Increased Expression of Yes-Associated Protein 1 in Hepatocellular Carcinoma with Stemness and Combined Hepatocellular-Cholangiocarcinoma

Gi Jeong Kim¹, Hyunki Kim¹*, Young Nyun Park^{1,2}*

1 Department of Pathology, Yonsei University College of Medicine, Seoul, Republic of Korea, 2 Integrated Genomic Research Center for Metabolic Regulation, Yonsei University College of Medicine, Seoul, Republic of Korea

Abstract

Combined hepatocellular-cholangiocarcinoma (cHC-CC) and some hepatocellular carcinomas (HCCs) express stemnessrelated markers, such as epithelial adhesion molecule (EpCAM) and keratin 19 (K19), the expression of which has been reported to be associated with more aggressive behavior therein than in HCCs without. Yes-associated protein 1 (YAP1), a potential oncogene, is known to promote stem cell proliferation. In the present study, YAP1 expression and clinicopathological features were evaluated and compared among three groups comprising 36 HCCs that expressed both EpCAM and K19, 64 HCCs that did not express EpCAM and K19, and 58 cHC-CCs, which consisted of 38 cases of the classical type and 20 cases of the intermediate-cell subtype. YAP1 expression was more frequently noted in EpCAM(+)/K19(+) HCCs (55.6%) and in cHC-CCs (67.2%) than in EpCAM(-)/K19(-) HCCs (17.2%) (P<0.001 for both). In cHC-CCs, YAP1 expression was observed in 63% of classical type cHC-CCs and in 75% of the intermediate subtype; moreover, such expression was correlated with poorer histological differentiation (P = 0.017) and was more frequently noted in transition zones than in HCC areas (P = 0.060). Disease-free and overall survival showed a statistically significant difference among the three groups: disease-free survival was highest for EpCAM(-)/K19(-) HCCs and lowest for cHC-CCs, with EpCAM(+)/K19(+) HCCs falling in between (P<0.05). Overall survival rate was lower in HCCs and cHC-CCs with YAP1 expression compared to those without (P=0.05), whereas disease-free survival showed no significant difference according to YAP1 expression. Increased YAP1 expression was more frequently found in cHC-CCs and HCCs with stemness than in HCCs without, and a YAP1 pathway is suggested to be involved in the obtainment stemness characteristics in HCCs and cHC-CCs.

Citation: Kim GJ, Kim H, Park YN (2013) Increased Expression of Yes-Associated Protein 1 in Hepatocellular Carcinoma with Stemness and Combined Hepatocellular-Cholangiocarcinoma. PLoS ONE 8(9): e75449. doi:10.1371/journal.pone.0075449

Editor: Motoyuki Otsuka, The University of Tokyo, Japan

Received July 4, 2013; Accepted August 15, 2013; Published September 24, 2013

Copyright: © 2013 Kim et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This study was supported by a grant of the Korea Health care technology R&D Project, Ministry of Health & Welfare, Republic of Korea (A120631). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: Young Nyun Park, MD, PhD who is a co-corresponding author of this manuscript, is a member of PLOS ONE editorial board. This does not alter the authors' adherence to all the PLOS ONE policies on sharing data and materials.

* E-mail: young0608@yuhs.ac (YNP); kimhyunki@yuhs.ac (HK)

Introduction

Hepatocellular carcinoma (HCC) is the sixth most common malignancy worldwide and the third greatest cause of cancerrelated mortality, especially in Asia and Africa.[1] Combined hepatocellular and cholangiocarcinoma (cHC-CC), an uncommon subtype that accounts for approximately 1% of all primary liver tumors, comprises morphologically and phenotypically mixed elements of HCC and cholangiocarcinoma (CC).[2,3] cHC-CCs can be categorized as classical type or subtypes with stem cell features. The latter can further be subcategorized into typical subtype, intermediate-cell subtype, and cholangiocellular subtype.[3]

Recent advances in the study of cancer stem cells have indicated that cancer stem cells play a critical role in tumor growth and the progression of HCCs, contributing to their ability to self-renew, differentiate, and generate metastatic tumors in local or distant organs.[4–8] HCCs expressing stemness-related markers, including epithelial cell adhesion molecule (EpCAM), keratin 19 (K19), CD90, and CD133, are known to exhibit more aggressive biological behavior and worse prognosis than HCCs that express no stemness-related markers. [5,9–11] As well, cHC-CCs, which also express stemness-related markers, have been reported to show more aggressive behavior than HCCs. [12,13]

Yes-associated protein 1 (YAP1) is a major downstream target of the Hippo-signaling pathway, an evolutionarily conserved pathway from Drosophila to humans that is known to control organ size during development.[14-16] Regulation of the Hippo-signaling pathway is known to be mediated by phosphorylation and subcellular localization of YAP1. Activation of the Hippo-signaling pathway induces phosphorylation of YAP1, which prevents the translocation thereof to the nucleus. Instead, phosphorylated YAP1 remains in the cytoplasm, where it is degraded by proteasomes. When the Hippo-signaling pathway is inactivated, dephosphorylated YAP1 is translocated to the nucleus where it interacts with transcription factors, eventually leading to the proliferation of cells to various organ systems.[17-20] One previous study using transgenic mice with liver-specific YAP1 overexpression revealed significant increases in liver size and number of primary liver tumors morphologically resembling cHC-CC in humans.[21]

To our knowledge, the expression of YAP1 has not been investigated in primary liver cancers with stemness features. In this study, YAP1 expression patterns and clinicopathological characteristics were compared among HCCs with and without stemnessrelated markers and cHC-CCs.

Materials and Methods

Case selection and clinicopathological analysis

A total of 158 cases of primary liver carcinoma showing the following features were studied: (1) 36 HCCs expressing both EpCAM and K19 [EpCAM(+)/K19(+)], (2) 64 HCCs without expression of both EpCAM and K19 [EpCAM(-)/K19(-)], and (3) 58 cHC-CCs. The cHC-CCs included 38 cases of classical type cHC-CC and 20 cases of the intermediate-cell subtype.

Tumor specimens were fixed in 10% buffered formalin and representative sections were submitted for histological examination. The histopathological variables recorded for each case included tumor size, multiplicity, differentiation according to a three-tiered grading system (well, moderately and poorly differentiated), vascular invasion, and intrahepatic metastasis. Clinical features including age, sex, etiology, and follow-up data were obtained from hospital charts. There were 55 cases with a history of preoperative treatment including 44 cases of transcatheter arterial chemoembolization (TACE), one case of concurrent chemoradiotherapy (CCRT), five cases of TACE and CCRT, one case of chemotherapy, and four cases of radiofrequency ablation. This study was approved by the Institutional Review Board of Severance Hospital, Yonsei University College of Medicine (Seoul, Korea). The Institutional Review Board waived the need for consent in this study (4-2013-0272).

Immunohistochemical staining

The expressions of YAP1, EpCAM, and K19 were evaluated by immunohistochemical staining in representative sections of formalin-fixed, paraffin-embedded (FFPE) tissues. Primary antibodies for YAP1 (1:100, Cell Signaling Technology, Danvers, MA, USA), EpCAM (1:1000, Calbiochem, Merck, Darmstadt, Germany), and K19 (1:100, Dako, Carpinteria, CA, USA) were used. Briefly, 4-µm-thick sections of FFPE tissues were deparaffinized and rehydrated. After treatment with a 3% hydrogen peroxide solution for 20 min to block endogenous peroxidases, the sections were pretreated in 10 mM citrate buffer (pH 6.0) in a microwave oven for 20 min for antigen retrieval. After incubation with the primary antibodies, the sections were then processed using the EnVisionTM Detection System (Dako) according to the manufacturer's instructions, and 3, 3'-diaminobenzidine tetrahydrochloride was used as a chromogen. All sections were counterstained with Mayer hematoxylin.

The immunoreactivities of YAP1, EpCAM, and K19 were evaluated by two independent observers (G. J. Kim and H. Kim). Conflicting cases were reviewed and discussed until a consensus was obtained. For the assessment of YAP1 expression, nuclear YAP1 expression of bile ductular epithelial cells was used as an internal positive control with moderate intensity. Non-tumor hepatocytes were used as an internal negative control. YAP1 expression was graded according to nuclear expression intensity: weak, moderate, or strong expression. Cases showing YAP1 expression in less than 5% of the tumor cells of any intensity grade or those of weak intensity were regarded as negative (no YAP1 expression), while cases showing moderate to strong intensities in more than 5% of the tumor cells were regarded as positive for YAP1 expression. For expression of EpCAM and K19, membranous or cytoplasmic expression in more than 5% of the tumor cells was considered positive. Bile ductular epithelial cells were used as an internal positive control for EpCAM and K19. All cHC-CCs were positive for both EpCAM and K19.

Statistical analyses

Statistical analyses were performed using SPSS software version 19.0 for Windows (SPSS Inc., Chicago, IL, USA). Fisher's exact test was used for analysis of categorical variables. Continuous variables were analyzed using one-way analysis of variance (ANOVA) or Student's t-test; these results are expressed as the mean \pm standard deviation. Histological grades were compared using the Mann-Whitney U test. On survival analysis, clinicopathologic variables were dichotomized and analyzed according to their effect on prognosis. Disease-free survival and overall survival analysis was performed using the Kaplan-Meier method, and differences between the groups were assessed using the log-rank test. Univariate and multivariate survival analyses were carried out using Cox proportional hazard regression models. Only variables significant on the univariate analysis of factors affecting survival were used in the multivariate analysis. Estimated relative risks of death were expressed as adjusted hazard ratios (HR) and corresponding 95% confidence intervals (95% CI). All P-values less than 0.05 were regarded as statistically significant.

Results

Clinicopathological features of EpCAM(-)/K19(-) HCCs, EpCAM(+)/K19(+) HCCs, and cHC-CCs

The clinicopathological characteristics of EpCAM(-)/K19(-)HCCs, EpCAM(+)/K19(+) HCCs, and cHC-CCs are summarized in Table 1. Both EpCAM(+)/K19(+) HCCs and cHC-CCs developed in patients of younger age than EpCAM(-)/K19(-)HCCs (P=0.001 and P=0.005, respectively). Tumor size was larger in the cHC-CCs than in HCCs (cHC-CCs vs. EpCAM(-)/ K19(-) HCCs, P<0.001; cHC-CCs vs. EpCAM(+)/K19(+) HCCs, P = 0.033). cHC-CCs also more frequently presented as a single lesion than EpCAM(-)/K19(-) HCCs and EpCAM(+)/ K19(+) HCCs (P=0.032 and P=0.002, respectively). Vascular invasion was more frequently observed in cHC-CCs than in HCCs (cHC-CCs vs. EpCAM(-)/K19(-) HCCs, P<0.001; cHC-CCs vs. EpCAM(+)/K19(+) HCCs, P = 0.025). Additionally, EpCAM(+)/ K19(+) HCCs and cHC-CCs exhibited poorer histological differentiation than EpCAM(-)/K19(-) HCCs (P<0.001 for both). Among cHC-CCs, the classical type was more frequently related to human hepatitis B virus or hepatitis C virus infection than the intermediated-cell subtype (P=0.001). There was no difference between the two types in terms of sex, age, tumor size, differentiation, et al. (Table S1).

YAP1 expression in EpCAM(-)/K19(-) HCCs, EpCAM(+)/ K19(+) HCCs, and cHC-CCs

YAP1 expression was found in 11/64 (17.2%) EpCAM(–)/ K19(–) HCCs, 20/36 (55.6%) EpCAM(+)/K19(+) HCCs, and 39/58 (67.2%) cHC-CCs (Table 1) (Figure 1). YAP1 expression was significantly, more frequently observed in EpCAM(+)/K19(+) HCCs and cHC-CCs than in EpCAM(–)/K19(–) HCCs (P<0.001 for both). There was no significant difference in YAP1 expression between EpCAM(+)/K19(+) HCCs and cHC-CCs.

In cHC-CCs, YAP1 expression was present in 24/38 (63.2%) classical type cHC-CCs and in 15/20 (75.0%) intermediate subtype cHC-CCs with stem cell features, a difference that was not statistically significant. YAP1 expression was further analyzed

Table 1. Clinicopathological features and YAP1 expression in HCCs and combined hepatocellular-cholangiocarcinomas (cHC-CCs).

	Group 1	Group 2	Group3			
	EpCAM(-)/K19(-) HCCs (%) (<i>n</i> =64)	EpCAM(+)/K19(+) HCCs (%) (<i>n</i> =36)	cHC-CCs (%) (<i>n</i> =58)	Group 1 vs. 2	Group 1 vs. 3	Group 2 vs. 3
Sex				0.892	0.810	0.943
Male	54 (84.4)	30 (83.3)	48 (82.8)			
Female	10 (15.6)	6 (16.7)	10 (17.2)			
Age (years)	59.6±9.7	52.4±11.8	54.4±10.5	0.001	0.005	0.378
Etiology				0.239	0.003	0.161
Non-viral	7 (10.9)	7 (19.4)	19 (32.8)			
HBV	50 (78.2)	28 (77.8)	35 (60.3)			
нси	7 (10.9)	1 (2.8)	4 (6.9)			
Tumor size (mm)	33.4±16.5	38.4±21.8	50.1 ± 30.6	0.204	<0.001	0.033
Differentiation				0.001	0.001	0.903
Well	27 (42.2)	3 (8.3)	10 (17.2)			
Moderate	30 (46.9)	26 (72.2)	33 (56.9)			
Poor	7 (10.9)	7 (19.5)	15 (25.9)			
Vascular invasion				0.199	<0.001	0.025
Absence	37 (57.8)	16 (44.4)	13 (22.4)			
Presence	27 (42.2)	20 (55.6)	45 (77.6)			
Multiplicity				0.251	0.032	0.002
Single	54 (84.4)	27 (75.0)	56 (96.6)			
Multiple	10 (15.6)	9 (25.0)	2 (3.4)			
Intrahepatic metastasis				0.617	0.003	0.071
Absence	62 (96.9)	34 (94.4)	46 (79.3)			
Presence	2 (3.1)	2 (5.6)	12 (20.7)			
Preoperative treatment				0.068	0.179	0.532
No	47 (73.4)	20 (55.6)	36 (62.1)			
Yes	17 (26.6)	16 (44.4)	22 (37.9)			
YAP1 expression*				<0.001	<0.001	0.255
Negative	53 (82.8)	16 (44.4)	19 (32.8)			
Positive	11 (17.2)	20 (55.6)	39 (67.2)			

HCC, hepatocellular carcinoma; cHC-CC, combined hepatocellular-cholangiocarcinoma

*Nuclear YAP1 expression with moderate to strong intensities in more than 5% of the tumor cells were regarded as positive. doi:10.1371/journal.pone.0075449.t001



Figure 1. EpCAM and K19 expression in hepatocellular carcinoma (HCC). (A-D) HCC expressing both EpCAM and K19. The expression of EpCAM was mainly membranous, and K19 showed cytoplasmic expression in tumor cells. Nuclear expression of YAP1 was noted. (E-H) HCC without expression of both EpCAM and K19. There was no nuclear expression of YAP1. (Original magnification, ×200). doi:10.1371/journal.pone.0075449.g001

Table 2. YAP1	expression in eac	n histologic compone	ent of combined he	patocellular-choland	aiocarcinomas (classic t	vpe).
		.			,	

YAP1 expression*	HCC area	CC area	Transition zone	Transition <i>vs.</i> HCC area	Transition <i>vs.</i> CC area	HCC vs. CC area
Positive	8 (27.6%)	12 (41.4%)	15 (51.7%)	0.060	0.430	0.269
Negative	21 (72.4%)	17 (58.6%)	14 (48.3%)			

HCC, hepatocellular carcinoma; CC, cholangiocarcinoma

*Nuclear YAP1 expression with moderate to strong intensities in more than 5% of the tumor cells were regarded as positive.

doi:10.1371/journal.pone.0075449.t002

in each histological component of classical type cHC-CCs (this analysis was performed in 29 cases of classical type cHC-CC due to a shortage of tissues). In doing so, positive expression was found in 8/29 (27.6%) HCC areas, 12/29 (41.4%) cholangiocarcinoma (CC) areas, and 15/29 (51.7%) transitional zones (Table 2) (Figure 2). YAP1 expression was more frequently recoded in transitional zones than in HCC areas (P=0.060), although this was not statistically significant. The intermediate-cell subtype of cHC-CCs with stem cell features predominantly consisted of tumor cells with intermediate features between hepatocytes and cholangiocytes, which showed no zonal pattern of YAP1 expression.

The clinicopathological characteristics of HCCs and cHC-CCs according to YAP1 expression are summarized in Table 3. Among EpCAM(+)/K19(+) HCCs, cases with YAP1 expression more frequently manifested as a single lesion than those that did not express YAP1 (P=0.005). Among cHC-CCs, the expression of YAP1 was associated with poorer differentiation (P=0.017), whereas both EpCAM(-)/K19(-) HCCs and EpCAM(+)/K19(+) HCCs showed no difference in tumor differentiation according to YAP1 expression. There were no differences in tumor size and vascular invasion according to YAP1 expression for all groups.

Survival analysis in EpCAM(-)/K19(-) HCCs, EpCAM(+)/ K19(+) HCCs, and cHC-CCs

Overall survival and disease-free survival were evaluated for 152 patients, including 61 cases of EpCAM(-)/K19(-) HCC, 35 cases of EpCAM(+)/K19(+) HCC, and 56 cases of cHC-CC. Six

patients who died within one month after an operation were excluded from the survival analysis to avoid any influence of perioperative mortality. The median follow-up time after surgical resection was 32.8 months (4.3–128.7) and 34 patients died of HCC or cHC-CC during follow-up. Disease-free survival showed a statistically significant difference among the three groups: disease-free survival rate was highest for EpCAM(–)/K19(–) HCCs and lowest for cHC-CCs, with EpCAM(+)/K19(+) HCCs falling in between (P=0.002) (Fig. 3A). Overall survival also revealed a statistically significant difference among the three groups (P<0.001) (Fig. 3B). Among the patients with cHC-CC, there was no difference between classical type and intermediate-cell subtype patients in overall survival and disease-free survival rate (Figure S1).

When primary liver cancers were divided into two groups according to YAP1 expression, there were 67 cases with YAP1 expression and 85 cases without. Disease-free survival showed no significant difference between these two groups, whereas overall survival rate was relatively lower in primary liver cancers with YAP1 expression compared to those that did not express YAP1 (P=0.050) (Fig. 3C and 3D).

Univariate analysis revealed that larger tumor size (\geq 4 cm) (P=0.006), history of preoperative treatment (P<0.001), vascular invasion (P<0.001), intrahepatic metastasis (P<0.001), and the histologic groups of cHC-CC and EpCAM(+)/K19(+) HCC (P=0.004) were adverse prognostic factors affecting disease-free survival after resection. In regards to overall survival, larger tumor size (\geq 4 cm) (P<0.001), vascular invasion (P<0.001), intrahepatic metastasis (P<0.001) and the histologic groups of cHC-CC and



Figure 2. YAP1 expression in combined hepatocellular cholangiocarcinoma (cHC-CC). (A-F) Pathological features and YAP1 expression are shown in each component of classical type combined hepatocellular cholangiocarcinoma, including a hepatocellular carcinoma (HCC) area (A, B), a cholangiocarcinoma (CC) area (C, D), and a transitional zone (E, F). YAP1 expression is evident in the nuclei of tumor cells in CC areas (D) and transitional zones (F) in contrast to weak nulcear YAP1 expression in HCC areas (B). (G-H) Intermediate subtype of cHC-CC with stem cell features showing strong nuclear YAP1 expression. (Original magnification, $\times 200$). doi:10.1371/journal.pone.0075449.q002

expression.
(AP1
tocellular-cholangiocarcinomas according to ${\sf Y}$
hepat
combined
CCs and
of H
haracteristics
thological c
. Clinicopat
Table 3.

	EpCAM(-)/K19(-)	HCCs		EpCAM(+)/K19(+) HCCs			cHC-CCs		
	YAP1 negative $(\%)$ $(n=53)$	YAP1 positive (%) (<i>n</i> =11)	م	YAP1 negative (%) (<i>n</i> =16)	YAP1 positive (%) (<i>n</i> =20)	ط	YAP1 negative (%) (<i>n</i> =19)	YAP1 positive (%) (<i>n</i> =39)	م
Sex			0.356			0.196			0.142
Male	46 (86.8)	8 (72.7)		15 (93.8)	15 (75.0)		18 (94.7)	30 (76.9)	
Female	7 (13.2)	3 (27.3)		1 (6.2)	5 (25.0)		1 (5.3)	9 (23.1)	
Age (years)	58.3±9.7	66.0±7.0	0.015	50.1±11.2	54.2±12.2	0.303	58.9±9.3	52.3±10.4	0.022
Tumor size (mm)	33.2±16.3	34.8±17.9	0.763	38.3±18.2	38.5±24.9	0.985	43.6±26.4	53.3±32.3	0.258
Etiology									
Non-viral	5 (9.4)	2 (18.2)		2 (12.5)	5 (25.0)		5 (26.3)	14 (35.9)	
Viral (HBV, HCV)	48 (90.6)	9 (81.8)		14 (87.5)	15 (75.0)		14 (73.7)	25 (64.1)	
Differentiation			0.517			0.085			0.017
Well	23 (43.4)	4 (36.4)		3 (18.8)	0 (0.0)		6 (31.6)	4 (10.3)	
Moderate	25 (47.2)	5 (45.5)		11 (68.8)	15 (75.0)		11 (57.9)	22 (56.4)	
Poor	5 (9.4)	2 (18.2)		2 (12.5)	5 (25.0)		2 (10.5)	13 (33.3)	
Vascular invasion			0.362			0.940			0.619
Absence	32 (60.4)	5 (45.5)		7 (43.7)	9 (45.0)		5 (26.3)	8 (20.5)	
Presence	21 (39.6)	6 (54.5)		9 (56.3)	11 (55.0)		14 (73.7)	31 (79.5)	
Multiplicity			1.000			0.005			1.000
Single	44 (83.0)	10 (90.9)		8 (50.0)	19 (95.0)		18 (94.7)	38 (97.4)	
Multiple	9 (17.0)	1 (9.1)		8 (50.0)	1 (5.0)		1 (5.3)	1 (2.6)	
Intrahepatic metastasis			1.000			0.190			0.733
Absence	51 (96.2)	11		14 (87.5)	20		16 (84.2)	30 (76.9)	
Presence	2 (3.8)	0		2 (12.5)	0		3 (15.8)	9 (23.1)	
Preoperative treatment			1.000			0.202			0.486
No	39 (73.6)	8 (72.7)		7 (43.8)	13 (65.0)		13 (68.4)	23 (59.0)	
Yes	14 (26.4)	3 (27.3)		9 (56.3)	7 (35.0)		6 (31.6)	16 (41.0)	
HCC, hepatocellular carcinoma; doi:10.1371/journal.pone.00754	cHC-CC, combined hepa 49.t003	atocellular-cholangiocar	cinoma						



Figure 3. Kaplan–Meier's plot analysis for disease-free survival and overall survival in HCCs and combined hepatocellularcholangiocarcinomas (cHC-CCs). Kaplan–Meier's plot analysis for disease-free survival (A) and overall survival (B) showing a significant difference among EpCAM(–)/K19(–) HCCs, EpCAM(+)/K19(+) HCCs, and cHC-CCs. Overall survival was relatively worse in HCCs and cHC-CCs with YAP1 expression (D), whereas there was no significant difference in disease-free survival between the two groups. doi:10.1371/journal.pone.0075449.q003

EpCAM(+)/K19(+) HCC (P=0.002) were revealed as adverse prognostic factors (Table 4).

Multivariable analysis indicated that history of preoperative treatment (HR = 2.063, P = 0.004) and vascular invasion (HR = 2.240, P = 0.007) were independent prognostic factors for disease-free survival after resection. For overall survival, larger tumor size (≥ 4 cm) (HR = 3.448, P = 0.008), vascular invasion (HR = 7.135, P = 0.009), and the histologic groups of cHC-CC and EpCAM(+)/K19(+) HCC (P = 0.034) were shown to be independent prognostic factors on multivariable analysis (Table 5).

Discussion

Among morphologically pure HCCs, cases that express stemness-related markers, such as EpCAM, K19, CD133, etc., have been reported to exhibit more aggressive clinicopathological features, including more frequent vascular invasion, increased angiogenesis, higher recurrence rate, and worse prognosis.[9,10,22,23] In this study, EpCAM(+)/K19(+) HCCs showed poorer histological differentiation, greater vascular invasion, and worse prognosis than EpCAM(-)/K19(-) HCCs.

cHC-CCs are rare primary liver tumors, and can be categorized into classical type cHC-CCs and subtypes with stem cell features.[3] Classical type cHC-CCs contain HCC areas, CC areas and transitional zones, which comprise tumor cells with intermediate morphology resembling stem/progenitor cells. Subtypes with stem cell features include the typical subtype, intermediate-cell subtype, and cholangiocellular subtype, and tumor cells that have phenotypical or immunophenotypical features of stem/progenitor cells are the main component.[3] The gene signatures associated with early liver development and stem cells have been reported to be significantly enriched in cHC-CC.[24] These features suggest that cHC-CC is closely associated with stemness. Moreover, cHC-CCs have been reported to exhibit aggressive characteristics of greater lymph node involvement, vascular invasion, and worse prognosis than HCC.[13,25-27] The present study also revealed that cHC-CCs show more aggressive characteristics of larger tumor size, more frequent vascular invasion and poorer differentiation than EpCAM(-)/K19(-) HCCs. Among these characteristics, larger tumor size and more frequent vascular invasion were also more frequently noted in cHC-CCs than in EpCAM(+)/K19(+) HCCs. Moreover, diseasefree survival and overall survival showed a statistically significant **Table 4.** Univariate analysis of disease-free and overall survival rate.

		Disease-fr	ee survival		Overall su	rvival	
Variable	Ν	HR	95% CI	Р	HR	95% CI	Ρ
Sex							
Female	25	1			1		
Male	127	1.021	0.546-1.909	0.946	1.670	0.588-4.745	0.336
Age (years)							
<50	43	1			1		
≥50	109	1.395	0.791–2.461	0.249	1.344	0.604–2.992	0.469
Etiology							
Viral (HBV, HCV)	121	1			1		
Non-viral	31	1.371	0.790-2.381	0.261	1.307	0.611-2.798	0.490
Tumor size							
<4 cm	83	1			1		
≥4 cm	69	1.974	1.213–3.211	0.006	6.925	2.859–16.769	<0.001
Multiplicity							
Single	134	1.000			1		
Multiple	18	1.437	0.731-2.824	0.292	1.251	0.381-4.101	0.712
Differentiation							
Well/moderate	124	1			1		
Poor	28	1.554	0.885–2.726	0.124	1.836	0.867–3.887	0.112
Preoperative treatment							
No	101	1			1		
Yes	51	2.307	1.422–3.744	<0.001	0.978	0.476-2.009	0.951
Vascular invasion							
Absence	63	1			1		
Presence	89	2.841	1.636–4.935	<0.001	14.769	3.534–61.715	<0.001
Intrahepatic metastasis							
Absence	136	1			1		
Presence	16	3.298	1.712–6.350	<0.001	4.563	2.035-10.232	<0.001
Histologic group							
EpCAM(-)/K19(-) HCCs	61	1		0.004	1		0.002
EpCAM(+)/K19(+) HCCs	35	2.148	1.092-4.226	0.026	16.533	2.091-130.707	0.008
cHC-CCs	56	2.812	1.530–5.167	0.001	29.442	3.953-219.286	0.001
YAP1 expression*							
Negative	85	1			1		
Positive	67	1.261	0.777-2.046	0.346	1.990	0.988-4.008	0.050

*Nuclear YAP1 expression with moderate to strong intensities in more than 5% of the tumor cells were regarded as positive.

doi:10.1371/journal.pone.0075449.t004

difference among cHC-CCs, HCCs with stemness, and HCCs without stemness: disease-free survival rate was highest in EpCAM(-)/K19(-) HCCs and lowest in cHC-CCs, with EpCAM(+)/K19(+) HCCs falling in between.

In a previous study, transgenic mice with liver-specific YAP1 overexpression were reported to develop primary liver tumors, which morphologically resembled human cHC-CCs.[21] In the present study, YAP1 expression was found in 67% of cHC-CCs, 56% of EpCAM(+)/K19(+) HCCs, and 17% of EpCAM(-)/K19(-) HCCs. Such expression was more frequently found in EpCAM(+)/K19(+) HCCs and cHC-CCs than in EpCAM(-)/K19(-) HCCs. In cHC-CCs, YAP1 expression was associated with poorer histological differentiation, and was more frequently

noted in transitional zones with features of stem/progenitor cells, compared to HCC areas, although this was not statistically significant.

YAP1 is known to have the ability to induce epithelial mesenchymal transition (EMT), the differentiation of polarized epithelial cells to contractile and motile mesenchymal cells.[28] EMT induction by ectopic expression of either Snail or Twist transcription factors was also reported to lead to cancer stem-cell properties in human breast cancer cells.[29] Interestingly, our previous study revealed that HCCs expressing K19 and/or EpCAM show upregulation of EMT-associated genes and more invasive characteristics than those without.[9] In this study, YAP1 expression was significantly higher in HCCs with

Table 5. Independent prognostic factors for disease-free and overall survival by multivariable analysis.

		Disease	-free survival		Overall su	rvival	
Variable	N	HR	95% CI	Р	HR	95% CI	Р
Tumor size							
<4 cm	83	1			1		
≥4 cm	69	1.356	0.781-2.354	0.277	3.448	1.378-8.624	0.008
Preoperative treatment							
No	101	1					
Yes	51	2.063	1.258-3.383	0.004			
Vascular invasion							
Absence	63	1			1		
Presence	89	2.240	1.249-4.015	0.007	7.135	1.646-30.927	0.009
Histologic group				0.133			0.034
EpCAM(-)/K19(-) HCCs	61	1			1		
EpCAM(+)/K19(+) HCCs	35	1.922	0.972-3.801	0.060	16.015	1.959–130.96	0.010
cHC-CCs	56	1.692	0.896-3.196	0.104	13.887	1.761-109.546	0.013

doi:10.1371/journal.pone.0075449.t005

stemness than in those without. Taken together, our results suggest that the Hippo-YAP1 pathway might be involved in the pathogenesis of liver cancers with stemness, such as EpCAM(+)/ K19(+) HCCs and cHC-CCs, which exhibit aggressive biological behavior. Additionally, YAP1 expression has been reported to be related to poor prognosis in several malignancies, including HCC, non-small cell lung cancer, gastric cancer and colorectal cancer.[30–33] In this study, overall survival rate was relatively lower in HCCs and cHC-CCs that expressed YAP1 compared to those that did not, whereas disease free survival showed no difference according to YAP1 expression. Also, YAP1 expression was revealed as a significant prognostic factor affecting overall survival on univariate analysis, but not on multivariate analysis.

In conclusion, this is the first study to provide clinicopathological evidence that YAP1 is more frequently expressed in HCCs expressing stemness-related markers (EpCAM and K19) and in cHC-CCs, compared to HCCs lacking such expression. Our findings suggest that YAP1 expression may contribute to the gain of stemness in HCCs and cHC-CCs, and could be a potential therapeutic target for treatment of these tumors.

References

- Jemal A, Center MM, DeSantis C, Ward EM (2010) Global patterns of cancer incidence and mortality rates and trends. Cancer Epidemiol Biomarkers Prev 19: 1893–1907.
- Goodman ZD, Ishak KG, Langloss JM, Sesterhenn IA, Rabin L (1985) Combined hepatocellular-cholangiocarcinoma. A histologic and immunohistochemical study. Cancer 55: 124–135.
- Theise ND, Nakashima O, Park YN, Nakamura Y (2010) Combined hepatocellular-cholangiocarcinoma. In: Bosman FT, Carneiro F, Hruban RH, Theise ND, editors. WHO classification of tumours of the digestive system. 4th ed. Lyon: International agency for research on cancer. pp. 225–227.
- Zhu Z, Hao X, Yan M, Yao M, Ge C, et al. (2010) Cancer stem/progenitor cells are highly enriched in CD133+CD44+ population in hepatocellular carcinoma. Int J Cancer 126: 2067–2078.
- Yamashita T, Honda M, Nakamoto Y, Baba M, Nio K, et al. (2013) Discrete nature of EpCAM+ and CD90+ cancer stem cells in human hepatocellular carcinoma. Hepatology 57: 1484–1497.
- Yang ZF, Ho DW, Ng MN, Lau CK, Yu WC, et al. (2008) Significance of CD90+ cancer stem cells in human liver cancer. Cancer Cell 13: 153–166.

Supporting Information

Figure S1 Kaplan–Meier's plot analysis for disease-free survival and overall survival in combined hepatocellular-cholangiocarcinomas (cHC-CCs). There was no difference between classical type and intermediate-cell subtype patients in disease-free survival (A) and overall survival rate (B). (TIF)

 Table S1
 Clinicopathological features and YAP1 expression in classical-type and intermediate-cell subtype combined hepatocel-lular-cholangiocarcinomas

 (DOCX)
 (DOCX)

Acknowledgments

The authors would like to thank Anthony Thomas Milliken (Medical Research Support Section, Yonsei University College of Medicine, Seoul, Korea) for his help with English editing.

Author Contributions

Conceived and designed the experiments: YNP HK. Performed the experiments: GJK. Analyzed the data: GJK HK. Contributed reagents/ materials/analysis tools: YNP. Wrote the paper: GJK HK YNP.

- Ma S, Tang KH, Chan YP, Lee TK, Kwan PS, et al. (2010) miR-130b Promotes CD133(+) liver tumor-initiating cell growth and self-renewal via tumor protein 53-induced nuclear protein 1. Cell Stem Cell 7: 694–707.
- Ma YC, Yang JY, Yan LN (2013) Relevant markers of cancer stem cells indicate a poor prognosis in hepatocellular carcinoma patients: a meta-analysis. Eur J Gastroenterol Hepatol.
- Kim H, Choi GH, Na DC, Ahn EY, Kim GI, et al. (2011) Human hepatocellular carcinomas with "Stemness"-related marker expression: keratin 19 expression and a poor prognosis. Hepatology 54: 1707–1717.
- Shan YF, Huang YL, Xie YK, Tan YH, Chen BC, et al. (2011) Angiogenesis and clinicopathologic characteristics in different hepatocellular carcinoma subtypes defined by EpCAM and alpha-fetoprotein expression status. Med Oncol 28: 1012–1016.
- Okamura D, Ohtsuka M, Kimura F, Shimizu H, Yoshidome H, et al. (2008) Ezrin expression is associated with hepatocellular carcinoma possibly derived from progenitor cells and early recurrence after surgical resection. Mod Pathol 21: 847–855.
- 12. Lee JH, Chung GE, Yu SJ, Hwang SY, Kim JS, et al. (2011) Long-term prognosis of combined hepatocellular and cholangiocarcinoma after curative

resection comparison with hepatocellular carcinoma and cholangiocarcinoma. J Clin Gastroenterol 45: $69{-}75.$

- Yin X, Zhang BH, Qiu SJ, Ren ZG, Zhou J, et al. (2012) Combined hepatocellular carcinoma and cholangiocarcinoma: clinical features, treatment modalities, and prognosis. Ann Surg Oncol 19: 2869–2876.
- Huang J, Wu S, Barrera J, Matthews K, Pan D (2005) The Hippo signaling pathway coordinately regulates cell proliferation and apoptosis by inactivating Yorkie, the Drosophila Homolog of YAP. Cell 122: 421–434.
- Kango-Singh M, Singh A (2009) Regulation of organ size: insights from the Drosophila Hippo signaling pathway. Dev Dyn 238: 1627–1637.
- Lian I, Kim J, Okazawa H, Zhao J, Zhao B, et al. (2010) The role of YAP transcription coactivator in regulating stem cell self-renewal and differentiation. Genes Dev 24: 1106–1118.
- Liu AM, Xu MZ, Chen J, Poon RT, Luk JM (2010) Targeting YAP and Hippo signaling pathway in liver cancer. Expert Opin Ther Targets 14: 855–868.
- Halder G, Dupont S, Piccolo S (2012) Transduction of mechanical and cytoskeletal cues by YAP and TAZ. Nat Rev Mol Cell Biol 13: 591–600.
- 19. Liu AM, Xu Z, Luk JM (2012) An update on targeting Hippo-YAP signaling in liver cancer. Expert Opin Ther Targets 16: 243–247.
- Lam-Himlin DM, Daniels JA, Gayyed MF, Dong J, Maitra A, et al. (2006) The hippo pathway in human upper gastrointestinal dysplasia and carcinoma: a novel oncogenic pathway. Int J Gastrointest Cancer 37: 103–109.
- Lee KP, Lee JH, Kim TS, Kim TH, Park HD, et al. (2010) The Hippo-Salvador pathway restrains hepatic oval cell proliferation, liver size, and liver tumorigenesis. Proc Natl Acad Sci U S A 107: 8248–8253.
- Durnez A, Verslype C, Nevens F, Fevery J, Aerts R, et al. (2006) The clinicopathological and prognostic relevance of cytokeratin 7 and 19 expression in hepatocellular carcinoma. A possible progenitor cell origin. Histopathology 49: 138–151.
- Yuan RH, Jeng YM, Hu RH, Lai PL, Lee PH, et al. (2011) Role of p53 and beta-catenin mutations in conjunction with CK19 expression on early tumor recurrence and prognosis of hepatocellular carcinoma. J Gastrointest Surg 15: 321–329.

- Coulouarn C, Cavard C, Rubbia-Brandt L, Audebourg A, Dumont F, et al. (2012) Combined hepatocellular-cholangiocarcinomas exhibit progenitor features and activation of Wnt and TGFbeta signaling pathways. Carcinogenesis 33: 1791–1796.
- Yu XH, Xu LB, Zeng H, Zhang R, Wang J, et al. (2011) Clinicopathological analysis of 14 patients with combined hepatocellular carcinoma and cholangiocarcinoma. Hepatobiliary Pancreat Dis Int 10: 620–625.
- Maeda T, Adachi E, Kajiyama K, Sugimachi K, Tsuneyoshi M (1995) Combined hepatocellular and cholangiocarcinoma: proposed criteria according to cytokeratin expression and analysis of clinicopathologic features. Hum Pathol 26: 956–964.
- Koh KC, Lee H, Choi MS, Lee JH, Paik SW, et al. (2005) Clinicopathologic features and prognosis of combined hepatocellular cholangiocarcinoma. Am J Surg 189: 120–125.
- Overholtzer M, Zhang J, Smolen GA, Muir B, Li W, et al. (2006) Transforming properties of YAP, a candidate oncogene on the chromosome 11q22 amplicon. Proc Natl Acad Sci U S A 103: 12405–12410.
- Mani SA, Guo W, Liao MJ, Eaton EN, Ayyanan A, et al. (2008) The epithelialmesenchymal transition generates cells with properties of stem cells. Cell 133: 704–715.
- Xu MZ, Yao TJ, Lee NP, Ng IO, Chan YT, et al. (2009) Yes-associated protein is an independent prognostic marker in hepatocellular carcinoma. Cancer 115: 4576–4585.
- Wang Y, Dong Q, Zhang Q, Li Z, Wang E, et al. (2010) Overexpression of yesassociated protein contributes to progression and poor prognosis of non-smallcell lung cancer. Cancer Sci 101: 1279–1285.
- Kang W, Tong JH, Chan AW, Lee TL, Lung RW, et al. (2011) Yes-associated protein 1 exhibits oncogenic property in gastric cancer and its nuclear accumulation associates with poor prognosis. Clin Cancer Res 17: 2130–2139.
 Wang L, Shi S, Guo Z, Zhang X, Han S, et al. (2013) Overexpression of YAP
- 33. Wang L, Shi S, Guo Z, Zhang X, Han S, et al. (2013) Overexpression of YAP and TAZ Is an Independent Predictor of Prognosis in Colorectal Cancer and Related to the Proliferation and Metastasis of Colon Cancer Cells. PLoS One 8: e65539.