5 to 25.2)	.82	66.9% (55.7 to 75.8)	.12	1.22 (0.69 to 2.15) .49
UnSup MRD-positive n = 50	40.5% (26.6 to 53.9)			12.2% (4.9 to 23)		57% (41.9 to 69.5)		
MRD < 0.2% n = 104	22.2% (14.7 to 30.7)	.001	2.39 (1.23 to 4.61) .01	16.5% (10.1 to 24.4)	.79	67.8 (57.8 to 75.9)	.037	1.73 (0.95 to 3.15) .075
MRD ≥ 0.2% n = 32	50% (31.5 to 66.4)			9.6% (2.4 to 23)		48.2 (30 to 64.3)		
UnSup MRD < 1% n = 103	21.4% (14 to 29.8)	< .001	2.52 (1.3 to 4.9) .006	17.8% (11 to 25.9)	.35	66.2% (56.1 to 74.6)	.11	1.41 (0.77 to 2.58) .28
UnSup MRD ≥ 1% n = 33	52% (33.3 to 67.8)			6.1% (1 to 17.9)		54% (35.5 to 69.2)		
UnSup-combined MRD-negative or equivocal n = 102	20.6% (13.3 to 28.9)	< .001	2.44 (1.25 to 4.74) .009	15.9% (9.5 to 23.7)	.86	68.2% (58.1 to 76.3)	.007	2.03 (1.13 to 3.63) .018
UnSup-combined MRD-positive n = 34	50.5% (32.2 to 66.2)			12% (3.7 to 25.5)		51.7% (33.7 to 67)		

NOTE. Conventional and unsupervised (computational) MRD comparisons are in transplanted patients. Adjusted results are the results of cox proportional hazard models adjusted for age, cytogenetic risk, *FLT3*-ITD presence, treatment arm, and HCT comorbidity.

Abbreviations: CIR, cumulative incidence of relapse; HR, hazard ratio; MRD, measurable residual disease (flow cytometric); TRM, transplant-related mortality; UnSup, unsupervised (computational) MRD analysis; UnSup-combined MRD, unsupervised MRD applying criteria of MRD-positive = aberrant blasts in at least 2 of the 3 antibody combinations (standard and stem cell), MRD-negative or equivocal = aberrant blasts in 0-1 of 3 antibody combinations.

combinations) to select patients with the most extensive immunophenotypic blast aberrancies. Most patients with test positivity by these criteria had conventional MRD levels ≥ 0.2% (Data Supplement). The 2-year CIR after transplant for patients with a positive test was 50.5% compared with 20.6% for patients with a negative or equivocal test (Gray's test *P* value < .001) (Fig 3D, Table 2), and the overall accuracy for relapse prediction was 73% (Data Supplement).

Chimerism and Transplant Outcome

To explore the contribution of a putative GVL effect to post-transplant outcome, we studied the impact of acquisition of full donor T-cell chimerism (FDTCC) on transplant outcome. Acquisition of FDTCC was similar in control and experimental arms and not affected by pretransplant MRD status (Data Supplement). Acquisition of FDTCC at 3 months post-transplant was associated with lower CIR (Gray's test *P* value < .001) with 2-year CIR in patients with FDTCC of 13.1% (95% CI, 7.3 to 20.5) compared with 44.8% (95% CI, 30.0 to 58.3) (Data Supplement). In patients with pretransplant MRD positivity, acquisition of FDTCC at 3 months post-transplant was associated with a

comparable outcome with that achieved by patients without detectable pretransplant MRD (Fig 4).

DISCUSSION

Strategies with the potential to reduce the risk of relapse in patients with AML or MDS transplanted using RIC include both intensification of the antitumor properties of the conditioning regimen³⁵ and optimization of the GVL effect.⁵ The cytoreductive properties of distinct RIC regimens vary considerably, and relapse rates ranging from 30% to 60% have been reported in patients using commonly adopted transplant protocols.^{3,36} In unrandomized phase II trials and retrospective registry data, the FLAMSA-Bu protocol, which incorporates additional cytoreductive chemotherapy prior to a fludarabine-based RIC regimen, has been reported to reduce relapse and improve outcome in high-risk AML or MDS and as a consequence has become widely adopted despite its attendant substantially increased inpatient stay and potential toxicity.⁵⁻⁷ Our data, however, show no impact on either relapse rate or survival in patients transplanted using this intensified regimen. Differences in control regimens and age-related FLAMSA-Bu dose adjustments constitute potential limitations to this analysis,

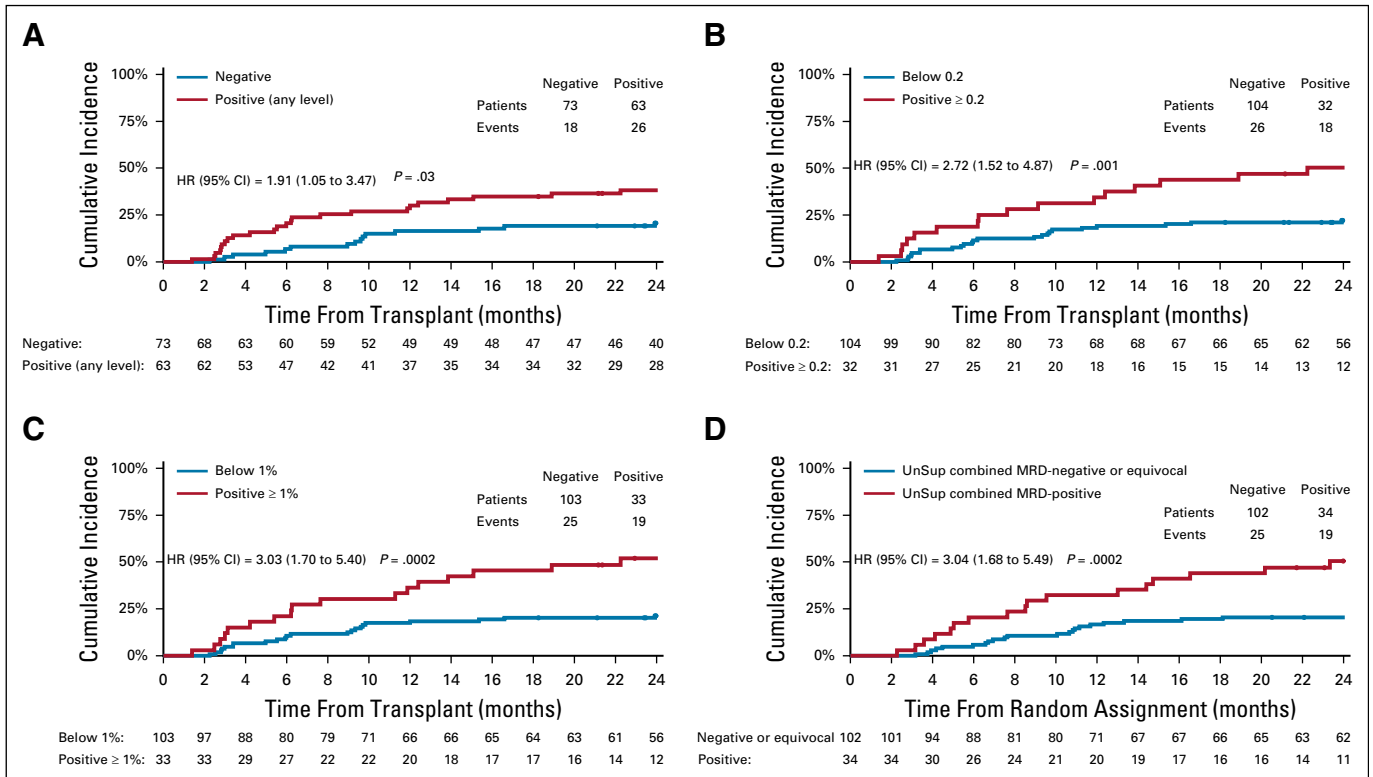


FIG 3. (A) CIR by conventional MFC-MRD status, (B) CIR by conventional MFC-MRD with 0.2% cutoff, (C) CIR by unsupervised (computational) MFC-MRD with 1% cutoff, (D) CIR by unsupervised (computational) combined MFC-MRD test status. Impact of flow cytometric MRD on outcomes of transplanted patients are shown with comparison of results from conventional and unsupervised (computational) analysis approaches. Unsupervised MFC-MRD cutoff is higher than conventional MFC-MRD as the former values summate all quantifiable nonoverlapping abnormal blast subpopulations from an antibody combination, whereas conventional MFC-MRD values are from a single LAIP. UnSup-combined MRD is unsupervised MRD applying criteria of MRD-positive = aberrant blasts in at least two of the three antibody combinations (standard and stem cell) and MRD-negative or equivocal = aberrant blasts in 0-1 of 3 antibody combinations. CIR, Cumulative incidence of relapse; LAIP, leukemia-associated immunophenotype; MFC-MRD, flow cytometric measurable residual disease; UnSup, unsupervised (computational) MRD analysis.

but we did not detect a differential effect on outcomes from any of these variables. Of particular note, FLAMSA-Bu did not result in improved survival in predefined subgroups including patients with an adverse-risk karyotype.

In exploratory studies, pretransplant MRD, measured using a widely used flow cytometric methodology, was prospectively examined as a prognostic determinant of transplant outcome. Pretransplant MRD status was identified as an important prognostic factor for relapse in multivariable analysis, confirming previous retrospective analyses. However, although the US CTN 0901 trial identified the presence of NGS-determined pretransplant MRD as a predictor of outcome in patients transplanted using a reduced intensity but not a MAC regimen,³¹ in the FIGARO trial, we observed no benefit accruing in MRD-positive patients from intensification of RI conditioning. Of interest, the risk of relapse after transplant in the RIC arm of US-CTN 0901 (48% at 18 months²) was strikingly higher than that observed in the FIGARO trial despite both trials using similar RIC regimens.

One of the major limitations of the widely used flow cytometric MRD assays has been the inevitable subjectivity from manual gating of immunophenotypic raw data. Using a novel unsupervised analysis approach as independent evaluation of conventional flow cytometric MRD, we were able to confirm the reproducibility of the prognostic significance of immunophenotypic pretransplant MRD in this older age group typically considered for RIC regimens. Incorporating comprehensive genetic information for genetic subtype MRD interpretation (such as *FLT3*-ITD^{29,31}), consideration of under-representation of MRD from hemodilution or hypoplasia and potentially MRD as a continuous variable will further progress refining and validating flow cytometric MRD thresholds for transplant decision making.

There is much debate concerning the benefit of an RIC allograft in patients with evidence of pretransplant MRD. It is therefore of interest that approximately 50% of FIGARO patients with evidence of pretransplant MRD did not relapse, confirming the validity of transplantation using an RIC regimen as a therapeutic strategy in high-risk

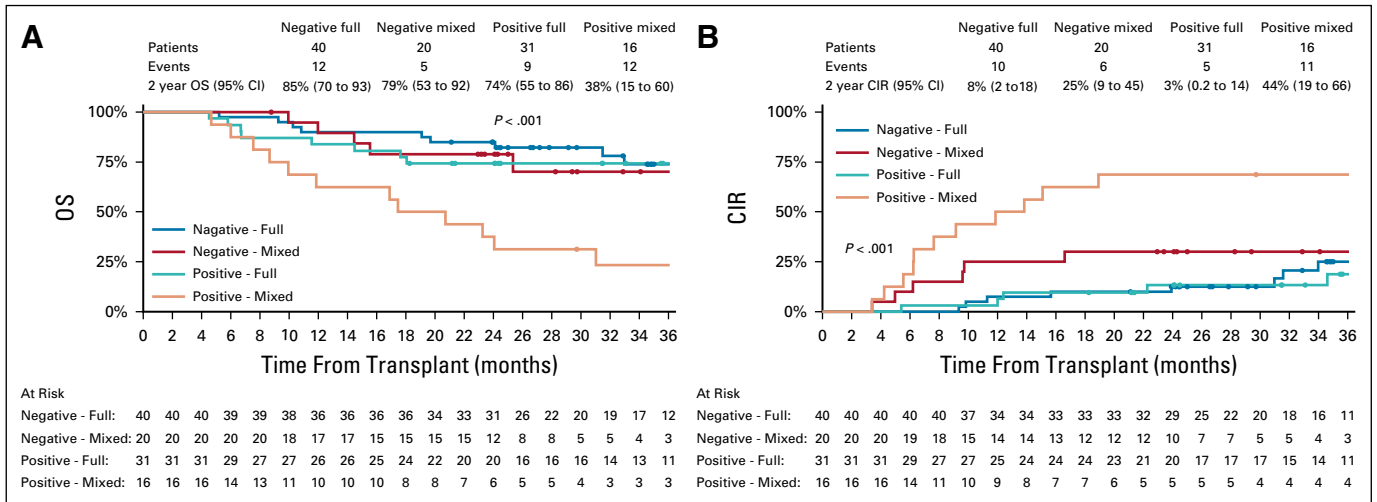


FIG 4. (A) OS by month 3 Chimerism with pretransplant MRD status and (B) CIR by month 3 Chimerism with pretransplant MRD status. Outcomes are for transplanted patients who were alive and relapse-free at day + 100. MRD status is by conventional flow MRD. Negative—full, pretransplant MRD-negative and month 3 full donor T-cell chimerism. Positive—full, pretransplant MRD-positive and month 3 full donor T-cell chimerism. Negative—mixed, pretransplant MRD-negative and month 3 mixed donor T-cell chimerism. Positive—mixed, pretransplant MRD-positive and month 3 mixed donor T-cell chimerism. CIR, Cumulative incidence of relapse; MRD, measurable residual disease; OS, overall survival.

AML—even in patients with detectable MRD. There is compelling evidence of a potent GVL effect in patients with AML allografted using an RIC regimen.³⁷ The observation that the adverse prognostic impact conferred by the presence of pretransplant MRD was mitigated by the acquisition of FDTCC at 3 months requires further prospective

examination and identifies optimization of the GVL effect as an important approach to improve outcome in patients transplanted using an RIC regimen. Such strategies include using a T replete graft, a rapid taper of post-transplant immunosuppression, or early administration of pharmacological agents such as azacitidine, decitabine,³⁸⁻⁴⁰ or DLI.

AFFILIATIONS

¹Centre for Clinical Haematology, Queen Elizabeth Hospital, Birmingham, United Kingdom

²Cancer Research UK Clinical Trials Unit, University of Birmingham, United Kingdom

³Institute of Immunology and Immunotherapy, University of Birmingham, United Kingdom.

⁴Addenbrookes Hospital, Cambridge, United Kingdom

⁵Churchill Hospital, Oxford, United Kingdom

⁶St James's Hospital, Leeds, United Kingdom

⁷Royal Liverpool University Hospital, United Kingdom

⁸Manchester Royal Infirmary, Manchester, United Kingdom

⁹Kings College Hospital, London, United Kingdom

¹⁰Bristol Haematology and Oncology Centre, United Kingdom

¹¹Leicester Royal Infirmary, United Kingdom

¹²Queen Elizabeth University Hospital, Glasgow, United Kingdom

¹³University Hospital Wales, United Kingdom

¹⁴Imperial College Hospital, London, United Kingdom

¹⁵Centre for Clinical Haematology, Nottingham, United Kingdom

¹⁶Department of Medical and Molecular Genetics, King's College, London, United Kingdom

CORRESPONDING AUTHOR

Charles Craddock, MD, Centre for Clinical Haematology, Queen Elizabeth Hospital, Birmingham B15 2TH; e-mail: Charles.Craddock@uhb.nhs.uk.

SUPPORT

Supported by Blood Cancer UK and Cure Leukemia. Pierre Fabre and Eurocept provided busulphan and amasacrine for trial purposes in addition to unrestricted educational support.

CLINICAL TRIAL INFORMATION

FIGARO

AUTHORS' DISCLOSURES OF POTENTIAL CONFLICTS OF INTEREST

Disclosures provided by the authors are available with this article at DOI <https://doi.org/10.1200/JCO.20.02308>.

AUTHOR CONTRIBUTIONS

Conception and Design: Charles Craddock, Aimee Jackson, Shamyala Siddique, Sandeep Nagra, Andrew Peniket, Eleni Tholouli, Sylvie D. Freeman, Keith Wheatley, Paresh Vyas, Ann Hunter

Administrative support: Shamyala Siddique

Provision of study material and patients: Sandeep Nagra, Ram Malladi, Andrew Peniket, Maria Gilleece, Rahuman Salim, Eleni Tholouli, Victoria Potter, Charles Crawley, Rachel Protheroe, Paresh Vyas, Ann Hunter, Anne Parker, Jenny Byrne

Collection and assembly of data: Charles Craddock, Aimee Jackson, Justin Loke, Andrea Hodgkinson, John Mason, Sandeep Nagra, Ram Malladi, Andrew Peniket, Maria Gilleece, Rahuman Salim, Eleni Tholouli, Victoria

Potter, Charles Crawley, Rachel Protheroe, Anne Parker, Keith Wilson, Jiri Pavlu, Jenny Byrne, Richard Dillon, Naeem Khan, Sylvie D. Freeman
Data analysis and interpretation: Charles Craddock, Aimee Jackson, Justin Loke, Shamyla Siddique, Andrea Hodgkinson, Georgia Andrew, Keith Wheatley, Richard Dillon, Nicholas McCarthy, Sylvie D. Freeman

Manuscript writing: All authors

Final approval of manuscript: All authors

Accountable for all aspects of the work: All authors

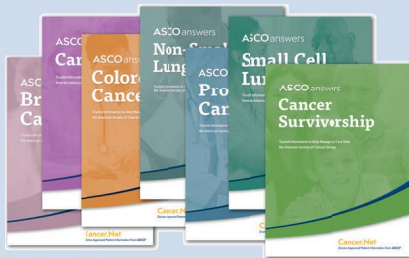
REFERENCES

- Jethava YS, Sica S, Savani B, et al: Conditioning regimens for allogeneic hematopoietic stem cell transplants in acute myeloid leukemia. *Bone Marrow Transpl* 52:1504-1511, 2017
- Scott BL, Pasquini MC, Logan BR, et al: Myeloablative versus reduced-intensity hematopoietic cell transplantation for acute myeloid leukemia and myelodysplastic syndromes. *J Clin Oncol* 35:1154-1161, 2017
- Beelen DW, Trenschele R, Stelljes M, et al: Treosulfan or busulfan plus fludarabine as conditioning treatment before allogeneic haemopoietic stem cell transplantation for older patients with acute myeloid leukaemia or myelodysplastic syndrome (MC-FludT. 14/L): A randomised, non-inferiority, phase 3 trial. *Lancet Haematol* 7:e28-e39, 2020
- Ciurea SO, Kongtim P, Varma A, et al: Is there an optimal conditioning for older patients with AML receiving allogeneic hematopoietic cell transplantation? *Blood* 135:449-452, 2020
- Schmid C, Schleuning M, Ledderose G, et al: Sequential regimen of chemotherapy, reduced-intensity conditioning for allogeneic stem-cell transplantation, and prophylactic donor lymphocyte transfusion in high-risk acute myeloid leukemia and myelodysplastic syndrome. *J Clin Oncol* 23:5675-5687, 2005
- Schmid C, Schleuning M, Schwerdtfeger R, et al: Long-term survival in refractory acute myeloid leukemia after sequential treatment with chemotherapy and reduced-intensity conditioning for allogeneic stem cell transplantation. *Blood* 108:1092-1099, 2006
- Malard F, Labopin M, Stuhler G, et al: Sequential intensified conditioning regimen allogeneic hematopoietic stem cell transplantation in adult patients with intermediate- or high-risk acute myeloid leukemia in complete remission: A study from the Acute Leukemia Working Party of the European Group for Blood and Marrow Transplantation. *Biol Blood Marrow Transpl* 23:278-284, 2017
- Maurillo L, Buccisano F, Del Principe M, et al: Toward optimization of postremission therapy for residual disease-positive patients with acute myeloid leukemia. *J Clin Oncol* 26:4944-4951, 2008
- Terwijn M, van Putten WL, Kelder A, et al: High prognostic impact of flow cytometric minimal residual disease detection in acute myeloid leukemia: Data from the HOVON/SAKK AML 42A study. *J Clin Oncol* 31:3889-3897, 2013
- Freeman SD, Hills RK, Virgo P, et al: Measurable residual disease at induction redefines partial response in acute myeloid leukemia and stratifies outcomes in patients at standard risk without NPM1 mutations. *J Clin Oncol* 36:1486-1497, 2018
- Balsat M, Renneville A, Thomas X, et al: Postinduction minimal residual disease predicts outcome and benefit from allogeneic stem cell transplantation in acute myeloid leukemia with NPM1 mutation: A study by the Acute Leukemia French Association Group. *J Clin Oncol* 35:185-193, 2017
- Ivey A, Hills RK, Simpson MA, et al: Assessment of minimal residual disease in standard-risk AML. *N Engl J Med* 374:422-433, 2016
- Jongen-Lavrencic M, Grob T, Hanekamp D, et al: Molecular minimal residual disease in acute myeloid leukemia. *N Engl J Med* 378:1189-1199, 2018
- Morita K, Kantarjian HM, Wang F, et al: Clearance of somatic mutations at remission and the risk of relapse in acute myeloid leukemia. *J Clin Oncol* 36:1788-1797, 2018
- Rucker FG, Agrawal M, Corbacioglu A, et al: Measurable residual disease monitoring in acute myeloid leukemia with t(8;21)(q22;q22.1): Results from the AML Study Group. *Blood* 134:1608-1618, 2019
- Schuurhuis GJ, Heuser M, Freeman S, et al: Minimal/measurable residual disease in AML: A consensus document from the European LeukemiaNet MRD Working Party. *Blood* 131:1275-1291, 2018
- Walter RB, Buckley SA, Pagel JM, et al: Significance of minimal residual disease before myeloablative allogeneic hematopoietic cell transplantation for AML in first and second complete remission. *Blood* 122:1813-1821, 2013
- Walter RB, Gooley TA, Wood BL, et al: Impact of pretransplantation minimal residual disease, as detected by multiparametric flow cytometry, on outcome of myeloablative hematopoietic cell transplantation for acute myeloid leukemia. *J Clin Oncol* 29:1190-1197, 2011
- Araki D, Wood BL, Othus M, et al: Allogeneic hematopoietic cell transplantation for acute myeloid leukemia: Time to move toward a minimal residual disease-based definition of complete remission? *J Clin Oncol* 34:329-336, 2016
- Oran B, Jorgensen JL, Marin D, et al: Pre-transplantation minimal residual disease with cytogenetic and molecular diagnostic features improves risk stratification in acute myeloid leukemia. *Haematologica* 102:110-117, 2017
- Anthias C, Dignan FL, Morilla R, et al: Pre-transplant MRD predicts outcome following reduced-intensity and myeloablative allogeneic hemopoietic SCT in AML. *Bone Marrow Transpl* 49:679-683, 2014
- Buccisano F, Maurillo L, Picocchi A, et al: Pre-transplant persistence of minimal residual disease does not contraindicate allogeneic stem cell transplantation for adult patients with acute myeloid leukemia. *Bone Marrow Transpl* 52:473-475, 2017
- Ustun C, Courville EL, DeFor T, et al: Myeloablative, but not reduced-intensity, conditioning overcomes the negative effect of flow-cytometric evidence of leukemia in acute myeloid leukemia. *Biol Blood Marrow Transpl* 22:669-675, 2016
- Gilleece MH, Labopin M, Yakoub-Agha I, et al: Measurable residual disease, conditioning regimen intensity, and age predict outcome of allogeneic hematopoietic cell transplantation for acute myeloid leukemia in first remission: A registry analysis of 2292 patients by the Acute Leukemia Working Party European Society of Blood and Marrow Transplantation. *Am J Hematol* 93:1142-1152, 2018
- Walter RB, Gyurkocza B, Storer BE, et al: Comparison of minimal residual disease as outcome predictor for AML patients in first complete remission undergoing myeloablative or nonmyeloablative allogeneic hematopoietic cell transplantation. *Leukemia* 29:137-144, 2015
- Getta BM, Devlin SM, Levine RL, et al: Multicolor flow cytometry and multigene next-generation sequencing are complementary and highly predictive for relapse in acute myeloid leukemia after allogeneic transplantation. *Biol Blood Marrow Transpl* 23:1064-1071, 2017
- Buckley SA, Wood BL, Othus M, et al: Minimal residual disease prior to allogeneic hematopoietic cell transplantation in acute myeloid leukemia: A meta-analysis. *Haematologica* 102:865-873, 2017

28. Guolo F, Minetto P, Clavio M, et al: Combining flow cytometry and WT1 assessment improves the prognostic value of pre-transplant minimal residual disease in acute myeloid leukemia. *Haematologica* 102:e348-e351, 2017
29. Dillon R, Hills R, Freeman S, et al: Molecular MRD status and outcome after transplantation in NPM1-mutated AML. *Blood* 135:680-688, 2020
30. Thol F, Gabdoulline R, Liebich A, et al: Measurable residual disease monitoring by NGS before allogeneic hematopoietic cell transplantation in AML. *Blood* 132:1703-1713, 2018
31. Hourigan CS, Dillon LW, Gui G, et al: Impact of conditioning intensity of allogeneic transplantation for acute myeloid leukemia with genomic evidence of residual disease. *J Clin Oncol* 38:1273-1283, 2020
32. Grimwade D, Hills RK, Moorman AV, et al: Refinement of cytogenetic classification in acute myeloid leukemia: Determination of prognostic significance of rare recurring chromosomal abnormalities among 5876 younger adult patients treated in the United Kingdom Medical Research Council Trials. *Blood* 116:354-365, 2010
33. Przepiorka D, Weisdorf D, Martin P, et al: 1994 consensus conference on acute GVHD grading. *Bone Marrow Transpl* 15:825-828, 1995
34. Shulman HM, Sullivan KM, Weiden PL, et al: Chronic graft-versus-host syndrome in man. A long-term clinicopathologic study of 20 Seattle patients. *Am J Med* 69:204-217, 1980
35. Rambaldi A, Grassi A, Masciulli A, et al: Busulfan plus cyclophosphamide versus busulfan plus fludarabine as a preparative regimen for allogeneic haemopoietic stem-cell transplantation in patients with acute myeloid leukaemia: An open-label, multicentre, randomised, phase 3 trial. *Lancet Oncol* 16:1525-1536, 2015
36. Blaise D, Tabrizi R, Boher JM, et al: Randomized study of 2 reduced-intensity conditioning strategies for human leukocyte antigen-matched, related allogeneic peripheral blood stem cell transplantation: Prospective clinical and socioeconomic evaluation. *Cancer* 119:602-611, 2013
37. Craddock C, Nagra S, Peniket A, et al: Factors predicting long-term survival after T-cell depleted reduced intensity allogeneic stem cell transplantation for acute myeloid leukemia. *Haematologica* 95:989-995, 2010
38. Craddock C, Jilani N, Siddique S, et al: Tolerability and clinical activity of post-transplantation azacitidine in patients allografted for acute myeloid leukemia treated on the RICAZA trial. *Biol Blood Marrow Transpl* 22:385-390, 2016
39. de Lima M, Oran B, Champlin RE, et al: CC-486 maintenance after stem cell transplantation in patients with acute myeloid leukemia or myelodysplastic syndromes. *Biol Blood Marrow Transpl* 24:2017-2024, 2018
40. Gao L, Zhang Y, Wang S, et al: Effect of rhG-CSF combined with decitabine prophylaxis on relapse of patients with high-risk MRD-negative AML after HSCT: An open-label, multicenter, randomized controlled trial. *J Clin Oncol* 38:4249-4259, 2020



ASCO Answers—The Ideal Take-Home Patient Education Resource



ASCO has created helpful resources to support your patients and their caregivers. **ASCO Answers** patient education materials provide trusted information on cancer types, diagnosis, treatment, side effects, and coping in three convenient formats: fact sheets, topic-specific booklets, and comprehensive, patient-friendly guides.

ASCO Answers can be purchased from the ASCO Store at cancer.net/estore. Free domestic shipping. Members save 20%.

Cancer.Net[™]

Doctor-Approved Patient Information from ASCO®

AUTHORS' DISCLOSURES OF POTENTIAL CONFLICTS OF INTEREST**Augmented Reduced-Intensity Regimen Does Not Improve Postallogeic Transplant Outcomes in Acute Myeloid Leukemia**

The following represents disclosure information provided by authors of this manuscript. All relationships are considered compensated unless otherwise noted. Relationships are self-held unless noted. I = Immediate Family Member, Inst = My Institution. Relationships may not relate to the subject matter of this manuscript. For more information about ASCO's conflict of interest policy, please refer to www.asco.org/rwc or ascopubs.org/jco/authors/author-center.

Open Payments is a public database containing information reported by companies about payments made to US-licensed physicians ([Open Payments](#)).

Charles Craddock

Honoraria: Abbvie, Celgene, Jazz Pharmaceuticals, Janssen, Pfizer, Daiichi Sankyo, Amgen, Astellas Pharma

Consulting or Advisory Role: Daiichi Sankyo, Abbvie, Janssen, Novartis, Bristol-Myers Squibb, Pfizer, Astellas Pharma, Daiichi Sankyo, eurocept

Speakers' Bureau: Abbvie, Janssen, Novartis, Roche, Bristol-Myers Squibb, Pfizer, Astellas Pharma, Daiichi Sankyo, Eurocept

Research Funding: Celgene, Jazz Pharmaceuticals, Kite Pharma, Jazz Pharmaceuticals

Expert Testimony: Daiichi Sankyo

Travel, Accommodations, Expenses: Celgene, Jazz Pharmaceuticals, Daiichi Sankyo

Justin Loke

Honoraria: Amgen, Janssen-Cilag

Travel, Accommodations, Expenses: Novartis, Daiichi Sankyo Europe GmbH

Shamyla Siddique

Honoraria: Celgene

Ram Malladi

Consulting or Advisory Role: Roche

Travel, Accommodations, Expenses: Amgen, Novartis, Gilead Sciences

Andrew Peniket

Consulting or Advisory Role: Jazz Pharmaceuticals

Speakers' Bureau: Merck

Maria Gilleece

Stock and Other Ownership Interests: GlaxoSmithKline

Consulting or Advisory Role: Jazz Pharmaceuticals

Speakers' Bureau: Jazz Pharmaceuticals

Travel, Accommodations, Expenses: Gilead sciences, Jazz Pharmaceuticals

Eleni Tholouli

Consulting or Advisory Role: Novartis, Gilead Sciences, Daiichi Sankyo, Jazz Pharmaceuticals, Astellas Pharma, Celgene, Pfizer

Speakers' Bureau: Novartis, Kite/Gilead, Jazz Pharmaceuticals, Pfizer, Janssen

Travel, Accommodations, Expenses: Jazz Pharmaceuticals, MSD

Victoria Potter

Consulting or Advisory Role: Eurocept, Jazz Pharmaceuticals

Speakers' Bureau: Pfizer

Charles Crawley

Travel, Accommodations, Expenses: Funding for attending ASH meeting in dec 2019

Keith Wheatley

Research Funding: Roche

Rachel Protheroe

Honoraria: Astellas Pharma, Jazz Pharmaceuticals, Gilead Sciences, Abbvie, Hartley Taylor Medical

Consulting or Advisory Role: Kiadis Pharma

Travel, Accommodations, Expenses: Astellas Pharma, Kite/Gilead, Jazz Pharmaceuticals

Pareesh Vyas

Stock and Other Ownership Interests: OxStem

Honoraria: Celgene, Pfizer, Jazz Pharmaceuticals, Abbvie, Daiichi Sankyo

Research Funding: Celgene, Forty Seven

Patents, Royalties, Other Intellectual Property: Patent for flow cytometric detection of leukaemic stem cells

Anne Parker

Consulting or Advisory Role: MSD, Pfizer, Gilead Sciences

Speakers' Bureau: Jazz Pharmaceuticals

Travel, Accommodations, Expenses: Gilead Sciences, MSD

Keith Wilson

Honoraria: Kite/Gilead, Novartis

Consulting or Advisory Role: Kite/Gilead

Speakers' Bureau: Kite/Gilead

Travel, Accommodations, Expenses: Kite/Gilead

Jiri Pavlu

Consulting or Advisory Role: Jazz Pharmaceuticals, Daiichi Sankyo

Speakers' Bureau: Jazz Pharmaceuticals

Travel, Accommodations, Expenses: Jazz Pharmaceuticals, Daiichi Sankyo

Jenny Byrne

Honoraria: Novartis Pharmaceuticals UK Ltd, ARIAD/Incyte, Jazz Pharmaceuticals

Richard Dillon

Honoraria: Abbvie, Pfizer, Novartis, Jazz Pharmaceuticals, Astellas Pharma

Consulting or Advisory Role: Abbvie, Novartis, Pfizer, Jazz Pharmaceuticals

Research Funding: Amgen, Abbvie

Naeem Khan

Research Funding: Oxford Biomedica

Sylvie D. Freeman

Speakers' Bureau: Jazz Pharmaceuticals, Novartis

Patents, Royalties, Other Intellectual Property: Vyas P, Goardon N, Freeman S: Detection of Acute Myeloid Leukaemia, 2011. US Patent Application 13/995347. Granted 2018

Travel, Accommodations, Expenses: BD Biosciences

No other potential conflicts of interest were reported.