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**Article** 

# Exposure to Porphyromonas gingivalis and Modifiable Risk **Factors Modulate Risk for Early Diabetic Retinopathy**

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**Purpose:** We hypothesized that exposure to *Porphyromonas gingivalis (Pg)* increases the risk for early diabetic retinopathy (DR) and that the risk can be modulated.

Methods: We identified 116 early DR cases, and 116 non-DR controls were selected randomly by frequency matching for age, sex, race, and education from the US Third National Health and Nutrition Examination Survey. DR was assessed using non-mydriatic fundus photographs and graded by trained graders using the Modified Airlie House Classification scheme and the Early Treatment for Diabetic Retinopathy Study severity scale. Serum Pg immunoglobulin G (IgG) antibody (Ab) was measured in enzymelinked immunosorbent assay units. Logistic regression was used to relate serum Pg IgG Ab levels to the risk for early DR.

**Results:** Per tenfold increase in Pg IgG Ab levels, there was an over 60% increased risk for early DR (odds ratio = 1.64; 95% confidence interval, 1.36-1.97), and a linear trend was noted for the estimated probabilities of early DR at various Pg IgG Ab levels (P for trend = 0.0053). The analysis also suggested that moderate alcohol consumption (less than 12 drinks in the past 12 months; P for interaction = 0.0003) and maintaining a normal serum glycated hemoglobulin level (HbA1c  $\leq$  5.7%; *P* for interaction < 0.0001) helped reduce the Pg-related DR risk.

**Conclusions:** The increased *Pg*-related DR risk could be alleviated by managing alcohol consumption and maintaining a normal blood glucose level.

Translational Relevance: Findings from this study provide new directions for developing novel therapeutics and prevention strategies for DR.

## Introduction

The global prevalence of diabetes mellitus is predicted to increase dramatically in the coming decades, from an estimated 382 million in 2013 to 592 million by 2035.<sup>1</sup> Patients with diabetes suffer many life-limiting and life-threatening complications. including macrovascular-related stroke, ischemic heart disease, and peripheral vascular disease and/or microvascular-related retinopathy, neuropathy, nephropathy, and periodontal disease.<sup>2</sup> Periodontitis is a frequent complication that affects up to 70% of the type 2 diabetic population.<sup>3</sup> As the most common microvascular complication of diabetes.<sup>4</sup> diabetic retinopathy (DR) is a burgeoning problem globally. The condition currently affects almost 100 million people worldwide. Estimates for 1990 and 2010 suggest that DR-related visual impairment and blindness increased by 64% and 27%, respectively.<sup>5</sup> DR falls into two broad categories: the earlier stage of nonproliferative diabetic retinopathy (NPDR) and the advanced stage of proliferative diabetic retinopathy. Current therapeutic paradigms for DR are invasive and destructive and focus on arresting the advanced stages of DR without curing the disease. Therefore, although there is an unmet need for noninvasive, nondestructive, longer lasting treatment options (because prevention

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is better than cure), early prevention strategies that address multiple risk factors are particularly needed for DR.<sup>6</sup>

Mucosal surfaces, including the oral mucosa, are colonized by a complex and dynamic microbial ecosystem that has important implications in human health and disease.<sup>7,8</sup> Porphyromonas gingivalis (Pg) is an asaccharolytic anaerobe frequently associated with periodontal disease.<sup>9</sup> In the oral cavity, this Gramnegative anaerobe colonizes the gingival sulcus in low numbers in health and comprises a significant proportion of the microbiota of the periodontal pocket in disease, contributing to a multispecies microbiome that eventually results in alveolar bone loss.<sup>10</sup> Recent studies suggest that, despite its low abundance, Pg may play a key role in periodontitis by contributing to a shift in the commensal microbiota and dysbiosis.<sup>7</sup> Pg elicits a systemic inflammatory response resulting in elevated levels of various inflammatory mediators.<sup>11</sup> This Pgrelated systemic inflammation has been suggested to increase the risk for several systemic diseases, such as atherosclerosis,<sup>12–14</sup> rheumatoid arthritis,<sup>15,16</sup> neurode-generative diseases,<sup>17–20</sup> and diabetes,<sup>21</sup> providing a vivid example of how disturbances in the commensal microbiome can impact other aspects of human health and disease in sites remote from the site of colonization. In addition, recent studies have identified Pg in the brain of Alzheimer's disease patients and demonstrated that oral Pg infection in mice resulted in brain colonization and increased production of a major component in amyloid plaques,<sup>22,23</sup> suggesting that Pg could access and spread into the human neural system and induce neuroinflammatory damages.

Our previous studies suggested a significant association between a Pg-dominant microbiota and increased diabetes-related mortality.<sup>24</sup> We therefore hypothesized that infection of Pg increases the risk for DR. In this study, we tested this hypothesis by relating serum immunoglobulin G (IgG) antibody (Ab) to Pg to the risk for early DR in a case-control study from a representative sample of the US population.

## Methods

#### **Study Cohort**

The Third National Health and Nutrition Examination Survey (NHANES III) was performed between 1988 and 1994 by the National Center for Health Statistics (NCHS) of the US Centers for Disease Control and Prevention (CDC). It is a cross-sectional nationwide health survey of non-institutionalized US residents 2 months of age and older using a stratified multistage probability sampling design to sample a representative cohort of the US general population. Of the 39,695 individuals included in the NHANES III survey, 33,994 (85.6%) participated in home interviews, which were used to collect data on demographic characteristics, socioeconomic status, family medical history, current medical conditions, and use of medications. All individuals who participated in a home interview were invited to visit a mobile examination center (MEC) for a medical examination, which consisted of a physical examination, gradable fundus photography, and collection of blood and urine samples for laboratory testing. Therefore, the fundus photographs and blood for IgG testing in this study were collected at the same time. A total of 16,575 participants 20 years of age or older were examined in the MEC.<sup>25</sup>

This study involved only the secondary data analysis of existing US national databases that are publicly available and have been de-identified. This research qualified for exemption of institutional review board human subjects approval under 45 CFR 46.101(b) (4) as specified by the Federal Regulations for Protection of Human Research Subjects. Thus, this is an exempt study and there was no need for institutional review board approval from our institutions.

#### **Case and Control Definition**

In NHANES III, DR was assessed in 9737 adults 40 years of age and older using non-stereoscopic, color 45° photographs centered between the optic nerve and the macula. The camera used was a Canon CR4-45NM non-mydriatic fundus camera (Canon Inc., Tokyo, Japan), which incorporates an infrared video camera to allow photographs to be taken in a darkened examination room without the use of dilating drops, allowing for dilation of the pupil, usually to 6 mm to 10 mm in diameter.<sup>26</sup> Trained graders at the University of Wisconsin Ophthalmic Epidemiology Reading Center used the Modified Airlie House Classification scheme and the Early Treatment for Diabetic Retinopathy Study severity scale to grade the photographs.<sup>27,28</sup>

Out of ~8000 eligible participants, we identified 116 early DR cases. We focused on the early stages because we consider early prevention to be the best strategy for this irreversible, visual-impairment disease when it progresses into later stages. Early DR included hard exudates, soft exudates, intraretinal microvascular abnormalities without microaneurysms, hemorrhages only without microaneurysms, microaneurysms only, and early or moderate NPDR. Among the persons not displaying signs of DR, we selected 116 matched controls (case:control = 1:1) by random selection using frequency matching in age, race, sex, and education.

#### Serum Pg IgG Antibody

The Ab measurement in the NHANES III was performed at the Forsyth Institute, Boston MA. Detailed experimental methods and a panel of oral bacterial strains used to prepare whole-cell antigenic extracts for determining the levels of IgG Ab by means of the "checkerboard" immunoassay can be found in the CDC, NCHS, and NHANES III Data Documentation.<sup>29,30</sup> To assess the level of Ab to Pg, a mixed suspension of Pg comprised of ATCC strains 33277 and 53978 (ATCC, Manassas, VA) was used. The serum levels of IgG Ab were reported in enzyme-linked immunosorbent assay units (EU).

#### **Statistical Methods**

Because the distribution of serum Pg IgG Ab levels is positively skewed, before further analysis was performed the Pg IgG Ab variable was  $log_{10}$  transformed to an approximately normal distribution.<sup>24</sup> This  $log_{10}$ (IgG Ab) variable was used in further analyses.

The following were covariates in our analyses: age, sex, race, education level, smoking status, alcohol consumption status, including alcohol drinker status (non-drinkers vs. drinkers) and alcohol drinking status among drinkers (moderate drinking, less than 12 drinks in the past 12 months; excessive drinking, at least 12 drinks in the past 12 months), body mass index (BMI, computed from weight and height;  $kg/m^2$ ), duration of diabetes, insulin use, hypertension, serum levels of C-reactive protein (CRP), two clinical periodontal measurements (mean number of tooth sites that bled on probing [mBOP] and mean clinical attachment loss [mCAL]), and glycated hemoglobin (HbA1c; Diabetes Control and Complications Trial percent [DCCT%]). Note that high HbA1c status was defined as HbA1c > 5.7 DCCT%, and normal HbA1cstatus was defined as HbA1c  $\leq 5.7$  DCCT%.<sup>31</sup>

Descriptive statistics for these covariates between cases and controls were calculated. To determine significance of differences, analysis of variance (ANOVA) for comparison of means of continuous variables and  $\chi^2$  tests for categorical variables were used. We also examined the correlations among serum *Pg* IgG Ab levels and these covariates using Pearson's correlation coefficient (*r*), Wilcoxon–Mann–Whitney tests, or Kruskal–Wallis tests, as appropriate.

To evaluate the association between Pg IgG Ab levels and early DR risk, logistic regression models were fitted by controlling for selected covariates. We used a hierarchical strategy in our model construction to examine the confounding effects from the covariates. Starting from an age-adjusted model (Model 1), we stepwise included the other covariates: Model 2 additionally adjusted for sex, race, education, and BMI; Model 3 additionally adjusted for smoking history and alcohol drinker/drinking status; Model 4 additionally adjusted for serum CRP and HbA1c levels, and hypertension history; and Model 5 additionally adjusted for the two clinical periodontitis measurements, mBOP and mCAL. We also used Model 5 to calculate the estimated probabilities for early DR at various Pg IgG Ab levels. For Models 3, 4, and 5, we did two sets of analyses (see footnotes in Table 3); one adjusted for alcohol drinker status (non-drinkers vs. drinkers) and the second adjusted for alcohol drinking status among drinkers (moderate drinking, less than 12 drinks in the past 12 months; excessive drinking, at least 12 drinks in the past 12 months). Forty-one non-drinkers, including 19 controls and 22 cases, were excluded from the analyses for alcohol drinking status (moderate vs. excessive). In the analysis for the first case, the smoking histories for the 41 non-drinkers were included in the analysis. In the analysis for the second case, the 41 non-drinkers were excluded from the analysis.

We further evaluated whether or not the effect of serum Pg IgG Ab levels on early DR risk varies by the status of the seven modifiable risk factors, including education levels, BMI, smoking history, alcohol drinker/drinking status, serum HbA1c status, mBOP, and mCAL. Seven models were constructed; in each model, an interaction term between serum Pg IgG Ab levels and the modifiable risk factor of interest was added to Model 5.

All analyses were performed using the SAS 9.4 SURVEY procedures (SAS Institute Inc., Cary, NC), which take into account the complex sampling design used in NHANES III to calculate unbiased point estimates, such as means and odds ratios (ORs), standard errors (SEs), and confidence intervals (CIs). We used P < 0.05 to denote statistical significance, and all tests were two-sided.

### Results

Because our controls were matched with cases for age, sex, race, and education, it is not surprising that the distributions of these four covariates were not significantly different between cases and controls (Table 1). The distribution for BMI; serum levels of CRP, HbA1c, and Pg IgG Ab; and the two clinical periodontitis measurements, mBOP and mCAL, were not significantly different, either. However, cases

Table 1.	Comparison	of Characteristics	of DR Cases Versu	s Frequenc	y-Matched Controls
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	Controls <sup>*</sup>	Cases <sup>*</sup>	
Characteristics	( <i>n</i> = 116)	( <i>n</i> = 116)	P <sup>†</sup>
Age (yr), mean (SE)	58.4 (0.17)	58.4 (0.15)	0.99
Male sex, <i>n</i> (%)	62 (48.3)	62 (47.2)	0.18
Race, n (%)			0.10
Non-Hispanic white	49 (80.8)	49 (82.9)	
Non-Hispanic black	37 (14.7)	37 (12.1)	
Hispanic	30 (4.5)	30 (5.0)	
Education, n (%)			0.23
<12 yr	57 (33.3)	57 (34.5)	
12 yr	41 (42.3)	41 (39.2)	
>12 yr	18 (24.4)	18 (26.3)	
BMI (kg/m²), mean (SE)	27.6 (0.06)	28.3 (0.12)	0.50
Smoking history, n (%)			< 0.0001
Non-smoker	50 (45.5)	47 (31.4)	
Former smoker	37 (35.1)	37 (44.0)	
Active smoker	29 (19.4)	32 (24.6)	
Alcohol drinking status (at least 12 drinks in the past 12 months), n (%	)		
Non-drinker	19 (6.2)	22 (8.1)	< 0.0001
No (moderate drinking)	47 (22.2) (50.7 <sup>‡</sup> )	44 (15.5) (37.0 <sup>‡</sup> )	
Yes (excessive drinking)	50 (21.6) (49.3 <sup>‡</sup> )	50 (26.4) (63.0 <sup>‡</sup> )	< 0.0001
Serum CRP level (mg/dL), mean (SE)	0.44 (0.001)	0.56 (0.022)	0.43
Serum HbA1c (DCCT%), mean (SE)	5.62 (0.012)	5.66 (0.011)	0.82
Serum HbA1c category, n (%)			
$\leq$ 5.7% (normal)	64 (61.68)	70 (67.75)	
5.7%–6.5% (prediabetic)	48 (35.80)	33 (20.04)	0.0005
>6.5% (diabetic)	4 (2.52)	13 (12.21)	< 0.0001
Ever diagnosed with diabetes or diabetic eye diseases, $n$ (%)			
No	116 (100)	116 (100)	
Yes	0 (0)	0 (0)	1.00
Ever diagnosed with hypertension, <i>n</i> (%)		.,	
No	69 (67.49)	59 (52.10)	
Yes	47 (32.51)	57 (47.90)	< 0.0001
Serum log <sub>10</sub> ( <i>Pg</i> lgG Ab) (EU), mean (SE)	2.32 (0.018)	2.59 (0.01)	0.12
mBOP, mean (SE)	0.044 (0.0009)	0.026 (0.001)	0.11
mCAL, mean (SE)	0.70 (0.006)	0.64 (0.017)	0.68

\*For categorical variables, sample sizes are raw numbers, and the percentages are weighted for the sampling design used in the NHANES III study.

<sup>†</sup>ANOVA was used for statistical tests of significance for continuous variables, and the Wald  $\chi^2$  test was used for all other categorical measures.

<sup>‡</sup>Forty-one non-drinkers, including 19 controls and 22 cases, were excluded from the analysis.

 $^{\text{S}}$ HbA1c > 5.7% vs. HbA1c  $\leq$  5.7%.

||HbA1c  $\leq$  5.7% vs. 5.7 <HbA1c  $\leq$  6.5% vs. HbA1c > 6.5%.

tended to be in the normal category (HbA1c  $\leq 5.7\%$  vs. HbA1c > 5.7%; P = 0.0005) or the diabetic category (HbA1c > 6.5% vs. HbA1c  $\leq 5.7\%$  and 5.7 < HbA1c  $\leq 6.5\%$ ; P < 0.0001) more than controls. Smoking history and alcohol drinking status were significantly different

(both P < 0.0001), and cases tended to be in higher categories for both exposures. Notably, none of our subjects had been diagnosed as either diabetic or having diabetic eye diseases. Cases were more likely to have a hypertension history than controls (P < 0.0001).

		Pearson's <i>r</i> with	
Continuous Covariates	n	log <sub>10</sub> ( <i>Pg</i> lgG Ab) (EU)	Р
Age (yr)	232	-0.20	0.002
BMI (kg/m <sup>2</sup> )	232	0.17	0.012
Serum CRP level (mg/dL)	232	0.08	0.23
Serum HbA1c (DCCT%)	232	0.052	0.43
mBOP	232	0.32	< 0.0001
mCAL	232	0.33	< 0.0001
		Median (IQR) for	
Categorical Covariates	n	log <sub>10</sub> ( <i>Pg</i> lgG Ab) (EU)	<b>P</b> *
Sex			
Male	124	2.50 (1.95–3.12)	0.06
Female	108	2.38 (1.78–2.92)	
Race			
Non-Hispanic white	98	2.12 (1.80–2.83)	< 0.0001
Non-Hispanic black	74	2.94 (2.50–3.47)	
Hispanic	60	3.14 (2.82–3.49)	
Education			
<12 yr	114	2.63 (1.91–3.18)	
12 yr	82	2.38 (1.62–2.97)	0.19
>12 yr	36	2.14 (1.96–2.77)	
Smoking history			
Non-smoker	97	2.76 (2.04–3.09)	
Former smoker	74	2.02 (1.85–2.77)	0.48
Active smoker	61	2.48 (1.75–3.19)	
Alcohol drinking status (at least 12 drinks in the past 12 months)			
Non-drinkers	41	2.55 (1.91–3.08)	0.42
No (moderate drinking)	91	2.28 (1.75–2.92)	0.22 <sup>†</sup>
Yes (excessive drinking)	100	2.19 (1.95–3.12)	
Ever diagnosed with hypertension			0.18
No	104	2.04 (1.85–2.82)	
Yes	128	2.66 (1.91–3.34)	

#### Table 2. Bivariate Associations Between Serum Pg IgG Ab Concentrations and Covariates

IQR, interquartile range.

\**P* values were obtained by Wilcoxon two-sample test for variables with two categories and from the Kruskal–Wallis test for variables with three or more categories.

<sup>†</sup>Forty-one non-drinkers were excluded from the analysis.

In our bivariate analysis (Table 2), age (r = -0.20; P = 0.002) was inversely correlated with serum Pg IgG Ab levels, whereas BMI (r = 0.17; P = 0.12), mBOP (r = 0.32; P < 0.0001), and mCAL (r = 0.33; P < 0.0001) were positively correlated. Males (P = 0.06) and Hispanics (P < 0.0001) tended to have higher levels of serum Pg IgG Ab. However, education level, smoking status, alcohol drinking status, hypertension history, and serum levels of CRP and HbA1c were not significantly correlated with serum Pg IgG Ab levels.

Next, using logistic regression analysis, we evaluated our primary interest in the association between serum Pg IgG Ab levels and risk for early DR (Table 3). Except for Model 5, the ORs per tenfold increase of PgIgG Ab levels in every higher hierarchical model were similar, which conferred over 30% increased risk for early DR. The difference in OR between Model 4 and Model 5 was over 20%, which is much higher than the OR difference (~10%) between any other two neighbor models in Table 3, implying a significant confounding effect from either one of the two periodontal

	Adjusted OR (95% CI) <sup>*</sup>					
Serum <i>Pg</i> IgG Level (EU)	Model 1	Model 2	Model 3	Model 4	Model 5	
Per tenfold increase	1.51 (1.41–1.62)	1.58 (1.46–1.70)	1.76 (1.59–1.95) <sup>†</sup>	1.71 (1.54–1.89) <sup>†</sup>	2.06 (1.80–2.36) <sup>†</sup>	
P for trend	<0.0001	<0.0001	<0.0001 <sup>†</sup> <0.0001 <sup>‡</sup>	<0.0001 <sup>†</sup> <0.0001 <sup>‡</sup>	<0.0001 <sup>†</sup> <0.0001 <sup>‡</sup>	

 Table 3.
 Logistic Analysis Relating Serum Pg IgG Ab Levels to Risk for Early DR

<sup>\*</sup>Model 1 was adjusted for age; Model 2 is Model 1 additionally adjusted for sex, race, education, and BMI; Model 3 is Model 2 additionally adjusted for smoking history and alcohol consumption status; Model 4 is Model 3 additionally adjusted for serum CRP and HbA1c levels and hypertension diagnosis; and Model 5 is Model 4 additionally adjusted for two clinical periodontitis measurements, mBOP and mCAL.

<sup>†</sup>Adjusted for alcohol drinker status (non-drinkers vs. drinkers).

<sup>‡</sup>Adjusted for alcohol drinking status among drinkers (moderate drinking, less than 12 drinks in the past 12 months; excessive drinking, at least 12 drinks in the past 12 months); 41 non-drinkers, including 19 controls and 22 cases, were excluded from the analysis.



Bubble area proportional to sampling weight

**Figure 1.** Logistic regression was used to calculate estimated probabilities for early DR at various *Pg* IgG Ab levels. The model was adjusted for age, sex, race, education, BMI, smoking history, alcohol drinking status, serum CRP and HbA1c levels, hypertension, mBOP, and mCAL. Forty-one non-drinkers were excluded from the analysis. The *Pg* IgG Ab levels were  $\log_{10}$  transformed. The estimated probabilities for early DR at various *Pg* IgG Ab levels and the fitted linear regression line are shown (regression coefficient  $\beta = 0.06192$ ; *P* for trend = 0.0053).

measurements, mBOP or mCAL, or from both. Further analyses indicated that mBOP was responsible for the majority of the confounding. All five models showed a significant trend (P < 0.0001) in the relationship between serum Pg IgG Ab levels and early DR risk. The estimated probabilities from Model 5 for Pgrelated early DR risk at various Pg IgG Ab levels and the fitted linear regression line among drinkers are shown in Figure 1 (regression coefficient  $\beta = 0.06192$ ; P for trend = 0.0053). The results from our interaction analysis indicated that *Pg*-related early DR risk significantly varied by alcohol drinking status among drinkers (moderate vs. excessive drinking; interaction  $\beta = 0.45$ ; *P* for interaction = 0.0003), serum HbA1c category (interaction  $\beta = 0.45$ ; *P* for interaction < 0.0001), and mBOP (interaction  $\beta =$ -3.44; *P* for interaction = 0.0003). Our analysis also indicated that *Pg*-related early DR risk did not significantly vary by alcohol drinker status (non-drinkers



**Figure 2.** *Pg*-related early DR risk significantly varied by the four combinations of alcohol drinking status (moderate or excessive drinking) and serum HbA1c status (high, HbA1c > 5.7 DCCT%; normal, HbA1c  $\leq$  5.7 DCCT%) (interaction  $\beta$  = 0.27; *P* for interaction < 0.0001). Shown are the estimated probabilities and fitted linear regression lines for (**A**) excessive drinking and high HbA1c ( $\beta$  = 0.11882; *P* for trend < 0.0001) vs. (**B**) excessive drinking and normal HbA1c ( $\beta$  = 0.00985; *P* for trend < 0.0001) vs. (**C**) moderate drinking and high HbA1c ( $\beta$  = 0.03317; *P* for trend < 0.0001) vs. (**D**) moderate drinking and normal HbA1c ( $\beta$  = 0.04695; *P* for trend < 0.0001). PGMX, a mixed suspension of *Pg* ATCC strains 33277 and 53978.

vs. drinkers; interaction  $\beta = 0.03$ ; P for interaction = 0.64). Further analysis showed that *Pg*-related early DR risk significantly varied for the four combinations of alcohol drinking status (moderate vs. excessive drinking) and serum HbA1c status (high, HbA1c > 5.7 DCCT% vs. normal, HbA1c  $\leq$  5.7 DCCT%) (interaction  $\beta = 0.27$ ; *P* for interaction < 0.0001). We also calculated the estimated probabilities and plotted the four fitted linear regression lines for excessive drinking and high HbA1c ( $\beta = 0.11882$ ; P for trend < 0.0001) (Fig. 2A) vs. excessive drinking and normal HbA1c ( $\beta$ = 0.00985; P for trend < 0.0001) (Fig. 2B) vs. moderate drinking and high HbA1c ( $\beta = 0.03317$ ; P for trend < 0.0001) (Fig. 2C) vs. moderate drinking and normal HbA1c ( $\beta = 0.04695$ ; P for trend < 0.0001) (Fig. 2D).

## Discussion

People with diabetes have increased risk for periodontitis and caries, and oral (periodontal) inflammation negatively impacts glycemic control by contributing to systemic inflammation in both diabetic and non-diabetic people.<sup>2,3,32</sup> Although several studies have related clinical periodontitis to DR,<sup>33–36</sup> little is known about the relationship between oral microbes and eye health, and, importantly, their interaction with the host immune response has not been investigated in the relationship. In this study, we showed that serum *Pg* IgG Ab levels were positively associated with the risk for early DR, adding to the accumulating evidence that, through immune responses, microbes in

the human body may have an impact on tissues and organs remote from their original habitat. Interestingly, we also found that moderate alcohol intake and normal HbA1c levels alleviated this *Pg*-related risk for early DR, suggesting that the *Pg*-related early DR risk could be attenuated by managing multiple modifiable risk factors for DR.

Previous studies have suggested that serum Ab is a better measure than clinical periodontitis for exposure to oral microbes.<sup>37</sup> The findings indicated that all Ab levels were significantly and strongly associated with carriage of the corresponding pathogens, but only weakly with the presence or number of teeth with periodontitis. It was also noted that individual Ab levels and the numbers of corresponding bacteria in saliva showed a positive association, independent of the severity of periodontitis. For many years, the Ab response to oral bacteria, particularly IgG, has been regarded as a reliable surrogate for systemic exposure to the organisms.<sup>37</sup> Interestingly, translocated oral bacteria, including Pg, directly impact the gut microbiome and possibly immune defense, resulting in IgG responses to translocated microbial antigens.<sup>38,39</sup> These findings suggest that the origin of serum IgG Ab to oral microbes is not limited to the oral cavity.

The complement classic pathway is activated when complement component Clq binds to the IgG constant region (Fc $\gamma$ ) attached to microbe surface antigens.40 The pathway leads to targeted lysis of the pathogenic surface through the assembly of membrane-penetrating pores known as the membrane attack complex (MAC). Interestingly, it has been well documented that the generation of undesirable quantities of MAC plays a significant role in the pathogenesis of DR.<sup>41-44</sup> However, although it has been demonstrated that the inflammatory and immune response triggered by Pg has not only local but also systemic effects, <sup>13,14,16,20,21,37,45</sup> the remote effects of complement activation induced by Pg on the retina remain to be determined. On the other hand, recent studies have indicated that, even in IgAdominated tissues, such as oral mucosa, commensal-IgG Fc $\gamma$  may cross-link Fc $\gamma$  receptors (Fc $\gamma$ Rs) on mononuclear phagocytes (MNPs), inducing IL-1 $\beta$ production, and that the MNP  $Fc\gamma R$  active:inhibitory  $(Fc\gamma R:Fc\gamma RIIB)$  ratio determines the magnitude of type 17 immunity and local inflammation.<sup>46</sup> Importantly, type 17 immunity plays an important role in maintaining mucosal barriers and contributing to pathogen clearance at mucosal surfaces, and it is implicated in autoimmune disorders. Furthermore, the loss of T helper 17 cell populations at mucosal surfaces has been linked to local inflammation and microbial translocation, leading to chronic inflammation at remote sites. These findings suggest that the IgG Ab levels are independent of clinical periodontitis and that serum Pg IgG Ab indicates a systemic response to this periodontal disease-associated pathogenic bacterium, lending biological support to our approach to study serum IgG Ab levels instead of the microbe per se.

Recent work has conceptualized DR as a disease of the neurovascular unit.<sup>6</sup> In addition to the component vascular cells (endothelial cells and pericytes), diverse retinal neuronal cell types, macroglial elements (Müller cells and astrocytes), and microglia, the neurovascular concept also suggests the importance of additional cell types, such as retinal pigment epithelium (RPE) and immune cells. Importantly, as diabetes progresses, the retina exhibits multiple elements of chronic, subclinical inflammation, including immune cell activation and production of inflammatory molecules, which play a pivotal role in modulating the constituent cells of the neurovascular unit and driving the progression of DR.<sup>6,47,48</sup> This new concept of DR pathogenesis lends further support to our strategy of using immunerelated biomarkers in epidemiological studies on DR.

In a recent in vitro study, Arjunan et al.<sup>49</sup> characterized Pg invasion in the human RPE cells, including vacuolar/cytosolic localization and prolonged survival by autophagy evasion within the RPE cells. These findings lend further support to the accumulating evidence that Pg may access and spread in the neural system.<sup>19</sup> Although infection has been suggested as a potential risk factor for DR and Pg has been related to several human metabolic disorders,<sup>50–53</sup> our study provides the first epidemiological evidence showing a significant association between Pg infection and risk for early DR.

In our interaction analysis, we found that, although either alcohol drinking status or serum HbA1c status had a synergistic action with serum Pg IgG Ab levels (interaction  $\beta > 0$ ), the effect of the combined action between mBOP and serum Pg IgG Ab levels is less than the sum of their individual effects (interaction  $\beta <$ 0). This phenomenon was not surprising, because it is probably due to the overlapping information provided by mBOP and serum Pg IgG Ab levels, both of which are related to periodontitis. Our confounding analysis indicating that the association between serum Pg IgG Ab levels and early DR risk was confounded by mBOP (Table 3) also provides support to this antagonistic interaction. Notably, although moderate drinking conferred lower Pg-related early DR risk than excessive drinking, non-drinkers did not gain additional benefit from not drinking alcohol. Furthermore, compared with either consuming moderate alcohol or maintaining a normal serum HbA1c level alone (Fig. 2B, 2C), our data suggest that their combined effect (Fig. 2D),

probably through eating a low-glycemic-index Mediterranean diet, 54-68 confers the lowest Pg-related early DR risk. Indeed, randomized nutritional intervention studies have demonstrated benefit of such a diet. including shifting the Pg-driven microbiota by decreasing the relative abundance of periodontopathogenic bacterial in the saliva and increasing levels of Streptococcus cristatus, which has been reported to be an antagonistic taxon inhibiting Pg gene expression.<sup>69–72</sup> It is also noteworthy that recent studies have found that prenylated flavonoids, which are present in wine and beer, 73, 74 suppress gingipains, Pg growth, and biofilm formation.<sup>75</sup> Furthermore, studies have suggested that Pg and related dysbiosis contribute to the development of insulin resistance and poor glycemic control.<sup>76–78</sup> Novel therapeutics and prevention strategies for DR may be developed from further mechanistic studies on these Pg-related mechanisms.

We found that Hispanics had higher serum levels of Pg IgG Ab (Table 2). Studies have also indicated that Hispanics have a higher risk of DR and that the increased risk could be attributed to both genetic background and environmental exposures, including diet.<sup>79</sup> Taking these data together, race was a potential confounder in the association of our interest; that is, between serum levels of Pg IgG Ab and risk for DR, we therefore adjusted race in our multivariable regression analyses. However, more research is needed to investigate the independent role of genetic background and diet in the association between serum levels of Pg IgG Ab and risk for DR.

The strengths of this study include its being a matched case-control study in a representative cohort of the US population, standardized collection of risk factor information, and photographic grading of maculopathy to minimize the influence of confounding factors and misclassifications. The cross-sectional nature of this study limits its strength in defining temporality; however, serum Pg IgG Ab is considered to reflect chronic exposure,<sup>18</sup> and the average age and prevalence of Pg infection are much younger and higher, respectively, than those for DR.<sup>80–83</sup> Furthermore, this study involved only secondary data analysis of existing U.S. national databases that are publicly available and have been de-identified. The NHANES III performed only Pg IgG measurement in the blood without measuring Pg IgA. It would be interesting to examine the association with Pg IgA Ab in future studies. In light of the importance of complement activation in Pg pathogenesis, complement factors, such as C1q, C2b, and C3a, should also be included in future studies. In this study cohort, we focused our interest on early DR because there were only a limited number of advanced DR cases which did not allow for meaningful analysis. Further study should investigate the associations with different categories of DR. We recognized that oral hygiene or socioeconomic factors (access to medical care) could be confounding factors in our analysis and therefore adjusted for education level in our multivariable regression analysis. However, residual confounding could still be an issue and remains to be further investigated.

In conclusion, we demonstrated a novel association between exposure to Pg and risk for early DR. Although the causality and detailed mechanism warrant further study, our findings may result in a significant impact on the therapeutic and prevention strategies for DR by virtue of the high prevalence of Pg infection in the human population.

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### References

- Guariguata L, Whiting DR, Hambleton I, Beagley J, Linnenkamp U, Shaw JE. Global estimates of diabetes prevalence for 2013 and projections for 2035. *Diabetes Res Clin Pract*. 2014;103(2):137– 149.
- Preshaw PM, Alba AL, Herrera D, et al. Periodontitis and diabetes: a two-way relationship. *Diabetologia*. 2012;55(1):21–31.
- 3. Preshaw PM, Bissett SM. Periodontitis and diabetes. *Br Dent J.* 2019;227(7):577–584.

- Antonetti DA, Klein R, Gardner TW. Diabetic retinopathy. N Engl J Med. 2012;366(13):1227– 1239.
- 5. Leasher JL, Bourne RR, Flaxman SR, et al. Global estimates on the number of people blind or visually impaired by diabetic retinopathy: a meta-analysis from 1990 to 2010. *Diabetes Care*. 2016;39(9):1643–1649.
- 6. Duh EJ, Sun JK, Stitt AW. Diabetic retinopathy: current understanding, mechanisms, and treatment strategies. *JCI Insight*. 2017;2(14):e93751.
- 7. Honda K. *Porphyromonas gingivalis* sinks teeth into the oral microbiota and periodontal disease. *Cell Host Microbe*. 2011;10(5):423–425.
- 8. Darveau RP. Periodontitis: a polymicrobial disruption of host homeostasis. *Nat Rev Microbiol*. 2010;8(7):481–490.
- 9. How KY, Song KP, Chan KG. *Porphyromonas gingivalis*: an overview of periodontopathic pathogen below the gum line. *Front Microbiol*. 2016;7:53.
- Boisvert H, Lorand L, Duncan MJ. Transglutaminase 2 is essential for adherence of *Porphyromonas gingivalis* to host cells. *Proc Natl Acad Sci USA*. 2014;111(14):5355–5360.
- Kamer AR, Craig RG, Dasanayake AP, Brys M, Glodzik-Sobanska L, de Leon MJ. Inflammation and Alzheimer's disease: possible role of periodontal diseases. *Alzheimers Dement*. 2008;4(4):242– 250.
- 12. Hussain M, Stover CM, Dupont A. *P. gingivalis* in periodontal disease and atherosclerosis scenes of action for antimicrobial peptides and complement. *Front Immunol.* 2015;6:45.
- Kozarov EV, Dorn BR, Shelburne CE, Dunn WAJ, Progulske-Fox A. Human atherosclerotic plaque contains viable invasive Actinobacillus actinomycetemcomitans and Porphyromonas gingivalis. Arterioscler Thromb Vasc Biol. 2005;25(3):e17–e18.
- Genco R, Offenbacher S, Beck J. Periodontal disease and cardiovascular disease: epidemiology and possible mechanisms. J Am Dent Assoc. 2002;133(suppl):14S–22S.
- Gómez-Bañuelos E, Mukherjee A, Darrah E, Andrade F. Rheumatoid arthritis-associated mechanisms of *Porphyromonas gingivalis* and *Aggregatibacter actinomycetemcomitans*. J Clin Med. 2019;8(9):1309.
- Rosenstein ED, Greenwald RA, Kushner LJ, Weissmann G. Hypothesis: the humoral immune response to oral bacteria provides a stimulus for the development of rheumatoid arthritis. *Inflammation*. 2004;28(6):311–318.
- 17. Sparks Stein P, Steffen MJ, Smith C, et al. Serum antibodies to periodontal pathogens are a risk fac-

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tor for Alzheimer's disease. *Alzheimers Dement*. 2012;8(3):196–203.

- Noble JM, Borrell LN, Papapanou PN, Elkind MS, Scarmeas N, Wright CB. Periodontitis is associated with cognitive impairment among older adults: analysis of NHANES-III. *J Neurol Neuro*surg Psychiatry. 2009;80(11):1206–1211.
- 19. Dominy SS, Lynch C, Ermini F, et al. *Porphyromonas gingivalis* in Alzheimer's disease brains: evidence for disease causation and treatment with small-molecule inhibitors. *Sci Adv.* 2019;5(1):eaau3333.
- 20. Olsen I, Kell DB, Pretorius E. Is *Porphyromonas* gingivalis involved in Parkinson's disease? *Eur J Clin Microbiol Infect Dis.* 2020;39(11):2013–2018.
- 21. Chapple IL, Genco R, Working Group 2 of the Joint EFP/AAP Workshop. Diabetes and periodontal diseases: consensus report of the Joint EFP/AAP Workshop on Periodontitis and Systemic Diseases. J Periodontol. 2013;84(4 suppl):S106–S112.
- 22. Dominy SS, Lynch C, Ermini F, et al. *Porphyromonas gingivalis* in Alzheimer's disease brains: evidence for disease causation and treatment with small-molecule inhibitors. *Sci Adv.* 2019;5(1):eaau3333.
- 23. Kantarci A, Tognoni CM, Yaghmoor W, et al. Microglial response to experimental periodontitis in a murine model of Alzheimer's disease. *Sci Rep.* 2020;10(1):18561.
- 24. Chiu CJ, Chang ML, Taylor A. Associations between periodontal microbiota and death rates. *Sci Rep.* 2016;6:35428.
- 25. Patrick PA, Visintainer PF, Shi Q, Weiss IA, Brand DA. Vitamin D and retinopathy in adults with diabetes mellitus. *Arch Ophthalmol.* 2012;130(6):756–760.
- 26. Plan and operation of the Third National Health and Nutrition Examination Survey, 1988-94. Series
  1: programs and collection procedures. *Vital Health Stat 1*. 1994;32:1–407.
- 27. National Technical Information Service. *Third National Health and Nutrition Examination Survey: Fundus Photograph Grading Protocol.* Springfield, VA: US Department of Health and Human Services; 1995.
- Early Treatment Diabetic Retinopathy Study Research Group. Grading diabetic retinopathy from stereoscopic color fundus photographs-an extension of the modified Airlie House classification: ETDRS report number 10. *Ophthalmology*. 1991;98(5 suppl):786–806.
- 29. Sakellari D, Socransky SS, Dibart S, Eftimiadi C, Taubman MA. Estimation of serum

antibody to subgingival species using checkerboard immunoblotting. *Oral Microbiol Immunol*. 1997;12(5):303–310.

- Dye BA, Herrera-Abreu M, Lerche-Sehm J, et al. Serum antibodies to periodontal bacteria as diagnostic markers of periodontitis. *J Periodontol*. 2009;80(4):634–647.
- American Diabetes Association. Standards of medical care in diabetes–2010. *Diabetes Care*. 2010;33(suppl 1):S11–S61.
- 32. Wolff RE, Wolff LF, Michalowicz BS. A pilot study of glycosylated hemoglobin levels in periodontitis cases and healthy controls. *J Periodontol*. 2009;80(7):1057–1061.
- 33. Veena HR, Natesh S, Patil SR. Association between diabetic retinopathy and chronic periodontitis-a cross-sectional study. *Med Sci* (*Basel*). 2018;6(4):104.
- 34. Song SJ, Lee SS, Han K, Park JB. Periodontitis is associated with diabetic retinopathy in non-obese adults. *Endocrine*. 2017;56(1):82–89.
- 35. Banthia R, Raje S, Banthia P, Saral SK, Singh P, Gupta S. Evaluation of the association between periodontal disease and diabetic retinopathy. *Gen Dent*. 2014;62(6):e28–e32.
- Amiri AA, Maboudi A, Bahar A, et al. Relationship between type 2 diabetic retinopathy and periodontal disease in Iranian adults. *N Am J Med Sci.* 2014;6(3):139–144.
- Pussinen PJ, Könönen E, Paju S, et al. Periodontal pathogen carriage, rather than periodontitis, determines the serum antibody levels. *J Clin Periodontol*. 2011;38(5):405–411.
- Olsen I, Yamazaki K. Can oral bacteria affect the microbiome of the gut? J Oral Microbiol. 2019;11(1):1586422.
- 39. Zeng MY, Cisalpino D, Varadarajan S, et al. Gut microbiota-induced immunoglobulin G controls systemic infection by symbiotic bacteria and pathogens. *Immunity*. 2016;44(3):647–658.
- 40. Dunkelberger JR, Song W-C. Complement and its role in innate and adaptive immune responses. *Cell Res.* 2010;20(1):34–50.
- 41. Chrzanowska M, Modrzejewska A, Modrzejewska M. New insight into the role of the complement in the most common types of retinopathy-current literature review. *Int J Ophthalmol.* 2018;11(11):1856–1864.
- 42. Xu H, Chen M. Targeting the complement system for the management of retinal inflammatory and degenerative diseases. *Eur J Pharmacol*. 2016;787:94–104.
- 43. Gerl VB, Bohl J, Pitz S, Stoffelns B, Pfeiffer N, Bhakdi S. Extensive deposits of complement

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C3d and C5b-9 in the choriocapillaris of eyes of patients with diabetic retinopathy. *Invest Ophthalmol Vis Sci.* 2002;43(4):1104–1108.

- 44. Zhang J, Gerhardinger C, Lorenzi M. Early complement activation and decreased levels of glycosylphosphatidylinositol-anchored complement inhibitors in human and experimental diabetic retinopathy. *Diabetes*. 2002;51(12):3499–3504.
- 45. Olsen I, Lambris JD, Hajishengallis G. *Porphyromonas gingivalis* disturbs host-commensal homeostasis by changing complement function. *J Oral Microbiol*. 2017;9(1):1340085.
- 46. Castro-Dopico T, Dennison TW, Ferdinand JR, et al. Anti-commensal IgG drives intestinal inflammation and type 17 immunity in ulcerative colitis. *Immunity*. 2019;50(4):1099–1114.e10.
- 47. Menini S, Iacobini C, Vitale M, Pugliese G. The inflammasome in chronic complications of diabetes and related metabolic disorders. *Cells*. 2020;9(8):1812.
- 48. Gui F, You Z, Fu S, Wu H, Zhang Y. Endothelial dysfunction in diabetic retinopathy. *Front Endocrinol (Lausanne)*. 2020;11:591.
- 49. Arjunan P, Swaminathan R, Yuan J, et al. Invasion of human retinal pigment epithelial cells by *Porphyromonas gingivalis* leading to vacuolar/cytosolic localization and autophagy dysfunction in-vitro. *Sci Rep.* 2020;10(1):7468.
- 50. Kallick CA. Diabetic retinopathy and *Ehrlichia*: the possible relationship. *Med Hypothesis Discov Innov Ophthalmol.* 2012;1(2):33– 36.
- 51. Makiura N, Ojima M, Kou Y, et al. Relationship of *Porphyromonas gingivalis* with glycemic level in patients with type 2 diabetes following periodontal treatment. *Oral Microbiol Immunol*. 2008;23(4):348–351.
- 52. Schara R, Skaleric E, Seme K, Skaleric U. Prevalence of periodontal pathogens and metabolic control of type 1 diabetes patients. *J Int Acad Periodontol.* 2013;15(1):29–34.
- 53. Simonsen JR, Järvinen A, Hietala K, et al. Bacterial infections as novel risk factors of severe diabetic retinopathy in individuals with type 1 diabetes [published online ahead of print September 14, 2020]. Br J Ophthalmol, https://doi.org/10. 1136/bjophthalmol-2020-316202.
- 54. Chiu CJ, Taylor A. Nutritional antioxidants and age-related cataract and maculopathy. *Exp Eye Res.* 2007;84(2):229–245.
- 55. Chiu CJ, Klein R, Milton RC, Gensler G, Taylor A. Does eating particular diets alter risk of age-related macular degeneration in users of the

Age-Related Eye Disease Study supplements? *Br J Ophthalmol.* 2009;93(9):1241–1246.

- 56. Chiu CJ, Milton RC, Klein R, Gensler G, Taylor A. Dietary compound score and risk of age-related macular degeneration in the Age-Related Eye Disease Study. *Ophthalmology*. 2009;116(5):939–946.
- Chiu CJ, Chang ML, Zhang FF, et al. The relationship of major American dietary patterns to agerelated macular degeneration. *Am J Ophthalmol.* 2014;158(1):118–127.
- 58. Chiu CJ, Robman L, McCarty CA, et al. Dietary carbohydrate in relation to cortical and nuclear lens opacities in the Melbourne Visual Impairment Project. *Invest Ophthalmol Vis Sci.* 2010;51(6):2897–2905.
- 59. Chiu CJ, Rabbani N, Rowan S, et al. Studies of advanced glycation end products and oxidation biomarkers for type 2 diabetes. *Biofactors*. 2018;44(3):281–288.
- 60. Chiu CJ, Morris MS, Rogers G, et al. Carbohydrate intake and glycemic index in relation to the odds of early cortical and nuclear lens opacities. *Am J Clin Nutr*. 2005;81(6):1411–1416.
- 61. Chiu CJ, Milton RC, Klein R, Gensler G, Taylor A. Dietary carbohydrate and progression of agerelated macular degeneration, a prospective study from the Age-Related Eye Disease Study. *Am J Clin Nutr*. 2007;86(4):1210–1218.
- 62. Chiu CJ, Milton RC, Gensler G, Taylor A. Association between dietary glycemic index and agerelated macular degeneration in nondiabetic participants in the Age-Related Eye Disease Study. *Am J Clin Nutr.* 2007;86(1):180–188.
- 63. Chiu CJ, Milton RC, Gensler G, Taylor A. Dietary carbohydrate and glycemic index in relation to cortical and nuclear lens opacities in the Age-Related Eye Disease Study. *Am J Clin Nutr.* 2006;83(5):1177–1184.
- 64. Chiu CJ, Liu S, Willett WC, et al. Informing food choices and health outcomes by use of the dietary glycemic index. *Nutr Rev.* 2011;69(4):231–242.
- 65. Chiu CJ, Hubbard LD, Armstrong J, et al. Dietary glycemic index and carbohydrate in relation to early age-related macular degeneration. *Am J Clin Nutr*. 2006;83(4):880–886.
- 66. Chiu CJ, Conley YP, Gorin MB, et al. Associations between genetic polymorphisms of insulinlike growth factor axis genes and risk for agerelated macular degeneration. *Invest Ophthalmol Vis Sci.* 2011;52(12):9099–9107.
- 67. Chiu C-J, Taylor A. Dietary hyperglycemia, glycemic index and metabolic retinal diseases. *Prog Retin Eye Res.* 2011;30(1):18–53.

- 68. Wong MYZ, Man REK, Fenwick EK, et al. Dietary intake and diabetic retinopathy: a systematic review. *PLoS One*. 2018;13(1):e0186582.
- 69. Laiola M, De Filippis F, Vitaglione P, Ercolini D. A Mediterranean diet intervention reduces the levels of salivary periodontopathogenic bacteria in overweight and obese subjects. *Appl Environ Microbiol.* 2020;86(12):e00777–20.
- Woelber JP, Bremer K, Vach K, et al. An oral health optimized diet can reduce gingival and periodontal inflammation in humans – a randomized controlled pilot study. *BMC Oral Health*. 2016;17(1):28.
- 71. Tennert C, Reinmuth AC, Bremer K, et al. An oral health optimized diet reduces the load of potential cariogenic and periodontal bacterial species in the supragingival oral plaque: a randomized controlled pilot study. *MicrobiologyOpen*. 2020;9(8):e1056.
- 72. Ajala O, English P, Pinkney J. Systematic review and meta-analysis of different dietary approaches to the management of type 2 diabetes. *Am J Clin Nutr*. 2013;97(3):505–516.
- 73. Arranz S, Chiva-Blanch G, Valderas-Martínez P, Medina-Remón A, Lamuela-Raventós RM, Estruch R. Wine, beer, alcohol and polyphenols on cardiovascular disease and cancer. *Nutrients*. 2012;4(7):759–781.
- 74. Boronat A, Soldevila-Domenech N, Rodríguez-Morató J, Martínez-Huélamo M, Lamuela-Raventós RM, de la Torre R. Beer phenolic composition of simple phenols, prenylated flavonoids and alkylresorcinols. *Molecules*. 2020;25(11):2582.
- 75. Kariu T, Nakao R, Ikeda T, Nakashima K, Potempa J, Imamura T. Inhibition of gingipains and *Porphyromonas gingivalis* growth and biofilm formation by prenyl flavonoids. *J Periodontal Res.* 2017;52(1):89–96.
- 76. Aemaimanan P, Amimanan P, Taweechaisupapong S. Quantification of key periodontal pathogens in insulin-dependent type 2 diabetic and non-diabetic patients with generalized chronic periodontitis. *Anaerobe*. 2013;22:64– 68.
- 77. Tian J, Liu C, Zheng X, et al. *Porphyromonas* gingivalis induces insulin resistance by increasing BCAA levels in mice. *J Dent Res.* 2020;99(7):839–846.
- 78. Mei F, Xie M, Huang X, et al. *Porphyromonas gingivalis* and its systemic impact: current status. *Pathogens*. 2020;9(11):944.
- Mora N, Kempen JH, Sobrin L. Diabetic retinopathy in Hispanics: a perspective on disease burden. *Am J Ophthalmol.* 2018;196:xviii–xxiv.

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- McClellan DL, Griffen AL, Leys EJ. Age and prevalence of *Porphyromonas gingivalis* in children. J Clin Microbiol. 1996;34(8):2017– 2019.
- 81. Nadkarni MA, Chhour KL, Browne GV, et al. Age-dependent changes in *Porphyromonas gingivalis* and *Prevotella* species/phylotypes in healthy gingiva and inflamed/diseased sub-gingival sites. *Clin Oral Investig.* 2015;19(4):911–919.
- Tuite-McDonnell M, Griffen AL, Moeschberger ML, Dalton RE, Fuerst PA, Leys EJ. Concordance of *Porphyromonas gingivalis* colonization in families. *J Clin Microbiol*. 1997;35(2):455–461.
- 83. Rafiei M, Kiani F, Sayehmiri K, et al. Prevalence of anaerobic bacteria (*P. gingivalis*) as major microbial agent in the incidence periodontal diseases by meta-analysis. *J Dent (Shiraz)*. 2018;19(3):232–242.