

## 

## Draft Genome Sequence of Mercury-Resistant *Pseudomonas* putida Strain DRA525

Kevin Drace,<sup>a</sup> Stephanie Giangiuli,<sup>b</sup> Claire LeSar,<sup>a</sup> Adam M. Kiefer<sup>b</sup>

<sup>a</sup>Department of Biology, Birmingham-Southern College, Birmingham, Alabama, USA <sup>b</sup>Department of Chemistry, Mercer University, Macon, Georgia, USA

**ABSTRACT** We report here the draft genome sequence of *Pseudomonas putida* strain DRA525, isolated from mercury-contaminated soil. This strain shows resistance to mercury and multiple antibiotics, and its genome sequence contains several gene sets known to confer resistance to heavy metals enzymatically and through multidrug efflux pumps.

Pseudomonas putida is generally associated with the soil environment and has been extensively studied due to its ability to tolerate and/or metabolize a variety of environmental contaminants, including some heavy metals (1, 2). *P. putida* strain DRA525 was isolated from soil sampled near an artisanal gold mining camp in Manica Province, Mozambique, where mercury is commonly used in the gold mining process. This strain was selected for further investigation because of its resistance to HgCl<sub>2</sub> (100  $\mu$ M) and multiple antibiotics, a commonly reported correlation (3–5). The draft genome sequence of *P. putida* strain DRA525 has the potential to provide information useful in addressing questions related to the correlation of heavy metal and antibiotic resistance and the contributions to the mercury cycle made by similar bacteria in highly contaminated environments.

The draft genome sequence of *P. putida* DRA525 was generated by ACGT, Inc., (Wheeling, IL) using the Illumina MiSeq platform derived from combined paired-end and mate-paired libraries of 3,105,539 and 5,194,923 total read pairs, respectively. Adaptors were trimmed and short reads filtered using Trim Galore version 0.3.7 (6) and Sickle version 1.33 (7). Trimmed and filtered reads were assembled using SPAdes version 3.5 (8) into a single closed contig sequence of 6,267,599 bases, with a G+C content of 63%. The draft assembly was annotated using the Rapid Annotations with Subsystems Technology (RAST) server (9), which identified 5,571 open reading frames, and 78 tRNAs were identified using tRNAscan-SE 2.0 (10).

The average nucleotide identity (ANI) was calculated through one-way ANI and two-way ANI between *P. putida* DRA525 and five *P. putida* strains, as previously described (11). The ANI values range from 86.8% (strain W619) to 96.1% (strain S16), with values of 89.6% (strain F1), 89.8% (strain KT2440), and 95.3% (strain HB3267) for the remaining comparisons. As expected, we confirmed the presence of the mercuric resistance (*mer*) operon (*merD*, *merA*, *merC*, *merP*, *merT*, and *merR*) in this strain. In addition, two copies of a gene set containing *arsH*, *acr3*, *arsC*, and *arsR* suggest arsenic resistance (12, 13), and multiple copies of genes related to the CzcC family of heavy-metal RND efflux outer membrane proteins suggest resistance to cadmium, cobalt, and zinc, among others (14). The MexE-MexF-OprN multidrug efflux system was also identified in the genome, which likely contributes to this strain's potential to tolerate the stress of heavy metals and multiple antibiotics (15, 16).

**Accession number(s).** This draft genome sequence has been deposited in GenBank under the accession number CP018743.

Received 26 March 2018 Accepted 26 March 2018 Published 3 May 2018

**Citation** Drace K, Giangiuli S, LeSar C, Kiefer AM. 2018. Draft genome sequence of mercuryresistant *Pseudomonas putida* strain DRA525. Genome Announc 6:e00370-18. https://doi .org/10.1128/genomeA.00370-18.

**Copyright** © 2018 Drace et al. This is an openaccess article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Kevin Drace, kmdrace@bsc.edu.

## REFERENCES

- Nelson KE, Weinel C, Paulsen IT, Dodson RJ, Hilbert H, Martins dos Santos VAP, Fouts DE, Gill SR, Pop M, Holmes M, Brinkac L, Beanan M, DeBoy RT, Daugherty S, Kolonay J, Madupu R, Nelson W, White O, Peterson J, Khouri H, Hance I, Lee PC, Holtzapple E, Scanlan D, Tran K, Moazzez A, Utterback T, Rizzo M, Lee K, Kosack D, Moestl D, Wedler H, Lauber J, Stjepandic D, Hoheisel J, Straetz M, Heim S, Kiewitz C, Eisen J, Timmis KN, Dusterhoft A, Tummler B, Fraser CM. 2002. Complete genome sequence and comparative analysis of the metabolically versatile *Pseudomonas putida* KT2440. Environ Microbiol 4:799–808. https://doi.org/10.1046/j .1462-2920.2002.00366.x.
- Loh KC, Cao B. 2008. Paradigm in biodegradation using *Pseudomonas putida*—a review of proteomics studies. Enzyme Microb Technol 43: 1–12. https://doi.org/10.1016/j.enzmictec.2008.03.004.
- Baker-Austin C, Wright MS, Stepanauskas R, McArthur JV. 2006. Coselection of antibiotic and metal resistance. Trends Microbiol 14: 176–182. https://doi.org/10.1016/j.tim.2006.02.006.
- Gullberg E, Albrecht LM, Karlsson C, Gullberg E, Albrecht LM, Karlsson C, Sandegren L, Andersson DI. 2014. Selection of a multidrug resistance plasmid by sublethal levels of antibiotics and heavy metals. mBio 5:e01918-14. https://doi.org/10.1128/mBio.01918-14.
- Di Cesare A, Eckert EM, D'Urso S, Bertoni R, Gillan DC, Wattiez R, Corno G. 2016. Co-occurrence of integrase 1, antibiotic and heavy metal resistance genes in municipal wastewater treatment plants. Water Res 94: 208–214. https://doi.org/10.1016/j.watres.2016.02.049.
- 6. Krueger F. 2015. Trim Galorel: a wrapper tool around Cutadapt and FastQC to consistently apply quality and adapter trimming to FastQ files. http://www.bioinformatics.babraham.ac.uk/projects/trim\_galore/.
- Joshi NA, Fass JN. 2011. Sickle: a sliding-window, adaptive, quality-based trimming tool for FastQ files (version 133). https://github.com/najoshi/ sickle.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new

genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.

- Aziz RK, Bartels D, Best A, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST Server: Rapid Annotations using Subsystems Technology. BMC Genomics 9:75–15. https://doi.org/10.1186/1471-2164-9-75.
- Lowe TM, Chan PP. 2016. tRNAscan-SE on-line: integrating search and context for analysis of transfer RNA genes. Nucleic Acids Res 44: W54–W57. https://doi.org/10.1093/nar/gkw413.
- Goris J, Konstantinidis KT, Klappenbach JA, Coenye T, Vandamme P, Tiedje JM. 2007. DNA-DNA hybridization values and their relationship to whole-genome sequence similarities. Int J Syst Evol Microbiol 57:81–91. https://doi.org/10.1099/ijs.0.64483-0.
- Fu HL, Meng Y, Ordóñez E, Villadangos AF, Bhattacharjee H, Gil JA, Mateos LM, Rosen BP. 2009. Properties of arsenite efflux permeases (Acr3) from Alkaliphilus metalliredigens and Corynebacterium glutamicum. J Biol Chem 284:19887–19895. https://doi.org/10.1074/jbc.M109 .011882.
- Tsai SL, Singh S, Chen W. 2009. Arsenic metabolism by microbes in nature and the impact on arsenic remediation. Curr Opin Biotechnol 20:659–667. https://doi.org/10.1016/j.copbio.2009.09.013.
- Hassan MT, van der Lelie D, Springael D, Römling U, Ahmed N, Mergeay M. 1999. Identification of a gene cluster, czr, involved in cadmium and zinc resistance in *Pseudomonas aeruginosa*. Gene 238:417–425. https:// doi.org/10.1016/S0378-1119(99)00349-2.
- Köhler T, Michéa-Hamzehpour M, Henze U, Gotoh N, Curty LK, Pechère JC. 1997. Characterization of MexE-MexF-OprN, a positively regulated multidrug efflux system of *Pseudomonas aeruginosa*. Mol Microbiol 23: 345–354. https://doi.org/10.1046/j.1365-2958.1997.2281594.x.
- Nies DH. 2003. Efflux-mediated heavy metal resistance in prokaryotes. FEMS Microbiol Rev 27:313–339. https://doi.org/10.1016/S0168-6445 (03)00048-2.