

Supplementary Information

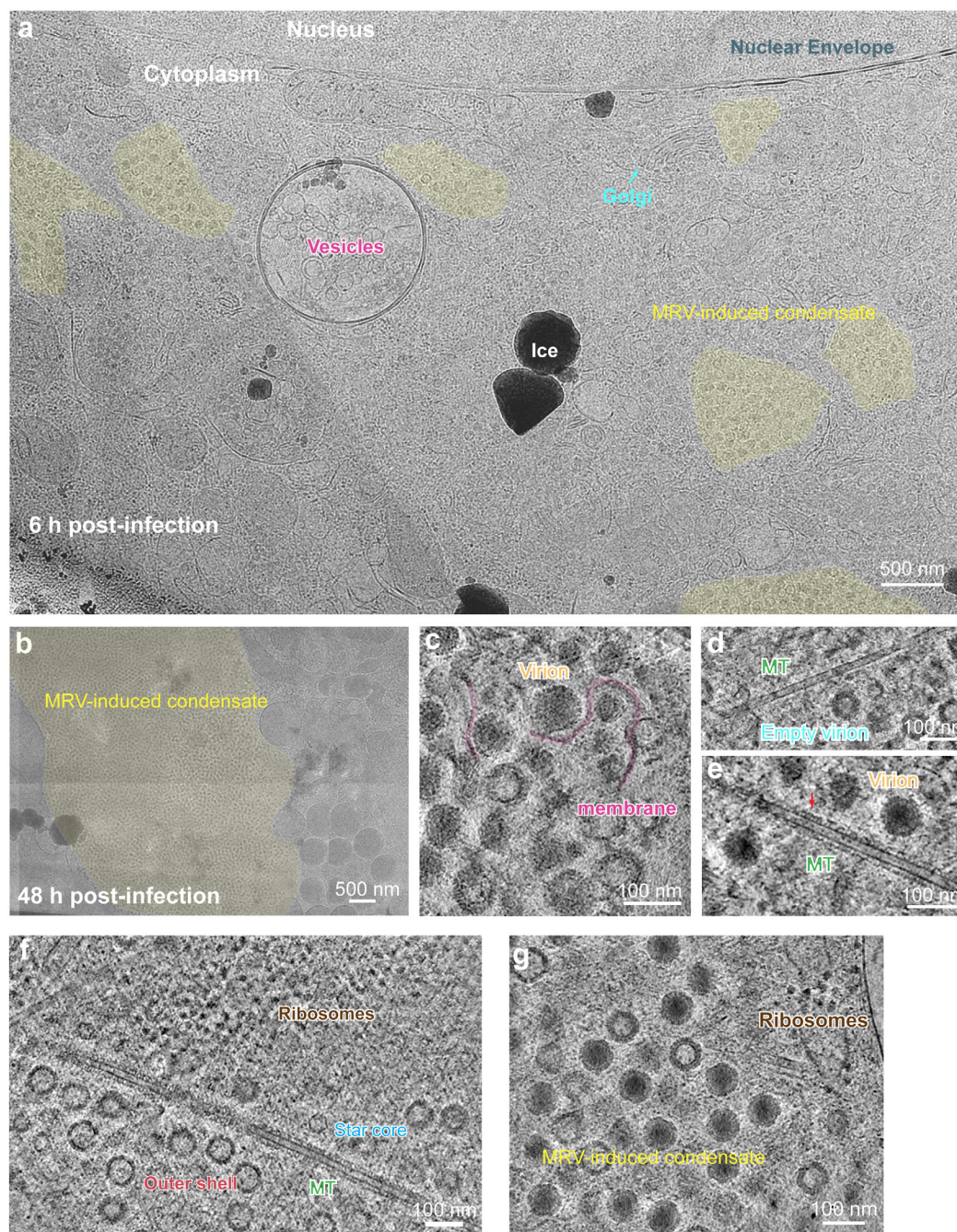


Fig. S1: MRV-induced cellular condensates captured by cryoET. **a**, Low magnification cryoEM image of the lamella from cells at 6 h post-infection. **b**, Low magnification cryoEM image of the lamella from cells at 48 h post-infection. **c-g**, Slice images from cryoET reconstructions showing different assembly intermediates and their interplay with host cell components, such as ribosomes, membranes, and microtubules (MT). The undecorated microtubule is shown in (d), and the decorated one is shown in (e). The decoration is indicated by a red arrow.

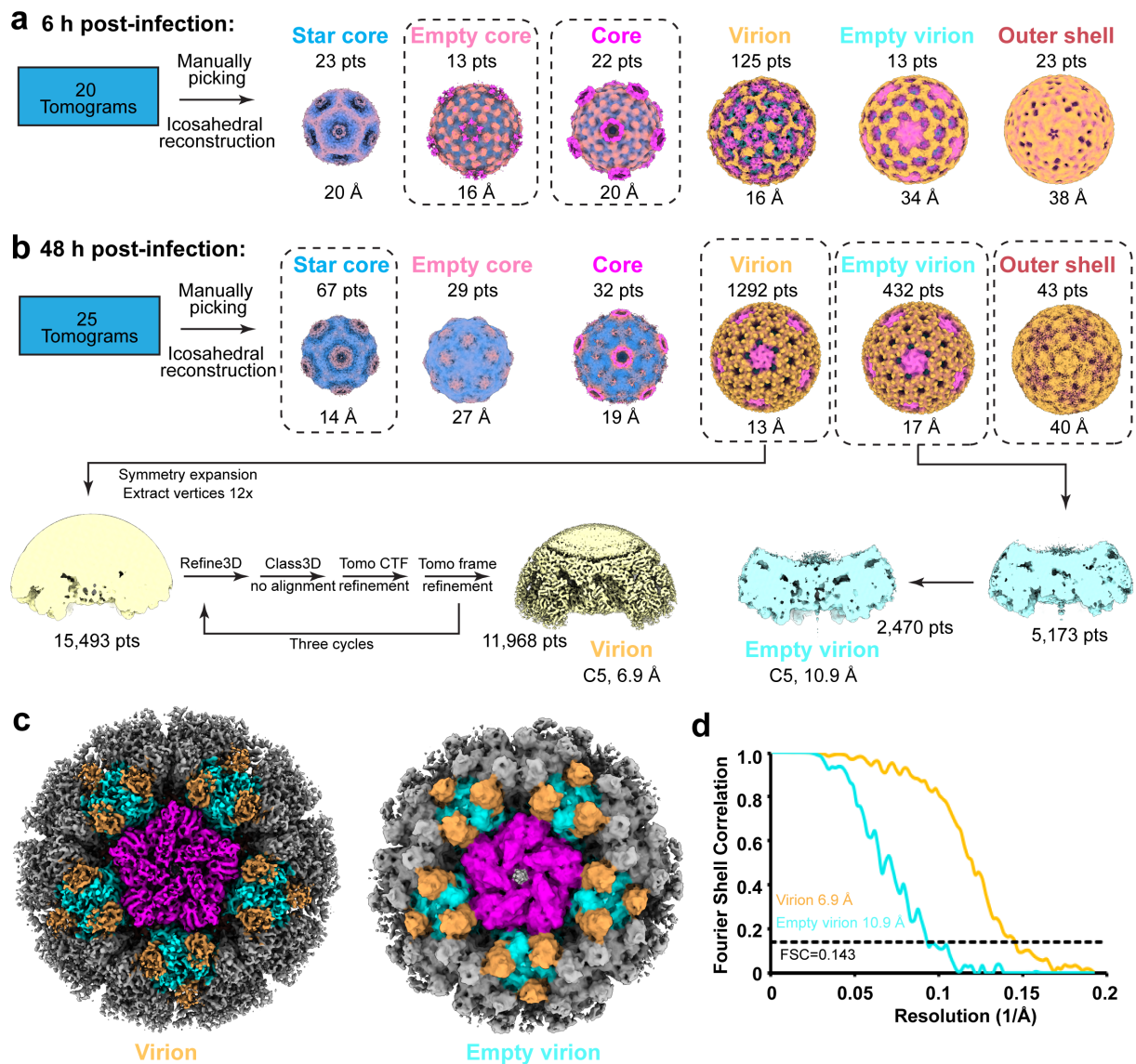


Fig. S2: In situ cryoET Data processing. **a**, Workflow of subtomogram averaging of cryoET data from cells at 6 h post MRV infection. Six assembly intermediates are captured. **b**, Workflow of subtomogram averaging of cryoET data from cells at 48 h post MRV infection. Six assembly intermediates are captured. Virion and empty virion were subjected to sub-particle reconstruction to improve resolution. Icosahedral reconstructions are colored by radius in a and b. **c**, Density map of virion and empty virion after sub-particle reconstruction from (b). **d**, Gold-standard Fourier shell correlation (FSC) curves for subtomogram averaging of virion and empty virion from (b).

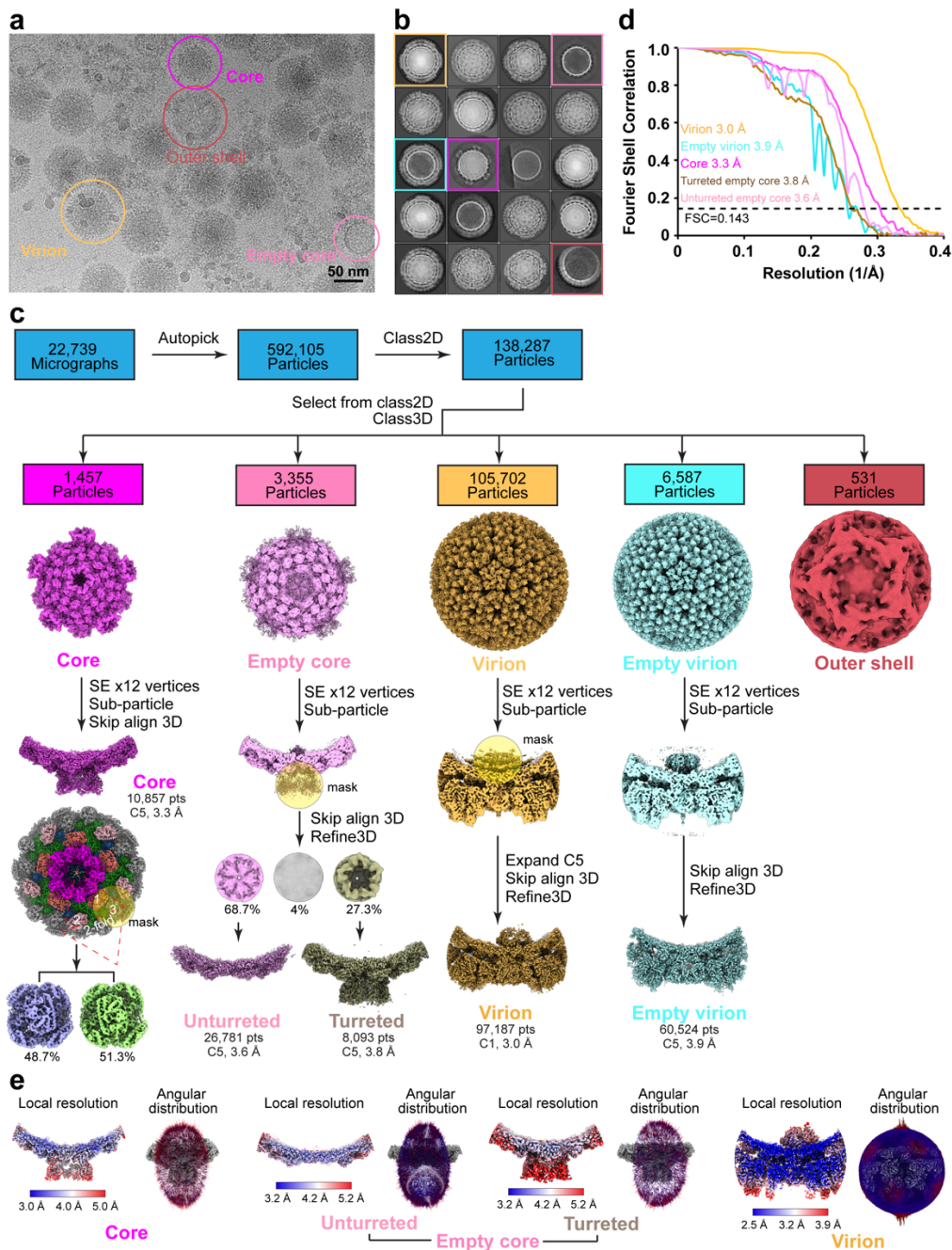


Fig. S3: CryoEM data processing of the MRV viral isolate from host cells at 6 h post-infection. **a**, Representative cryoEM image of viral isolate from host cells. Different assembly intermediates are indicated by circles and labelled. **b**, Selected 2D class averages of the cryoEM particles. Colored boxes showing different assembly intermediates. **c**, Flow chart of cryoEM data processing showing five assembly intermediates. Yellow circles on sub-particle reconstructions of core, empty core and virion show the masked area of focused classification. **d**, Gold-standard Fourier shell correlation (FSC) curves for cryoEM maps from sub-particle reconstructions. **e**, Maps colored to local resolution and representations of the angular distribution of particles used in the final reconstruction.

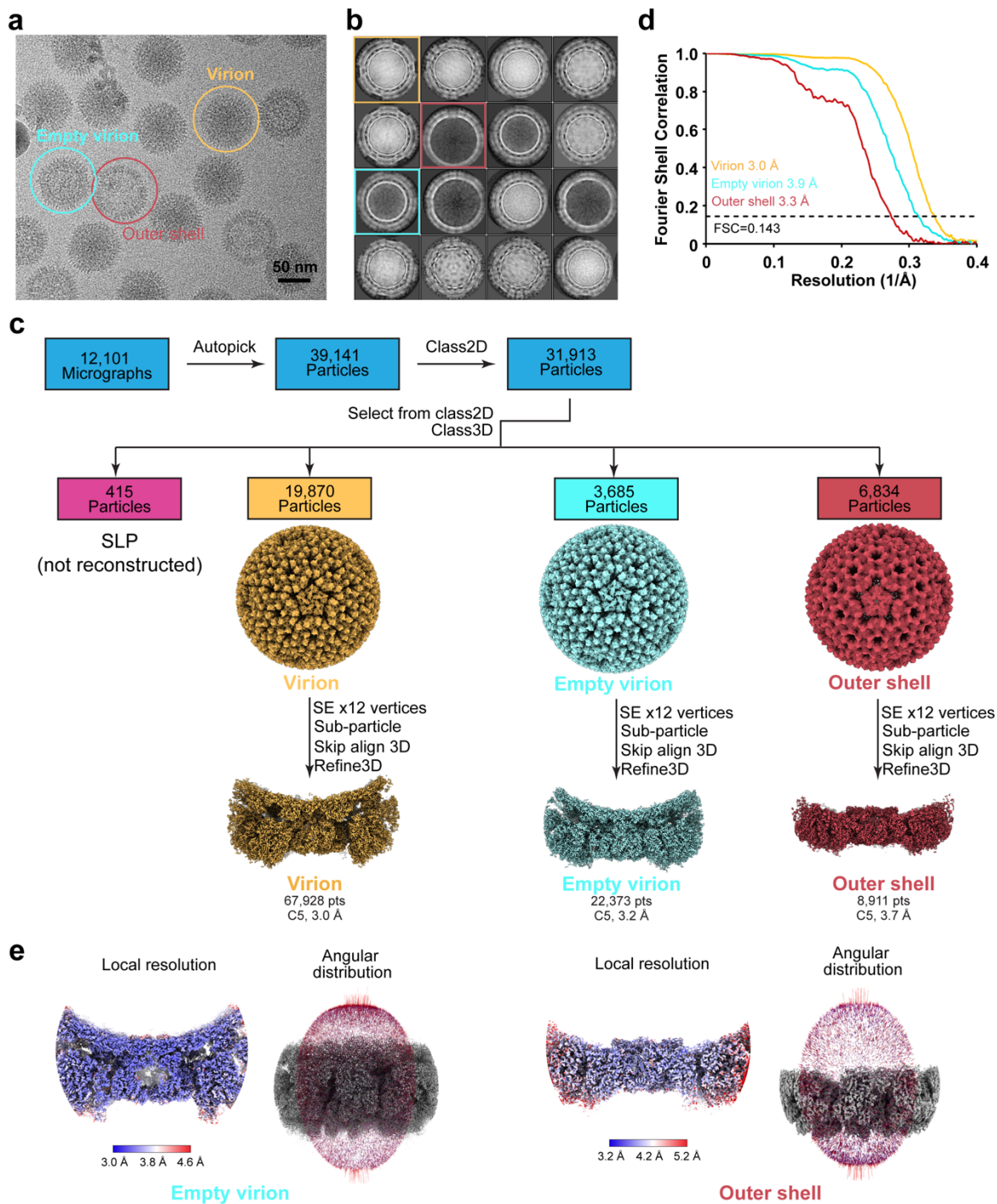
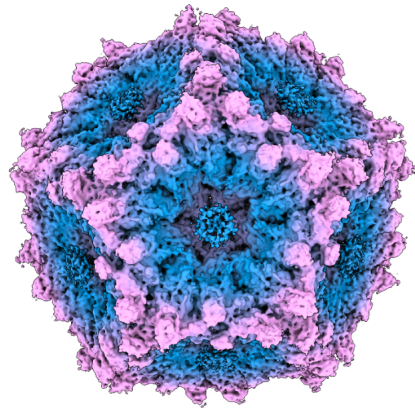


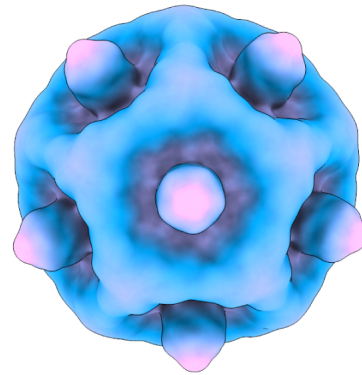
Fig. S4: CryoEM data processing of the MRV viral isolate from host cells at 48 h post-infection. **a**, Representative cryoEM image of viral isolate from host cells. **b**, Selected 2D class averages of the cryoEM particles. Colored boxes showing different assembly intermediates. **c**, Flow chart of cryoEM data processing showing three assembly intermediates. **d**, Gold-standard Fourier shell correlation (FSC) curves for cryoEM maps from sub-particle reconstructions. **e**, Maps colored to local resolution and representations of the angular distribution of particles used in the final reconstruction.

a Mammalian reovirus



EMD-22165

b Rotavirus



EMD-16762

Fig. S5: Star-shaped SLP of mammalian reovirus (MRV) and rotavirus from previous studies. CryoEM maps are colored by radius.

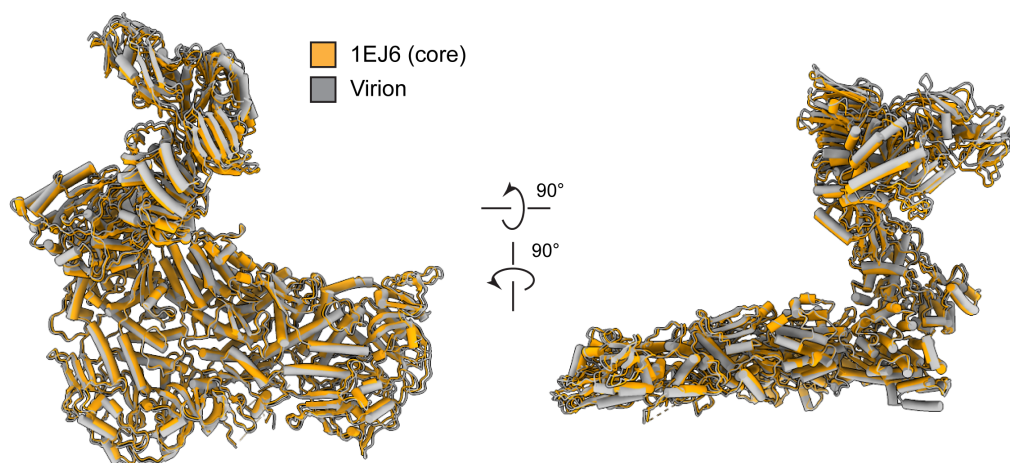


Fig. S6: Structure superposition of $\lambda 2$ from core (published crystal structure) and virion (reported in this study). The two structures are almost identical but different from the cryoEM structure of MRV core isolated from cell cytoplasm reported in this study.

Supplementary Table

Table S1. Summary for CryoET data collection and subtomogram averaging of cells at 6 h post-infection

	Star core	Core EMD-47313	Empty core EMD-47314	Virion	Empty virion	Outer shell
Magnification			42,000			
Voltage (kV)			300			
Pixel size (Å)			1.965			
Total dose (e⁻/Å²)			100			
Defocus range (μm)			-3.5 to -5			
Acquisition scheme			Dose symmetric scheme -60° - 40° or -40° - 60°			
Tilt series number			20			
Symmetry imposed	I3	I3	I3	I3	I3	I3
Particle number	23	13	22	125	13	23
Map resolution (Å)	20	16	20	16	34	38
FSC threshold	0.143	0.143	0.143	0.143	0.143	0.143

Table S2. Summary for CryoET data collection and subtomogram averaging of cells at 48 h post-infection

	Star core	Core	Empty core	Virion	Empty virion	Outer shell
	EMD-47315			EMD-47316 (EMD-47317)	EMD-47318	EMD-47319
Magnification				33,000		
Voltage (kV)				300		
Pixel size (Å)				2.6		
Total dose (e⁻/Å²)				100		
Defocus range (µm)				-3.5 to -5		
Acquisition scheme				Dose symmetric scheme -60° - 40° or -40° - 60°		
Tilt series number				25		
Symmetry imposed	I3	I3	I3	I3 (C5)*	I3 (C5)*	I3
Particle number	67	29	32	1292 (11,968)	432 (2,470)	43
Map resolution (Å)	14	27	19	13 (6.9)	17 (10.9)	40
FSC threshold	0.143	0.143	0.143	0.143	0.143	0.143

*Numbers in the parentheses are results from sub-particle reconstructions.

Table S3. CryoEM data collection, refinement and validation statistics of MRV isolate from cells at 6 h post-infection

	Core	Unturreted empty core	Turreted empty core	Virion	Empty virion
EMDB PDB	EMD-46053 9CYX	EMD-47321	EMD-47322	EMD-46054 9CYY	
Data collection and processing					
Magnification			81,000		
Voltage (kV)			300		
Electron exposure (e ⁻ /Å ²)			50		
Defocus range (μm)			-1.8 to -2.6		
Pixel size (Å)			1.1		
Symmetry imposed	C5	C5	C5	C1	C5
Particle number	10,857	26,781	8,093	97,187	60,524
Map resolution (Å)	3.3	3.6	3.8	3.0	3.9
FSC threshold	0.143	0.143	0.143	0.143	0.143
Refinement					
Model resolution (Å)	3.5			3.0	
FSC threshold	0.5			0.5	
Model composition					
Non-hydrogen atoms	34529			145753	
Protein residues	4377			18565	
Nucleotide	0			0	
<i>B</i> factors (Å ²)					
Protein	32			110	
Ligand					
R.m.s. deviations					
Bond lengths (Å)	0.006			0.003	
Bond angles (°)	0.746			0.587	
Validation					
MolProbity score	1.63			1.41	
Clashscore	6.01			5.33	
Poor rotamers (%)	0.34			0.14	
Ramachandran plot					
Favored (%)	95.66			97.39	
Allowed (%)	4.34			2.61	
Disallowed (%)	0.00			0	

Table S4. CryoEM data collection, refinement and validation statistics of MRV isolate from cells at 48 h post-infection

	Virion	Empty virion	Outer shell
EMDB PDB		EMD-47320	EMD-46049 9CYT
Data collection and processing			
Magnification		81,000	
Voltage (kV)		300	
Electron exposure (e ⁻ /Å ²)		50	
Defocus range (μm)		-1.8 to -2.6	
Pixel size (Å)		1.1	
Symmetry imposed	C5	C5	C5
Particle number	67,928	22,373	8,911
Map resolution (Å)	3.0	3.2	3.7
FSC threshold	0.143	0.143	0.143
Refinement			
Model resolution (Å)			3.6
FSC threshold			0.5
Model composition			
Non-hydrogen atoms			33997
Protein residues			4390
Nucleotide			0
<i>B</i> factors (Å ²)			
Protein			14.7
Ligand			
R.m.s. deviations			
Bond lengths (Å)			0.004
Bond angles (°)			0.621
Validation			
MolProbity score			1.62
Clashscore			6.22
Poor rotamers (%)			0.16
Ramachandran plot			
Favored (%)			96.02
Allowed (%)			3.98
Disallowed (%)			0