# Prevalence and determinants of iron deficiency in cardiac amyloidosis

Antoine Jobbé-Duval<sup>1</sup>, Mélanie Bézard<sup>2</sup>, Stéphane Moutereau<sup>3</sup>, Mounira Kharoubi<sup>2</sup>, Silvia Oghina<sup>2</sup>, Amira Zaroui<sup>2</sup>, Arnault Galat<sup>2</sup>, Coraline Chalard<sup>2</sup>, Elisabeth Hugon-Vallet<sup>1</sup>, Francois Lemonnier<sup>4</sup>, Damien Eyharts<sup>2</sup>, Elsa Poulot<sup>5</sup>, Pascale Fanen<sup>6</sup>, Benoit Funalot<sup>6</sup>, Valérie Molinier-Frenkel<sup>7</sup>, Vincent Audard<sup>8</sup>, Luc Hittinger<sup>2</sup>, Marc Antoine Delbarre<sup>2</sup>, Emmanuel Teiger<sup>2</sup> and Thibaud Damy<sup>2\*</sup>

<sup>1</sup>Heart Failure and Transplant Department, 'Louis Pradel' Cardiologic Hospital, Hospices Civils de Lyon, Lyon, France; <sup>2</sup>Department of Cardiology, French Referral Centre for Cardiac Amyloidosis, Cardiogen Network, GRC Amyloid Research Institute, DHU A-TVB, InsermU955, Henri Mondor Teaching Hospital, APHP, 51 Avenue Marechal de Lattre de Tassigny, Creteil, 94000, France; <sup>3</sup>Department of Biochemistry, Henri Mondor Teaching Hospital, APHP, Creteil, France; <sup>4</sup>Department of Haematology, Henri Mondor Teaching Hospital, APHP, Creteil, France; <sup>5</sup>Department of Pathology, Henri Mondor Teaching Hospital, APHP, Creteil, France; <sup>6</sup>Department of Genetics, Henri Mondor Teaching Hospital, APHP, Creteil, France; <sup>7</sup>Department of Immunobiology, Henri Mondor Teaching Hospital, APHP, Creteil, France; and <sup>8</sup>Department of Nephrology, Henri Mondor Teaching Hospital, APHP, Creteil, France; <sup>1</sup>Department of Immunobiology, Henri Mondor Teaching Hospital, APHP, Creteil, France; and <sup>8</sup>Department of Nephrology, Henri

### Abstract

**Aims** Iron deficiency (ID) is common in patient with chronic heart failure (HF) and has been widely studied. In contrast, data concerning ID in cardiac amyloidosis (CA) are limited. Amyloidosis is a severe and fatal systemic disease, characterized by an accumulation of amyloid fibrils in various tissues/organs, including nerves, kidneys, gastrointestinal tract, and heart. Amyloid deposits in the heart eventually cause HF. The main subtypes of CA are light chain (AL), hereditary transthyretin (ATTRv), and wild-type transthyretin (ATTRwt). We performed this study to determine the prevalence, clinical outcome (all-cause mortal-ity), and determinants of ID among the three main subtypes of CA.

**Methods and results** Iron deficiency status were analysed in 816 CA patients enrolled at the French Referral Centre for Cardiac Amyloidosis: 271 (33%) had AL, 164 (20%) ATTRv, and 381 (47%) ATTRwt. ID affected 49% of CA patients, 45% with AL, 58% with ATTRv, and 48% with ATTRwt. We identified ATTR status (ATTRv P = 0.003, ATTRwt P = 0.037), diabetes (P = 0.003), aspirin treatment (P = 0.009), haemoglobin levels (P = 0.006), and altered global longitudinal strain (P = 0.02) as independent ID determinants. There is no difference in all-cause mortality considering ID status.

**Conclusions** Iron deficiency is common in patients with CA, irrespective of the subtype. Patients seem more likely to have ID if diagnosed with ATTR, if diabetic, and/or treated with aspirin. In CA, the benefit of intravenous iron therapy, for ID, on morbidity and mortality needs further study.

Keywords Amyloidosis; Iron deficiency; Heart failure

Received: 22 February 2021; Revised: 23 December 2021; Accepted: 17 January 2022

\*Correspondence to: Thibaud Damy, Department of Cardiology, French Referral Centre for Cardiac Amyloidosis, Cardiogen Network, GRC Amyloid Research Institute, DHU A-TVB, InsermU955, Henri Mondor Teaching Hospital, APHP, 51 Avenue Marechal de Lattre de Tassigny, 94000 Créteil, France. Tel: +33 (0)1.49.81.22.53. Email: thibaud.damy@aphp.fr

## Introduction

Worldwide, iron deficiency (ID) is a very common nutritional disorder, even in industrialized nations where it often occurs with cardiovascular disease.<sup>1</sup> ID also frequently occurs with chronic diseases, like chronic heart failure<sup>2</sup> and chronic kidney disease.<sup>3</sup> There are two types of ID: functional ID when serum ferritin levels are between 100 and 299  $\mu$ g/L and

transferrin saturation is below 20%, and absolute ID when serum ferritin levels drop below 100  $\mu$ g/L.<sup>4,5</sup> In patients with functional ID, iron is inadequately distributed for proper tissue function: often from chronic inflammation with iron retention (sequestration), by macrophages and hepatocytes, that is frequently associated with decreased iron absorption.<sup>6</sup> Absolute ID can result from chronic blood loss and inadequate iron intake, or develops from functional ID.<sup>6</sup>

© 2022 The Authors. ESC Heart Failure published by John Wiley & Sons Ltd on behalf of European Society of Cardiology.

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.

Moreover, absolute ID is associated with specific diseases. In heart failure (HF) gastrointestinal iron malabsorption, decreased duodenal iron transfer from gut oedema, and macrophage iron sequestration from inflammation (with high circulating levels of pro-inflammatory mediators) leads to ID.7 Approximately 50-62% of HF patients have ID.<sup>2,8,9</sup> ID occurs irrespective of the left ventricular ejection fraction (LVEF). Indeed, a single-centre, prospective study reported that 53% of HF patients had ID: among these 50% had HF with reduced (HFrEF), 61% had HF with mildly reduced (HFmrEF), and 64% had HF with preserved ejection fractions (HFpEF).<sup>9</sup> Overall, ID prevalence tends to increases as diastolic function deteriorates.<sup>10</sup> Moreover, ID in HF is a strong, independent predictor of mortality<sup>2</sup> and is associated with reduced exercise capacity, and diminished guality of life.<sup>8–10</sup> Recent studies have reported that iron therapy with carboxymaltose ferric improves quality of life and exercise capacity in HFrEF patients and reduces heart failure re-hospitalization in patients with acute heart failure (LVEF <50%).<sup>4,11-14</sup> The impact of iron therapy on HFpEF patients with ID is being evaluated in the FAIR HFpEF study (NCT03074591).

Recently, several studies have shown that cardiac amyloidosis (CA) is an underestimated cause of HF.<sup>15–17</sup> A study in elderly patients found that wild-type TTR amyloidosis (ATTRwt) represented 13% of HFpEF hospitalizations.<sup>16</sup> Amyloidosis is a severe, progressive systemic disease characterized by deposits of misfolded, insoluble, toxic proteins (amyloid fibrils) in the extracellular matrix of various organs, including the heart.<sup>18</sup> Numerous proteins are implicated in CA, among these immunoglobulin light chains (ALs) and transthyretin (TTR) are the most frequent. The three main types of CA are AL, wild-type TTR amyloidosis (ATTRwt), and hereditary TTR amyloidosis (ATTRv).<sup>19</sup> ATTRv results from a genetic variant of the TTR gene, which produces unstable amyloidogenic TTR.<sup>20-22</sup> In CA, the thickening and increased stiffness of the cardiac walls from amyloid deposits leads to HF, conduction disorders, atrial arrhythmias, and eventually cardiovascular death.

In CA patients besides HF because of oedema in organs, amyloid fibrils also infiltrate the extracellular matrix of the gastrointestinal tract, vessels, and the nervous system.<sup>23,24</sup> Nervous system infiltration causes gastroparesis and dysphagia that leads to malnutrition. Vascular vessels infiltration may lead to bleeding and ID. Gastrointestinal amyloid infiltration may aggravate the HF malabsorption and amplify ID associated with HF. We hypothesized that pathophysiological mechanisms observed in CA and associated with HF lead to ID. Thus, amyloid infiltration severity may correlate to ID severity. To our knowledge, ID prevalence and determinants have not yet been assessed in patients with CA.

The aim of our study was to evaluate ID prevalence and its association with clinical, biological, and imaging characteristics in CA patients with AL, ATTRv, and ATTRwt.

## Methods

#### **Study population**

This retrospective cohort study was conducted in the French Referral Centre for Cardiac Amyloidosis at the Henri Mondor Teaching Hospital (Creteil) from August 2010 to March 2020. All consecutive patients with confirmed CA (with either AL, ATTRv, or ATTRwt amyloidosis) and baseline iron status were prospectively included. Patients had a comprehensive medical evaluation at arrival with clinical and laboratory assessments, including complete blood count, bilirubin, albumin, serum N-terminal pro-B-type natriuretic peptide (NT-proBNP), high-sensitivity cardiac troponin T, and serum ID tests, electrocardiography, echocardiography, and <sup>99m</sup>Tc-hydroxy-methyl-diphosphonate (HMDP) scintigraphy.

#### Definition of cardiac amyloidosis and its type

Cardiac amyloidosis was suspected when the interventricular septum thickness (IVST) was >12 mm, measured by echocardiography, in the absence of other known cause of cardiac hypertrophy. AL amyloidosis was diagnosed by an excess of serum and/or urinary free-light chains (FLCs) and AL deposits assessed by immunohistochemistry or immunofluorescence, with a specific anti-FLC antibody, on an extra-cardiac or endomyocardial biopsy. CA severity was assessed using cardiac biomarkers: NT-proBNP and troponin T.<sup>25</sup> ATTR amyloidosis was diagnosis using the method previously described.<sup>26</sup> Genetic sequencing of patients with ATTR amyloidosis was performed to differentiate ATTRwt and ATTRv subtypes.

# Biological tests and assessment of iron deficiency status and anaemia

Iron deficiency was diagnosed using the baseline serum levels of ferritin and transferrin. ID was diagnosed when patients had serum ferritin <100 µg/L (absolute ID), or serum ferritin between 100 and 299 µg/L with transferrin saturation <20% (functional ID).<sup>27,28</sup> Anaemia was defined as haemoglobin levels <13 g/dL for men and <12 g/dL for women, as recommended by the World Health Organization (World Health Organization/United Nations University/UNICEF. Iron Deficiency Anaemia, Assessment, Prevention and Control: A Guide for Programme Managers. Geneva: WHO; 2001).

#### Clinical follow-up

Patient follow-up visits were performed according to standard of care. Patients usually consulted every 6–9 months. During these visit, standard clinical evaluations were performed.

#### Ethical considerations

**Statistical analysis** 

All patients provided oral consent to be included in the Amyloidosis Network registry. The study was approved by the local ethics committee and data were recorded electronically in accordance with the French CNIL (*Commission National de l'Informatique et des Libertés*; N°1846564 v0). percentages. Frequencies for quantitative variables were compared using the  $\chi^2$  test with Pearson's correction. Continuous data were compared using the Mann–Whitney test for two groups and the Kruskal–Wallis test for more than two groups. Follow-up data were obtained from medical files, or if required, by contacting the patients' families. The study assessed all-cause mortality in patients with or without ID.

The statistical analyses were performed using the SPSS software (version 19.0 for Windows 2010 SPSS Inc.). A *P* value below 0.05 was considered as statistically significant.

Continuous variables were expressed as median with interquartile range (IQR) and dichotomous data as numbers with Logistical regression was used to assess the association between ID and baseline characteristics.

#### Table 1 Baseline characteristics of patients according to amyloidosis status

N (%)	All 816 (100)	AL 271 (33)	ATTRv	ATTRwt	Р
N (%)	816 (100)	271 (33)	164 (20)	381 (47)	P
Clinical characteristics					
Age at inclusion, years	76 (67; 82)	67 (59; 75)	72 (66; 78)	81 (76; 85)	<0.001
Gender, women n (%)	210 (26)	103 (38)	55 (34)	52 (14)	< 0.001
BMI, kg/m <sup>2</sup>	25 (22; 27)	23 (21; 26)	24 (22; 27)	25 (23; 28)	< 0.001
CV risk factors					
Diabetes, n (%)	151 (19)	45 (17)	32 (20)	74 (19)	0.615
Hypertension, n (%)	438 (54)	107 (40)	91 (56)	240 (63)	< 0.001
Dyslipidaemia, n (%)	259 (32)	73 (27)	42 (26)	144 (38)	0.068
CV characteristics	200 (02)		.= (= 0)	(20)	0.000
NYHA Class III–IV vs. I–II, n (%)	342 (42)	130 (48)	69 (42)	143 (38)	0.003
Heart rate, beats/min	76 (68; 86)	82 (73; 94)	75 (66; 84)	74 (67; 81)	< 0.001
Systolic blood pressure, mmHg	122 (108; 137)	112 (101; 127)	121 (108; 133)	129 (115; 143)	<0.001
Atrial fibrillation, n (%)	213 (26)	30 (11)	32 (20)	151 (40)	< 0.001
Ischaemic heart disease, n (%)	150 (18)	38 (14)	12 (7)	100 (26)	<0.001
Echocardiography characteristics	54 (42, 62)				
LVEF, %	51 (42; 60)	55 (45; 62)	49 (36; 59)	50 (41; 58)	< 0.001
IVST, mm	17 (15; 19)	15 (14; 17)	18 (15; 20)	18 (15; 20)	< 0.001
GL Strain, %	-10 (-8; -13)	-10 (-8; -14)	-10 (-8; -13)	-10 (-8; -13)	0.452
Blood parameters					
NT-proBNP, ng/mL	3550 (1740; 6735)	5074 (2257; 10982)	2525 (1065; 5368)	3247 (1734; 5802)	<0.001
Troponin T HS, ng/mL	71 (45; 106)	83 (53; 136)	67 (40; 96)	66 (43; 95)	<0.001
eGFR, mL/min/1.73 m <sup>2</sup>	50 (37; 67)	55 (38; 75)	54 (38; 68)	48 (37; 60)	0.002
Haemoglobin, g/dL	13.1 (11.8; 14.1)	12.7 (10.9; 13.8)	12.9 (11.8; 13.7)	13.4 (12.2; 14.4)	< 0.001
Mean corpuscular volume, fL	92 (87; 95)	91 (87; 95)	90 (85; 94)	92 (88; 96)	< 0.001
Ferritin, µg/L	204 (109; 328)	225 (118; 360)	139 (72; 283)	210 (124; 314)	<0.001
Transferrin saturation, %	18 (13; 24)	18 (12; 24)	17 (13; 25)	18 (13; 24)	0.560
Iron deficiency treatment <sup>a</sup>					
Oral iron during follow-up, n (%)	30 (8)	4 (3)	7 (8)	19 (10)	0.069
I.V. iron during follow-up, n (%)	191 (48)	57 (47)	44 (47)	90 (50)	0.883
Anticoagulant and antiplatelet treatr		57 (47)	++ (+/)	50 (50)	0.001
Aspirin, n (%)	213 (26)	75 (28)	34 (21)	104 (27)	0.220
Clopidogrel, n (%)	46 (6)	12 (4)	6 (4)	28 (7)	0.220
	. ,				< 0.132
Oral anticoagulants, n (%)	434 (53)	111 (41)	74 (45)	249 (65)	< 0.00
Heart failure treatment	246 (26)	42 (46)	54 (22)	120 (21)	
ACE inhibitor, n (%)	216 (26)	42 (16)	54 (33)	120 (31)	< 0.001
ARB, n (%)	121 (15)	25 (9)	29 (18)	67 (18)	0.007
Digoxin, n (%)	14 (2)	2 (1)	1 (1)	11 (3)	0.055
Selective beta-blocker, n (%)	221 (27)	63 (23)	48 (29)	110 (29)	0.223
Non-selective beta-blocker, n (%)	16 (2)	5 (2)	1 (1)	10 (3)	0.297
Calcium antagonist, <i>n</i> (%)	222 (27)	62 (23)	45 (27)	115 (30)	0.123
Amiodarone, <i>n</i> (%)	159 (19)	38 (14)	31 (19)	90 (24)	0.010
Loop diuretic, <i>n</i> (%)	595 (73)	197 (73)	107 (65)	291 (76)	0.030
Thiazide diuretic, n (%)	278 (34)	69 (25)	61 (37)	148 (39)	0.001
Vasodilator, n (%)	25 (3)	8 (3)	5 (3)	12 (3)	0.991

ACE, angiotensin-converting enzyme; ARB, angiotensin II receptor blocker; BMI, body mass index; eGFR, estimated glomerular filtration rate; GL, global longitudinal; I.V., intravenous; IVST, interventricular septum thickness; LVEF, left ventricular ejection fraction; NT-proBNP, N-terminal pro-B-type natriuretic peptide; NYHA, New York Heart Association.

Values are median (interquartile range).

 $^{\circ}N = 400.$ 

1317

The Kruskal–Wallis test was used to analyse the association between malabsorption and ferritin levels in the CA population. Ferritin levels were compared with albumin tertiles. Similarly, in AL patients, where hepatic dysfunction is frequent and can affect ferritin production, we compared ferritin levels to bilirubin tertiles (T1–T2 vs. T3).

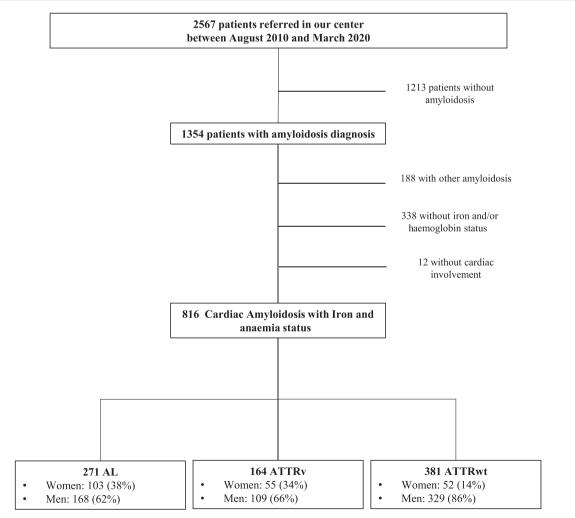
# Results

### **Patient characteristics**

From August 2010 to March 2020, of the 2567 patients that presented at the French Referral Centre with suspected CA, 816 patients had CA with iron status and were enrolled in the study: 271 (33%) with AL, 164 (20%) with ATTRv, and

381 (47%) with ATTRwt (Table 1 and Figure 1). The median age in the overall population was 76 years (IQR: 67-82): 67 years (IQR: 59-75) for patients with AL, 72 years (IQR: 66-78) with ATTRv, and 81 years (IQR: 76-85) with ATTRwt. Women represented 38% of AL, 34% of ATTRv, and 14% of ATTRwt patients. The baseline characteristics of the overall population and according to subtypes of CA are shown in Table 1. Overall, patient age, cardiovascular risk factors, clinical laboratory, and echocardiography variables differed significantly between CA subtypes (Table 1). Median age, hypertension, and ischaemic heart disease were significantly higher in ATTRwt patients (P < 0.001). While New York Heart Association (NYHA) status (P = 0.003), NT-proBNP and highsensitivity troponin T levels (P < 0.001) were significantly higher in AL patients. ATTRwt patients had more atrial fibrillations (P < 0.001) and received significantly more oral anticoagulant (P < 0.001) than AL and ATTRv patients.

Figure 1 Flowchart.



# Prevalence of iron deficiency among cardiac amyloidosis patients

Iron deficiency was diagnosed in 400 CA patients (49%): 123 (45%) with AL, 95 (58%) with ATTRv, and 182 (48%) with ATTRwt (*Table 2, Figures 2* and *3A*). The baseline characteris-

tics of the overall population and according to the presence or not of ID are shown in *Table 2*. The median ferritin levels in patients with ID were 113  $\mu$ g/L compared with 326  $\mu$ g/L in those without ID. Similarly, the median transferrin saturation levels were 14% in ID patients and 23% in those without ID.

 Table 2
 Baseline characteristics of patient according to iron status

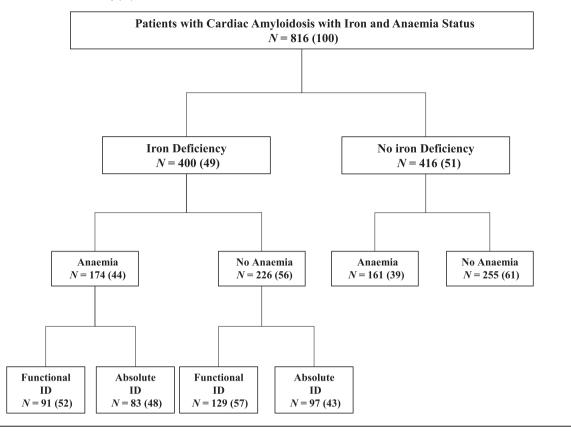
Cite (a)         Cite (a)           Clinical characteristics         Age at inclusion, years         76 (68; 83)         76 (67; 82)           Gender, women $n$ (%)         125 (31)         85 (20)           BMI, kg/m <sup>4</sup> 25 (22; 27)         24 (22; 27)           CV risk factors         219 (55)         219 (53)           Diabetes, $n$ (%)         219 (55)         219 (53)           Dyslipidaemia, $n$ (%)         137 (34)         122 (29)           CV characteristics         NYHA Class III-IV vs I-II, $n$ (%)         179 (48)         163 (43)           Heart rate, beats/min         75 (68; 85)         77 (68; 87)           Systolic blood pressure, mmHg         122 (108; 137)         121 (107; 139)           Atrial fibrillation, $n$ (%)         166 (29)         107 (28)           Ischaemic heart disease, $n$ (%)         84 (21)         66 (16)           Amyloidosis status         Itage (48)         199 (52)           Echocardiography characteristics         LVEF, %         50 (40; 59)         52 (43; 60)           LVEF <40%, $n$ (%)         182 (48)         199 (52)         Echocardiography characteristics           LVEF <40%, $n$ (%)         183 (48)         210 (54)         112 (29)           LVEF <40%, $n$ (%)         108 (23)		All	CA	
$\begin{array}{c c c c c c c c c c c c c c c c c c c $				P value
Age at inclusion, years76 (68; 83)76 (67; 82)Gender, women n (%)125 (31)85 (20)BMI, kg/m²25 (22; 27)24 (22; 27)CV risk factors7 (14)Hypertension, n (%)219 (55)219 (53)Dyslipidaemia, n (%)137 (34)122 (29)CV characteristics77 (68; 87)NYHA Class III-IV vs.I-II, n (%)179 (48)163 (43)Heart rate, beats/min75 (68; 85)77 (68; 87)Systolic blood pressure, mmHg122 (108; 137)121 (107; 139)Atrial fibrillation, n (%)106 (29)107 (28)Ischaemic heart disease, n (%)84 (21)66 (16)Amyloidosis status7475 (66; 63)Atrin (%)123 (45)148 (55)ATTRv, n (%)95 (56)69 (44)ATTRv, n (%)182 (48)199 (52)Echocardiography characteristics1212 (29)LVEF <40%, n (%)				
Gender, women $n$ (%)125 (31)85 (20)BML, kg/m²25 (22; 27)24 (22; 27)CV risk factors $35$ (22; 27)24 (22; 27)Diabetes, $n$ (%)94 (24)57 (14)Hypertension, $n$ (%)137 (34)122 (29)CV characteristics $137$ (34)122 (29)CV characteristics $179$ (48)163 (43)Heart rate, beats/min75 (68; 85)77 (68; 87)Systolic blood pressure, mmHg122 (108; 137)121 (107; 139)Atrial fibrillation, $n$ (%)106 (29)107 (28)Ischaemic heart disease, $n$ (%)84 (21)66 (16)Amyloidosis status $44$ (21)66 (16)Amyloidosis status $123$ (45)148 (55)ATTRv, $n$ (%)95 (56)69 (44)ATTRv, $n$ (%)182 (48)199 (52)Echocardiography characteristics $UVEF$ (40%, $n$ (%)188 (48)LVEF (43s: $112$ (29) $112$ (29)LVEF 40-S0%, $n$ (%)183 (48)210 (54)UVEF 40-S0%, $n$ (%)183 (48)210 (54)IVST, mm17 (15; 20)17 (15; 19)GL strain, % $-10$ ( $-7$ , $-12$ ) $-11$ ( $-8$ ; $-13$ )Biology variables $393$ (1810; 6659)3555 (1688; 6736)Troponin T HS, $ng/mL$ 240 (38; 67)51 (37; 67)Haemoglobin, $g/dL$ 122 (31)91 (22)Transfermi saturation, %°113 (63; 194)326 (21; 520)Transfermi saturation, %°122 (31)91 (22)Transfermi saturation, %°122 (31)91 (22)<		76 (68: 83)	76 (67: 82)	0.212
BMI, kg/m²       25 (22; 27)       24 (22; 27)         CV risk factors       57 (14)         Diabetes, n (%)       219 (55)       219 (53)         Dyslipidaemia, n (%)       137 (34)       122 (29)         CV characteristics       71 (68; 87)       77 (68; 87)         NYHA Class III-IV vs I-II, n (%)       179 (48)       163 (43)         Heart rate, beats/min       75 (68; 85)       77 (68; 87)         Systolic blood pressure, mmHg       122 (108; 137)       121 (107; 139)         Atrial fibrillation, n (%)       123 (45)       148 (55)         Anyloidosis status       74 (48)       199 (52)         Echocardiography characteristics       122 (48)       199 (52)         Echocardiography characteristics       122 (48)       199 (52)         LVEF <40%, n (%)	5			< 0.001
CV risk factors       94 (24)       57 (14)         Diabetes, n (%)       219 (55)       219 (53)         Dyslipidaemia, n (%)       137 (34)       122 (29)         CV characteristics       NYHA Class III-IV vs I-II, n (%)       179 (48)       163 (43)         Heart rate, beats/min       75 (68; 85)       77 (68; 87)       55         Systolic blood pressure, mmHg       122 (108; 137)       121 (107; 139)       141 (107; 139)         Atrial fibrillation, n (%)       106 (29)       107 (28)       15         Ischaemic heart disease, n (%)       84 (21)       66 (16)         Amyloidosis status       122 (45)       148 (55)       148 (55)         ATTRv, n (%)       95 (56)       69 (44)       41         ATTRv, n (%)       182 (48)       199 (52)       Echocardiography characteristics         LVEF, %       50 (40; 59)       52 (43; 60)       112 (29)       112 (29)         LVEF 40%, n (%)       183 (48)       210 (54)       140 (51)         LVEF 40%, n (%)       183 (48)       210 (54)       137 (57)         UVEF 40%, n (%)       183 (48)       210 (54)       153 (15)         UVEF 40%, n (%)       183 (48)       210 (54)       165         UVEF 40%, n (%)       183 (48) <td></td> <td></td> <td></td> <td>0.285</td>				0.285
$\begin{array}{llllllllllllllllllllllllllllllllllll$		23 (22, 27)	2 (22, 27)	0.205
Hypertension, $n$ (%)219 (55)219 (53)Dyslipidaemia, $n$ (%)137 (34)122 (29)CV characteristics		94 (24)	57 (14)	< 0.001
$\begin{array}{c c c c c c c c c c c c c c c c c c c $				0.297
CV characteristics       179 (48)       163 (43)         NYHA Class III-IV vs I-II, n (%)       179 (48)       163 (43)         Heart rate, beats/min       75 (68, 85)       77 (68, 87)         Systolic blood pressure, mmHg       122 (108; 137)       121 (107; 139)         Attral fibrillation, n (%)       106 (29)       107 (28)         Ischaemic heart disease, n (%)       84 (21)       66 (16)         Amyloidosis status				0.336
NYHA Class III-IV vs I-II, n (%)         179 (48)         163 (43)           Heart rate, beats/min         75 (68, 85)         77 (68, 87)           Systolic blood pressure, mmHg         122 (108; 137)         121 (107; 139)           Atrial fibrillation, n (%)         106 (29)         107 (28)           Ischaemic heart disease, n (%)         84 (21)         66 (16)           Amyloidosis status         123 (45)         148 (55)           ATTRv, n (%)         95 (56)         69 (44)           ATTRv, n (%)         182 (48)         199 (52)           Echocardiography characteristics         124 (48)         199 (52)           LVEF, %         50 (40; 59)         52 (43; 60)           LVEF          60, n (%)         183 (48)         210 (54)           LVEF          70, n (%)         108 (29)         112 (29)           LVEF          170, n (%)         108 (29)         117 (5; 19)           GL strain, %         -10 (-7; -12)         -11 (-8; -13)           Biology variables         17 (15; 20)         17 (15; 19)           GL strain, %         -10 (27; -12)         -11 (-8; -13)           Miony ng/mL         3493 (1810; 6659)         3655 (1688; 6736)           Troponin T HS, ng/mL         129 (115; 13.9)         13.				
$\begin{array}{c c c c c c c c c c c c c c c c c c c $		179 (48)	163 (43)	0.204
Systolic blood pressure, mmHg       122 (108; 137)       121 (107; 139)         Atrial fibrillation, n (%)       106 (29)       107 (28)         Ischaemic heart disease, n (%)       84 (21)       66 (16)         Amyloidosis status				0.171
Atrial fibrillation, $n$ (%)106 (29)107 (28)Ischaemic heart disease, $n$ (%)84 (21)66 (16)Amyloidosis status69 (44)AL, $n$ (%)123 (45)148 (55)ATTRw, $n$ (%)95 (56)69 (44)ATTRw, $n$ (%)182 (48)199 (52)Echocardiography characteristics106 (29)12 (29)LVEF, %50 (40; 59)52 (43; 60)LVEF, %108 (29)112 (29)LVEF <20%, $n$ (%)183 (48)210 (54)IVST, mm17 (15; 20)17 (15; 19)GL strain, %-10 (-7; -12)-11 (-8; -13)Biology variablesNT-proBNP, ng/mL3493 (1810; 6659)3655 (1688; 6736)Troponin T HS, ng/mL22 (45; 106)68 (45; 105)eGFR, mL/min/1.73 m²50 (38; 67)51 (37; 67)Haemoglobin, g/dL12.9 (11.5; 13.9)13.3 (12.0; 14.3)Mean corpuscular volume, $\mu m^3$ 90 (85; 94)93 (89; 97)Anaemia, $n$ (%)174 (44)161 (39)Ferritin, $\mu g/L^a$ 13 (63; 194)326 (211; 520)Transferrin saturation, %a14 (10; 17)23 (20; 28)Anticoagulant, and or antiplatelet treatments22 (31)91 (22)Clopidogrel, $n$ (%)26 (7)20 (55)Treatment of CHF $122$ (31)91 (22)AcE inhibitor, $n$ (%)20 (53)225 (55)Treatment of CHF34 (2)6 (2)ARB, $n$ (%)88 (2)62 (2)				0.803
Amyloidosis statusAL, $n$ (%)123 (45)148 (55)ATTRv, $n$ (%)95 (56)69 (44)ATTRv, $n$ (%)182 (48)199 (52)Echocardiography characteristics124 (48)199 (52)LVEF, %50 (40; 59)52 (43; 60)LVEF class:124 (29)112 (29)LVEF <40-50%, $n$ (%)108 (29)112 (29)LVEF >50%, $n$ (%)108 (29)17 (15; 19)GL strain, %-10 (-7; -12)-11 (-8; -13)Biology variables125 (45; 106)68 (45; 105)NT-proBNP, $ng/mL$ 3493 (1810; 6659)3655 (1688; 6736)Troponin T HS, $ng/mL$ 72 (45; 106)68 (45; 105)eGFR, $mL/min/1.73$ m²50 (38; 67)51 (37; 67)Haemoglobin, $g/dL$ 12.9 (11.5; 13.9)13.3 (12.0; 14.3)Mean corpuscular volume, $\mu m^3$ 90 (85; 94)93 (89; 97)Anaemia, $n$ (%)113 (63; 194)326 (211; 520)Transferrin saturation, $\%^a$ 12.2 (31)91 (22)Clopidogrel, $n$ (%)26 (7)20 (5)Oral anticoagulants, $n$ (%)26 (7)20 (5)Treatment of CHFTTACE inhibitor, $n$ (%)68 (17)53 (13)Digoxin, $n$ (%)68 (17)53 (13)Digoxin, $n$ (%)8 (2)6 (2)				0.812
Amyloidosis statusAL, $n$ (%)123 (45)148 (55)ATTRv, $n$ (%)95 (56)69 (44)ATTRv, $n$ (%)182 (48)199 (52)Echocardiography characteristics124 (48)199 (52)LVEF, %50 (40; 59)52 (43; 60)LVEF class:124 (29)112 (29)LVEF <40-50%, $n$ (%)108 (29)112 (29)LVEF >50%, $n$ (%)108 (29)17 (15; 19)GL strain, %-10 (-7; -12)-11 (-8; -13)Biology variables125 (45; 106)68 (45; 105)NT-proBNP, $ng/mL$ 3493 (1810; 6659)3655 (1688; 6736)Troponin T HS, $ng/mL$ 72 (45; 106)68 (45; 105)eGFR, $mL/min/1.73$ m²50 (38; 67)51 (37; 67)Haemoglobin, $g/dL$ 12.9 (11.5; 13.9)13.3 (12.0; 14.3)Mean corpuscular volume, $\mu m^3$ 90 (85; 94)93 (89; 97)Anaemia, $n$ (%)113 (63; 194)326 (211; 520)Transferrin saturation, $\%^a$ 12.2 (31)91 (22)Clopidogrel, $n$ (%)26 (7)20 (5)Oral anticoagulants, $n$ (%)26 (7)20 (5)Treatment of CHFTTACE inhibitor, $n$ (%)68 (17)53 (13)Digoxin, $n$ (%)68 (17)53 (13)Digoxin, $n$ (%)8 (2)6 (2)	Ischaemic heart disease, n (%)	84 (21)	66 (16)	0.085
ATRv, n (%)95 (56)69 (44)ATRv, n (%)182 (48)199 (52)Echocardiography characteristicsEchocardiography characteristicsLVEF, %50 (40; 59)52 (43; 60)LVEF class:67 (17)LVEF 40-50%, n (%)108 (29)112 (29)LVEF ≥50%, n (%)183 (48)210 (54)IVST, mm17 (15; 20)17 (15; 19)GL strain, %-10 (-7; -12)-11 (-8; -13)Biology variables710 (55)68 (45; 105)NT-proBNP, ng/mL24 43; 106)68 (45; 105)eGFR, mL/min/1.73 m²50 (38; 67)51 (37; 67)Haemoglobin, g/dL12.9 (11.5; 13.9)13.3 (12.0; 14.3)Mean corpuscular volume, μm³90 (85; 94)93 (89; 97)Anatenia, n (%)174 (44)161 (39)Ferritin, µg/L³133 (63; 194)326 (211; 520)Transferin saturation, %³142 (10; 17)23 (20; 28)Anticoagulant and or antiplatelet treatments209 (53)225 (55)Aspirin, n (%)209 (53)225 (55)Treatment of CHF101 (26)115 (28)ARB, n (%)68 (17)53 (13)Digoxin, n (%)8 (2)6 (2)				0.032
ATRv, n (%)95 (56)69 (44)ATRv, n (%)182 (48)199 (52)Echocardiography characteristicsEchocardiography characteristicsLVEF, %50 (40; 59)52 (43; 60)LVEF class:67 (17)LVEF 40-50%, n (%)108 (29)112 (29)LVEF ≥50%, n (%)183 (48)210 (54)IVST, mm17 (15; 20)17 (15; 19)GL strain, %-10 (-7; -12)-11 (-8; -13)Biology variables710 (55)68 (45; 105)NT-proBNP, ng/mL24 43; 106)68 (45; 105)eGFR, mL/min/1.73 m²50 (38; 67)51 (37; 67)Haemoglobin, g/dL12.9 (11.5; 13.9)13.3 (12.0; 14.3)Mean corpuscular volume, μm³90 (85; 94)93 (89; 97)Anatenia, n (%)174 (44)161 (39)Ferritin, µg/L³133 (63; 194)326 (211; 520)Transferin saturation, %³142 (10; 17)23 (20; 28)Anticoagulant and or antiplatelet treatments209 (53)225 (55)Aspirin, n (%)209 (53)225 (55)Treatment of CHF101 (26)115 (28)ARB, n (%)68 (17)53 (13)Digoxin, n (%)8 (2)6 (2)	ÁL, n (%)	123 (45)	148 (55)	
Echocardiography characteristics       50 (40; 59)       52 (43; 60)         LVEF, %       50 (40; 59)       52 (43; 60)         LVEF <40%, n (%)				
LVEF, %       50 (40; 59)       52 (43; 60)         LVEF class:	ATTRwt, n (%)	182 (48)	199 (52)	
LVEF class:       LVEF 40%, n (%)       86 (23)       67 (17)         LVEF 40~50%, n (%)       108 (29)       112 (29)         LVEF $\geq$ 50%, n (%)       183 (48)       210 (54)         IVST, mm       17 (15; 20)       17 (15; 19)         GL strain, % $-10 (-7; -12)$ $-11 (-8; -13)$ Biology variables       72 (45; 106)       68 (45; 105)         NT-proBNP, ng/mL       72 (45; 106)       68 (45; 105)         eGFR, mL/min/1.73 m <sup>2</sup> 50 (38; 67)       51 (37; 67)         Haemoglobin, g/dL       12.9 (11.5; 13.9)       13.3 (12.0; 14.3)         Mean corpuscular volume, $\mu$ m <sup>3</sup> 90 (85; 94)       93 (89; 97)         Anaemia, n (%)       174 (44)       161 (39)         Ferritin, $\mu g/L^3$ 113 (63; 194)       326 (211; 520)         Transferrin saturation, % <sup>3</sup> 14 (10; 17)       23 (20; 28)         Anticoagulant and or antiplatelet treatments       209 (53)       225 (55)         Aprim, n (%)       209 (53)       225 (55)         Treatment of CHF       420       15 (28)         ARB, n (%)       68 (17)       53 (13)         Digoxin, n (%)       8 (2)       6 (2)	Echocardiography characteristics			
LVEF <40%, n (%)86 (23)67 (17)LVEF 40-50%, n (%)108 (29)112 (29)LVEF $\geq$ 50%, n (%)183 (48)210 (54)IVST, mm17 (15; 20)17 (15; 19)GL strain, %-10 (-7; -12)-11 (-8; -13)Biology variables		50 (40; 59)	52 (43; 60)	0.069
LVEF <40%, n (%)86 (23)67 (17)LVEF 40-50%, n (%)108 (29)112 (29)LVEF $\geq$ 50%, n (%)183 (48)210 (54)IVST, mm17 (15; 20)17 (15; 19)GL strain, %-10 (-7; -12)-11 (-8; -13)Biology variables	LVEF class:			0.129
LVEF ≥50%, n (%)       183 (48)       210 (54)         IVST, mm       17 (15; 20)       17 (15; 19)         GL strain, %       -10 (-7; -12)       -11 (-8; -13)         Biology variables       3493 (1810; 6659)       3655 (1688; 6736)         NT-proBNP, ng/mL       72 (45; 106)       68 (45; 105)         eGFR, mL/min/1.73 m²       50 (38; 67)       51 (37; 67)         Haemoglobin, g/dL       12.9 (11.5; 13.9)       13.3 (12.0; 14.3)         Mean corpuscular volume, μm³       90 (85; 94)       93 (89; 97)         Anaemia, n (%)       174 (44)       161 (39)         Ferritin, μg/L³       113 (63; 194)       326 (211; 520)         Transferrin saturation, %³       14 (10; 17)       23 (20; 28)         Anticoagulant and or antiplatelet treatments       Aspirin, n (%)       220 (5)         Oral anticoagulants, n (%)       209 (53)       225 (55)         Treatment of CHF       ACE inhibitor, n (%)       101 (26)       115 (28)         ARB, n (%)       8 (2)       6 (2)		86 (23)	67 (17)	
IVST, mm17 (15; 20)17 (15; 19)GL strain, % $-10 (-7; -12)$ $-11 (-8; -13)$ Biology variables $-10 (-7; -12)$ $-11 (-8; -13)$ NT-proBNP, ng/mL3493 (1810; 6659)3655 (1688; 6736)Troponin T HS, ng/mL72 (45; 106)68 (45; 105)eGFR, mL/min/1.73 m²50 (38; 67)51 (37; 67)Haemoglobin, g/dL12.9 (11.5; 13.9)13.3 (12.0; 14.3)Mean corpuscular volume, $\mu$ m³90 (85; 94)93 (89; 97)Anaemia, $n (%)$ 174 (44)161 (39)Ferritin, $\mu$ g/L³113 (63; 194)326 (211; 520)Transferrin saturation, %³14 (10; 17)23 (20; 28)Anticoagulant and or antiplatelet treatments $Aspirin, n (%)$ 26 (7)Aspirin, $n (%)$ 26 (7)20 (5)Oral anticoagulants, $n (\%)$ 209 (53)225 (55)Treatment of CHF $ARB, n (\%)$ 101 (26)115 (28)ARB, $n (\%)$ 8 (2)6 (2)	LVEF 40–50%, n (%)	108 (29)	112 (29)	
GL strain, % $-10(-7; -12)$ $-11(-8; -13)$ Biology variablesNT-proBNP, ng/mL3493 (1810; 6659)3655 (1688; 6736)Troponin T HS, ng/mL72 (45; 106)68 (45; 105)eGFR, mL/min/1.73 m²50 (38; 67)51 (37; 67)Haemoglobin, g/dL12.9 (11.5; 13.9)13.3 (12.0; 14.3)Mean corpuscular volume, $\mu$ m³90 (85; 94)93 (89; 97)Anaemia, n (%)174 (44)161 (39)Ferritin, $\mu g/L^a$ 113 (63; 194)326 (211; 520)Transferrin saturation, %a14 (10; 17)23 (20; 28)Anticoagulant and or antiplatelet treatments26 (7)20 (5)Aspirin, n (%)26 (7)20 (5)Oral anticoagulants, n (%)209 (53)225 (55)Treatment of CHF401 (26)115 (28)ARB, n (%)68 (17)53 (13)Digoxin, n (%)8 (2)6 (2)	LVEF ≥50%, <i>n</i> (%)	183 (48)	210 (54)	
Biology variables NT-proBNP, ng/mL3493 (1810; 6659)3655 (1688; 6736)Troponin T HS, ng/mL72 (45; 106)68 (45; 105)eGFR, mL/min/1.73 m²50 (38; 67)51 (37; 67)Haemoglobin, g/dL12.9 (11.5; 13.9)13.3 (12.0; 14.3)Mean corpuscular volume, $\mu$ m³90 (85; 94)93 (89; 97)Anaemia, n (%)174 (44)161 (39)Ferritin, $\mu g/L^a$ 113 (63; 194)326 (211; 520)Transferrin saturation, %a14 (10; 17)23 (20; 28)Anticoagulant and or antiplatelet treatments Aspirin, n (%)26 (7)20 (5)Oral anticoagulants, n (%)26 (7)20 (5)Treatment of CHF ACE inhibitor, n (%)101 (26)115 (28)ARB, n (%)68 (17)53 (13)Digoxin, n (%)8 (2)6 (2)	IVST, mm	17 (15; 20)	17 (15; 19)	0.828
NT-proBNP, ng/mL $3493 (1810; 6659)$ $3655 (1688; 6736)$ Troponin T HS, ng/mL72 (45; 106)68 (45; 105)eGFR, ml/min/1.73 m²50 (38; 67)51 (37; 67)Haemoglobin, g/dL12.9 (11.5; 13.9)13.3 (12.0; 14.3)Mean corpuscular volume, $\mu$ m³90 (85; 94)93 (89; 97)Anaemia, n (%)174 (44)161 (39)Ferritin, $\mu$ g/L°113 (63; 194)326 (211; 520)Transferrin saturation, %°14 (10; 17)23 (20; 28)Anticoagulant and or antiplatelet treatments26 (7)20 (5)Aspirin, n (%)26 (7)20 (5)Oral anticoagulants, n (%)209 (53)225 (55)Treatment of CHF401 (26)115 (28)ARB, n (%)68 (17)53 (13)Digoxin, n (%)8 (2)6 (2)	GL strain, %	-10 (-7; -12)	-11 (-8; -13)	0.022
Troponin T HS, ng/mL72 (45; 106)68 (45; 105)eGFR, mL/min/1.73 m²50 (38; 67)51 (37; 67)Haemoglobin, g/dL12.9 (11.5; 13.9)13.3 (12.0; 14.3)Mean corpuscular volume, $\mu$ m³90 (85; 94)93 (89; 97)Anaemia, n (%)174 (44)161 (39)Ferritin, $\mu g/L^a$ 113 (63; 194)326 (211; 520)Transferrin saturation, %a14 (10; 17)23 (20; 28)Anticoagulant and or antiplatelet treatments91 (22)Clopidogrel, n (%)26 (7)20 (5)Oral anticoagulants, n (%)209 (53)225 (55)Treatment of CHF72 (a)101 (26)115 (28)ARB, n (%)68 (17)53 (13)Digoxin, n (%)8 (2)6 (2)	Biology variables			
eGFR, mL/min/1.73 m²50 (38; 67)51 (37; 67)Haemoglobin, g/dL12.9 (11.5; 13.9)13.3 (12.0; 14.3)Mean corpuscular volume, $\mu$ m³90 (85; 94)93 (89; 97)Anaemia, n (%)174 (44)161 (39)Ferritin, $\mu g/L^a$ 113 (63; 194)326 (211; 520)Transferrin saturation, %a14 (10; 17)23 (20; 28)Anticoagulant and or antiplatelet treatments26 (7)20 (5)Oral anticoagulants, n (%)26 (7)20 (5)Oral anticoagulants, n (%)209 (53)225 (55)Treatment of CHF4CE inhibitor, n (%)101 (26)115 (28)ARB, n (%)68 (17)53 (13)Digoxin, n (%)8 (2)6 (2)	NT-proBNP, ng/mL		3655 (1688; 6736)	0.988
$\begin{array}{cccccc} & 12.9 & (11.5; 13.9) & 13.3 & (12.0; 14.3) \\ & \text{Mean corpuscular volume, } \mu\text{m}^3 & 90 & (85; 94) & 93 & (89; 97) \\ & \text{Anaemia, } n & (\%) & 174 & (44) & 161 & (39) \\ & \text{Ferritin, } \mu\text{g/L}^a & 113 & (63; 194) & 326 & (211; 520) \\ & \text{Transferrin saturation, } \%^a & 14 & (10; 17) & 23 & (20; 28) \\ & \text{Anticoagulant and or antiplatelet treatments} & & & \\ & \text{Aspirin, } n & (\%) & 122 & (31) & 91 & (22) \\ & \text{Clopidogrel, } n & (\%) & 26 & (7) & 20 & (5) \\ & \text{Oral anticoagulants, } n & (\%) & 209 & (53) & 225 & (55) \\ & \text{Treatment of CHF} & & & \\ & \text{ACE inhibitor, } n & (\%) & 101 & (26) & 115 & (28) \\ & \text{ARB, } n & (\%) & 8 & (2) & 6 & (2) \\ \end{array}$				0.519
Mean corpuscular volume, $\mu m^3$ 90 (85; 94)93 (89; 97)Anaemia, n (%)174 (44)161 (39)Ferritin, $\mu g/L^a$ 113 (63; 194)326 (211; 520)Transferrin saturation, $\%^a$ 14 (10; 17)23 (20; 28)Anticoagulant and or antiplatelet treatments23 (20; 28)Aspirin, n (%)122 (31)91 (22)Clopidogrel, n (%)26 (7)20 (5)Oral anticoagulants, n (%)209 (53)225 (55)Treatment of CHF4CE inhibitor, n (%)101 (26)115 (28)ARB, n (%)68 (17)53 (13)Digoxin, n (%)8 (2)6 (2)		50 (38; 67)	51 (37; 67)	0.996
Anaemia, $n$ (%)174 (44)161 (39)Ferritin, $\mu g/L^a$ 113 (63; 194)326 (211; 520)Transferrin saturation, $\%^a$ 14 (10; 17)23 (20; 28)Anticoagulant and or antiplatelet treatments23 (20; 28)Aspirin, $n$ (%)122 (31)91 (22)Clopidogrel, $n$ (%)26 (7)20 (5)Oral anticoagulants, $n$ (%)209 (53)225 (55)Treatment of CHF4CE inhibitor, $n$ (%)101 (26)115 (28)ARB, $n$ (%)68 (17)53 (13)Digoxin, $n$ (%)8 (2)6 (2)		12.9 (11.5; 13.9)	13.3 (12.0; 14.3)	0.002
Ferritin, $\mu g/L^a$ 113 (63; 194)326 (211; 520)Transferrin saturation, $\%^a$ 14 (10; 17)23 (20; 28)Anticoagulant and or antiplatelet treatments23 (20; 28)Aspirin, $n$ (%)122 (31)91 (22)Clopidogrel, $n$ (%)26 (7)20 (5)Oral anticoagulants, $n$ (%)209 (53)225 (55)Treatment of CHF400 (53)225 (55)ARB, $n$ (%)68 (17)53 (13)Digoxin, $n$ (%)8 (2)6 (2)	Mean corpuscular volume, µm²			< 0.001
Transferrin saturation, $\%^a$ 14 (10; 17)23 (20; 28)Anticoagulant and or antiplatelet treatmentsAspirin, $n$ (%)122 (31)Clopidogrel, $n$ (%)26 (7)Oral anticoagulants, $n$ (%)209 (53)225 (55)Treatment of CHFACE inhibitor, $n$ (%)101 (26)ARB, $n$ (%)68 (17)Digoxin, $n$ (%)8 (2)6 (2)				0.164
Anticoagulant and or antiplatelet treatments         Aspirin, $n$ (%)       122 (31)       91 (22)         Clopidogrel, $n$ (%)       26 (7)       20 (5)         Oral anticoagulants, $n$ (%)       209 (53)       225 (55)         Treatment of CHF       7       7         ACE inhibitor, $n$ (%)       101 (26)       115 (28)         ARB, $n$ (%)       68 (17)       53 (13)         Digoxin, $n$ (%)       8 (2)       6 (2)				<0.001
Aspirin, $n$ (%)122 (31)91 (22)Clopidogrel, $n$ (%)26 (7)20 (5)Oral anticoagulants, $n$ (%)209 (53)225 (55)Treatment of CHF101 (26)115 (28)ARB, $n$ (%)68 (17)53 (13)Digoxin, $n$ (%)8 (2)6 (2)			23 (20; 28)	<0.001
Clopidogrel, $n$ (%)26 (7)20 (5)Oral anticoagulants, $n$ (%)209 (53)225 (55)Treatment of CHF101 (26)115 (28)ARB, $n$ (%)68 (17)53 (13)Digoxin, $n$ (%)8 (2)6 (2)				
Oral anticoagulants, n (%)         209 (53)         225 (55)           Treatment of CHF         101 (26)         115 (28)           ARB, n (%)         68 (17)         53 (13)           Digoxin, n (%)         8 (2)         6 (2)				0.004
ACE inhibitor, n (%)         101 (26)         115 (28)           ARB, n (%)         68 (17)         53 (13)           Digoxin, n (%)         8 (2)         6 (2)				0.286
ACE inhibitor, n (%)       101 (26)       115 (28)         ARB, n (%)       68 (17)       53 (13)         Digoxin, n (%)       8 (2)       6 (2)	5 , , , ,	209 (53)	225 (55)	0.655
ARB, n (%)     68 (17)     53 (13)       Digoxin, n (%)     8 (2)     6 (2)				
Digoxin, n (%) 8 (2) 6 (2)	, , ,			0.465
				0.081
Selective peta-plocker, $n$ (%) 11/(30) 104 (25)				0.649
				0.157
Non-selective beta-blocker, $n$ (%) 7 (2) 9 (2)	, , ,			0.678
Calcium antagonist, $n$ (%) 114 (29) 108 (26)				0.388
Amiodarone, n (%) 75 (19) 84 (20)				0.629
Loop diuretic, n (%) 304 (77) 291 (71)				0.036
Thiazide diuretic, n (%)         140 (35)         138 (34)           Vasodilator, n (%)         11 (3)         14 (3)	, , ,			0.544 0.620

ACE, angiotensin-converting enzyme; ARB, angiotensin II receptor blocker; BMI, body mass index; eGFR, estimated glomerular filtration rate; GL, global longitudinal; ID, iron deficiency; I.V., intravenous; IVST, interventricular septum thickness; LVEF, left ventricular ejection fraction; NT-proBNP, N-terminal pro-B-type natriuretic peptide; NYHA, New York Heart Association.

Values are median (interquartile range).

P: ID vs no ID in each category: All, AL, ATTRv, ATTRwt.

<sup>a</sup>By definition.



Interestingly, median ferritin levels were significantly lower in ATTRv patients (139  $\mu$ g/L) compared with those with AL (225  $\mu$ g/L) and ATTRwt (210  $\mu$ g/L) (*Table 1*).

Iron deficiency was functional in 67 patients with AL (55%), 40 with ATTRv (42%), and 113 with ATTRwt (62%) (*Figure 3* (*B*)). It is noteworthy that ID was predominantly absolute in ATTRv patients (58%) (*Figure 3B*). The prevalence of anaemia was similar in patients with and without ID. Indeed, 174 patients (44%) with ID had anaemia vs. 161 (39%) without ID (P = 0.164) (*Table 2*).

# Relationship between iron deficiency and clinical and biological imaging characteristics

In the overall population, there were no differences between patients with and without ID in terms of NYHA III–IV (48% vs. 43%; P = 0.2), NT-proBNP levels (3493 pg/mL vs. 3655 pg/mL; P = 0.988), nor high-sensitivity troponin T (72 ng/mL vs. 68 ng/mL; P = 0.519) (*Table 2, Figure 4A–C*). In terms of echocardiography characteristics, LVEF (50% in ID patients vs. 52% in non-ID patients; P = 0.669) and median IVST (17 mm in both groups; P = 0.828) were similar. In contrast, the left ventricular global longitudinal

strain (GLS) was increased in patients with ID compared with those without ID (-10% vs. -11%; *P* = 0.022) (*Table 2*).

In ATTRwt subtype, ID patients presented more dyspnoea, with 46% classified NYHA III-IV compared with 34% of non-ID patients (P = 0.023) (*Table 3*). There were no other differences in the subtypes concerning symptoms, cardiac biomarkers, or echocardiography attributes.

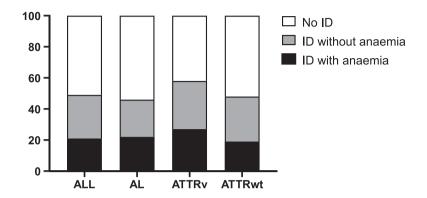
In the overall population, the C-reactive protein (CRP) levels were significantly higher in patients with functional ID, 4.3 ng/mL, compared with 2.8 ng/mL in those with absolute ID (P = 0.001) (supporting information, Table S1).

# Determinants of iron deficiency in cardiac amyloidosis

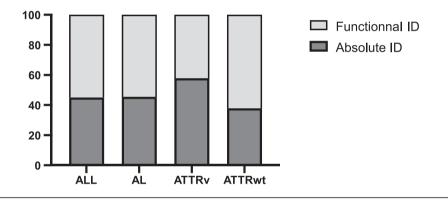
In multivariate analysis, by logistic regression (*Table 4*, *Figure 5*), the following variables were found to be significantly associated with ID: female sex, amyloidosis subtype (ATTRv/ATTRwt vs. AL), diabetes status, aspirin treatment, haemoglobin levels, and altered GLS.

Figure 3 Prevalence of iron deficiency (ID) among amyloid population. (A) Prevalence of iron deficiency with or without anaemia according to the type of cardiac amyloidosis. (B) Prevalence of absolute and functional iron deficiency according to the type of cardiac amyloidosis.

(A) Prevalence of iron deficiency with or without anaemia according to the type of cardiac amyloidosis



(B) Prevalence of absolute and functional iron deficiency according to the type of cardiac amyloidosis



#### Impact on outcome

During a mean follow-up 17 months, a total of 299 deaths occurred (*Figure S1*) illustrates the freedom from all-cause mortality depending on the presence or not of ID. The difference in mortality was not significant different in CA patients with or without ID (log rank = 0.785).

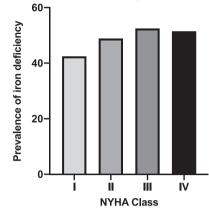
### **Discussion**

We found a high prevalence of ID in the CA patients (49%), which is comparable with that found in HF patients (50–62%).<sup>2,8,9</sup> ID was more frequent in ATTRv amyloidosis patients compared with those with ATTRwt and AL. The prevalence of ID was significantly associated with diabetes, aspirin treatment, altered GLS, and lower haemoglobin levels.

# High prevalence of iron deficiency in cardiac amyloidosis

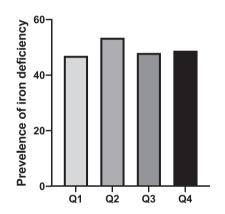
Iron deficiency incidence in CA is similar to that observed in HF patients and is not specific to a CA subtype. Thus, ID cannot be used for diagnosis. Multiple assessments, including electrocardiography, transthoracic echocardiogram, magnetic resonance imagery, and biological assessments (clonal expressions), should be used to diagnose CA. It is crucial that CA be diagnosis rapidly so that appropriate treatment, specific to the amyloidosis subtype, can be implemented. CA is a severe and debilitating disease with limited treatments to improve prognosis and symptoms.

In HF, several mechanisms cause inflammation and malabsorption of iron that results in ID.<sup>6</sup> Functional ID results from chronic inflammation with iron sequestration by macrophages.<sup>6</sup> Elevated levels of interleukin-6 are associated with inflammatory iron sequestration in HF and increased iron metabolism.<sup>29</sup> HF is also associated with structural and Figure 4 (A) Prevalence of iron deficiency according to New York Heart Association (NYHA) class. (B) Prevalence of iron deficiency according to quartile of N-terminal pro-B-type natriuretic peptide (NT-proBNP). (C) Prevalence of iron deficiency according to quartile of troponin T HS.

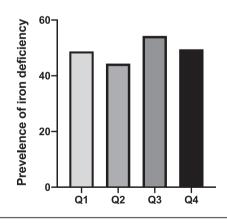


(A) Prevalence of iron deficiency according to NYHA class

(B) Prevalence of iron deficiency according to quartile of NT-proBNP



(C) Prevalence of iron deficiency according to quartile of troponin T HS



functional changes in the gut that result in peritoneal oedema and malabsorption, causing absolute ID.<sup>30</sup> We analysed albumin levels classified by tertiles according to ID status. Patients with ID had significantly decreased albumin levels, affirming the malabsorption hypothesis (*Figure S2*, T1 vs. T2 vs. T3, P = 0.022). In our study, we observed a more pronounced systemic inflammation (higher levels of CRP) in CA patients with functional ID than those with absolute ID. Thus, in patients with absolute ID, we suspect that other mechanisms specific to amyloidosis, such as gastrointestinal bleeding, might be involved.<sup>23</sup> Recently, a study confirmed, by duodenal biopsy,

		AL		ATTRV	Kv		ATTRW	Rwt	
N (%)	ID 123 (45)	No ID 148 (55)	٩	ID 95 (58)	No ID 69 (42)	ط	ID 182 (48)	No ID 199 (52)	٩
Clinical characteristics Age at inclusion, years	68 (55; 75)	67 (60; 74)	0.845	74 (68; 79)	71 (63; 78)	0.131	82 (76; 86)	81 (76; 84)	0.124
Gender, women <i>n</i> (%) BMI, kg/m <sup>2</sup>	61 (49) 24 (22; 27)	42 (28) 23 (21; 27)	<0.001 0.248	33 (35) 25 (22; 27)	22 (32) 24 (21; 27)	0.702 0.879	31 (17) 25 (22; 28)	21 (11) 25 (23; 28)	0.066 0.979
CV risk factors	(81) CC	73 (16)	U EUE	(36) 10	(21) 8	000	1967	(21) 90	000
Hypertension, n (%)	43 (35)	64 (43)	0.165	59 (62)	32 (46)	0.045	117 (64)	123 (62)	0.617
Dyslipidaemia, n (%) CV characteristics	31 (25)	42 (28)	0.590	24 (25)	18 (26)	0.948	82 (45)	62 (31)	0.029
NYHA Class III-IV vs. I-II, n (%)	60 (53)	70 (56)	0.650	41 (45)	28 (44)	0.988	78 (46)	65 (34)	0.023
Heart rate, beats/min	81 (71; 94)	82 (74; 93)	0.543	75 (67; 82)	74 (65; 84)	0.700	73 (67; 80)	75 (66; 81)	0.361
Systolic blood pressure, mmHg Atrial fibrillation 2 (%)	112 (101; 128) 12 (11)	111 (100; 126) 18 (13)	0.639	126 (108; 139) 21 (23)	115 (107; 130) 11 (10)	0.078	72 (114; 140) 72 (11)	130 (116; 145) 78 (72)	0.093
Ischaemic heart disease, n (%)	20 (17)	18 (12)	0.176	9 (10)	3 (4)	0.141	55 (30)	45 (23)	0.173
Echocardiography characteristics	(63.44) 63	EE (16, 62)		101.36/01	(03.7C/ 03	0 E 10	E0 /40. E0)	(4) E0	0 1 00
LVET, // IVST_mm	15 (14· 17)	15 (13·17)	0.254	17 (15·21)	18 (16·20)	64C.0	17 (15, 20)	18 (16: 20)	0.713
GL strain, %	10 (8; 13)	11 (9; 14)	0.320	10 (8; 12)	10 (8; 14)	0.150	10 (7; 12)	10 (8; 13)	0.116
Biology variables									
NT-proBNP, ng/mL Transmin T US	4960 (2036; 10 541)	5 227 (2394; 13 099)	0.634	2553 (1253; 5404)	2150 (799; 5440)	0.296	3310 (1929; 6368)	3233 (1556; 5605)	0.388
eGFR. mL/min/1.73 m <sup>2</sup>	62 (40, 122) 58 (41: 77)	64 (30, 131) 52 (35: 72)	0.162	51 (38: 65)	67 (36; 79) 58 (36; 79)	0.157	7 0 (40, 30) 47 (35: 58)	(ce (11) 20 48 (37: 61)	0.500
Haemoglobin, g/dL	12.4 (10.9; 13.6)	12.8 (10.8; 13.9)	0.320	12.7 (11.5; 13.6)	13.0 (11.8; 13.9)	0.259	13.2 (11.9; 14.2)	13.6 (12.6; 14.7)	0.001
Mean corpuscular volume,  m <sup>2</sup>	90 (85; 94)	96 (88; 97)	0.001	88 (84; 92)	92 (89; 96)	< 0.001	91 (86; 94)	94 (90; 97)	<0.001
Anaemia, <i>n</i> (%) Ferritin uc/l <sup>a</sup>	59 (48) 118 (59·185)	/6 (51) 343 (757: 578)	6/5/0//	44 (46) 81 (56·141)	26 (38) 308 (183: 482)	0.278	/1 (39) 176 (75: 208)	59 (30) 307 (205- 511)	20.0 200.0
Transferrin saturation. % <sup>a</sup>	13 (9: 17)	22 (18:27)	<0.001	14 (10: 17)	25 (21: 31)	<0.001	14 (11: 18)	23 (20: 28)	<0.001
Anticoagulant and or antiplatelet treatments	atments								
Aspirin, n (%)	37 (31)	38 (26)	0.350	27 (29)	7 (10)	0.004	58 (32)	46 (24)	0.068
Clopidogrel, n (%)	5 (4)	7 (5)	0.825	5 (5)	1 (1)	0.191	16 (9)	12 (6)	0.322
Ural anti-coagulants, <i>ח</i> (%) דרמשלאמיל השב	45 (38)	(44) 00	0.241	41 (44)	33 (48)	0.63/	123 (68)	126 (64)	0.499
ACE inhibitor. n (%)	16 (13)	26 (18)	0.343	27 (29)	27 (39)	0.178	58 (32)	62 (32)	0.961
ARB, n (%)	11 (9)	14 (10)	0.935	21 (23)	8 (12)	0.071	36 (20)		0.313
Digoxin, n (%)	1 (1)	1 (1)	0.881	1 (1)	0 (0)	0.388	4 (2)	7 (4)	0.427
Selective beta-blocker, n (%)	28 (23)	35 (24)	0.952	29 (31)		0.615	60 (33)	50 (26)	0.111
Non-selective beta-blocker, $n$ (%)	3 (3)	2 (1)	0.490	(0) 0		0.244	4 (2)	6 (3)	0.601
Calcium antagonist, <i>n</i> (%)	(19)	39 (26)	0.165	34 (37)	11 (16)	0.004	(12)/5	(05) 85	21/.0
Amiodarone, n (%)	14 (12)	24 (16)	0.288	18 (19)	13 (19)	0.934	43 (24)	47 (24)	0.936
Loop diuretic, <i>n</i> (%) Thisside diuretic <i>n</i> (%)	(9/) 1.6 (0/) cc	(77) 901	0.437	(10) 77 (30) 60	(07) CO	0.230	74 (81)	(72) (73) (73) (73) (73) (73) (73)	533 0
Vasodilator. n (%)	3 (3)	5 (3)	0.674	(oc) cc		0.299	4 (2)	(9C) 4 ( 8 (4)	0.297

ESC Heart Failure 2022; **9**: 1314–1327 DOI: 10.1002/ehf2.13818

		Univariate		Multivariate			
Variables	Ν	Hazard ratio	CI	P value	Hazard ratio	Cl	P value
Women	816	1.770	1.287; 2.434	< 0.001	1.919	1.360; 2.707	< 0.001
Amyloidosis type	271		AL reference			AL reference	
ATTRv	164	1.657	1.120; 2.451	0.012	1.849	1.227; 2.785	0.003
ATTRwt	381	1.100	0.805; 1.504	0.548	1.438	1.022; 2.023	0.037
Diabetes	816	1.935	1.346; 2.780	< 0.001	1.752	1.204; 2.548	0.003
Aspirin	808	1.581	1.153; 2.168	0.004	1.555	1.119; 2.161	0.009
Haemoglobin	816	0.892	0.827; 0.961	0.003	0.895	0.826; 0.969	0.006
Absolute GLS	816	0.970	0.941; 1.000	0.049	0.963	0.933; 0.994	0.020

Table 4 Univariate and multivariate logistic regression of the iron deficiency determinants in cardiac amyloidosis

CI, confidence interval; GLS, global longitudinal strain.

Figure 5 Iron d	leficiency's c	leterminants in	cardiac amy	loidosis.
-----------------	----------------	-----------------	-------------	-----------

Variables	Ν		Hazard ratio		P-Value
AL	271	-		Reference	
ATTRv	164	-	₽	1.849 (1.227; 2.785)	0.003
ATTRwt	381			1.438 (1.022; 2.023)	0.037
Women	816	4		1.919 (1.360; 2.707)	<0.001
Diabetes	816	4	<b>⊢−−−</b> −	1.752 (1.204; 2.548)	0.003
Aspirin	808	4	<b>├</b> ─── <b>■</b> ────┤	1.555 (1.119; 2.161)	0.009
Haemoglobin	816	4		0.895 (0.826; 0.969)	0.006
GLS	816	-		0.963 (0.933; 0.994)	0.020
		0	1 2	3	

submucosal amyloid deposits of AL and TTR fibrils in systemic amyloidosis.<sup>24</sup> As the duodenum is the predominant site of iron absorption, this infiltration may limit iron absorption in the intestine, leading to ID. In addition, intestinal bleeding, a major cause of ID, is associated with amyloidosis.<sup>31</sup> Amyloid deposits on blood vessel walls may cause erosive changes and mucosal friability leading to intestinal bleeding.<sup>31</sup> Aspirin treatment can accentuate this gastrointestinal frailty.<sup>32</sup>

The AL amyloidosis, contrary to ATTR, is often associated with hepatic amyloid infiltration.<sup>33</sup> This liver involvement includes hyperbilirubinemia and hepatic insufficiency.<sup>34</sup> In our AL CA population, we analysed bilirubin levels classified by tertiles: patients with ID had higher bilirubin levels (data not shown, T1 vs. T2 vs. T3, P = 0.031).

# Variations of iron deficiency prevalence and severity among cardiac amyloidosis types

Iron deficiency severity was different in the CA subtypes. ID was more severe (with lower ferritin levels) in ATTRv while HF severity appears to be less severe in the same patients (with lower NT-proBNP levels). The increased severity of ID

resulting from the prolonged exposure of the intestinal tract to amyloid protein in ATTRv amyloidosis due to early onset because of its genetic aetiology. This extended exposure can lead to chronic intestinal bleeding in ATTRv patients that exacerbates ID. ATTRv amyloidosis is also associated with gastroparesis, leading to malnutrition, attributed to autonomic dysfunction by nerve infiltrations.<sup>23</sup> In contrast, these gastrointestinal autonomic dysfunctions are poorly described in AL and ATTRwt amyloidosis.

### Clinical and imaging variables associated with iron deficiency in amyloidosis

Several characteristics were significantly associated with ID, including diabetes, aspirin treatment, and GLS. Diabetes was present in 24% of patients with ID compared with 14% in those without ID (P < 0.001). The association between ID and diabetes has already been described.<sup>2,9,35</sup> Indeed, Praveen *et al.* reported that 14/89 (16%) diabetic patients had ID<sup>35</sup> and identified a significantly increased inflammatory response in these patients. They suggest that chronic inflammation in diabetic patients leads to macrophage iron

sequestration (functional ID) and intestinal malabsorption (absolute ID).

Aspirin treatment was identified as a determinant of ID in our CA population (HR = 1.55, P = 0.009). As previously described, chronic use of aspirin increases the risk of gastrointestinal bleeding. A randomized control trial reported a 60% increase in gastrointestinal bleeding in patients using aspirin.<sup>32</sup>

Altered GLS was also associated with ID (P = 0.049). Altered GLS is an established independent prognosis marker in CA and is strongly associated with cardiac amyloid burden.<sup>36,37</sup> Data regarding a potential link between iron status and LV function are limited. Iron participates in several enzymatic reactions implicated in cellular respiration, oxidative phosphorylation, citric acid cycle, and production of reactive oxygen species.<sup>38,39</sup> Indeed, mitochondrial and left ventricular dysfunction have been associated with ID in an animal model.<sup>40</sup> Therefore, ID could be associated with amyloid infiltration and toxicity, leading to cardiomyocyte dysfunction in CA patients.

Iron therapy for HF patients with ID is beneficial, particularly in HFrEF and HFpEF patients.<sup>4,41–44</sup> The benefits include fewer cardiovascular-related hospitalizations, with improved heart function, exercise capacity, and quality of life. HF patients with anaemia benefit more from intravenous iron therapy than those without anaemia. Intravenous iron therapy reduces heart failure hospitalization but not cardiovascular mortality in HFrEF patients with ID.<sup>15</sup> Considering the prevalence of ID in CA, the benefit of iron therapy in these patients needs to be assessed.

## **Study limitations**

This study has several limitations. Our study was designed as an observational study and not a randomized controlled trial. HF is strongly associated with ID. Thus, in CA patients, the severity of HF may interfere with ID prevalence. Indeed, ID was associated with ATTRv patients that had less severe HF signs compared with AL or ATTRwt patients. Unfortunately, we did not collect data concerning the cardiac hepcidin–ferroportin axis shown to be essential for cardiomyocyte iron homeostasis and heart function. We spared patients from invasive cardiac biopsies that would have allowed us to assess cardiac iron concentration. In HFrEF, cardiac iron concentrations have been shown to be unrelated to biomarkers of systemic iron status or anaemia.<sup>45</sup> However, cardiac iron concentrations are associated with disease severity. It would have been interesting to see if this association also exists in CA.

Our retrospective study design does not allow us to further assess mortality in patients with and without ID, found to be not significantly different. A prospective study is needed to confirm our result. Finally, our study does not address the therapeutic use of iron in CA. The benefit of intravenous iron therapy in CA needs to be evaluated in a randomized, placebo-controlled trial.

## Conclusions

Iron deficiency is frequent in patients with CA and more prevalent in ATTRv compared with AL and ATTRwt amyloidosis. ID was independently associated with female gender, amyloidosis type, diabetes, aspirin treatment, lower haemoglobin levels, and altered left ventricular GLS. The presence of ID does not seem to be associated with increased mortality. Prospective studies are needed to assess whether ID affects morbidity and mortality, as well as the benefit of iron in CA patients.

### **Conflict of interest**

T.D. has received grant and/or consultant fees from Pfizer, Alnylam, Vifor, Akcea, Novartis, Resmed, Bayer, Astra-Zeneca, Sanofi-Aventis. A.J.D. has received grant and/or consultant fees from Novartis, Amicus, Sanofi-Genzyme, Boehringer Ingelheim.

## Supporting information

Additional supporting information may be found online in the Supporting Information section at the end of the article.

Figure S1: Survival according iron status. Figure S2: Ferritin levels according to albumin. Table S1: CRP levels according ID status and CA subtype.

## References

- 1. Camaschella C. Iron deficiency. *Blood* 2019; **133**: 30–39.
- Klip IT, Comin-Colet J, Voors AA, Ponikowski P, Enjuanes C, Banasiak W, Lok DJ, Rosentryt P, Torrens A,

Polonski L, Van Veldhuisen DJ. Iron deficiency in chronic heart failure: an international pooled analysis. *Am Heart J* 2013; **165**: 575–582.e3.  Macdougall IC, Bock AH, Carrera F, Eckardt KU, Gaillard C, van Wyck D, Roubert B, Nolen JG, Roger SD, FIND-CKD Study Investigators. FIND-CKD: a randomized trial of intravenous ferric carboxymaltose versus oral iron in patients with chronic kidney disease and iron deficiency anaemia. *Nephrol Dial Transplant* 2014; **29**: 2075–2084.

- Anker SD, Comin Colet J, Filippatos G, Willenheimer R, Dickstein K, Drexler H, Lüscher TF, Bart B, Banasiak W, Niegowska J, Kirwan BA, Mori C, von Eisenhart Rothe B, Pocock SJ, Poole-Wilson PA, Ponikowski P. Ferric carboxymaltose in patients with heart failure and iron deficiency. N Engl J Med 2009; 361: 2436–2448.
- 5. Authors/Task Force Members. McMurray JJV, Adamopoulos S, Anker SD, Auricchio A, Böhm M, Dickstein K, Falk V, Filippatos G, Fonseca C, Gomez-Sanchez MA, Jaarsma T, Køber L, Lip GYH, Maggioni AP, Parkhomenko A, Pieske BM, Popescu BA, Rønnevik PK, Rutten FH, Schwitter J, Seferovic P, Stepinska J, Trindade PT, Voors AA, Zannad F, Zeiher A, ESC Committee for Practice Guidelines (CPG), Bax JJ, Baumgartner H, Ceconi C, Dean V, Deaton C, Fagard R, Funck-Brentano C, Hasdai D, Hoes A, Kirchhof P, Knuuti J, Kolh P, McDonagh T, Moulin C, Popescu BA, Reiner Ž, Sechtem U, Sirnes PA, Tendera M, Torbicki A, Vahanian A, Windecker S, Document Reviewers, McDonagh T, Sechtem U, Bonet LA, Avraamides P, Ben Lamin HA, Brignole M, Coca A, Cowburn P, Dargie H, Elliott P, Flachskampf FA, Guida GF, Hardman S, Iung B, Merkely B, Mueller C, Nanas JN, Nielsen OW, Ørn S, Parissis JT, Ponikowski P. ESC guidelines for the diagnosis and treatment of acute and chronic heart failure 2012: the task force for the diagnosis and treatment of acute and chronic heart failure 2012 of the European Society of Cardiology. Developed in collaboration with the Heart Failure Association (HFA) of the ESC. Eur J Heart Fail 2012; 14: 803-869.
- Anand IS, Gupta P. Anemia and iron deficiency in heart failure: current concepts and emerging therapies. *Circulation* 2018; 138: 80–98.
- Jankowska EA, von Haehling S, Anker SD, Macdougall IC, Ponikowski P. Iron deficiency and heart failure: diagnostic dilemmas and therapeutic perspectives. *Eur Heart J* 2013; 34: 816–829.
- 8. Yeo TJ, Yeo PSD, Ching-Chiew Wong R, Ong HY, Leong KTG, Jaufeerally F, Sim D, Santhanakrishnan R, Lim SL, M.Y. Chan M, Chai P, Low AF, Ling LH, Ng TP, Richards AM, Lam CSP. Iron deficiency in a multi-ethnic Asian population with and without heart failure: prevalence, clinical correlates, functional significance and prognosis: iron deficiency in Asian heart failure. *Eur J Heart Fail* 2014; **16**: 1125–1132.
- Martens P, Nijst P, Verbrugge FH, Smeets K, Dupont M, Mullens W. Impact of iron deficiency on exercise capacity and outcome in heart failure with reduced, mid-range and preserved ejection fraction. *Acta Cardiol* 2018; 73: 115–123.

- Bekfani T, Pellicori P, Morris D, Ebner N, Valentova M, Sandek A, Doehner W, Cleland JG, Lainscak M, Schulze PC, Anker SD, von Haehling S. Iron deficiency in patients with heart failure with preserved ejection fraction and its association with reduced exercise capacity, muscle strength and quality of life. *Clin Res Cardiol* 2019; **108**: 203–211.
- Okonko DO, Grzeslo A, Witkowski T, Mandal AKJ, Slater RM, Roughton M, Foldes G, Thum T, Majda J, Banasiak W, Missouris CG, Poole-Wilson PA, Anker SD, Ponikowski P. Effect of intravenous iron sucrose on exercise tolerance in anemic and nonanemic patients with symptomatic chronic heart failure and iron deficiency. J Am Coll Cardiol 2008; **51**: 103–112.
- Toblli JE, Lombraña A, Duarte P, di Gennaro F. Intravenous iron reduces NT-pro-brain natriuretic peptide in anemic patients with chronic heart failure and renal insufficiency. J Am Coll Cardiol 2007; 50: 1657–1665.
- 13. Ponikowski P, Kirwan BA, Anker SD, McDonagh T, Dorobantu M, Drozdz J, Fabien V, Filippatos G, Göhring UM, Keren A, Khintibidze I, Kragten H, Martinez FA, Metra M, Milicic D, Nicolau JC, Ohlsson M, Parkhomenko A, Pascual-Figal DA, Ruschitzka F, Sim D, Skouri H, van der Meer P, Lewis BS, Comin-Colet J, von Haehling S, Cohen-Solal A, Danchin N, Doehner W, Dargie HJ, Motro M, Butler J, Friede T, Jensen KH, Pocock S, Jankowska EA, Azize G, Fernandez A, Zapata GO, Garcia Pacho P, Glenny A, Ferre Pacora F, Parody ML, Bono J, Beltrano C, Hershson A, Vita N, Luquez HA, Cestari HG, Fernandez H, Prado A, Berli M, García Durán R, Thierer J, Diez M, Lobo Marquez L, Borelli RR, Hominal MÁ, Metra M, Ameri P, Agostoni P, Salvioni A, Fattore L, Gronda E, Ghio S, Turrini F, Uguccioni M, di Biase M, Piepoli M, Savonitto S, Mortara A, Terrosu P, Fucili A, Boriani G, Midi P, Passamonti E, Cosmi F, van der Meer P, van Bergen P, van de Wetering M, Al-Windy NYY, Tanis W, Meijs M, Groutars RGEJ, The HKS, Kietselaer B, van Kesteren H, Beelen DPW, Heymeriks J, van de Wal R, Schaap J, Emans M, Westendorp P, Nierop PR, Nijmeijer R, Manintveld OC, Dorobantu М, Darabantiu DA Zdrenghea D, Toader DM, Petrescu L, Militaru C, Crisu D, Tomescu MC, Stanciulescu G, Rodica Dan A, Iosipescu LC, Serban DL, Drozdz J, Szachniewicz J, Bronisz M, Tycińska A, Wozakowska-Kaplon B, Mirek-Bryniarska E, Gruchała M. Nessler J. Straburzvńska-Migai E. Mizia-Stec K, Szelemej R, Gil R, Gasior M, Gotsman I, Halabi M, Shochat M, Shechter M, Witzling V, Zukermann R, Arbel Y, Flugelman M, Ben-Gal T, Zvi V, Kinany W, Weinstein JM, Atar S, Goland S, Milicic D, Horvat D, Tušek S, Udovicic M, Šutalo K, Samodol A, Pesek K, Artuković M, Ružić A, Šikić J, McDonagh

T, Trevelvan J, Wong YK, Gorog D, Ray R, Pettit S, Sharma S, Kabir A, Hamdan H, Tilling L, Baracioli L, Nigro Maia L, Dutra O, Reis G, Pimentel Filho P, Saraiva JF, Kormann A, dos Santos F, Bodanese L, Almeida D, Precoma D, Rassi S, Costa F, Kabbani S, Abdelbaki K, Abdallah C, Arnaout MS, Azar R, Chaaban S, Raed O, Kiwan G, Hassouna B, Bardaji A, Zamorano J, del Prado S, Gonzalez Juanatev JR. Ga Bosa Oieda FI, Gomez Bueno M, Molina BD, Pascual Figal DA, Sim D, Yeo TJ, Loh SY, Soon D, Ohlsson M, Smith JG, Gerward S, Khintibidze I, Lominadze Z, Chapidze G, Emukhvari N, Khabeishvili G, Chumburidze V, Paposhvili Κ, Т, Khabeishvili Shaburishvili G, Parhomenko O, Kraiz I, Koval O, Zolotaikina V, Malynovsky Y, Vakaliuk I, Rudenko L, Tseluyko V, Stanislavchuk M. Ferric carboxymaltose for iron deficiency at discharge after acute heart failure: a multicentre, double-blind, randomised, controlled trial. Lancet 2020; 396: 1895-1904.

- 14. Bolger AP, Bartlett FR, Penston HS, O'Leary J, Pollock N, Kaprielian R, Chapman CM. Intravenous iron alone for the treatment of anemia in patients with chronic heart failure. J Am Coll Cardiol 2006; 48: 1225–1227.
- Mohammed SF, Mirzoyev SA, Edwards WD, Dogan A, Grogan DR, Dunlay SM, Roger VL, Gertz MA, Dispenzieri A, Zeldenrust SR, Redfield MM. Left ventricular amyloid deposition in patients with heart failure and preserved ejection fraction. *JACC Heart Fail* 2014; 2: 113–122.
- 16. González-López E, Gallego-Delgado M, Guzzo-Merello G, de Haro-del Moral FJ, Cobo-Marcos M, Robles C, Bornstein B, Salas C, Lara-Pezzi E, Alonso-Pulpon L, Garcia-Pavia P. Wild-type transthyretin amyloidosis as a cause of heart failure with preserved ejection fraction. Eur Heart J 2015; 36: 2585–2594.
- Lindmark K, Pilebro B, Sundström T, Lindqvist P. Prevalence of wild type transtyrethin cardiac amyloidosis in a heart failure clinic. *ESC Heart Fail* 2020; 8: ehf2.13110.
- Damy T, Maurer MS, Rapezzi C, Planté-Bordeneuve V, Karayal ON, Mundayat R, Suhr OB, Kristen AV. Clinical, ECG and echocardiographic clues to the diagnosis of TTR-related cardiomyopathy. *Open Heart* 2016; 3: e000289.
- Damy T, Jaccard A, Guellich A, Lavergne D, Galat A, Deux JF, Hittinger L, Dupuis J, Frenkel V, Rigaud C, Plante-Bordeneuve V, Bodez D, Mohty D. Identification of prognostic markers in transthyretin and AL cardiac amyloidosis\*. *Amyloid* 2016; 23: 194–202.
- 20. Gillmore JD, Maurer MS, Falk RH, Merlini G, Damy T, Dispenzieri A, Wechalekar AD, Berk JL, Quarta CC, Grogan M, Lachmann HJ, Bokhari S, Castano A, Dorbala S, Johnson GB, Glaudemans AWJM, Rezk T, Fontana

M, Palladini G, Milani P, Guidalotti PL, Flatman K, Lane T, Vonberg FW, Whelan CJ, Moon JC, Ruberg FL, Miller EJ, Hutt DF, Hazenberg BP, Rapezzi C, Hawkins PN. Nonbiopsy diagnosis of cardiac transthyretin amyloidosis. *Circulation* 2016; **133**: 2404–2412.

- Damy T, Kristen AV, Suhr OB, Maurer MS, Planté-Bordeneuve V, Yu CR, Ong ML, Coelho T, Rapezzi C, THAOS Investigators. Transthyretin cardiac amyloidosis in continental Western Europe: an insight through the transthyretin amyloidosis outcomes survey (THAOS). Eur Heart J 2019: ehz173.
- Mohty D, Damy T, Cosnay P, Echahidi N, Casset-Senon D, Virot P, Jaccard A. Cardiac amyloidosis: updates in diagnosis and management. *Arch Cardiovasc Dis* 2013; **106**: 528–540.
- Obici L, Suhr OB. Diagnosis and treatment of gastrointestinal dysfunction in hereditary TTR amyloidosis. *Clin Auton Res* 2019; 29: 55–63.
- Iida T, Yamano H, Nakase H. Systemic amyloidosis with gastrointestinal involvement: diagnosis from endoscopic and histological views: endoscopic views of systemic amyloidosis. J Gastroenterol Hepatol 2018; 33: 583–590.
- Dispenzieri A, Merlini G, Comenzo RL. Amyloidosis: 2008 BMT Tandem Meetings (February 13–17, San Diego). Biol Blood Marrow Transplant 2008; 14: 6–11.
- 26. Béquignon E, Guellich A, Bartier S, Raynal M, Prulière-Escabasse V, Canouï-Poitrine F, Coste A, Damy T. How your ears can tell what is hidden in your heart: wild-type transthyretin amyloidosis as potential cause of sensorineural hearing loss inelderly-AmyloDEAFNESS pilot study. Amyloid 2017; 24: 96–100.
- 27. Manito N, Cerqueiro JM, Comín-Colet J, García-Pinilla JM, González-Franco A, Grau-Amorós J, Peraira JR, Manzano L. Documento de consenso de la Sociedad Española de Cardiología y la Sociedad Española de Medicina Interna sobre el diagnóstico y tratamiento del déficit de hierro en la insuficiencia cardíaca. *Rev Clin Esp* 2017; 217: 35–45.
- McDonagh T, Damy T, Doehner W, Lam CSP, Sindone A, van der Meer P, Cohen-Solal A, Kindermann I, Manito N, Pfister O, Pohjantähti-Maaroos H, Taylor J, Comin-Colet J. Screening, diagnosis and treatment of iron deficiency in chronic heart failure: putting the 2016 European Society of Cardiology heart failure guidelines into clinical practice. *Eur J Heart Fail* 2018; 20: 1664–1672.
- 29. Markousis-Mavrogenis G, Tromp J, Ouwerkerk W, Devalaraja M, Anker SD, Cleland JG, Dickstein K, Filippatos GS, Harst P, Lang CC, Metra M, Ng LL, Ponikowski P, Samani NJ, Zannad F, Zwinderman AH, Hillege HL, Veldhuisen DJ, Kakkar R, Voors AA, Meer P. The clinical significance of interleukin-6 in heart failure: results

from the BIOSTAT-CHF study. *Eur J Heart Fail* 2019; **21**: 965–973.

- Sandek A, Bauditz J, Swidsinski A, Buhner S, Weber-Eibel J, von Haehling S, Schroedl W, Karhausen T, Doehner W, Rauchhaus M, Poole-Wilson P. Altered intestinal function in patients with chronic heart failure. *J Am Coll Cardiol* 2007; **50**: 1561–1569.
- Cowan AJ, Skinner M, Seldin DC, Berk JL, Lichtenstein DR, O'Hara CJ, Doros G, Sanchorawala V. Amyloidosis of the gastrointestinal tract: a 13-year, single-center, referral experience. *Haematologica* 2013; 98: 141–146.
- 32. Mahady SE, Margolis KL, Chan A, Polekhina G, Woods RL, Wolfe R, Nelson MR, Lockery JE, Wood EM, Reid C, Ernst ME, Murray A, Thao LTP, McNeil JJ. Major GI bleeding in older persons using aspirin: incidence and risk factors in the ASPREE randomised controlled trial. Gut 2021; 70: 717–724.
- 33. Abe R, Katoh N, Takahashi Y, Takasone K, Yoshinaga T, Yazaki M, Kametani F, Sekijima Y. Distribution of amyloidosis subtypes based on tissue biopsy site—consecutive analysis of 729 patients at a single amyloidosis center in Japan. Pathol Int 2020; 71: pin.13041.
- Rosenzweig M, Comenzo RL. Liver and gastrointestinal involvement. *Hematol* Oncol Clin North Am 2020; 34: 1081–1090.
- 35. Praveen M, Jain N, Raizada N, Sharma S, Narang S, Madhu SV. Anaemia in patients with type 2 diabetes mellitus without nephropathy is related to iron deficiency. *Diabetes Metab Syndr Clin Res Rev* 2020; 14: 1837–1840.
- Quarta CC, Solomon SD, Uraizee I, Kruger J, Longhi S, Ferlito M, Gagliardi C, Milandri A, Rapezzi C, Falk RH. Left ventricular structure and function in transthyretin-related versus light-chain cardiac amyloidosis. *Circulation* 2014; 129: 1840–1849.
- 37. Ternacle J, Bodez D, Guellich A, Audureau E, Rappeneau S, Lim P, Radu C, Guendouz S, Couetil JP, Benhaiem N, Hittinger L, Dubois-Randé JL, Plante-Bordeneuve V, Mohty D, Deux JF, Damy T. Causes and consequences of longitudinal LV dysfunction assessed by 2D strain echocardiography in cardiac amyloidosis. JACC Cardiovasc Imaging 2016; 9: 126–138.
- Lakhal-Littleton S. Mechanisms of cardiac iron homeostasis and their importance to heart function. *Free Radic Biol Med* 2019; 133: 234–237.
- Jankowska EA, Ponikowski P. Molecular changes in myocardium in the course of anemia or iron deficiency. *Heart Fail Clin* 2010; 6: 295–304.
- 40. Dong F, Zhang X, Culver B, Chew HG Jr, Kelley RO, Ren J. Dietary iron deficiency induces ventricular dilation, mitochondrial ultrastructural aberrations and cytochrome c release: involvement of nitric oxide synthase and protein tyro-

sine nitration. *Clin Sci* 2005; **109**: 277–286.

- 41. Zhang J, Hu S, Jiang Y, Zhou Y. Efficacy and safety of iron therapy in patients with chronic heart failure and iron deficiency: a systematic review and meta-analysis based on 15 randomised controlled trials. *Postgrad Med J* 2020: postgradmedj-2019–137342.
- 42. Anker SD, Kirwan BA, van Veldhuisen DJ, Filippatos G, Comin-Colet J, Ruschitzka F, Lüscher TF, Arutyunov GP, Motro M, Mori C, Roubert B. Effects of ferric carboxymaltose on hospitalisations and mortality rates in iron-deficient heart failure patients: an individual patient data meta-analysis. *Eur J Heart Fail* 2018; **20**: 125–133.
- 43. Ponikowski P, Voors AA, Anker SD, Bueno H. Cleland JGF. Coats AJS. Falk V, González-Juanatey JR, Harjola VP, Jankowska EA, Jessup M, Linde C, Nihoyannopoulos P, Parissis JT, Pieske B, Riley JP, Rosano GMC, Ruilope LM, Ruschitzka F, Rutten FH, van der Meer P, ESC Scientific Document Group. 2016 ESC guidelines for the diagnosis and treatment of acute and chronic heart failure: the task force for the diagnosis and treatment of acute and chronic heart failure of the European Society of Cardiology (ESC) developed with the special contribution of the Heart Failure Association (HFA) of the ESC. Eur Heart J 2016; 37: 2129-2200.
- McDonagh TA, Metra M, Adamo M, 44. Gardner RS, Baumbach A, Böhm M, Burri H, Butler J, Čelutkiene J, Chioncel O, Cleland JGF, Coats AJS, Crespo-Leiro MG, Farmakis D, Gilard M, Heymans S, Hoes AW, Jaarsma T, Jankowska EA, Lainscak M, Lam CSP, Lyon AR, McMurray JJV, Mebazaa A, Mindham R, Muneretto C, Francesco Piepoli M, Price S, Rosano GMC, Ruschitzka F, Kathrine Skibelund A, ESC Scientific Document Group, de Boer RA, Christian Schulze P, Abdelhamid M, Aboyans V, Adamopoulos S, Anker SD, Arbelo E, Asteggiano R, Bauersachs J, Bayes-Genis A, Borger MA, Budts W, Cikes M, Damman K, Delgado V, Dendale P, Dilaveris P, Drexel H, Ezekowitz J, Falk V, Fauchier L, Filippatos G, Fraser A, Frey N, Gale CP, Gustafsson F, Harris J, Iung B, Janssens S, Jessup M, Konradi A, Kotecha D, Lambrinou E, Lancellotti P, Landmesser U, Leclercq C, Lewis BS, Leyva F, Linhart A, Løchen ML, Lund LH, Mancini D, Masip J, Milicic D, Mueller C, Nef H, Nielsen JC, Neubeck L, Noutsias M, Petersen SE, Sonia Petronio A, Ponikowski P, Prescott E, Rakisheva A, Richter DJ, Schlvakhto E, Seferovic P, Senni M, Sitges M, Sousa-Uva M, Tocchetti CG, Touyz RM, Tschoepe C, Waltenberger J, Adamo M, Baumbach A, Böhm M, Burri H, Čelutkienė J, Chioncel O, Cleland JGF, Coats AJS, Crespo-Leiro MG, Farmakis D, Gardner RS, Gilard M, Heymans S, Hoes AW, Jaarsma T, Jankowska EA,

Lainscak M, Lam CSP, Lyon AR, McMurray JJV, Mebazaa A, Mindham R, Muneretto C, Piepoli MF, Price S, Rosano GMC, Ruschitzka F, Skibelund AK. 2021 ESC guidelines for the diagnosis and treatment of acute and chronic heart failure. *Eur Heart J* 2021; **42**: 3599–3726.

 Hirsch VG, Tongers J, Bode J, Berliner D, Widder JD, Escher F, Mutsenko V, Chung B, Rostami F, Guba-Quint A, Giannitsis E, Schultheiss HP, Vogt C, Bauersachs J, Wollert KC, Kempf T. Cardiac iron concentration in relation to systemic iron status and disease severity in non-ischaemic heart failure with reduced ejection fraction. *Eur J Heart Fail* 2020; **22**: 2038–2046.