

## Human T2R38 Bitter Taste Receptor Expression and COVID-19: From Immunity to Prognosis

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### Abstract

**Background:** Bitter taste-sensing type 2 receptor (T2Rs or TAS2Rs) found on ciliated epithelial cells and solitary chemosensory cells have a role in respiratory tract immunity. T2Rs have shown protection against SARS-CoV-2 by enhancing the innate immune response. The purpose of this review is to outline the current sphere of knowledge regarding this association.

**Methods:** A narrative review of the literature was done by searching (T2R38 OR bitter taste receptor) AND (COVID-19 OR SARS-CoV-2) keywords in PubMed and google scholar.

**Results:** T2R38, an isoform of T2Rs encoded by the *TAS2R38* gene, may have a potential association between phenotypic expression of T2R38 and prognosis of COVID-19. Current studies suggest that due to different genotypes and widespread distributions of T2Rs within the respiratory tract and their role in innate immunity, treatment protocols for COVID-19 and other respiratory diseases may change accordingly. Based on the phenotypic expression of T2R38, it varies in innate immunity and host response to respiratory infection, systemic symptoms and hospitalization.

**Conclusion:** This review reveals that patients' innate immune response to SARS-COV-2 could be influenced by T2R38 receptor allelic variations.

**Keywords:** Bitter taste receptors (T2Rs), Coronavirus disease, COVID-19, Infection, T2R38

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### Introduction

The innate immune system is crucial in fighting against Severe Acute Respiratory Syndrome Coronavirus (SARS-CoV-2), as is the case with most infections<sup>1</sup>. Inhaled pathogens and particles naturally pose a constant threat to the respiratory system. Mucociliary Clearance (MCC), which consists of two components, including mucus production and mucus transport, is the primary physical defense against such irritants<sup>2-4</sup>. Debris-laden mucus is transported to the oropharynx by ciliary beating and removed by swallowing or expectoration<sup>2,5</sup>. Various environmental and host stimuli increase ciliary beating rate *via* various secondary messenger routes, such as intracellular Ca<sup>2+</sup> and Nitric Oxide (NO) production<sup>2,6-9</sup>. The wider literature suggests bitter taste-sensing type 2 receptor (T2Rs or TAS2Rs),

found on ciliated epithelial cells and solitary chemosensory cells, have a function in sinonasal immunity<sup>2,10,11</sup>. T2R38 is one of the numerous T2R isoforms found in human motile cilia<sup>1,2</sup>. When T2R38 is stimulated by Propylthiouracil and phenylcarbamide, NO is produced and antimicrobial peptides are released<sup>12</sup>. NO increases MCC and kills pathogens in the human respiratory tract mucosa<sup>12,13</sup>. Two common haplotypes result from three single-nucleotide variants in the gene which encodes T2R38, including the functional variant Proline-Alanine-Valine (PAV) and the nonfunctional variant Alanine-Valine-Isoleucine (AVI)<sup>1,14,15</sup>. When compared to those homozygous for nonfunctional T2R38 (AVI/AVI) and heterozygous for this receptor (PAV/AVI), individuals with functional T2R38 (PAV/

PAV), had fewer gram-negative upper respiratory infections and a better quality of life<sup>16,17</sup>. Several studies have investigated the association between T2R38 phenotype expression and COVID-19 severity and prognosis<sup>1,16,18</sup>. The purpose of this review is to outline the current sphere of knowledge regarding this association.

### Materials and Methods

This study was a narrative review of literature which was done by searching (T2R38 OR bitter taste receptor) AND (COVID-19 OR SARS-CoV-2) keywords. Electronic databases including PubMed and Google scholar was searched to find articles investigating the association of human T2R38 bitter taste receptor expression and COVID-19.

### Results

#### What is T2R38?

The role of bitter taste is protective against toxin ingestion. The bitter taste sense arise from G-Protein Coupled Receptors (GPCRs) found in type II taste receptor cells in the oral taste buds T2Rs, present on extra-oral ciliated epithelial cells and solitary chemosensory cells (first line defence elements in tracheobronchial pathway) that have been linked to sinonasal innate immunity<sup>19-21</sup>. T2Rs are GPCRs which take on various isoforms. There are 25 known T2Rs in the human body. T2R38 is a type of T2Rs that responds to chemicals with thiourea (N-C=S). Furthermore, these receptors can respond to other compounds containing (N=C=S). T2R38 is also one such isoform populating motile cilia in human mucosal linings, including airways, human placenta, and colon<sup>21</sup>. T2R38 is encoded by the *TAS2R38* gene in the human genome expressed as two types of haplotypes, the functional variant PAV and the non-functional variant AVI.

#### Role of T2R38 in innate immunity

T2Rs have little influence on taste perception within the extra-oral airway. When T2Rs bind to their specific agonists, they respond to a plethora of bitter chemicals, including phenylthiocarbamide (PTC), denatonium

benzoate, strychnine, quinine and caffeine<sup>22,23</sup>. Ciliated sinonasal epithelial cells are essential for the upper respiratory tract immunity's initial line of defense. To maintain a healthy sinonasal tract, effective MCC necessitates the coordinated ciliary-driven movement of airway surface fluids, which contains mucus-trapped pathogenic organisms and detritus. Stasis of sinonasal secretions and resulting local inflammation occur when MCC is hindered propagating a susceptibility to infection<sup>4,24-27</sup>.

Calcium-driven NO generation is a T2R38 innate immunological response. This calcium and NO signaling incorporates two classical components of the taste signaling cascade using a phospholipase C isoform (PLC2)<sup>28,29</sup>, and the TRPM5 ion channel in type II taste cells. NO damages the intracellular components of infectious bacteria, and raises ciliary beat frequency by acting on protein kinase G and guanylyl cyclase, resulting in increased MCC activity<sup>29,30</sup>. The clearance of mucus-trapped pathogens and dispersion of antimicrobial chemicals in response to pathogens is accelerated by ciliary beat frequency<sup>30,31</sup>. Solitary chemosensory cells are nonciliated epithelial cells which express both sweet (T1R2/3) and bitter (T2R) receptors. Although stimulation of T2Rs on ciliated epithelial cells causes a Ca<sup>2+</sup>-dependent NO response. T2R stimulation on solitary chemosensory cells results in Ca<sup>2+</sup> propagation throughout gap junctions into ciliated cells. This releases antimicrobial compounds such as defensins 1 and 2, lactoferrin and others<sup>20,32</sup>. Current literature explores oral taste sensitivity to measure extra-oral taste receptor function, where basic taste test may show changes in sinonasal immune functions<sup>32,33</sup>. The role of T2R38 receptor against pathogens is illustrated in figure 1.

#### Role of T2R38 in respiratory infections and diseases

Quorum Sensing (QS) is a mechanism by which *Pseudomonas aeruginosa* (*P. aeruginosa*) uses signal molecules to regulate gene expression according to population density. Molecules are sensed by the ciliary airway T2R38 receptors, prompting changes in NO

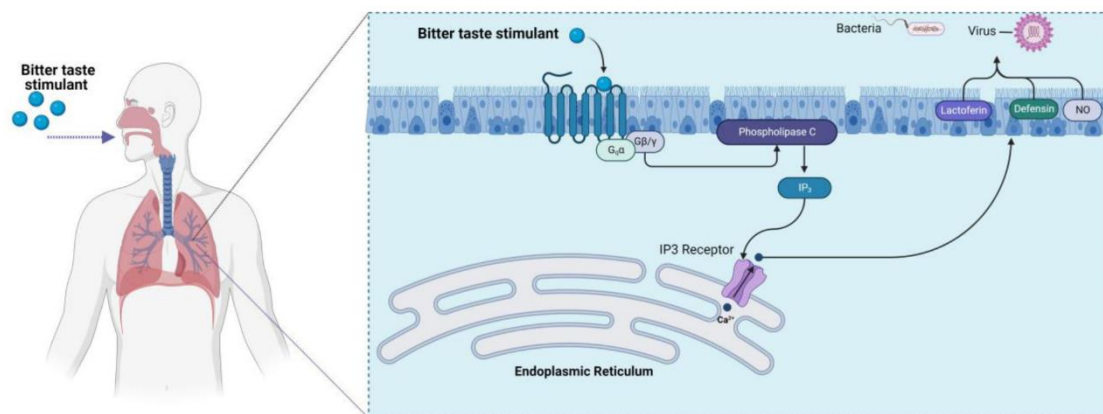


Figure 1. The role of T2R38 receptor against pathogens.

production and ciliary beat frequency, possibly leading to enhanced bacterial clearance<sup>2</sup>. Castaldo *et al*<sup>34</sup> demonstrated frequency of PAV allele was significantly reduced in Cystic Fibrosis (CF) patients with nasal polyps requiring surgery and also in CF patients with chronic pulmonary *P. aeruginosa* colonization. This confirms the relationship between the altered function of the T2R38 receptor and the risk of lower and upper airway infections and chronic Rhino-Sinusitis (CRS). AVI/AVI genotype is also an independent risk factor for patients with medically recalcitrant CRS<sup>2,35</sup>. Adapa *et al* demonstrated that patients with failing medical treatment for CRS undergoing Functional Endoscopic Sinus Surgery (FESS) revealed frequency of AVI/AVI genotype which was higher in this cohort than in the control population. They also compared the distribution of age, sex, aspirin sensitivity, diabetes, asthma, allergies, nasal polyposis and smoking status in CRS patients requiring FESS. Here, PAV/PAV was over-represented in CRS patients with allergies, asthma, aspirin sensitivity, nasal polyposis and diabetes. Evidence from mouse and human studies suggests that TAS2Rs expressed on airway smooth muscles promote muscle relaxation and bronchodilation, and also regulates migration of white blood cells especially eosinophils<sup>35-37</sup>. TAS2R has the potential to be considered as a therapeutic target in obstructive lung disease. Further research is necessary to determine which particular TAS2R receptors are of importance<sup>36</sup>. There is paucity within the literature showing correlation between T2R38 genotype and chronic lung conditions.

#### Role of T2R38 in coronaviruses and COVID-19

T2R38 is activated by phenylthiocarbamide and propylthiouracil<sup>38</sup>. NO is generated when T2R38 is stimulated by agonists, which promotes MCC, and destroys pathogenic material in the human respiratory mucosa<sup>10</sup>. Åkerström *et al*<sup>13</sup> discovered that NO suppresses SARS-CoV replication by two separate pathways. NO and its derivatives reduce the palmitoylation of newly generated spike proteins, affecting its fusion with its cognate receptor, ACE-2. The cellular entry receptor for SARS-CoV-2 is Angiotensin-Converting Enzyme 2 (ACE-2), is widely present in the human lower respiratory tract, as well as other tissues<sup>39</sup>. NO and its metabolites diminish viral RNA production during the initial phases of viral replication explained by the cysteine proteases encoded in Orf1a gene of SARS-CoV genome sequence. Phenotypic perception of phenylthiouracil bitter sensitivity throughout the course of a person's life has been assessed, and found that sensitivity to the T2R38 agonist, phenylthiouracil, decreases as they get older<sup>40,41</sup>. This phenomenon appears to be more common in heterozygotes. Davies *et al*<sup>42</sup> estimated that susceptibility to COVID-19 infection in individuals younger than 20 years is approximately half that of adult people more than 20 years of age. Those symptomatic accounted for 21% of those aged 10 to 19 years, and 69% in those aged 70 years

and older. This used an age-structured mathematical formulation of data from 6 countries illustrating T2R phenotypic expression declines with ageing. A retrospective, cross-sectional study evaluating 100 adult patients positive for SARS-CoV showed that all 21 patients (100%) requiring hospital admission were classified as non-tasters (AVI/AVI), whereas 79 non-hospitalised patients with mild to moderate symptoms were classified as tasters (PAV/AVI) and no supertasters (PAV/PAV). This may necessitate a potential association between outcome and clinical course of COVID-19 and phenotypic expression of T2R38 in relation to hospitalization<sup>16</sup>.

#### Discussion

In the extra-oral airway, there are bitter taste receptors on various cells with T2Rs present in the epithelial ciliated cells, which are involved in innate immunity when attached to their agonist. Activation of calcium-based T2R38 produces NO that plays three vital roles through its biocidal function:

1. it damages the components inside the microbial cells,
2. increases Ciliary Beat Frequency (CBF), and
3. increases MCC<sup>43-46</sup>. It has been proposed that evaluating treatment regimens for COVID-19 patients and other respiratory tract infections in accordance with their *TAS2R38* alleles may lead to a beneficial course of action<sup>18</sup>. Akerstrom *et al*<sup>47</sup> showed that NO and its derivatives could reduce the initial production of viral RNA by inhibiting spike protein. Many antibiotics, including macrolides, affect the host's innate immune response<sup>48,49</sup>. One of these off-target effects of bitter-tasting antibiotics, notably Azithromycin, activates T2Rs<sup>50,51</sup>. Taste status when using Azithromycin in COVID-19 management has not been assessed profoundly. Quinine derivatives<sup>52</sup> are known agonists of T2Rs, however their effectiveness against COVID-19 has not been thoroughly examined. The effects of Azithromycin were associated with the taste status (measured by the T2R38 phenotype), but Azithromycin was not recognized as a T2R38 agonist<sup>53</sup>. Jaggupilli *et al*<sup>54</sup>, showed the highest bitterness score recorded *via* E-tongue activates Taste 2 Receptor Member 4 (T2R4). In another research evaluating drug responses based on T2R38, Caly *et al* purported that Ivermectin reduced the viral load up to 5,000-fold in culture within 48 *hr*<sup>55</sup>. However, the mechanism of Ivermectin against COVID-19 remained unknown. Patients display a variety of clinical features of COVID-19 depending on their different genetic structures. Taha *et al* demonstrated there is a clear correlation between the *T2R38* phenotype and severe acute respiratory syndrome coronavirus (SARS-CoV-2) symptoms duration and severity<sup>1,16</sup>. Supertasters with high levels of *T2R38* expression showed localized symptoms in the upper respiratory tract. Tasters with moderate levels of *T2R38* displayed local and some mild to moderate generalized symptoms such as mild fever. In contrast, non-taster

Table 1. Summary of studies about the role of T2R38 receptor in COVID-19

Author	Measured outcome	Result
Barham <i>et al</i> (16) 2020	Clinical course of SARS-CoV-2	All patients requiring hospitalization were reported as nontasters, whereas all other patients, not requiring admission were reported as tasters (p<0.001)
Risso <i>et al</i> (56) 2022	Assessing the frequency of PAV and AVI alleles in COVID-19 patients with severe or non-severe symptoms compared to healthy subjects	No significant relationship between TAS2R38 genotypes and presence or the severity of COVID-19 infection
Castaldo <i>et al</i> (34) 2020	Sinonasal disease severity and pulmonary P. a colonization in CF patients compared to controls	Frequency of the PAV allele was significantly lower in CF patients with nasal polyposis requiring surgery and, in CF patients with chronic pulmonary P. a colonization
Adappa <i>et al</i> (35) 2014	Correlation of TAS2R38 genotype with CRS	The frequency of PAV/PAV genotype is lower in patients failing medical therapy, necessitating FESS compared to controls (p=0.0383)

people, who did not express *T2R38* alleles, showed more severe infections with generalized symptoms. Intrinsic immunity, in the form of T2R38, acts as a shield against upper respiratory tract pathogens<sup>53</sup>. Supertasters include 25% of the population and account for the majority of asymptomatic carriers. Non-taster people account for 25% of the population and usually have severe symptoms. 50% of the population are tasters and can often recover from COVID-19 infection<sup>1</sup>. Recent studies have demonstrated that possessing *T2R38* alleles (taster) may operate as a protective factor against SARS-CoV-2 infection, while non-tasters may represent a risk factor. According to Risso *et al*'s research<sup>56</sup> that investigated the frequency of tasters and non-tasters in 54 COVID-19 patients in comparison to healthy individuals, a direct correlation between *T2R38* phenotypes and vulnerability to show mild or severe symptoms of SARS-CoV-2 infection based on *T2R38* allele presentation could not be found. A summary of recent studies regarding association of T3R38 receptor and COVID-19 is available in table 1.

### Conclusion

This review reveals that patients' innate immune response to SARS-COV-2 was influenced by T2R38 receptor allelic variations. The clinical outcome of patients with SARS-COV-2 infection was correlated with the T2R phenotype. To lessen the severe clinical symptoms linked to the deteriorating respiratory system, cytokine storm, hyper-reactive immune responses, and renal and cardiovascular issues on the SARS-COV-2 patients, prospective bitter agonists of natural origin or derived bio-active should be targeted. This study suggests that COVID-19 treatment protocols may be modified based on T2R phenotypic expression because T2Rs are widely distributed in the respiratory tract and a variety of treatment protocols currently available around the world have yielded conflicting results regarding their efficacy. However, sufficient clinical trials must be conducted to demonstrate the effectiveness

of targeting the bitter taste receptors to control the SARS-COV-2 outbreak and its associated pathogenicity on the worldwide population. Additionally, the wide diversity of the *T2SR38* gene polymorphism and its potential link to mortality raises questions about the effectiveness of vaccine development programs among various racial groups.

### Conflict of Interest

The authors declare that there is no conflict of interest.

### References

1. Barham HP, Taha MA, Broyles ST, Stevenson MM, Zito BA, Hall CA. Association between bitter taste receptor phenotype and clinical outcomes among patients with COVID-19. *JAMA Netw Open* 2021;4(5):e2111410-e.
2. Lee RJ, Xiong G, Kofonow JM, Chen B, Lysenko A, Jiang P, et al. T2R38 taste receptor polymorphisms underlie susceptibility to upper respiratory infection. *J Clin Invest* 2012;122(11):4145-59.
3. Sade J, Eliezer N, Silberberg A, Nevo A. The role of mucus in transport by cilia. *Am Rev Respir Dis* 1970; 102(1):48-52.
4. Sleigh MA, Blake JR, Liron N. The propulsion of mucus by cilia. *Am Rev Respir Dis* 1988;137(3):726-41.
5. Antunes MB, Gudis DA, Cohen NA. Epithelium, cilia, and mucus: their importance in chronic rhinosinusitis. *Immunol Allergy Clin North Am* 2009;29(4):631-43.
6. Korngreen A, Priel Z. Simultaneous measurement of ciliary beating and intracellular calcium. *Biophys J* 1994; 67(1):377.
7. Salathe M, Bookman RJ. Coupling of [Ca<sup>2+</sup>]<sub>i</sub> and ciliary beating in cultured tracheal epithelial cells. *J Cell Sci* 1995;108(Pt 2):431-40.
8. Schipor I, Palmer JN, Cohen AS, Cohen NA. Quantification of ciliary beat frequency in sinonasal epithelial cells using differential interference contrast microscopy and high-speed digital video imaging. *Ame J Rhinol* 2006;20(1):124-7.

9. Uzlaner N, Priel Z. Interplay between the NO pathway and elevated [Ca<sup>2+</sup>]<sub>i</sub> enhances ciliary activity in rabbit trachea. *J Physiol* 1999;516(1):179-90.
10. Lee RJ, Kofonow JM, Rosen PL, Siebert AP, Chen B, Doghramji L, et al. Bitter and sweet taste receptors regulate human upper respiratory innate immunity. *J Clin Invest* 2014;124(3):1393-405.
11. Workman AD, Palmer JN, Adappa ND, Cohen NA. The role of bitter and sweet taste receptors in upper airway immunity. *Curr Allergy Asthma Rep* 2015;15(12):1-8.
12. Parsa S, Mogharab V, Ebrahimi M, Ahmadi SR, Shahi B, Mehramiz NJ, et al. COVID-19 as a worldwide selective event and bitter taste receptor polymorphisms: An ecological correlational study. *Int J Biol Macromol* 2021; 177:204-10.
13. Åkerström S, Gunalan V, Keng CT, Tan Y-J, Mirazimi A. Dual effect of nitric oxide on SARS-CoV replication: viral RNA production and palmitoylation of the S protein are affected. *Virology* 2009;395(1):1-9.
14. Bufe B, Breslin PA, Kuhn C, Reed DR, Tharp CD, Slack JP, et al. The molecular basis of individual differences in phenylthiocarbamide and propylthiouracil bitterness perception. *Curr Biol* 2005;15(4):322-7.
15. Meyerhof W, Batram C, Kuhn C, Brockhoff A, Chudoba E, Bufe B, et al. The molecular receptive ranges of human TAS2R bitter taste receptors. *Chem Senses* 2010; 35(2):157-70.
16. Barham HP, Taha MA, Hall CA, editors. Does phenotypic expression of bitter taste receptor T2R38 show association with COVID-19 severity? *Int Forum Allergy Rhinol* 2020 2020 Nov;10(11):1255-7.
17. Maina IW, Workman AD, Cohen NA. The role of bitter and sweet taste receptors in upper airway innate immunity: recent advances and future directions. *World J Otorhinolaryngology Head Neck Surg* 2018;4(3):200-8.
18. Taha MA, Hall CA, Shortess CJ, Rathbone RF, Barham HP. Treatment protocol for COVID-19 based on T2R phenotype. *Viruses* 2021;13(3):503.
19. Tizzano M, Gulbransen BD, Vandenbeuch A, Clapp TR, Herman JP, Sibhatu HM, et al. Nasal chemosensory cells use bitter taste signaling to detect irritants and bacterial signals. *Proc Natl Acad Sci USA* 2010;107(7):3210-5.
20. Barham HP, Cooper SE, Anderson CB, Tizzano M, Kingdom TT, Finger TE, et al., editors. Solitary chemosensory cells and bitter taste receptor signaling in human sinonasal mucosa. *Int Forum Allergy Rhinol* 2013 Jun;3(6):450-7.
21. Tran HTT, Herz C, Ruf P, Stetter R, Lamy E. Human T2R38 bitter taste receptor expression in resting and activated lymphocytes. *Front Immunol* 2018:2949.
22. Brockhoff A, Behrens M, Massarotti A, Appendino G, Meyerhof W. Broad tuning of the human bitter taste receptor hTAS2R46 to various sesquiterpene lactones, clerodane and labdane diterpenoids, strychnine, and denatonium. *J Agric Food Chem* 2007;55(15):6236-43.
23. Hansen JL, Reed DR, Wright MJ, Martin NG, Breslin PA. Heritability and genetic covariation of sensitivity to PROP, SOA, quinine HCl, and caffeine. *Chem Senses* 2006;31(5):403-13.
24. Parker D, Prince A. Innate immunity in the respiratory epithelium. *Am J Respir Cell Mol Biol* 2011;45(2):189-201.
25. Kato A, Schleimer RP. Beyond inflammation: airway epithelial cells are at the interface of innate and adaptive immunity. *Current Opin Immunol* 2007;19(6):711-20.
26. Patel NN, Kohanski MA, Maina IW, Triantafillou V, Workman AD, Tong CC, et al., editors. Solitary chemosensory cells producing interleukin-25 and group-2 innate lymphoid cells are enriched in chronic rhinosinusitis with nasal polyps. *International Forum of Allergy & Rhinology* 2018 May 9;10.1002/alr.22142.
27. Kohanski MA, Workman AD, Patel NN, Hung L-Y, Shtraks JP, Chen B, et al. Solitary chemosensory cells are a primary epithelial source of IL-25 in patients with chronic rhinosinusitis with nasal polyps. *J Allergy Clin Immunol* 2018;142(2):460-9. e7.
28. Mennella JA, Spector AC, Reed DR, Coldwell SE. The bad taste of medicines: overview of basic research on bitter taste. *Clin Ther* 2013;35(8):1225-46.
29. Yamamoto K, Ishimaru Y, editors. Oral and extra-oral taste perception. *Semin Cell Dev Biol* 2013 Mar;24(3): 240-6.
30. Iwata S, Yoshida R, Ninomiya Y. Taste transductions in taste receptor cells: basic tastes and moreover. *Curr Pharm Des* 2014;20(16):2684-92.
31. Sollai G, Melis M, Pani D, Cosseddu P, Usai I, Crnjar R, et al. First objective evaluation of taste sensitivity to 6-n-propylthiouracil (PROP), a paradigm gustatory stimulus in humans. *Sci Rep* 2017;7:40353.
32. Mosimann BL, White MV, Hohman RJ, Goldrich MS, Kaulbach HC, Kaliner MA. Substance P, calcitonin gene-related peptide, and vasoactive intestinal peptide increase in nasal secretions after allergen challenge in atopic patients. *J Allergy Clin Immunol* 1993;92(1 Pt 1): 95-104.
33. Farquhar DR, Kovatch KJ, Palmer JN, Shofer FS, Adappa ND, Cohen NA. Phenylthiocarbamide taste sensitivity is associated with sinonasal symptoms in healthy adults. *Int Forum Allergy Rhinol* 2015 Feb;5(2):111-8.
34. Castaldo A, Cerneria G, Iacotucci P, Cimbalo C, Gelzo M, Comegna M, et al. TAS2R38 is a novel modifier gene in patients with cystic fibrosis. *Sci Rep* 2020;10(1):1-6.
35. Adappa ND, Zhang Z, Palmer JN, Kennedy DW, Doghramji L, Lysenko A, et al. The bitter taste receptor T2R38 is an independent risk factor for chronic rhinosinusitis requiring sinus surgery. *Int Forum Allergy Rhinol* 2014 Jan;4(1):3-7.
36. Liggett SB. Bitter taste receptors in the wrong place: novel airway smooth muscle targets for treating asthma. *Trans Am Clin Climatol Assoc* 2014;125:64-74; discussion 74-5.
37. Sharma P, Yi R, Nayak AP, Wang N, Tang F, Knight MJ, et al. Bitter taste receptor agonists mitigate features of allergic asthma in mice. *Sci Rep* 2017;7(1):1-14.

38. Kim U, Drayna D. Genetics of individual differences in bitter taste perception: lessons from the PTC gene. *Clin Genet* 2005;67(4):275-80.
39. Walls AC, Park Y-J, Tortorici MA, Wall A, McGuire AT, Veelsler D. Structure, function, and antigenicity of the SARS-CoV-2 spike glycoprotein. *Cell* 2020;181(2):281-92. e6.
40. Mennella JA, Pepino MY, Duke FF, Reed DR. Age modifies the genotype-phenotype relationship for the bitter receptor TAS2R38. *BMC Genet* 2010;11:60.
41. Whissell-Buechy D. Effects of age and sex on taste sensitivity to phenylthiocarbamide (PTC) in the Berkeley Guidance sample. *Chemical Senses* 1990;15(1):39-57.
42. Davies NG, Klepac P, Liu Y, Prem K, Jit M, CMMID COVID-19 working group, et al. Age-dependent effects in the transmission and control of COVID-19 epidemics. *Nat Med* 2020;26(8):1205-11.
43. Lee RJ, Xiong G, Kofonow JM, Chen B, Lysenko A, Jiang P, et al. T2R38 taste receptor polymorphisms underlie susceptibility to upper respiratory infection. *J Clin Invest* 2012;122(11):4145-59.
44. Parker D, Prince A. Innate immunity in the respiratory epithelium. *Am J Respir Cell Mol Biol* 2011;45(2):189-201.
45. Zhang Y, Hoon MA, Chandrashekar J, Mueller KL, Cook B, Wu D, et al. Coding of sweet, bitter, and umami tastes: different receptor cells sharing similar signaling pathways. *Cell* 2003;112(3):293-301.
46. Iwata S, Yoshida R, Ninomiya Y. Taste transductions in taste receptor cells: basic tastes and moreover. *Curr Pharm Des* 2014;20(16):2684-92.
47. Akerstrom S, Gunalan V, Keng CT, Tan YJ, Mirazimi A. Dual effect of nitric oxide on SARS-CoV replication: viral RNA production and palmitoylation of the S protein are affected. *Virology* 2009;395(1):1-9.
48. Culic O, Erakovic V, Parnham MJ. Anti-inflammatory effects of macrolide antibiotics. *Eur J Pharmacol* 2001;429(1-3):209-29.
49. Good JT, Jr., Rollins DR, Martin RJ. Macrolides in the treatment of asthma. *Curr Opin Pulm Med* 2012;18(1):76-84.
50. Rollins DR, Beuther DA, Martin RJ. Update on infection and antibiotics in asthma. *Curr Allergy Asthma Rep* 2010;10(1):67-73.
51. Gao X, Ray R, Xiao Y, Ishida K, Ray P. Macrolide antibiotics improve chemotactic and phagocytic capacity as well as reduce inflammation in sulfur mustard-exposed monocytes. *Pulm Pharmacol Ther* 2010;23(2):97-106.
52. Savarino A, Boelaert JR, Cassone A, Majori G, Cauda R. Effects of chloroquine on viral infections: an old drug against today's diseases? *Lancet Infect Dis* 2003;3(11):722-7.
53. Taha MA, Hall CA, Shortess CJ, Rathbone RF, Barham HP. Treatment protocol for COVID-19 based on T2R phenotype. *Viruses* 2021;13(3):503.
54. Jaggupilli A, Singh N, De Jesus VC, Gounni MS, Dhanaraj P, Chelikani P. Chemosensory bitter taste receptors (T2Rs) are activated by multiple antibiotics. *FASEB J* 2019;33(1):501-17.
55. Caly L, Druce JD, Catton MG, Jans DA, Wagstaff KM. The FDA-approved drug ivermectin inhibits the replication of SARS-CoV-2 in vitro. *Antiviral Res* 2020;178:104787.
56. Risso D, Carmagnola D, Morini G, Pellegrini G, Canciani E, Antinucci M, et al. Distribution of TAS2R38 bitter taste receptor phenotype and haplotypes among COVID-19 patients. *Sci Rep* 2022;12(1):7381.