





Complete Genome Sequences of Two Vibrio natriegens **Bacteriophages**

Meghan T. Harris, a.b Tereasa Ching Ho, a Harry Fruchtman, Mira E. Garin, Victor Kubatin, Tiger Lu, Lingyan Xue, a Michael T. Marr II^{a,b}

^aDepartment of Biology, Brandeis University, Waltham, Massachusetts, USA

ABSTRACT Vibrio natriegens, a fast-growing Gram-negative bacterium, is gaining interest as a platform for rapid biotechnology applications and metabolic engineering. Only a few bacteriophages that infect this bacterium have been identified. Here, we describe the isolation and characterization of two V. natriegens bacteriophages isolated from Hatches Creek, Wellfleet, Massachusetts.

ibrio natriegens is a Gram-negative moderately halophilic marine bacterium first isolated in the 1950s (1). With a doubling time of less than 10 minutes under optimal conditions, it is the fastest-growing bacterium (2). It has garnered interest as a biotechnology platform for protein production, molecular biology, and metabolic engineering (3-6). Identifying bacteriophage that infect V. natriegens has the potential to aid in these efforts, as molecular genetic tools are often derived from bacterio-

Environmental samples from Cape Cod, Massachusetts, were tested for bacteriophage by spotting them onto Luria-Bertani Vibrio natriegens broth (LB-Vn) top agar (1% tryptone, 0.5% yeast extract, 3% NaCl, 4 mM KCl, 20 mM MgSO₄, 4 mM CaCl₂, and 0.5% agar) containing V. natriegens (ATCC 14048). V. natriegens was prepared by a 1:100 dilution of an overnight culture in LB-Vn liquid medium, followed by aerobic growth at 37°C for 4 to 6 hours. Zones of clearing were observed for one sample of filtered water from a mixture of sediment and brackish water taken from Hatches Creek in Wellfleet, Massachusetts. The clear spot was harvested by scraping and then suspending it in phage buffer (50 mM Tris [pH 7.5], 100 mM NaCl, 8 mM MgSO₄, 5 mM CaCl₂, and 0.01% gelatin). Individual plaques were obtained by serial dilution of this sample in phage buffer and two rounds of plaque purification on LB-Vn top agar plates containing V. natriegens.

Genome sequences were obtained by a combination of methods. Two well-separated plaques were subjected to direct sequencing using a Nextera kit (8). Phage genomic DNA was also purified from high-titer stocks prepared from webbed plates using the Promega Wizard DNA cleanup system followed by Nextera tagmentation. Libraries prepared directly from plaques were sequenced on a NextSeq 500 instrument with a 75-cycle output. Libraries created from purified DNA were sequenced on a MiSeq instrument using 150-bp reads. Libraries were initially sequenced and analyzed individually. All three libraries yielded the same two phage genomes. To increase the confidence and coverage of the genomes, the library sequences were combined and reassembled. The combined library contains 140,432,773 total reads. Reads were processed online using Galaxy and the Center for Phage Technology (CPT) Galaxy. Reads were quality checked and trimmed using FastQC (Galaxy version 0.72+galaxy1) and Trim Galore! (Galaxy version 0.6.3). Trimmed reads were aligned to the *V. natriegens* genome (9) using Bowtie 2 (Galaxy version 2.3.4.3+galaxy0) Citation Harris MT, Ho TC, Fruchtman H, Garin ME, Kubatin V, Lu T, Xue L, Marr MT, II. 2020. Complete genome sequences of two Vibrio natriegens bacteriophages. Microbiol Resour Announc 9:e01133-20. https://doi.org/10.1128/

Editor Simon Roux, DOE Joint Genome

Copyright © 2020 Harris et al. This is an openaccess article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Michael T. Marr II, mmarr@brandeis.edu.

Received 29 September 2020 Accepted 15 October 2020 Published 5 November 2020

bRosenstiel Basic Medical Sciences Research Center, Waltham, Massachusetts, USA



(10). Unaligned reads were used to assemble the bacteriophage genome sequences using Unicycler (Galaxy version 0.4.8.0) (11). The Unicycler output contained two separate scaffolds. Both scaffolds were considered to represent complete bacteriophage genomes based on the following two lines of evidence: (i) the Unicycler output flagged both genomes as "circular," indicating overlapping reads across the entire scaffold, and (ii) manual permutation and read mapping with Bowtie 2 showed consistent coverage across the original scaffold origin. The genomes were annotated using Prokka (Galaxy version 1.14.5) (12).

The genome of bacteriophage VH1_2019 is 246,059 bp with a GC content of 42.6%. The sequence was assembled with an average base coverage of $31,443\times$. It is a member of the T4 *Myoviridae* family. The genome contains 381 open reading frames (ORFs) and 27 tRNA genes. BLASTn searches using NCBI Virus indicate that VH1_2019 is closely related to the broad-host-range vibriophage KVP40 (13). The two viruses are 98% identical over 97% of the length of the VH1_2019 sequence.

The genome of bacteriophage VH2_2019 is 81,721 bp with a GC content of 48%. It was sequenced with an average base coverage of $2,318\times$. It is a member of the *Siphoviridae* family. The genome contains 108 ORFs. NCBI Virus BLASTn analysis indicates that VH2_2019 is most closely related to vibriophage vB_VpS_PG28. The two viruses are 88% identical over 94% of the length of the VH2_2019 sequence.

Data availability. The annotated genome sequences for VH1_2019 and VH2_2019 were deposited under GenBank accession numbers MN794232 and MN794238, respectively. The raw sequencing reads were deposited in the Sequence Read Archive (SRA) under BioProject number PRJNA663890.

ACKNOWLEDGMENT

Funding for this work was provided by the biology department at Brandeis University.

REFERENCES

- Payne WJ. 1958. Studies on bacterial utilization of uronic acids. Ill. Induction of oxidative enzymes in a marine isolate. J Bacteriol 76:301–307. https://doi.org/10.1128/JB.76.3.301-307.1958.
- Eagon RG. 1962. Pseudomonas natriegens, a marine bacterium with a generation time of less than 10 minutes. J Bacteriol 83:736–737. https:// doi.org/10.1128/JB.83.4.736-737.1962.
- Weinstock MT, Hesek ED, Wilson CM, Gibson DG. 2016. Vibrio natriegens as a fast-growing host for molecular biology. Nat Methods 13:849–851. https://doi.org/10.1038/nmeth.3970.
- Des Soye BJ, Davidson SR, Weinstock MT, Gibson DG, Jewett MC. 2018. Establishing a high-yielding cell-free protein synthesis platform derived from Vibrio natriegens. ACS Synth Biol 7:2245–2255. https://doi.org/10 .1021/acssynbio.8b00252.
- Wiegand DJ, Lee HH, Ostrov N, Church GM. 2018. Establishing a cell-free Vibrio natriegens expression system. ACS Synth Biol 7:2475–2479. https://doi.org/10.1021/acssynbio.8b00222.
- Ellis GA, Tschirhart T, Spangler J, Walper SA, Medintz IL, Vora GJ. 2019. Exploiting the feedstock flexibility of the emergent synthetic biology chassis Vibrio natriegens for engineered natural product production. Mar Drugs 17:679. https://doi.org/10.3390/md17120679.

- Salmond GP, Fineran PC. 2015. A century of the phage: past, present and future. Nat Rev Microbiol 13:777–786. https://doi.org/10.1038/nrmicro3564.
- Kot W. 2018. Genome sequencing of dsDNA-containing bacteriophages directly from a single plaque. Methods Mol Biol 1681:179–184. https://doi .org/10.1007/978-1-4939-7343-9_13.
- Wang Z, Lin B, Hervey WJ, Vora GJ. 2013. Draft genome sequence of the fastgrowing marine bacterium Vibrio natriegens strain ATCC 14048. Genome Announc 1:e00589-13. https://doi.org/10.1128/genomeA.00589-13.
- Langmead B, Salzberg SL. 2012. Fast gapped-read alignment with Bowtie 2. Nat Methods 9:357–359. https://doi.org/10.1038/nmeth.1923.
- 11. Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. PLoS Comput Biol 13:e1005595. https://doi.org/10.1371/journal.pcbi.1005595.
- Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. Bioinformatics 30:2068–2069. https://doi.org/10.1093/bioinformatics/btu153.
- Miller ES, Heidelberg JF, Eisen JA, Nelson WC, Durkin AS, Ciecko A, Feldblyum TV, White O, Paulsen IT, Nierman WC, Lee J, Szczypinski B, Fraser CM. 2003.
 Complete genome sequence of the broad-host-range vibriophage KVP40: comparative genomics of a T4-related bacteriophage. J Bacteriol 185: 5220–5233. https://doi.org/10.1128/jb.185.17.5220-5233.2003.