

# The Hong Kong consensus recommendations on the diagnosis and management of pancreatic cystic lesions

# Tan-To Cheung<sup>1</sup><sup>^</sup>, Yuk Tong Lee<sup>2</sup>, Raymond Shing-Yan Tang<sup>3</sup><sup>^</sup>, Wong Hoi She<sup>1</sup><sup>^</sup>, Kai Chi Cheng<sup>4</sup><sup>^</sup>, Chin Cheung Cheung<sup>5</sup>, Keith Wan Hang Chiu<sup>6</sup><sup>^</sup>, Kenneth Siu Ho Chok<sup>1</sup><sup>^</sup>, Wing Sun Chow<sup>7</sup><sup>^</sup>, Tak Wing Lai<sup>8</sup>, Wai-Kay Seto<sup>9,10</sup><sup>^</sup>, Thomas Yau<sup>7</sup><sup>^</sup>

<sup>1</sup>Department of Surgery, Queen Mary Hospital, The University of Hong Kong, Hong Kong, China; <sup>2</sup>Gastroenterologist in private practice, Hong Kong, China; <sup>3</sup>Department of Medicine and Therapeutics, The Chinese University of Hong Kong, Hong Kong, China; <sup>4</sup>Department of Surgery, Kwong Wah Hospital, Hong Kong, China; <sup>5</sup>Department of Surgery, Tuen Mun Hospital, Hong Kong, China; <sup>6</sup>Department of Radiology & Imaging, Queen Elizabeth Hospital, Hong Kong, China; <sup>7</sup>Department of Medicine, The University of Hong Kong, Hong Kong, China; <sup>8</sup>Department of Surgery, Princess Margaret Hospital, Hong Kong, China; <sup>9</sup>Department of Medicine, School of Clinical Medicine, The University of Hong Kong, Hong Kong, Hong Kong, Hong Kong, China; <sup>10</sup>State Key Laboratory of Liver Research, The University of Hong Kong, Hong Kong, China

*Contributions:* (I) Conception and design: TT Cheung; (II) Administrative support: TT Cheung; (III) Provision of study materials or patients: KC Cheng, TT Cheung, KWH Chiu, KSH Chok, WS Chow, WK Seto, WH She, RSY Tang, T Yau; (IV) Collection and assembly of data: KC Cheng, TT Cheung, KWH Chiu, KSH Chok, WS Chow, WK Seto, WH She, RSY Tang, T Yau; (V) Data analysis and interpretation: All authors; (VI) Manuscript writing: All authors; (VII) Final approval of manuscript: All authors.

*Correspondence to:* Tan-To Cheung, MBBS, MS. Clinical Professor, Chief of Hepatobiliary and Pancreatic Surgery, Department of Surgery, Queen Mary Hospital, The University of Hong Kong, 102 Pokfulam Road, Hong Kong, China. Email: tantocheung@hotmail.com.

**Background:** The finding of pancreatic cystic lesions (PCL) on incidental imaging is becoming increasingly common. International studies report a prevalence of 2.2–44.7% depending on the population, imaging modality and indication for imaging, and the prevalence increases with age. Patients with PCL are at risk of developing pancreatic cancer, a disease with a poor prognosis. This publication summarizes recommendations for the diagnosis and management of PCL and post-operative pancreatic exocrine insufficiency (PEI) from a group of local specialists.

**Methods:** Clinical evidence was consolidated from narrative reviews and consensus statements formulated during two online meetings in March 2022. The expert panel included gastroenterologists, hepatobiliary surgeons, oncologists, radiologists, and endocrinologists.

**Results:** Patients with PCL require careful investigation and follow-up due to the risk of malignant transformation of these lesions. They should undergo clinical investigation and pancreas-specific imaging to classify lesions and understand the risk profile of the patient. Where indicated, patients should undergo pancreatectomy to excise PCL. Following pancreatectomy, patients are at risk of PEI, leading to gastrointestinal dysfunction and malnutrition. Therefore, such patients should be monitored for symptoms of PEI, and promptly treated with pancreatic enzyme replacement therapy (PERT). Patients with poor response to PERT may require increases in dose, addition of a proton pump inhibitor, and/or further investigation, including tests for pancreatic function. Patients are also at risk of new-onset diabetes mellitus after pancreatectomy; they should be screened and treated with insulin if indicated.

**Conclusions:** These statements are an accurate summary of our approach to the diagnosis and management of patients with PCL and will be of assistance to clinicians treating these patients in a similar clinical landscape.

Keywords: Pancreatic cystic lesions; pancreatic endocrine insufficiency; pancreatic enzyme replacement therapy

ORCID: Tan-To Cheung, 0000-0002-2633-5883; Raymond Shing-Yan Tang, 0000-0002-2433-8036; Wong Hoi She, 0000-0003-2049-3140; Kai Chi Cheng, 0000-0002-6440-7825; Keith Wan Hang Chiu, 0000-0002-7930-1193; Kenneth Siu Ho Chok, 0000-0001-7921-3807; Wing Sun Chow, 0000-0003-1625-6716; Wai-Kay Seto, 0000-0002-9012-313X; Thomas Yau, 0000-0002-5221-9755.

Submitted Oct 07, 2022. Accepted for publication Feb 10, 2023. Published online Jul 06, 2023. doi: 10.21037/hbsn-22-471

View this article at: https://dx.doi.org/10.21037/hbsn-22-471

# Introduction

Pancreatic cystic lesions (PCL) comprise a diverse group of neoplasms and are mostly diagnosed incidentally during radiographic scans for non-pancreatic indications (1). They may be benign, but can also progress to pancreatic cancer-a disease with limited treatment options and poor outcomes (2). Therefore, identifying and monitoring patients at risk of malignant disease is critical. Patients with PCL at low risk of malignant disease are managed with surveillance, but partial or total pancreatectomy is indicated for those with higher-risk PCL (3,4). Following pancreatectomy, patients are at risk of pancreatic exocrine insufficiency (PEI) and consequent gastrointestinal (GI) dysfunction and malnutrition (5), as well as new-onset diabetes mellitus (NODM) (6); these can be managed via pancreatic enzyme replacement therapy (PERT) (7,8) and insulin replacement, respectively (9).

### Highlight box

### Key findings

- Patients with pancreatic cystic lesions (PCL) should be monitored and undergo pancreas-specific imaging;
- Endoscopic ultrasound can be considered when worrisome features are identified;
- Medically fit patients at high risk of pancreatic cancer should undergo pancreatectomy;
- After pancreatectomy, patients should be monitored for the development of pancreatic exocrine insufficiency (PEI);
- Patients with PEI should be treated with pancreatic enzyme replacement therapy (PERT).

### What is known and what is new?

- Previous guidelines focused on either PCL diagnosis and presurgical management or post-surgical management;
- Our recommendations offer comprehensive advice on both preand post-surgical management, adapted to the clinical landscape of East Asia.

### What is the implication, and what should change now?

- These guidelines summarize the latest data on PCL management, focusing on current practice in Hong Kong;
- Future research topics include optimizing PCL monitoring, improving pancreatectomy outcomes, and optimizing PERT dosing in Asian patients.

The incidence of PCL in European and North American populations is increasing (10), and although formal data are not available, clinical experience suggests the incidence of PCL is also increasing in Hong Kong. Registry data from 2019 show that pancreatic cancer had the fifth highest mortality among cancer types in the territory (11). The challenges of correctly diagnosing PCL patients were illustrated in a recent retrospective study that compared preoperative versus final diagnosis and found that in 22% of patients, the pathology did not correlate between the two stages (12). Updated, evidence-based guidelines may reduce the risk of unnecessary surgeries, and although international guidelines for management of PCL and complications subsequent to pancreatectomy are available (13), there is a need for guidance that is tailored to the clinical landscape of Hong Kong. Furthermore, there is a need in East Asia for a broad guidance document that covers multiple aspects of the diagnosis and management of patients with PCL, PEI, and NODM. To meet these needs and assist physicians treating patients with PCL, a group of local experts formulated consensus statements to guide healthcare professionals in the diagnosis and management of PCL.

### Literature review and consensus methodology

Two online meetings including gastroenterologists, surgeons, oncologists, radiologists, and endocrinologists were convened in March 2022. The selection of experts included physicians from both public hospitals and private practice and was representative of the specialists involved in the diagnosis and management of patients with PCL and subsequent PEI. Prior to the meeting, selected experts performed a narrative review of literature and formulated consensus statements on the diagnosis and management of PCL, PEI, and post-pancreatectomy diabetes mellitus (Table 1; Table S1). To integrate the diverse medical expertise of the authors into consensus statements in an adaptive, anonymized and unbiased manner, the Delphi methodology was used under the supervision of a medical writer (Figure S1). Draft consensus statements and their supporting data were presented to the committee and anonymously evaluated using a Likert scale (1, accept completely; 2, accept with some reservations; 3, accept

Summary			

The Polyment of Harrady Provession Strategy			
Item	Specification		
Date of search	Up to March 2022		
Databases and other sources searched	PubMed		
Search terms used	See Table S1		
Inclusion and exclusion criteria	Inclusion: English language		
	Exclusion: Studies in model organisms, non-English language publications		
Selection process	Relevance as assessed by reviewing author for each topic (listed in Table S1)		

with major reservations; 4, reject with reservations; 5, reject completely). Voting was anonymous, and consensus was defined as  $\geq$ 80% of participants voting to accept a statement 'completely' or 'with some reservations'. Where this threshold was not achieved, statements were revised and voting repeated until a consensus was met. Evidence supporting the statements was evaluated using the Oxford Centre for Evidence-Based Medicine's 2011 Levels of Evidence (14) and this manuscript was prepared in alignment with the CREDES reporting checklist (available at https://hbsn.amegroups.com/article/view/10.21037/ hbsn-22-471/rc) (15).

### Prevalence and classification of PCL

Statement 1: Incidental pancreatic cystic lesions detected on crosssectional imaging performed for non-pancreatic indications are common, and their prevalence increases with age (Level 2)

Statement 2: Pancreatic cystic lesions can be either neoplastic or non-neoplastic cysts. Neoplastic cysts can be broadly classified as serous or mucinous pancreatic cystic lesions, or cystic degeneration of solid tumors of the pancreas

Statement 3: Mucinous pancreatic cystic lesions such as intraductal papillary mucinous neoplasms (main duct type, branch duct type, or mixed type) and mucinous cystic neoplasms are considered pre-malignant lesions with variable malignant potential (Level 1)

In epidemiological studies from the USA, Europe, and Korea, estimates of the prevalence of PCL range from 2.2% to 44.7% (*Table 2*), with many studies reporting increasing prevalence of PCL with increasing age (16-20). The authors' clinical experience suggests that the prevalence of PCL in Hong Kong is within this range, and that prevalence of PCL is increasing, consistent with the aging trend in the population of the territory (21). The wide range in prevalence rates reported is explained by differences in the methods of calculation, imaging indication, and imaging modality used; for example, studies that use magnetic resonance imaging (MRI) generally report higher incidences than studies using computed tomography (CT) scans (22). The very high prevalence seen in the study of Girometti and colleagues, 44.7%, is likely due to the use of magnetic resonance cholangiopancreatography (MRCP) (18), which is usually performed for patients with suspected bile duct or pancreatic pathology.

PCL can be classified as either neoplastic or nonneoplastic cysts, and neoplastic cysts can be further classified as serous PCL, mucinous PCL, or cystic degeneration of solid pancreatic tumors. Intraductal papillary mucinous neoplasm (IPMN), a common PCL subtype, is also classified according to its involvement with the main pancreatic duct (MD) or one of the branch ducts (BD) (10). Common mixed solid and cystic lesions may include cystic degeneration of neuroendocrine tumors, solid pseudopapillary tumors, adenocarcinoma, and mucinous cystic lesions with a solid component. The scheme from the European Study Group on Cystic Tumours of the Pancreas (ESGCTP), which broadly classifies cysts by epithelial/ nonepithelial and neoplastic/non-neoplastic status based on the World Health Organization criteria (3), is a useful guide for cyst classification.

Accurate classification of PCL is important because the risk of malignant transformation varies among subtypes. For IPMN, in an extensive review performed by Tanaka and colleagues to inform a 2012 guideline publication (23), the chance of malignant transformation was >62.2% for MD IPMN, >24.4% for BD IPMN, and >57.6% for mixed-type IPMN (24-45). In a review of published surgical cases of resected MD IPMN, included in a 2017 guideline (4), invasive carcinoma and high-grade dysplasia were found in 61.6% of subjects (26,30-35,38-44). Risk of malignant transformation is generally lower in BD IPMN than MD

Study and location	Population, imaging modality, and indication	PCL prevalence	Comments
Laffan <i>et al.</i> USA (19)	Single-center study using contrast-enhanced multi- detector CT for non-pancreatic indications; n=2,832	2.6%	Increasing prevalence associated with older age and Asian ethnicity
de Jong <i>et al.</i> Germany (17)	Single-center study using MRI for a preventive medical examination; n=2,803	2.4%	Increasing prevalence associated with older age
Lee <i>et al.</i> USA (20)	Single-center retrospective review of all abdominal MRI scans performed; n=616	13.5%	Increasing prevalence associated with older age
Chang <i>et al.</i> Korea (16)	Single-center study reviewing CT scans from health screening examinations; n=21,745	2.2%	Increasing prevalence associated with older age
Girometti <i>et al.</i> Italy (18)	Single-center study reviewing MRCP scans of patients with unsuspected/unknown pancreatic disease; n=152	44.7%	Prevalence correlated with age ≥60 years and history of hepatobiliary autoimmune disease

Table 2 Summary of prevalence data of PCL (16-20)

PCL, pancreatic cystic lesion; CT, computed tomography; MRI, magnetic resonance imaging; MRCP, magnetic resonance cholangiopancreatography.

IPMN; in seven surgical series of resected BD IPMN, invasive carcinoma and high-grade dysplasia were found in 31.1% of subjects, and among natural history studies, this rate was even lower (1.4–6.9%) (4,46-52). Treating physicians must be aware that pancreatic cancer can develop from the IPMN itself or from parenchyma not involved in the cystic lesion—a phenomenon referred to as a 'field defect' (53).

### **Detection and diagnosis of PCL**

Statement 4: In patients with pancreatic cystic lesions, newonset or worsening diabetes may be associated with underlying pancreatic adenocarcinoma

Guidelines for the diagnosis of PCL are available from various international groups, most notably, the International Association of Pancreatology (IAP), the American Gastroenterological Association (AGA), the American College of Gastroenterology (ACG), the ESGCTP, and the American College of Radiology (ACR) (3,4,54-56). Mass screening for PCL is not recommended (22,57), because even a highly specific test would likely be subject to a high rate of false positives due to the low incidence in the overall population.

Genetic risk factors for pancreatic cancer include mutations in genes such as *BRCA1*, *BRCA2*, *MLH1* and others, but these mutations are present in fewer than 10% of pancreatic cancer cases, and the most commonly found mutations (e.g., *BRCA1* and *BRCA2*) increase the risk of pancreatic ductal adenocarcinoma (PDAC) moderately, by around two- to six-fold (58). Reports suggest 2-4% of patients who are finally diagnosed with PDAC present with symptoms that mimic acute pancreatitis (59), and a systematic review and meta-analysis found the lifetime risk of pancreatic cancer in patients with chronic pancreatitis was elevated 16-fold versus those without chronic pancreatitis (60). However, the majority of patients who present with PCL do not have a history of pancreatitis (61). Patients with NODM have a 6- to 8-fold increase in risk of underlying PDAC, and among patients with NODM, 3-year pancreatic cancer incidence is ~1% (62). The incidence of PCL in patients with NODM is unclear, and models using clinical characteristics to stratify NODM patients according to their risk of pancreatic cancer are limited by their low predictive value (63). Several studies have evaluated prospective screening for pancreatic cancer in subjects with NODM (64-66), but results suggest further selection methods (e.g., biomarkers) need to be identified to improve the diagnostic yield (67). Other reported risk factors associated with increased risk of pancreatic cancer include low dietary intake of whole grains (68), higher prevalence of smoking, alcohol consumption, physical inactivity and obesity (69), and the presence of metabolic syndrome (70).

# Role of CT/MRI in the diagnosis and management of PCL

Statement 5: Contrast-enhanced computed tomography and magnetic resonance imaging are the imaging modalities of choice for diagnostic workup and surveillance of pancreatic cystic lesions (Level 5)

Statement 6: When available, a specific pancreatic protocol fulfilling minimal technical and reporting standards for structural cross-sectional imaging should be used in the workup and surveillance of pancreatic cystic lesions (Level 5)

The main radiographic modalities used to detect PCL are CT, MRI, and transabdominal ultrasound (71). The accuracy in identifying specific subtypes of PCL and differentiating malignant from benign lesions also varies with modality. MRI and CT have similar performance for distinguishing benign from malignant PCL and distinguishing between subtypes (72,73), but a comparison by Sainani and colleagues suggested MRI with MRCP may be more sensitive than CT for identifying a connection between PCL and the pancreatic duct as well as the presence of an enhancing mural nodule or internal septations, and for detecting multifocal disease (45,73). European guidelines recommend the use of CT for detection of parenchymal, mural or central calcification, and where assessment of vascular involvement is required (3). Transabdominal ultrasound imaging provides useful information on cyst site and size but expert opinion is that it is limited by operator-dependency and suboptimal visualization of the pancreas (74). Smaller PCL (<10 mm) can be difficult to characterize by cross-sectional imaging (75). Overall, MRI is the preferred choice, where available, because CT exposes patients to ionizing radiation, which is undesirable due to the associated risk of cancer (76), particularly in younger PCL patients who may need long-term surveillance requiring repeated imaging.

There is no universal standardized protocol for crosssectional imaging of the pancreas. Single-phase noncontrast CT scans or MRI alone have limited diagnostic value. Pancreas-specific protocols for CT imaging specify the use of intravenous contrast, multi-phase acquisition and thin slices (77,78) and, for MRI, imaging with either 1.5T or 3T is acceptable (79,80). Guidelines from the ACR recommend the inclusion of five elements in a radiology report: MD size, the presence of 'worrisome features' and/ or 'high-risk stigmata,' growth on the indexed lesion on serial imaging, and multiplicity of PCL (55).

# The role of endoscopic ultrasound in diagnosis and management of PCL

Statement 7: Endoscopic ultrasound with fine needle aspiration for cyst fluid analysis and tissue acquisition is useful to differentiate between neoplastic and non-neoplastic pancreatic cystic lesions

Statement 8: Endoscopic ultrasound should be considered in patients with suspected mucinous cystic neoplasms and intraductal papillary mucinous neoplasms on cross-sectional imaging with worrisome features Statement 9: Endoscopic ultrasound-guided fine needle aspiration for fluid analysis or tissue acquisition should be considered if the result would change management

The role of endoscopic ultrasound (EUS) is adjunctive to other imaging modalities used to make the initial diagnosis; it can detect features of concern, and EUS-guided fine needle aspiration (FNA) permits sampling of cystic fluid for biochemical and cyto-pathological evaluation (81). Furthermore, EUS can provide high-resolution images and information to guide subsequent management, such as lesion size, number and location, communication of lesion with MD, and presence of mural nodules. Detection of early signs of malignant transformation—e.g., nodules with vascular flow—is also possible with EUS techniques (81).

Although EUS is minimally invasive, guidelines from international expert groups limit the indication to higherrisk patients. In AGA guidance, EUS-FNA is indicated for patients with at least two of the following high-risk features: cyst size  $\geq$ 3 cm, MD dilation, or the presence of a mural nodule/solid component (56). The IAP guidelines indicate EUS if imaging shows 'worrisome features' (4), and ESGCTP recommends EUS should be performed if the PCL has clinical or radiological features of concern in the initial imaging (3).

Biomarker analysis of EUS-FNA samples can differentiate mucinous versus non-mucinous lesions, but EUS alone cannot reliably differentiate malignant from benign lesions (82). In Hong Kong, typical analyses from EUS-FNA samples may include carcinoembryonic antigen (CEA), glucose, amylase and cytology; DNA analysis is not routinely performed. The diagnostic utility of these markers is supported by numerous studies. For example, in a meta-analysis of 10 studies, the sensitivity and specificity of EUS-FNA cytology for diagnosing mucinous versus non-mucinous PCN were 42% and 99%, respectively (83). A meta-analysis of eight studies concluded intracyst glucose could differentiate mucinous and non-mucinous PCN with 91% sensitivity and 86% specificity (84). Furthermore, analysis of cytologic samples from EUS-FNA may assist diagnosis and management in some patients and reduce unnecessary surgeries (12). A retrospective analysis of 585 patients undergoing pancreatic resection concluded EUS with cytologic sampling improved the accuracy of diagnosis of patients with PCL (12). 'Through-the-needle' EUSguided biopsy in patients with PCL has also been shown to be feasible and clinically useful in a meta-analysis (85).

Although FNA can be performed with EUS, it should only be performed if the FNA or cyst fluid analysis will 
 Table 3 Worrisome features in patients with IPMN (3,4,54,56)

Clinical history

• NODM

History of pancreatitis

Radiological

- Mural nodule <5 mm</li>
- Cyst size ≥3 cm
- MD diameter 5–9 mm
- Rapidly increasing size of cyst (5 mm in 2 years)

Biochemical

• Elevated CA 19-9

IPMN, intraductal papillary mucinous neoplasm; NODM, newonset diabetes mellitus; MD, main duct; CA 19-9, cancer antigen 19-9.

change clinical management (3). In patients with crosssectional imaging or EUS showing obvious malignant transformation of the PCL indicative of surgical resection, EUS-FNA may not be necessary. Needle tract seeding—tumor cell implantation along the needle tract—has been reported as a very rare but serious complication of EUS-FNA (86). A meta-analysis of 10 studies (n=13,238) found a pooled rate of needle tract seeding of 0.3%, and the authors concluded that needle tract seeding is very unlikely to affect outcomes and should not be a reason to discourage EUS-FNA (86).

EUS-guided needle-based confocal laser endomicroscopy (EUS nCLE) is an emerging technique that enables realtime microscopic imaging during ultrasound-guided EUS-FNA (87). Several studies have demonstrated that EUS nCLE imaging is highly accurate and reliable for riskstratification of pancreatic cysts (88-90), and EUS nCLE may have better specificity and sensitivity for diagnosing highgrade dysplasia than current guideline algorithms (Fukuoka and AGA) (89). A 2022 consensus statement concluded that EUS nCLE could improve the diagnosis of PCL and noted that it should be systematically considered when EUS-FNA is indicated (87). Currently EUS nCLE is not widely used in Hong Kong and local experience is limited.

### **Recommendations for surveillance**

Statement 10: A bistory of pancreatitis, new onset of diabetes mellitus, mural nodule sized <5 mm, cyst size  $\geq$ 3 cm, main pancreatic duct diameter 5–9 mm, rapid increasing size of cyst (5 mm in 2 years) and elevated cancer antigen 19-9 level in patients with intraductal papillary mucinous neoplasms are worrisome features

Statement 11: Surveillance for neoplastic pancreatic cystic lesions should be lifelong, as long as the patient is fit for surgery

Statement 12: Patients with intraductal papillary mucinous neoplasms who are fit for surgery should receive regular followup with structural imaging to pick up malignant transformation changes

Diverse guidelines make references to 'worrisome features'-i.e., features associated with a higher risk of malignancy-when guiding management of patients with PCL. The definition of worrisome features in patients with IPMN defined in this publication (Table 3) is largely informed by those of the IAP, ESGCTP, AGA, ACG (3,4,54,56). Similar to the ESGCTP guidelines, we denote NODM and elevated cancer antigen 19-9 (CA 19-9) as worrisome features; however, for cyst size, the threshold is aligned with that of other international organizations ( $\geq 3 vs. \geq 4 cm$ for ESGCTP) (13). Cyst size  $\geq$ 30 mm is associated with a higher risk of high-grade dysplasia or malignancy (3), and data from multiple studies of IPMN patients show MD dilatation  $\geq 5$  mm is associated with a higher rate of malignancy or high-grade dysplasia (91-93). Several studies have found elevated serum CA 19-9 (>37 U/mL) to be associated with increased risk of invasive carcinoma or high-grade dysplasia (94-96).

Patients with PCL should be regularly monitored due to their increased risk of pancreatic cancer compared with the general population (97). The risk of malignant transformation because of PCL going undetected must be carefully balanced against the cost, inconvenience, and invasiveness of surveillance. Recommendations for duration and interval of monitoring of IPMN vary among expert groups, but in general, patients with larger cysts and higher-risk features should receive more intensive followup. Guidance from the AGA specifies, in the absence of concerning EUS-FNA findings, MRI surveillance at 1 year then every 2 years, with surveillance discontinued at 5 years or if the patient is no longer eligible for surgery (56). The IAP recommends lifelong surveillance with CT/MRCP every 2 years, and initial short-term follow-up of PCL, increasing in frequency with larger cyst size, with patients who have cysts >3 cm receiving 3-6 monthly follow-up with MRI alternating with EUS (4). Guidelines from the ESGCTP recommend surveillance with MRI or EUS 6-monthly for 1 year then annual (lifelong) monitoring if no risk factors are present (3). Our recommendation is that patients with IPMN or mucinous cystic neoplasm (MCN) with no indications for surgery or worrisome features

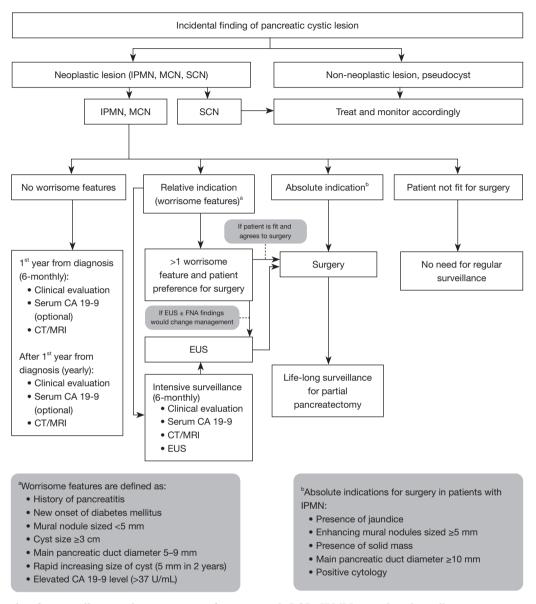


Figure 1 Algorithm for surveillance and management of patients with PCL. IPMN, intraductal papillary mucinous neoplasm; MCN, mucinous cystic neoplasm; SCN, serous cystic neoplasm; CA 19-9, cancer antigen 19-9; CT, computed tomography; MRI, magnetic resonance imaging; EUS, endoscopic ultrasound; FNA, fine needle aspiration; PCL, pancreatic cystic lesions.

receive monitoring (including clinical evaluation, MRI/CT imaging, and CA 19-9 tests) at 6–12 months, and 1-yearly intervals thereafter (*Figure 1*). Patients with worrisome features but no absolute indication for surgery should receive 6-monthly surveillance.

# Management and prognosis of PCL

Statement 13: Surgery may be offered to fit patients with

### symptomatic neoplastic pancreatic cystic lesions

Statement 14: Patients with main pancreatic duct type or mixed-type intraductal papillary mucinous neoplasms should be considered for surgery as there is a considerable risk of cancer formation

Statement 15: Presence of jaundice, enhancing mural nodules sized  $\geq 5$  mm, presence of solid mass, main pancreatic duct diameter  $\geq 10$  mm and positive cytology in patients with intraductal papillary mucinous neoplasms are absolute indications for surgery More than 90% of PCL may be classed as one of serous cystadenoma, IPMN (BD, MD or mixed type), or MCNs; all other types are rare, making up the remaining 10% (98-101). Serous cystic neoplasm has distinct morphological characteristics and, although considered benign, required resection in two-thirds of patients in a European study, even though the risk of transformation was <1% (102). Post-operative mortality among pancreatectomy patients has declined as techniques have matured. A US study found 30-day mortality rates halved from 6% in 1991 to 3% in 2005 (103). Another study of US patients who received pancreaticoduodenectomy (PDD) between 2014 and 2018 reported a mortality rate of 0.8% (104), and Hong Kong Hospital Authority data from 2018 to 2019 show a 30-day mortality rate of ~1% (105).

Indications for surgery vary among international guidelines; the absolute indications for surgery suggested here are aligned with ESGCTP guidelines: jaundice, enhancing mural nodules  $\geq 5$  mm, presence of solid mass, MD diameter  $\geq 10$  mm or positive cytology (3). Similar indications are endorsed by the 2017 IAP guideline (4). Patients with more than one worrisome feature may also be candidates for surgery, subject to EUS findings and patient preferences. Our recommendations for the surveillance and management of patients with PCL are summarized in *Figure 1*.

Enrollment in clinical trials is a potential option for patients unfit for surgery, and EUS-guided chemoablation or radiofrequency ablation are promising options in clinical development that may be suitable for selected patients unfit for pancreatectomy (106,107).

# Systemic management and adjuvant treatment for cancer formation

Statement 16: When a pancreatic cystic lesion manifests as malignant transformation, surgery is recommended for fit patients

Statement 17: Adjuvant chemotherapy is beneficial in the majority of patients with resectable pancreatic ductal cancer after malignant transformation of a pancreatic cystic lesion (Level 1)

Pancreatic cancer is extremely difficult to manage, resulting in high mortality; US data (2012–2018) show only 12% of patients have localized disease at diagnosis; most have regional (30%) or metastatic (52%) disease (108). Survival at 5 years ranges from 44% for those diagnosed with localized disease to 3% for those diagnosed with metastatic disease (108).

### Cheung et al. Management of PCL in Hong Kong

Numerous studies have shown beneficial effects of chemotherapy in patients with PDAC who have resected or metastatic disease (Table 4) (109-117). FOLFIRINOX (oxaliplatin + irinotecan + leucovorin + fluorouracil) or modified FOLFIRINOX regimens are considered to be the current standard of care for these patients (118). Adjuvant chemotherapy is administered to a broader population of patients with pancreatic cancer, compared with other cancer types, possibly due to its poor prognosis. However, chemotherapy is not suitable for all patients; the decision should be guided by disease stage and the patient's clinical status following surgery. Ideally, chemotherapy should be initiated within a few weeks of surgery, but should patients need a longer time to recover, chemotherapy initiation can be delayed by up to 12 weeks from surgery without a negative effect on outcomes, according to an analysis of patients with PDAC (119). Chemotherapy is beneficial in many patients with PDAC, but a review of 361 published cases with gastroenteropancreatic neuroendocrine neoplasms by Lania and colleagues concluded that the current evidence does not support its use in patients with pancreatic neuroendocrine tumors (120)

# Common adverse effects and management of patients following pancreatectomy

Statement 18: New-onset diabetes mellitus occurs in up to onequarter of patients after partial pancreatectomy (Level 2)

Statement 19: The incidence of pancreatic exocrine insufficiency following pancreaticoduodenectomy is higher than with distal pancreatectomy (Level 2)

Statement 20: After resection of intraductal papillary mucinous neoplasm, surveillance should be continued for as long as the patient remains fit for surgery (Level 5)

Incidence of NODM has been reported in patients after various forms of pancreatectomy. A study of 25 patients who received distal pancreatectomy (DP) and islet auto-transplantation found six patients (24%) had NODM at a median of 185 days (121). Among 31 patients with spleen-preserving DP, a post-operative incidence of NODM of 16% (5/31) was reported (122), and in a retrospective matched-pairs study, 14 of 50 patients developed NODM after DP (123). Lee *et al.* investigated 188 consecutive patients undergoing DP, or spleen-preserving DP; in this study, 20 (11%) patients developed NODM (124). Comparable rates have been seen in larger studies. A US 2022 database study identified 311 nondiabetic patients who underwent pancreatectomy and

Study	Setting	Comparators (n) and OS	
ESPAC-4 (113)	Resected pancreatic cancer	Gemcitabine (n=366): 25.5 months	
	(adjuvant)	Gemcitabine + capecitabine (n=364): 28.0 months	
		HR: 0.82 (95% CI: 0.68–0.98; P=0.032)	
PRODIGE 24–ACCORD (111)	Resected pancreatic cancer (adjuvant)	mFOLFIRINOX (n=247): 54.4 months	
		Gemcitabine (n=246): 35.0 months	
		HR: 0.64 (95% CI: 0.48–0.86; P=0.003)	
	1 Metastatic pancreatic cancer	FOLFIRINOX (n=171): 11.1 months	
(110)		Gemcitabine (n=171): 6.8 months	
		HR: 0.57 (95% CI: 0.45–0.73; P<0.001)	
APACT (109,115)	Resected pancreatic cancer (adjuvant)	nab-paclitaxel + gemcitabine (n=432): 40.5 months	
		Gemcitabine (n=434): 36.2 months	
		HR: 0.82 (95% CI: 0.680–0.996; P=0.045)	
MPACT (117)	Metastatic pancreatic cancer	nab-paclitaxel + gemcitabine (n=431): 8.5 months	
		Gemcitabine (n=430): 6.7 months	
		HR: 0.72 (95% CI: 0.62–0.83; P<0.001)	
PREOPANC (116)	Resectable/borderline resectable	Preoperative gemcitabine + RT + adjuvant gemcitabine (n=119): 16.0 months	
	pancreatic cancer (perioperative)	Adjuvant gemcitabine: 14.53 months	
		HR: 0.78 (95% CI: 0.58–1.05; P=0.096)	
Alliance A021501 (112)	Resectable/borderline resectable	Neoadjuvant mFOLFIRINOX (n=70): 31.0 months	
	pancreatic cancer (neoadjuvant)	Neoadjuvant mFOLFIRINOX + RT (n=56): 17.1 months	
SWOG-s1505 (114)	Resectable pancreatic cancer	Perioperative mFOLFIRINOX (n=55): 23.2 months	
	(perioperative)	Perioperative gemcitabine + nab-paclitaxel (n=47): 23.6 months	

Table 4 Summary of overall survival in studies of chemotherapy in patients with PDAC (109-117)

PDAC, pancreatic ductal adenocarcinoma; OS, overall survival; HR, hazard ratio; CI, confidence interval; mFOLFIRINOX, modified FOLFIRINOX; nab-paclitaxel, albumin-bound paclitaxel; FOLFIRINOX, oxaliplatin + irinotecan + leucovorin + fluorouracil; RT, radiotherapy.

reported a NODM incidence of 20.2% at 24 months (125). Multivariable analysis revealed older age, obesity, hypertension and cardiovascular (CV) disease to be independent predictors of NODM (125). A meta-analysis of 476 patients who underwent DP for benign or potentially malignant lesions reported a  $\geq 6$  months incidence of NODM of 14% (126). Furthermore, a meta-analysis including 1,121 patients who underwent pancreatoduodenectomy reported a mean weighted overall proportion with NODM of 16% (127). Some studies have suggested a higher incidence of NODM in patients undergoing DP than with central pancreatectomy (CP) (128-130), but a matched-pairs analysis of 100 patients who underwent DP or CP did not find a significant difference in rates of NODM between

groups (123).

The incidence of PEI reported following pancreatectomy varies considerably due to different definitions and study types, but data suggest a higher incidence of PEI after PDD compared with DP. These include a systematic review of nine studies (n=673) that reported pre-operative incidences of PEI in 44% of patients undergoing PDD and 20% prior to DP, and post-operative (6-month) rates of 74% (range, 36–100%) and 67–80%, respectively (131). Another review has reported a general trend of higher rates of PEI after PDD than after DP, reporting an incidence of 35–100% following PDD, 19–80% following DP, and 12% following CP (132). Risk factors for PEI include Caucasian race, lower body mass index (BMI), family history of diabetes

Study	Patient population	Recurrence rates
He et al. (137)	N=130, non-invasive IPMN	Any new IPMN:
		1-year: 4%
		5-year: 25%
		10-year: 62%
		IPMN requiring surgery:
		1-year: 1.6%
		5-year: 14%
		10-year: 18%
		Invasive IPMN:
		1-year: 0%
		5-year: 7%
		10-year: 38%
Kang <i>et al.</i> (136)	N=298, non-invasive IPMN	Non-invasive IPMN: 2.0% (median follow-up 44.4 months)
		Invasive IPMN: 3.4% (median follow-up 44.4 months)
Marchegiani <i>et al.</i> (135)	N=299, non-invasive IPMN	Non-invasive IPMN: 9.3% (median follow-up 58 months)
		Invasive IPMN: 2% (median follow-up 58 months)

Table 5 Rates of postoperative recurrence in patients with IPMN (135-137)

IPMN, intraductal papillary mucinous neoplasm.

mellitus, steatorrhea, elevated pre-operative bilirubin, ductobstructive pancreatic pathology, and a history of acute pancreatitis (133,134).

Pancreatic cysts arise due to a 'field effect' in the remnant pancreas tissue that predisposes patients to recurrence of IPMN and new-onset PDAC (53); therefore, long-term postoperative surveillance is essential. Several large studies have provided data on postoperative recurrence rates of IPMN in large populations (*Table 5*) (135-137). A study of 195 patients who underwent pancreatectomy for IPMN reported cumulative 5- and 10-year incidence rates of PDAC of 4.5% and 5.9%, respectively (138). Predictors of recurrent IPMN and PDAC include high-grade dysplasia in resected specimens, margin-positive resection, family history of PDAC, and gastric and pancreatobiliary subtypes of IPMN (137,138).

Because the risk of progression of IPMN does not decrease over time after resection (137), surveillance should continue, providing the patient is fit for surgery. We suggest surveillance includes cross-sectional imaging every 6–12 months, with 6-monthly imaging recommended for high-risk groups (e.g., family history of PDAC, surgical margin positive for high-grade dysplasia and non-intestinal subtype of IPMN). Invasive IPMN should receive the same follow-up as PDAC.

### **Diagnosis of pancreatic exocrine insufficiency**

Statement 21: For patients with pancreatic cystic lesions, postpancreatectomy pancreatic exocrine insufficiency diagnosis should be based on suggestive clinical and laboratory findings

PEI occurs when secretion of pancreatic enzymes in the intestinal lumen is below the threshold level required for normal digestion, leading to impaired absorption of essential nutrients, including fat, liposoluble vitamins and antioxidants, and severe maldigestion (139,140). Symptoms of PEI vary with severity but may include diarrhea, abdominal pain or functional bowel disorders in undiagnosed patients, and steatorrhea, flatulence, weight loss and deficiencies in liposoluble vitamins such as vitamin D and other nutrients (139,140). These symptoms decrease patients' quality of life and may lead to CV events and malnutrition-related complications (141-143). A notable consequence of PEI is osteoporosis, with a meta-analysis

of 513 patients with acute pancreatitis (most of whom had PEI) reporting a pooled prevalence rate of osteoporosis and osteopenia of 65% (144).

Conditions with a high prevalence of PEI include chronic pancreatitis, pancreatic cancer and pancreatic surgery (145), and pathogenesis of PEI may involve insufficient stimulation secretion, reduced pancreatic function or enzyme production due to chronic pancreatitis or pancreatectomy, and obstruction of pancreatic ducts (139). Asynchrony of GI secretions after pancreatobiliary or GI surgery may also contribute to PEI (146).

Although the optimal method of PEI diagnosis is not defined, diagnosis is usually based on patient-reported changes in bowel function, weight loss, and other patient characteristics. Symptoms alone may lead to under- or over-diagnosis (139). Serum nutritional markers may be of assistance (140), but physicians need to investigate other causes of deficiencies. Imaging may identify structural causes (139), but PEI can occur in patients with a morphologically normal pancreas.

Direct functional tests of the pancreas involve the collection of stimulated pancreatic secretions, namely the secretin-cholecystokinin stimulation test, or an endoscopic pancreatic function test (139,146), but these tests are invasive and costly, limiting their clinical use. Indirect functional tests on blood, fecal or breath samples are cheaper and simpler but are less sensitive and specific (145). Coefficient of fat absorption measured by the 72-hour fecal fat test is considered to be the 'gold standard' test for fat malabsorption but requires the patient to follow a standardized diet for 5 days prior and a 3-day hospital stay (147,148). Compliance therefore tends to be poor. A more convenient alternative is the fecal elastase-1 (FE-1) test. FE-1 is an exocrine-specific pancreatic enzyme reflecting pancreatic exocrine function and is not degraded in the bowel lumen (7); FE-1 can be measured using an enzymelinked immunosorbent assay (149). A system for staging PEI as 'mild', 'moderate', or 'severe' based on FE-1 levels, coefficient of fat absorption and other patient characteristics has been proposed (150), but these definitions are arbitrary. A meta-analysis found FE-1 tests had a sensitivity of 77% and a specificity of 88% versus direct pancreatic function tests (149). In Hong Kong, FE-1 testing is not widely available and therefore is rarely used. The <sup>13</sup>C-mixed triglyceride breath test (13C-MTBT) measures pancreatic

function and digestion using a triglyceride substrate that uses carbon dioxide as a metabolite (140). Sensitivity is high, but the test is not routinely available in Hong Kong. In Hong Kong, the usual clinical practice is to diagnose postpancreatectomy PEI based on patient-reported symptoms and initiate PERT. Imaging and laboratory tests such as FE-1 may be used in the follow-up of patients who do not respond to initial treatment.

### **Management of PEI**

Statement 22: Pancreatic enzyme replacement therapy should be given in symptomatic patients with pancreatic exocrine insufficiency (Level 5)

Statement 23: In patients with suboptimal response after pancreatic enzyme replacement therapy, consider increasing the dose or adding a proton pump inhibitor (Level 5)

Statement 24: Monitoring of the nutritional status of the patients after pancreatic enzyme replacement therapy should be based on clinical parameters, with blood tests as an adjunct (Level 5)

Patients diagnosed with suspected PEI are advised to avoid tobacco and alcohol use as these are risk factors for pancreatitis (151-153). The Pancreatic Exocrine Insufficiency Questionnaire (PEI-Q) is an 18-item patient questionnaire developed by European clinicians (with support from Abbott) to assist diagnosis and monitoring of patients with PEI (154,155). The PEI-Q calculates a score based on three domains: abdominal symptoms, bowel movements, and impact on patients' quality of life, and the reliability of this instrument has been demonstrated in a validation study of 162 European patients with PEI (155). The PEI-Q can provide useful insights to a clinician treating patients with PEI and may inform better decision making (155), and a certified Chinese translation of the PEI-Q is available.\*

The mainstay of therapy in patients with PEI is PERT, formulated as pH-sensitive, enteric-coated minimicrospheres of lipase, protease, and amylase that protect enzymes from gastric acidity and allow them to disintegrate rapidly at pH 5.5 in the duodenum (146). The efficacy of PERT has been demonstrated in several randomized studies in patients with PEI from chronic pancreatitis and pancreatic surgery (156-159).

Patients need to be instructed on the correct use of PERT for it to be effective, most importantly taking

<sup>\*</sup> The certified Chinese translation of the PEI-Q is available upon request from Abbott Laboratories Ltd., Hong Kong.

capsules with meals and snacks, and spacing multiple doses throughout a meal. European guidelines recommend a lipase dose of 40,000-50,000 Pharmacopeia units (PhU) per meal-which is approximately 10% of the physiologically secreted dose of post-meal lipase in the duodenumand half this dose for snacks (151,160). A minimum of 30,000 PhU lipase is suggested per meal (~3 capsules per meal) (151). The UK consensus guidelines recommend a minimum starting dose of 50,000 PhU lipase for main meals and 25,000 for snacks (7). Australasian guidelines recommend 25,000-40,000 PhU lipase to be taken with food and this should be individualized based on bodyweight and titrated based on weight gain and bowel symptoms (8). Studies of PERT dosing specific to Asian populations are not available, but based on local clinical experience, the lower bodyweight of Asian patients, and lower dietary fat content versus Western populations, a smaller starting dose may be appropriate. The typical approach in Hong Kong is to start patients on low doses (e.g., 20,000-30,000 PhU per meal) and step up dosage until symptoms resolve.

Efficacy of PERT can be assessed by resolution of maldigestion symptoms and in non-responders, the use of pancreatic function tests may prove valuable (fat absorption, <sup>13</sup>C-MTBT). In patients with poor response, increases of PERT dose or the addition of proton pump inhibitors (PPIs) to reduce gastric acid and thereby decrease degradation of enzymes should be considered. It is important that nutritional deficiencies are resolved for treatment to be considered a success. Clinical evaluation for malnutrition, including screening patients for deficiencies of calcium, zinc, and liposoluble vitamins, should therefore be considered.

Our suggested approach to initiating PERT and dose adjustment is summarized in Figure 2. Briefly, PERT should be initiated at 20,000-30,000 PhU/meal in postpancreatectomy patients with GI symptoms, although considerably higher doses may be needed depending on the severity of a patient's symptoms and composition of meals. The dose should be increased in one-capsule (10,000 PhU) increments until symptoms resolve. Assessment of symptoms with an instrument such as the PEI-Q is suggested at diagnosis, and during follow-up, to evaluate response to therapy. Patients should be clinically evaluated for nutritional deficiencies, with blood tests for nutritional deficiencies suggested for patients with poor response to therapy. Compliance should be checked, and addition of a PPI is suggested for patients with poor response. Maximum doses in post-pancreatectomy patients have

not been defined, but in cystic fibrosis, a daily maximum dose of 10,000 lipase units per kg bodyweight is recommended (161).

# Management of diabetes in post-pancreatectomy patients

Statement 25: Clinicians should screen for diabetes following partial pancreatectomy, and subsequent management should follow standard of care (Level 5)

Statement 26: Multiple daily injection or continuous subcutaneous insulin infusion are the mainstays of insulin replacement therapy after total pancreatectomy (Level 2)

The management of diabetes arising from pancreatic diseases, referred to as 'type 3c diabetes' in some literature, is mostly adapted from recommendations for type 1 diabetes, as there are few studies specific to this population to guide treatment (9). Following pancreatectomy, low levels of insulin, glucagon, and other pancreatic polypeptides contribute to rapid fluctuations in glucose levels, sometimes described as 'brittle diabetes' (162). However, studies suggest total daily insulin and basal insulin requirements (excluding prandial insulin) are significantly lower in patients who have undergone total pancreatectomy than in type 1 diabetes (162-164). Therefore, patients and their physicians should be aware that there may be an increased risk of hypoglycemia in this subtype of diabetes compared with other diabetes types (165).

Metabolic outcomes after total pancreatectomy were evaluated in a case series including 141 patients who received pancreatectomy between 1985 and 2006 (166). When surveyed in 2007, responses from 47 patients showed a mean glycosylated hemoglobin (HbA<sub>1c</sub>) of 7.5%, with 89% of patients on a complex insulin regimen (≥3 insulin doses per day) (166). Hypoglycemia was experienced by 37 (79%) patients and severe hypoglycemia by 15 (41%) patients (166). A literature review of studies of perioperative management of endocrine insufficiency after total pancreatectomy found that ~80% of patients develop hypoglycemia episodes, and 40% develop severe hypoglycemia, leading to mortality in 0-8% of cases and morbidity in 25-45% of cases (162). These episodes can be reduced with patient education by nutritionists and endocrinologists before surgery, and re-evaluation to ensure the patient has the appropriate understanding, support, and resources.

Data on the prevalence of NODM following pancreatectomy are available from numerous studies. A retrospective cohort

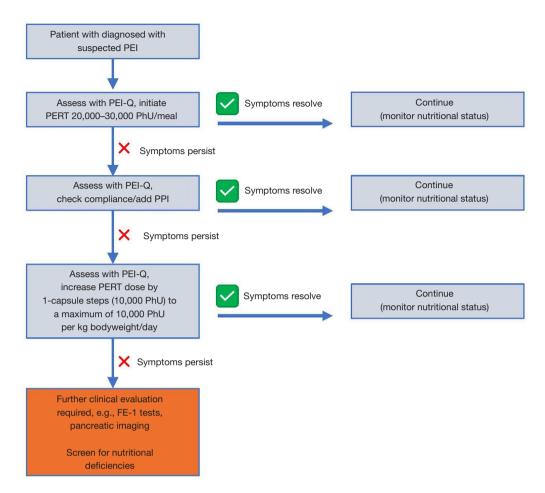


Figure 2 Dosing and monitoring of PERT in patients with PEI after pancreatectomy. PEI, pancreatic exocrine insufficiency; PEI-Q, Pancreatic Exocrine Insufficiency Questionnaire; PPI, proton pump inhibitor; PERT, pancreatic enzyme replacement therapy; PhU, pharmacopeia units; FE-1, fecal elastase 1.

study of 1,717 patients after pancreatectomy (median followup 18 months) found 20% had postoperative endocrine insufficiency, requiring introduction or escalation of pharmacologic intervention; NODM was reported in 217 (12.6%) patients-62.7% of whom needed insulin (134). Risk factors for diabetes in this population included male gender, increased BMI, tobacco use, family/personal history of diabetes, and PDD (134). Longer-term data are available from a study of 80 patients with median follow-up of 9.5 years (167). In this population, 12.5% had diabetes mellitus before surgery and 28.6% had NODM after surgery (21.9% after excluding patients with total pancreatectomy) (167). Of the 30 patients with diabetes mellitus, 22 (73.3%) needed insulin and 12 manifested microvascular complications (167). Predictors of postpancreatectomy NODM have been evaluated in a

retrospective cohort study of Japanese patients, 18.4% (125/681) of whom had NODM at 1–12 months (168). Predictors of NODM included BMI, HbA<sub>1c</sub> prior to surgery, blood glucose level, and indication for surgery (168). A systematic review based on 36 articles assessed the literature for type 3c diabetes, including patients with PDD (n=5,636), DP (n=3,922), and CP (n=315) (6). Rates of NODM (median onset 3–15 months) were 9–24% after PDD, 3–40% after DP, and 0–14% after CP, and surgical site, higher preoperative HbA<sub>1c</sub>, fasting plasma glucose and larger pancreatic resection volume had the strongest associations (6).

The optimal form of insulin replacement following pancreatectomy is not well defined, with data limited to small case series and observational studies (162,169,170). Consistent improvements in  $HbA_{1c}$  levels from continuous

subcutaneous insulin infusion (CSII) versus multiple daily injection (MDI) insulin have not been reported, although the former may be associated with lower rates of hypoglycemia. For example, a study that compared CSII with MDI insulin in 39 patients following total pancreatectomy reported no significant differences in median HbA<sub>1c</sub> between groups (7.3% vs. 8.1%; P=0.16), but severe hypoglycemia rates were lower among patients receiving CSII compared with MDI (17% vs. 52%; P=0.02) (170). Despite the higher rate of severe hypoglycemia, no significant differences in quality of life were reported between groups (170).

In accordance with US guidance, we suggest that, following pancreatectomy for PCL, patients should be screened for diabetes using the criteria of fasting plasma glucose (FPG)  $\geq$ 126 mg/dL (7.0 mmol/L),  $\geq$ HbA<sub>1c</sub> 6.5% (48 mmol/moL) or 2-hour plasma glucose  $\geq$ 200 mg/dL (11.1 mmol/L) following a 75 g oral glucose tolerance test (9), and insulin replacement should be initiated where indicated.

# Conclusions

PCL are becoming increasingly common in Hong Kong, because of its aging population, and have the potential to give rise to pancreatic cancer, a disease with few treatment options and poor prognosis (3,19,57). We recommend patients with suspected PCL in incidental imaging receive careful follow-up, and those with PCL should undergo pancreas-specific imaging, including EUS if indicated. EUS combined with FNA has a central role in differentiating neoplastic and non-neoplastic PCL and detecting highrisk features. Patients with higher-risk features should be monitored more intensively than those without. Where indicated, patients with symptomatic neoplastic PCL who are fit for surgery should undergo pancreatectomy, and following pancreatectomy, patients should be monitored for symptoms of PEI. Treatment with PERT should be initiated for patients who report GI symptoms consistent with PEI, and patients should be monitored for resolution of symptoms. Increased doses of PERT and addition of PPIs should be considered for patients who do not respond to initial therapy, and pancreatic imaging and further clinical investigation may be needed to evaluate patients who continue to respond poorly. The nutritional status of patients on PERT should be monitored, and serum tests for nutritional deficiencies should be considered, especially in patients with prolonged poor response. Following pancreatectomy, patients should also be screened for postoperative diabetes, and insulin should be initiated if needed. Physicians should be aware that insulin-treated patients with type 3c diabetes tend to have lower insulin requirements and a higher risk of hypoglycemia than patients with type 1 diabetes mellitus. Topics that may be of interest for future research include PERT dose optimization in East Asian patients and optimizing the safety and efficacy of insulin therapy in patients with type 3c diabetes.

With these consensus statements, we have aimed to capture the contemporary approach to diagnosis and management of PCL and PEI in Hong Kong, and we hope this document serves as a useful guide to clinicians treating these diseases both in Hong Kong and abroad.

## **Acknowledgments**

The authors would like to thank Dr Alister Smith and Isabelle Roper (MIMS Hong Kong) for assistance with the consensus meetings, medical writing, and project management of the manuscript submission.

*Funding:* The consensus meetings to develop these statements, medical writing, and project management of the manuscript were supported by an independent educational grant from Abbott Laboratories Ltd., Hong Kong.

### Footnote

*Reporting Checklist:* The authors have completed the CREDES reporting checklist. Available at: https://hbsn.amegroups.com/article/view/10.21037/hbsn-22-471/rc

*Conflicts of Interest:* All authors have completed the ICMJE uniform disclosure form (available at https://hbsn.amegroups.com/article/view/10.21037/hbsn-22-471/coif). TTC, TY, and KSHC have received an advisory board honorarium from Abbott. WKS has received speaker's fees from Mylan, AbbVie, AstraZeneca and Gilead, an advisory board honorarium from Abbott, research funding from Gilead, and he is an advisory board member for Gilead, AbbVie, and Abbott. YTL has received an advisory board honorarium from Abbott, honoraria for lectures from Cook Medical, and is President of the Hong Kong Society of Endosonography. The other authors have no conflicts of interest to declare.

*Ethical Statement:* The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

*Open Access Statement:* This is an Open Access article distributed in accordance with the Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International License (CC BY-NC-ND 4.0), which permits the non-commercial replication and distribution of the article with the strict proviso that no changes or edits are made and the original work is properly cited (including links to both the formal publication through the relevant DOI and the license). See: https://creativecommons.org/licenses/by-nc-nd/4.0/.

# References

- Ferrone CR, Correa-Gallego C, Warshaw AL, et al. Current trends in pancreatic cystic neoplasms. Arch Surg 2009;144:448-54.
- Siegel R, Naishadham D, Jemal A. Cancer statistics, 2012. CA Cancer J Clin 2012;62:10-29.
- European Study Group on Cystic Tumours of the Pancreas. European evidence-based guidelines on pancreatic cystic neoplasms. Gut 2018;67:789-804.
- Tanaka M, Fernández-Del Castillo C, Kamisawa T, et al. Revisions of international consensus Fukuoka guidelines for the management of IPMN of the pancreas. Pancreatology 2017;17:738-53.
- 5. Phillips ME. Pancreatic exocrine insufficiency following pancreatic resection. Pancreatology 2015;15:449-55.
- Wu L, Nahm CB, Jamieson NB, et al. Risk factors for development of diabetes mellitus (Type 3c) after partial pancreatectomy: A systematic review. Clin Endocrinol (Oxf) 2020;92:396-406.
- Phillips ME, Hopper AD, Leeds JS, et al. Consensus for the management of pancreatic exocrine insufficiency: UK practical guidelines. BMJ Open Gastroenterol 2021;8:e000643.
- 8. Working Party of the Australasian Pancreatic Club; Smith RC, Smith SF, et al. Summary and recommendations from the Australasian guidelines for the management of pancreatic exocrine insufficiency. Pancreatology 2016;16:164-80.
- American Diabetes Association Professional Practice Committee. 2. Classification and Diagnosis of Diabetes: Standards of Medical Care in Diabetes—2022. Diabetes Care 2022;45:S17-38.
- Keane MG, Afghani E. A Review of the Diagnosis and Management of Premalignant Pancreatic Cystic Lesions. J Clin Med 2021;10:1284.
- Hong Kong Cancer Registry. Leading Cancer Sites in Hong Kong in 2019. Available online: https://www3. ha.org.hk/cancereg/pdf/top10/rank\_2019.pdf

- Giannone F, Crippa S, Aleotti F, et al. Improving diagnostic accuracy and appropriate indications for surgery in pancreatic cystic neoplasms: the role of EUS. Gastrointest Endosc 2022;96:648-56.e2.
- Hasan A, Visrodia K, Farrell JJ, et al. Overview and comparison of guidelines for management of pancreatic cystic neoplasms. World J Gastroenterol 2019;25:4405-13.
- Oxford Centre for Evidence-Based Medicine. Oxford Centre for Evidence-Based Medicine 2011 Levels of Evidence. Available online: https://www.cebm.net/wpcontent/uploads/2014/06/CEBM-Levels-of-Evidence-2.1.pdf
- Jünger S, Payne SA, Brine J, et al. Guidance on Conducting and REporting DElphi Studies (CREDES) in palliative care: Recommendations based on a methodological systematic review. Palliat Med 2017;31:684-706.
- Chang YR, Park JK, Jang JY, et al. Incidental pancreatic cystic neoplasms in an asymptomatic healthy population of 21,745 individuals: Large-scale, single-center cohort study. Medicine (Baltimore) 2016;95:e5535.
- de Jong K, Nio CY, Hermans JJ, et al. High prevalence of pancreatic cysts detected by screening magnetic resonance imaging examinations. Clin Gastroenterol Hepatol 2010;8:806-11.
- Girometti R, Intini S, Brondani G, et al. Incidental pancreatic cysts on 3D turbo spin echo magnetic resonance cholangiopancreatography: prevalence and relation with clinical and imaging features. Abdom Imaging 2011;36:196-205.
- Laffan TA, Horton KM, Klein AP, et al. Prevalence of unsuspected pancreatic cysts on MDCT. AJR Am J Roentgenol 2008;191:802-7.
- Lee KS, Sekhar A, Rofsky NM, et al. Prevalence of incidental pancreatic cysts in the adult population on MR imaging. Am J Gastroenterol 2010;105:2079-84.
- 21. Census and Statistics Department Hong Kong Special Administrative Region. 2021 Population Census – Summary Results. Available online: https://www.censtatd.gov.hk/en/EIndexbySubject. html?scode=600&pcode=B1120106
- 22. Kromrey ML, Bülow R, Hübner J, et al. Prospective study on the incidence, prevalence and 5-year pancreatic-related mortality of pancreatic cysts in a population-based study. Gut 2018;67:138-45.
- Tanaka M, Fernández-del Castillo C, Adsay V, et al. International consensus guidelines 2012 for the management of IPMN and MCN of the pancreas. Pancreatology 2012;12:183-97.

- 24. Bournet B, Kirzin S, Carrère N, et al. Clinical fate of branch duct and mixed forms of intraductal papillary mucinous neoplasia of the pancreas. J Gastroenterol Hepatol 2009;24:1211-7.
- 25. Crippa S, Fernández-Del Castillo C, Salvia R, et al. Mucin-producing neoplasms of the pancreas: an analysis of distinguishing clinical and epidemiologic characteristics. Clin Gastroenterol Hepatol 2010;8:213-9.
- 26. Hwang DW, Jang JY, Lee SE, et al. Clinicopathologic analysis of surgically proven intraductal papillary mucinous neoplasms of the pancreas in SNUH: a 15-year experience at a single academic institution. Langenbecks Arch Surg 2012;397:93-102.
- Jang JY, Kim SW, Lee SE, et al. Treatment guidelines for branch duct type intraductal papillary mucinous neoplasms of the pancreas: when can we operate or observe? Ann Surg Oncol 2008;15:199-205.
- 28. Kanno A, Satoh K, Hirota M, et al. Prediction of invasive carcinoma in branch type intraductal papillary mucinous neoplasms of the pancreas. J Gastroenterol 2010;45:952-9.
- Kawamoto S, Lawler LP, Horton KM, et al. MDCT of intraductal papillary mucinous neoplasm of the pancreas: evaluation of features predictive of invasive carcinoma. AJR Am J Roentgenol 2006;186:687-95.
- 30. Kim SC, Park KT, Lee YJ, et al. Intraductal papillary mucinous neoplasm of the pancreas: clinical characteristics and treatment outcomes of 118 consecutive patients from a single center. J Hepatobiliary Pancreat Surg 2008;15:183-8.
- Lee SY, Lee KT, Lee JK, et al. Long-term follow up results of intraductal papillary mucinous tumors of pancreas. J Gastroenterol Hepatol 2005;20:1379-84.
- 32. Mimura T, Masuda A, Matsumoto I, et al. Predictors of malignant intraductal papillary mucinous neoplasm of the pancreas. J Clin Gastroenterol 2010;44:e224-9.
- 33. Nagai K, Doi R, Kida A, et al. Intraductal papillary mucinous neoplasms of the pancreas: clinicopathologic characteristics and long-term follow-up after resection. World J Surg 2008;32:271-8; discussion 279-80.
- 34. Nara S, Onaya H, Hiraoka N, et al. Preoperative evaluation of invasive and noninvasive intraductal papillary-mucinous neoplasms of the pancreas: clinical, radiological, and pathological analysis of 123 cases. Pancreas 2009;38:8-16.
- 35. Ohno E, Hirooka Y, Itoh A, et al. Intraductal papillary mucinous neoplasms of the pancreas: differentiation of malignant and benign tumors by endoscopic ultrasound findings of mural nodules. Ann Surg 2009;249:628-34.
- 36. Rodriguez JR, Salvia R, Crippa S, et al. Branch-duct

intraductal papillary mucinous neoplasms: observations in 145 patients who underwent resection. Gastroenterology 2007;133:72-9; quiz 309-10.

- 37. Sadakari Y, Ienaga J, Kobayashi K, et al. Cyst size indicates malignant transformation in branch duct intraductal papillary mucinous neoplasm of the pancreas without mural nodules. Pancreas 2010;39:232-6.
- Salvia R, Fernández-del Castillo C, Bassi C, et al. Mainduct intraductal papillary mucinous neoplasms of the pancreas: clinical predictors of malignancy and long-term survival following resection. Ann Surg 2004;239:678-85; discussion 685-7.
- Schmidt CM, White PB, Waters JA, et al. Intraductal papillary mucinous neoplasms: predictors of malignant and invasive pathology. Ann Surg 2007;246:644-51; discussion 651-4.
- Schnelldorfer T, Sarr MG, Nagorney DM, et al. Experience with 208 resections for intraductal papillary mucinous neoplasm of the pancreas. Arch Surg 2008;143:639-46; discussion 646.
- Serikawa M, Sasaki T, Fujimoto Y, et al. Management of intraductal papillary-mucinous neoplasm of the pancreas: treatment strategy based on morphologic classification. J Clin Gastroenterol 2006;40:856-62.
- 42. Sohn TA, Yeo CJ, Cameron JL, et al. Intraductal papillary mucinous neoplasms of the pancreas: an updated experience. Ann Surg 2004;239:788-97; discussion 797-9.
- 43. Sugiyama M, Izumisato Y, Abe N, et al. Predictive factors for malignancy in intraductal papillary-mucinous tumours of the pancreas. Br J Surg 2003;90:1244-9.
- 44. Suzuki Y, Atomi Y, Sugiyama M, et al. Cystic neoplasm of the pancreas: a Japanese multiinstitutional study of intraductal papillary mucinous tumor and mucinous cystic tumor. Pancreas 2004;28:241-6.
- 45. Waters JA, Schmidt CM, Pinchot JW, et al. CT vs MRCP: optimal classification of IPMN type and extent. J Gastrointest Surg 2008;12:101-9.
- 46. Aso T, Ohtsuka T, Matsunaga T, et al. "High-risk stigmata" of the 2012 international consensus guidelines correlate with the malignant grade of branch duct intraductal papillary mucinous neoplasms of the pancreas. Pancreas 2014;43:1239-43.
- Fritz S, Klauss M, Bergmann F, et al. Pancreatic main-duct involvement in branch-duct IPMNs: an underestimated risk. Ann Surg 2014;260:848-55; discussion 855-6.
- 48. Goh BK, Thng CH, Tan DM, et al. Evaluation of the Sendai and 2012 International Consensus Guidelines based on cross-sectional imaging findings performed for the initial triage of mucinous cystic lesions of the pancreas:

731

a single institution experience with 114 surgically treated patients. Am J Surg 2014;208:202-9.

- Jang JY, Park T, Lee S, et al. Validation of international consensus guidelines for the resection of branch ducttype intraductal papillary mucinous neoplasms. Br J Surg 2014;101:686-92.
- Nguyen AH, Toste PA, Farrell JJ, et al. Current recommendations for surveillance and surgery of intraductal papillary mucinous neoplasms may overlook some patients with cancer. J Gastrointest Surg 2015;19:258-65.
- 51. Roch AM, Ceppa EP, DeWitt JM, et al. International Consensus Guidelines parameters for the prediction of malignancy in intraductal papillary mucinous neoplasm are not properly weighted and are not cumulative. HPB (Oxford) 2014;16:929-35.
- 52. Sahora K, Mino-Kenudson M, Brugge W, et al. Branch duct intraductal papillary mucinous neoplasms: does cyst size change the tip of the scale? A critical analysis of the revised international consensus guidelines in a large singleinstitutional series. Ann Surg 2013;258:466-75.
- 53. Uehara H, Nakaizumi A, Ishikawa O, et al. Development of ductal carcinoma of the pancreas during follow-up of branch duct intraductal papillary mucinous neoplasm of the pancreas. Gut 2008;57:1561-5.
- Elta GH, Enestvedt BK, Sauer BG, et al. ACG Clinical Guideline: Diagnosis and Management of Pancreatic Cysts. Am J Gastroenterol 2018;113:464-79.
- 55. Megibow AJ, Baker ME, Morgan DE, et al. Management of Incidental Pancreatic Cysts: A White Paper of the ACR Incidental Findings Committee. J Am Coll Radiol 2017;14:911-23.
- 56. Vege SS, Ziring B, Jain R, et al. American Gastroenterological Association Institute guideline on the diagnosis and management of asymptomatic neoplastic pancreatic cysts. Gastroenterology 2015;148:819-22; quize12-3.
- US Preventive Services Task Force; Owens DK, Davidson KW, et al. Screening for Pancreatic Cancer: US Preventive Services Task Force Reaffirmation Recommendation Statement. JAMA 2019;322:438-44.
- Hu C, Hart SN, Polley EC, et al. Association Between Inherited Germline Mutations in Cancer Predisposition Genes and Risk of Pancreatic Cancer. JAMA 2018;319:2401-9.
- Munigala S, Kanwal F, Xian H, et al. Increased risk of pancreatic adenocarcinoma after acute pancreatitis. Clin Gastroenterol Hepatol 2014;12:1143-50.e1.
- 60. Kirkegård J, Mortensen FV, Cronin-Fenton D. Chronic Pancreatitis and Pancreatic Cancer Risk: A Systematic

Review and Meta-analysis. Am J Gastroenterol 2017;112:1366-72.

- 61. Spinelli KS, Fromwiller TE, Daniel RA, et al. Cystic pancreatic neoplasms: observe or operate. Ann Surg 2004;239:651-7; discussion 657-9.
- Sharma A, Kandlakunta H, Nagpal SJS, et al. Model to Determine Risk of Pancreatic Cancer in Patients With New-Onset Diabetes. Gastroenterology 2018;155:730-9.e3.
- Khan S, Safarudin RF, Kupec JT. Validation of the ENDPAC model: Identifying new-onset diabetics at risk of pancreatic cancer. Pancreatology 2021;21:550-5.
- 64. Ogawa Y, Tanaka M, Inoue K, et al. A prospective pancreatographic study of the prevalence of pancreatic carcinoma in patients with diabetes mellitus. Cancer 2002;94:2344-9.
- 65. Illés D, Terzin V, Holzinger G, et al. New-onset type 2 diabetes mellitus--A high-risk group suitable for the screening of pancreatic cancer? Pancreatology 2016;16:266-71.
- 66. Damiano J, Bordier L, Le Berre JP, et al. Should pancreas imaging be recommanded in patients over 50 years when diabetes is discovered because of acute symptoms? Diabetes Metab 2004;30:203-7.
- 67. Mizuno S, Nakai Y, Ishigaki K, et al. Screening Strategy of Pancreatic Cancer in Patients with Diabetes Mellitus. Diagnostics (Basel) 2020;10:572.
- Schacht SR, Olsen A, Dragsted LO, et al. Whole-Grain Intake and Pancreatic Cancer Risk-The Danish, Diet, Cancer and Health Cohort. J Nutr 2021;151:666-74.
- 69. Huang J, Lok V, Ngai CH, et al. Worldwide Burden of, Risk Factors for, and Trends in Pancreatic Cancer. Gastroenterology 2021;160:744-54.
- Park JH, Han K, Hong JY, et al. Changes in Metabolic Syndrome Status are Associated With Altered Risk of Pancreatic Cancer: A Nationwide Cohort Study. Gastroenterology 2022;162:509-20.e7.
- 71. Putzer D, Jaschke W. Radiological evaluation of focal pancreatic lesions. Dig Dis 2015;33:91-8.
- 72. Lee HJ, Kim MJ, Choi JY, et al. Relative accuracy of CT and MRI in the differentiation of benign from malignant pancreatic cystic lesions. Clin Radiol 2011;66:315-21.
- 73. Sainani NI, Saokar A, Deshpande V, et al. Comparative performance of MDCT and MRI with MR cholangiopancreatography in characterizing small pancreatic cysts. AJR Am J Roentgenol 2009;193:722-31.
- 74. Brugge WR. Diagnosis and management of cystic lesions of the pancreas. J Gastrointest Oncol 2015;6:375-88.
- 75. Bhosale P, Balachandran A, Tamm E. Imaging of benign

# Cheung et al. Management of PCL in Hong Kong

and malignant cystic pancreatic lesions and a strategy for follow up. World J Radiol 2010;2:345-53.

- 76. Sodickson A, Baeyens PF, Andriole KP, et al. Recurrent CT, cumulative radiation exposure, and associated radiation-induced cancer risks from CT of adults. Radiology 2009;251:175-84.
- Almeida RR, Lo GC, Patino M, et al. Advances in Pancreatic CT Imaging. AJR Am J Roentgenol 2018;211:52-66.
- 78. Schueller G, Schima W, Schueller-Weidekamm C, et al. Multidetector CT of pancreas: effects of contrast material flow rate and individualized scan delay on enhancement of pancreas and tumor contrast. Radiology 2006;241:441-8.
- Isoda H, Kataoka M, Maetani Y, et al. MRCP imaging at 3.0 T vs. 1.5 T: preliminary experience in healthy volunteers. J Magn Reson Imaging 2007;25:1000-6.
- Kim SY, Byun JH, Lee SS, et al. Biliary tract depiction in living potential liver donors: intraindividual comparison of MR cholangiography at 3.0 and 1.5 T. Radiology 2010;254:469-78.
- van Huijgevoort NCM, Del Chiaro M, Wolfgang CL, et al. Diagnosis and management of pancreatic cystic neoplasms: current evidence and guidelines. Nat Rev Gastroenterol Hepatol 2019;16:676-89.
- 82. Brugge WR, Lewandrowski K, Lee-Lewandrowski E, et al. Diagnosis of pancreatic cystic neoplasms: a report of the cooperative pancreatic cyst study. Gastroenterology 2004;126:1330-6.
- 83. Gillis A, Cipollone I, Cousins G, et al. Does EUS-FNA molecular analysis carry additional value when compared to cytology in the diagnosis of pancreatic cystic neoplasm? A systematic review. HPB (Oxford) 2015;17:377-86.
- McCarty TR, Garg R, Rustagi T. Pancreatic cyst fluid glucose in differentiating mucinous from nonmucinous pancreatic cysts: a systematic review and meta-analysis. Gastrointest Endosc 2021;94:698-712.e6.
- 85. Balaban VD, Cazacu IM, Pinte L, et al. EUS-through-theneedle microbiopsy forceps in pancreatic cystic lesions: A systematic review. Endosc Ultrasound 2021;10:19-24.
- 86. Facciorusso A, Crinò SF, Gkolfakis P, et al. Needle Tract Seeding after Endoscopic Ultrasound Tissue Acquisition of Pancreatic Lesions: A Systematic Review and Meta-Analysis. Diagnostics (Basel) 2022;12:2113.
- Napoleon B, Krishna SG, Marco B, et al. Confocal endomicroscopy for evaluation of pancreatic cystic lesions: a systematic review and international Delphi consensus report. Endosc Int Open 2020;8:E1566-81.
- 88. Krishna SG, Hart PA, DeWitt JM, et al. EUS-guided

confocal laser endomicroscopy: prediction of dysplasia in intraductal papillary mucinous neoplasms (with video). Gastrointest Endosc 2020;91:551-63.e5.

- 89. Machicado JD, Chao WL, Carlyn DE, et al. High performance in risk stratification of intraductal papillary mucinous neoplasms by confocal laser endomicroscopy image analysis with convolutional neural networks (with video). Gastrointest Endosc 2021;94:78-87.e2.
- Machicado JD, Napoleon B, Lennon AM, et al. Accuracy and agreement of a large panel of endosonographers for endomicroscopy-guided virtual biopsy of pancreatic cystic lesions. Pancreatology 2022;22:994-1002.
- 91. Seo N, Byun JH, Kim JH, et al. Validation of the 2012 International Consensus Guidelines Using Computed Tomography and Magnetic Resonance Imaging: Branch Duct and Main Duct Intraductal Papillary Mucinous Neoplasms of the Pancreas. Ann Surg 2016;263:557-64.
- 92. Hackert T, Fritz S, Klauss M, et al. Main-duct Intraductal Papillary Mucinous Neoplasm: High Cancer Risk in Duct Diameter of 5 to 9mm. Ann Surg 2015;262:875-80; discussion 880-1.
- Abdeljawad K, Vemulapalli KC, Schmidt CM, et al. Prevalence of malignancy in patients with pure main duct intraductal papillary mucinous neoplasms. Gastrointest Endosc 2014;79:623-9.
- 94. Fritz S, Hackert T, Hinz U, et al. Role of serum carbohydrate antigen 19-9 and carcinoembryonic antigen in distinguishing between benign and invasive intraductal papillary mucinous neoplasm of the pancreas. Br J Surg 2011;98:104-10.
- 95. Kim JR, Jang JY, Kang MJ, et al. Clinical implication of serum carcinoembryonic antigen and carbohydrate antigen 19-9 for the prediction of malignancy in intraductal papillary mucinous neoplasm of pancreas. J Hepatobiliary Pancreat Sci 2015;22:699-707.
- 96. Wang W, Zhang L, Chen L, et al. Serum carcinoembryonic antigen and carbohydrate antigen 19-9 for prediction of malignancy and invasiveness in intraductal papillary mucinous neoplasms of the pancreas: A meta-analysis. Biomed Rep 2015;3:43-50.
- Munigala S, Gelrud A, Agarwal B. Risk of pancreatic cancer in patients with pancreatic cyst. Gastrointest Endosc 2016;84:81-6.
- Fernández-del Castillo C, Targarona J, Thayer SP, et al. Incidental pancreatic cysts: clinicopathologic characteristics and comparison with symptomatic patients. Arch Surg 2003;138:427-3; discussion 433-4.
- Fernández-del Castillo C, Warshaw AL. Cystic tumors of the pancreas. Surg Clin North Am 1995;75:1001-16.

# 732

- 100.Kosmahl M, Pauser U, Peters K, et al. Cystic neoplasms of the pancreas and tumor-like lesions with cystic features: a review of 418 cases and a classification proposal. Virchows Arch 2004;445:168-78.
- 101. Sarr MG, Murr M, Smyrk TC, et al. Primary cystic neoplasms of the pancreas. Neoplastic disorders of emerging importance-current state-of-the-art and unanswered questions. J Gastrointest Surg 2003;7:417-28.
- 102. Jais B, Rebours V, Malleo G, et al. Serous cystic neoplasm of the pancreas: a multinational study of 2622 patients under the auspices of the International Association of Pancreatology and European Pancreatic Club (European Study Group on Cystic Tumors of the Pancreas). Gut 2016;65:305-12.
- 103. Mayo SC, Gilson MM, Herman JM, et al. Management of patients with pancreatic adenocarcinoma: national trends in patient selection, operative management, and use of adjuvant therapy. J Am Coll Surg 2012;214:33-45.
- 104. Aizpuru M, Starlinger P, Nagorney DM, et al. Contemporary outcomes of pancreaticoduodenectomy for benign and precancerous cystic lesions. HPB (Oxford) 2022;24:1416-24.
- 105.Hospital Authority (Hong Kong). Surgical Outcomes Monitoring and Improvement Programme. 2018-2019 Report. Hong Kong.
- 106. Olsen GA, Aalling L, Karstensen JG. Endoscopic ultrasound-guided ablation is a promising treatment for pancreatic cystic neoplasms - a systematic review. Dan Med J 2020;67:A01200019.
- 107. Choi JH, Seo DW, Song TJ, et al. Long-term outcomes after endoscopic ultrasound-guided ablation of pancreatic cysts. Endoscopy 2017;49:866-73.
- 108. National Cancer Institute. SEER Cancer Stat Facts: Pancreatic Cancer. Available online: https://seer.cancer. gov/statfacts/html/pancreas.html
- 109. Clinical Trials.gov. Nab-paclitaxel and Gemcitabine vs Gemcitabine Alone as Adjuvant Therapy for Patients With Resected Pancreatic Cancer (the "Apact" Study) (apact). NCT01964430. Available online: https://clinicaltrials.gov/ ct2/show/results/NCT01964430
- 110. Conroy T, Desseigne F, Ychou M, et al. FOLFIRINOX versus gemcitabine for metastatic pancreatic cancer. N Engl J Med 2011;364:1817-25.
- 111. Conroy T, Hammel P, Hebbar M, et al. FOLFIRINOX or Gemcitabine as Adjuvant Therapy for Pancreatic Cancer. N Engl J Med 2018;379:2395-406.
- 112.Katz MHG, Shi Q, Meyers JP, et al. Alliance A021501: Preoperative mFOLFIRINOX or mFOLFIRINOX plus hypofractionated radiation therapy (RT) for borderline

resectable (BR) adenocarcinoma of the pancreas. J Clin Oncol 2021;39:Abstract 377.

- 113. Neoptolemos JP, Palmer DH, Ghaneh P, et al. Comparison of adjuvant gemcitabine and capecitabine with gemcitabine monotherapy in patients with resected pancreatic cancer (ESPAC-4): a multicentre, open-label, randomised, phase 3 trial. Lancet 2017;389:1011-24.
- 114. Sohal DPS, Duong M, Ahmad SA, et al. Efficacy of Perioperative Chemotherapy for Resectable Pancreatic Adenocarcinoma: A Phase 2 Randomized Clinical Trial. JAMA Oncol 2021;7:421-7.
- 115. Tempero MA, Reni M, Riess H, et al. APACT: phase III, multicenter, international, open-label, randomized trial of adjuvant nab-paclitaxel plus gemcitabine (nab-P/ G) vs gemcitabine (G) for surgically resected pancreatic adenocarcinoma. J Clin Oncol 2019;37:Abstract 4000.
- 116. Versteijne E, Suker M, Groothuis K, et al. Preoperative Chemoradiotherapy Versus Immediate Surgery for Resectable and Borderline Resectable Pancreatic Cancer: Results of the Dutch Randomized Phase III PREOPANC Trial. J Clin Oncol 2020;38:1763-73.
- 117. Von Hoff DD, Ervin T, Arena FP, et al. Increased survival in pancreatic cancer with nab-paclitaxel plus gemcitabine. N Engl J Med 2013;369:1691-703.
- 118.National Comprehensive Cancer Network. NCCN Clinical Practice Guidelines. Pancreatic Adenocarcinoma. V2.2022. Available online: https://www.nccn.org/ professionals/physician\_gls/pdf/pancreatic.pdf
- 119. Valle JW, Palmer D, Jackson R, et al. Optimal duration and timing of adjuvant chemotherapy after definitive surgery for ductal adenocarcinoma of the pancreas: ongoing lessons from the ESPAC-3 study. J Clin Oncol 2014;32:504-12.
- 120. Lania A, Ferraù F, Rubino M, et al. Neoadjuvant Therapy for Neuroendocrine Neoplasms: Recent Progresses and Future Approaches. Front Endocrinol (Lausanne) 2021;12:651438.
- 121.Balzano G, Maffi P, Nano R, et al. Diabetes-free survival after extended distal pancreatectomy and islet auto transplantation for benign or borderline/malignant lesions of the pancreas. Am J Transplant 2019;19:920-8.
- 122. Dumitrascu T, Dima S, Stroescu C, et al. Clinical value of spleen-preserving distal pancreatectomy: a case-matched analysis with a special emphasis on the postoperative systemic inflammatory response. J Hepatobiliary Pancreat Sci 2014;21:654-62.
- 123. DiNorcia J, Ahmed L, Lee MK, et al. Better preservation of endocrine function after central versus

### Cheung et al. Management of PCL in Hong Kong

distal pancreatectomy for mid-gland lesions. Surgery 2010;148:1247-54; discussion 1254-6.

- 124. Lee SE, Jang JY, Hwang DW, et al. Clinical efficacy of organpreserving pancreatectomy for benign or low-grade malignant potential lesion. J Korean Med Sci 2010;25:97-103.
- 125. Firkins SA, Hart PA, Porter K, et al. Incidence and Risk Factors for New-Onset Diabetes Mellitus After Surgical Resection of Pancreatic Cystic Lesions: A MarketScan Study. Pancreas 2022;51:427-34.
- 126. De Bruijn KM, van Eijck CH. New-onset diabetes after distal pancreatectomy: a systematic review. Ann Surg 2015;261:854-61.
- 127. Scholten L, Mungroop TH, Haijtink SAL, et al. New-onset diabetes after pancreatoduodenectomy: A systematic review and meta-analysis. Surgery 2018;S0039-6060(18)30081-3.
- 128. Hirono S, Tani M, Kawai M, et al. A central pancreatectomy for benign or low-grade malignant neoplasms. J Gastrointest Surg 2009;13:1659-65.
- 129. Shikano T, Nakao A, Kodera Y, et al. Middle pancreatectomy: safety and long-term results. Surgery 2010;147:21-9.
- 130. Xiang GM, Tan CL, Zhang H, et al. Central pancreatectomy for benign or borderline lesions of the pancreatic neck: a single centre experience and literature review. Hepatogastroenterology 2012;59:1286-9.
- 131. Tseng DS, Molenaar IQ, Besselink MG, et al. Pancreatic Exocrine Insufficiency in Patients With Pancreatic or Periampullary Cancer: A Systematic Review. Pancreas 2016;45:325-30.
- 132. Chaudhary A, Domínguez-Muñoz JE, Layer P, et al. Pancreatic Exocrine Insufficiency as a Complication of Gastrointestinal Surgery and the Impact of Pancreatic Enzyme Replacement Therapy. Dig Dis 2020;38:53-68.
- 133. Hallac A, Aleassa EM, Rogers M, et al. Exocrine pancreatic insufficiency in distal pancreatectomy: incidence and risk factors. HPB (Oxford) 2020;22:275-81.
- 134.Kusakabe J, Anderson B, Liu J, et al. Long-Term Endocrine and Exocrine Insufficiency After Pancreatectomy. J Gastrointest Surg 2019;23:1604-13.
- 135.Marchegiani G, Mino-Kenudson M, Ferrone CR, et al. Patterns of Recurrence After Resection of IPMN: Who, When, and How? Ann Surg 2015;262:1108-14.
- 136. Kang MJ, Jang JY, Lee KB, et al. Long-term prospective cohort study of patients undergoing pancreatectomy for intraductal papillary mucinous neoplasm of the pancreas: implications for postoperative surveillance. Ann Surg 2014;260:356-63.
- 137. He J, Cameron JL, Ahuja N, et al. Is it necessary to

follow patients after resection of a benign pancreatic intraductal papillary mucinous neoplasm? J Am Coll Surg 2013;216:657-65; discussion 665-7.

- 138. Miyasaka Y, Ohtsuka T, Tamura K, et al. Predictive Factors for the Metachronous Development of High-risk Lesions in the Remnant Pancreas After Partial Pancreatectomy for Intraductal Papillary Mucinous Neoplasm. Ann Surg 2016;263:1180-7.
- 139. Lindkvist B. Diagnosis and treatment of pancreatic exocrine insufficiency. World J Gastroenterol 2013;19:7258-66.
- 140. Roeyen G, Berrevoet F, Borbath I, et al. Expert opinion on management of pancreatic exocrine insufficiency in pancreatic cancer. ESMO Open 2022;7:100386.
- 141. de la Iglesia D, Vallejo-Senra N, López-López A, et al. Pancreatic exocrine insufficiency and cardiovascular risk in patients with chronic pancreatitis: A prospective, longitudinal cohort study. J Gastroenterol Hepatol 2019;34:277-83.
- 142. Sikkens EC, Cahen DL, Koch AD, et al. The prevalence of fat-soluble vitamin deficiencies and a decreased bone mass in patients with chronic pancreatitis. Pancreatology 2013;13:238-42.
- 143. Shintakuya R, Uemura K, Murakami Y, et al. Sarcopenia is closely associated with pancreatic exocrine insufficiency in patients with pancreatic disease. Pancreatology 2017;17:70-5.
- 144. Duggan SN, Smyth ND, Murphy A, et al. High prevalence of osteoporosis in patients with chronic pancreatitis: a systematic review and meta-analysis. Clin Gastroenterol Hepatol 2014;12:219-28.
- 145. Diéguez-Castillo C, Jiménez-Luna C, Prados J, et al. State of the Art in Exocrine Pancreatic Insufficiency. Medicina (Kaunas) 2020;56:523.
- 146. Dominguez-Muñoz JE. Diagnosis and treatment of pancreatic exocrine insufficiency. Curr Opin Gastroenterol 2018;34:349-54.
- 147.Korostensky M, Martin SR, Swain M, et al. Elimination of 72-Hour Quantitative Fecal Fat Testing by Restriction, Laboratory Consultation, and Evaluation of Specimen Weight and Fat Globules. J Appl Lab Med 2018;3:357-65.
- 148. Pezzilli R, Andriulli A, Bassi C, et al. Exocrine pancreatic insufficiency in adults: a shared position statement of the Italian Association for the Study of the Pancreas. World J Gastroenterol 2013;19:7930-46.
- 149. Vanga RR, Tansel A, Sidiq S, et al. Diagnostic Performance of Measurement of Fecal Elastase-1 in Detection of Exocrine Pancreatic Insufficiency: Systematic Review and Metaanalysis. Clin Gastroenterol Hepatol 2018;16:1220-8.e4.

- 150.Khan A, Vege SS, Dudeja V, et al. Staging exocrine pancreatic dysfunction. Pancreatology 2022;22:168-72.
- 151. Löhr JM, Dominguez-Munoz E, Rosendahl J, et al. United European Gastroenterology evidence-based guidelines for the diagnosis and therapy of chronic pancreatitis (HaPanEU). United European Gastroenterol J 2017;5:153-99.
- 152. Aune D, Mahamat-Saleh Y, Norat T, et al. Tobacco smoking and the risk of pancreatitis: A systematic review and meta-analysis of prospective studies. Pancreatology 2019;19:1009-22.
- 153. Samokhvalov AV, Rehm J, Roerecke M. Alcohol Consumption as a Risk Factor for Acute and Chronic Pancreatitis: A Systematic Review and a Series of Metaanalyses. EBioMedicine 2015;2:1996-2002.
- 154.Johnson CD, Arbuckle R, Bonner N, et al. Qualitative Assessment of the Symptoms and Impact of Pancreatic Exocrine Insufficiency (PEI) to Inform the Development of a Patient-Reported Outcome (PRO) Instrument. Patient 2017;10:615-28.
- 155. Johnson CD, Williamson N, Janssen-van Solingen G, et al. Psychometric evaluation of a patient-reported outcome measure in pancreatic exocrine insufficiency (PEI). Pancreatology 2019;19:182-90.
- 156. Halm U, Löser C, Löhr M, et al. A double-blind, randomized, multicentre, crossover study to prove equivalence of pancreatin minimicrospheres versus microspheres in exocrine pancreatic insufficiency. Aliment Pharmacol Ther 1999;13:951-7.
- 157. Safdi M, Bekal PK, Martin S, et al. The effects of oral pancreatic enzymes (Creon 10 capsule) on steatorrhea: a multicenter, placebo-controlled, parallel group trial in subjects with chronic pancreatitis. Pancreas 2006;33:156-62.
- 158. Whitcomb DC, Lehman GA, Vasileva G, et al. Pancrelipase delayed-release capsules (CREON) for exocrine pancreatic insufficiency due to chronic pancreatitis or pancreatic surgery: A double-blind randomized trial. Am J Gastroenterol 2010;105:2276-86.
- 159. Thorat V, Reddy N, Bhatia S, et al. Randomised clinical trial: the efficacy and safety of pancreatin enteric-coated minimicrospheres (Creon 40000 MMS) in patients with pancreatic exocrine insufficiency due to chronic pancreatitis--a double-blind, placebo-controlled study. Aliment Pharmacol Ther 2012;36:426-36.
- 160.Keller J, Layer P. Human pancreatic exocrine response to nutrients in health and disease. Gut 2005;54 Suppl 6:vi1-28.
- 161. Stallings VA, Stark LJ, Robinson KA, et al. Evidence-

based practice recommendations for nutrition-related management of children and adults with cystic fibrosis and pancreatic insufficiency: results of a systematic review. J Am Diet Assoc 2008;108:832-9.

- 162. Maker AV, Sheikh R, Bhagia V, et al. Perioperative management of endocrine insufficiency after total pancreatectomy for neoplasia. Langenbecks Arch Surg 2017;402:873-83.
- 163. Muller WA, Brennan MF, Tan MH, et al. Studies of glucagon secretion in pancreatectomized patients. Diabetes 1974;23:512-6.
- 164. Vigili de Kreutzenberg S, Maifreni L, Lisato G, et al. Glucose turnover and recycling in diabetes secondary to total pancreatectomy: effect of glucagon infusion. J Clin Endocrinol Metab 1990;70:1023-9.
- 165.Nosadini R, del Prato S, Tiengo A, et al. Insulin sensitivity, binding, and kinetics in pancreatogenic and type I diabetes. Diabetes 1982;31:346-55.
- 166. Parsaik AK, Murad MH, Sathananthan A, et al. Metabolic and target organ outcomes after total pancreatectomy: Mayo Clinic experience and meta-analysis of the literature. Clin Endocrinol (Oxf) 2010;73:723-31.
- 167.Mayeux SE, Kwon W, Rosario VL, et al. Long-term health after pancreatic surgery: the view from 9.5 years. HPB (Oxford) 2021;23:595-600.
- 168. Yamamoto-Kataoka S, Shimizu S, Yamazaki H, et al. Development of a preoperative prediction model for new-onset diabetes mellitus after partial pancreatectomy: A retrospective cohort study. Medicine (Baltimore) 2021;100:e26311.
- 169. Liu A, Carmichael KA, Schallom ME, et al. Retrospective review of postoperative glycemic control in patients after distal pancreatectomy. Int J Surg 2017;41:86-90.
- 170. Struyvenberg MR, Fong ZV, Martin CR, et al. Impact of Treatments on Diabetic Control and Gastrointestinal Symptoms After Total Pancreatectomy. Pancreas 2017;46:1188-95.

**Cite this article as:** Cheung TT, Lee YT, Tang RSY, She WH, Cheng KC, Cheung CC, Chiu KWH, Chok KSH, Chow WS, Lai TW, Seto WK, Yau T. The Hong Kong consensus recommendations on the diagnosis and management of pancreatic cystic lesions. HepatoBiliary Surg Nutr 2023;12(5):715-735. doi: 10.21037/hbsn-22-471