



Data Article

Data on the docking of phytoconstituents of betel plant and matcha green tea on SARS-CoV-2



Fatimawali ^{a,*}, Rizky Ramadhan Maulana ^a,
 Axl Laurens Lukas Windah ^a, Irma Febrianti Wahongan ^a,
 Sefren Geiner Tumilaar ^a, Ahmad Akroman Adam ^b,
 Billy Johnson Kepel ^c, Widdhi Bodhi ^c, Trina Ekawati Tallei ^d

^a Pharmacy Study Program, Faculty of Mathematics and Natural Sciences, Sam Ratulangi University, Manado 95115, Indonesia

^b Dentistry Study Program, Faculty of Medicine, Sam Ratulangi University, Manado 95115, Indonesia

^c Department of Chemistry, Faculty of Medicine, Sam Ratulangi University, Manado 95115, Indonesia

^d Department of Biology, Faculty of Mathematics and Natural Sciences, Sam Ratulangi University, Manado 95115, Indonesia

ARTICLE INFO

Article history:

Received 25 October 2020

Revised 19 March 2021

Accepted 7 April 2021

Available online 14 April 2021

Keywords:

Betel
 Matcha green tea
 Phytoconstituent
 In silico
 Docking
 SARS-CoV-2
 Antiviral

ABSTRACT

Betel (*Piper betle* L.) and green tea (*Camellia sinensis* (L) O. Kuntze) have been used for a long time as traditional medicine. The docking of phytoconstituents contained in the betel plant was evaluated against M^{pro}, and matcha green tea was evaluated against five target receptors of SARS-CoV-2 as follows: spike ectodomain structure (open state), receptor-binding domain (RDB), main protease (M^{pro}), RNA-dependent RNA polymerase (RdRp), dan papain-like protease (PL^{pro}). The evaluation was carried out based on the value of binding-free energy and the types of interactions of the amino acids at the receptors that interact with the ligands.

© 2021 The Authors. Published by Elsevier Inc.
 This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>)

* Corresponding author.

E-mail addresses: fatimawali@unsrat.ac.id (Fatimawali), trina_tallei@unsrat.ac.id (T.E. Tallei).

Specifications Table

Subject	Biological sciences
Specific subject area	Bioinformatics, in silico analysis, molecular docking
Type of data	Tables and Figures
How data were acquired	AutoDock Vina and Biovia Discovery Studio Visualizer 2020
Data format	Raw and analyzed
Parameters for data collection	<p>Direct URL to the data for betel plant: http://dx.doi.org/10.17632/s72rcpk82b.1</p> <p>Direct URL to the data for matcha green tea: http://dx.doi.org/10.17632/4dn4svm3jb.1</p> <p>In the drug discovery setting, Lipinski's rule of 5 predicts that poor absorption or permeation is more likely when there are more than 5 H-bond donors, 10 H-bond acceptors, the molecular weight is greater than 500, and the calculated Log P (CLog P) is greater than 5.</p> <p>The docking score was obtained based on the most negative Gibbs' free energy of binding generated using autodock Vina.</p> <p>The interactions between receptors' amino acid residues and the ligands were visualized using Biovia Discovery Studio 2020.</p>
Description of data collection	Betel plant phytoconstituents were obtained from GC-MS analysis; Phytoconstituents of matcha were collected from published articles listed in the references.
Data source location	The receptors' structures were retrieved from https://www.rcsb.org/ The ligands' structures were retrieved from https://pubchem.ncbi.nlm.nih.gov/
Data accessibility	<p>Repository name: Mendeley Data</p> <p>Data identification number for betel plant: http://dx.doi.org/10.17632/s72rcpk82b.1</p> <p>Data identification number for matcha green tea: http://dx.doi.org/10.17632/4dn4svm3jb.1</p> <p>Direct URL to the data for betel plant: https://data.mendeley.com/datasets/s72rcpk82b/1</p> <p>Direct URL to the data for matcha green tea: https://data.mendeley.com/datasets/4dn4svm3jb/1</p>
Related research article	T.E. Tallei, S.G. Tumilaar, A.A. Adam, Fatimawali, Evaluasi potensi polifenol Matcha sebagai agen anti-SARS-CoV-2 menggunakan pendekatan penambatan molekul, in: K. Wikantika, F.M. Dwivany, M.F. Ghazali, L.F. Yusman, C. Novianti (Eds.), ForMIND Bunga Rampai 2020, ITB Press, Bandung, 2020, pp. 147–155.

Value of the Data

- The data provide information on the results of GC-MS analysis of various phytoconstituents contained in betel plant (leaf and fruit parts).
- The data provide information on the interactions of various betel leaf and fruit as well as matcha green tea phytoconstituents on important enzyme and proteins of SARS-CoV-2, i.e.: spike ectodomain structure (open state) (PDB code: 6VYB), receptor-binding domain (RDB) (PDB code: 6YLA), main protease (M^{PRO}) (PDB code: 6LU7), RNA-dependent RNA polymerase (RdRp) (PDB code: 6M71), and papain-like protease (PLP^{PRO}) (PDB code: 6WX4).
- The data may be useful to researchers working on COVID-19 drug discovery and development;
- The data provide promising phytoconstituents for betel and matcha green tea which could serve as potential clues for the development of future therapeutics for COVID-19.

1. Data Description

Plants are sources of phytomedicine which has the potential to be developed as antiviral agents for SARS-COV-2, as has been reported by previous studies [1,2]. Betel leaf and fruit contain many phytoconstituents which reveal their uses for various therapeutic purposes. The plant or its parts can be used for the treatment of various disorders in humans such as diabetes, fungal

Table 1

Lipinski's rule of five value of betel leaf and fruit phytoconstituents.

Compound name	Molecular weight	No. H-bond acceptors	No. H-bond donors	log P	Molar refractivity	No. of violations
(5 β)Pregnane-3,20 β -diol, 14 α ,18 α -[4-methyl-3-oxo-(1-oxa-4-azabutane-1,4-diy)]-, diacetate	489	6	0	5.962	144.653	2
N1-Benzyl-N2-(bezylidenyln-benzylamino)-	403	0	1	4.276	117.477	0
25-Norisopropyl-9,19-cyclolanostan-22-en-24-one, 3-acetoxy-24-phenyl-4,4,14-trimethyl-	516	3	0	8.118	166.353	3
Milbemycin B, 6,28-anhydro-15-chloro-25-isopropyl-13-dehydro-5-O-demethyl-4-methyl-1H-2,8a-Methanocyclopenta[a]cyclopropa[e]cycloden-11-one,	590	7	1	7.752	169.584	3
1a,2,5,5a,6,9,10,10a-octahydro-5,5a,6-trihydroxy-1,4-bis(hydroxymethyl)-1,7,9-trimethyl-, [1S-(1a,1aa,2a,5 β ,5a β ,6 β ,8aa,9a,10aa)]-	364	6	3	3.593	100.307	0
1H-2,8a-methanocyclopenta[a]cyclopropa[e]cycloden-6-yl ester, [1aR-(1aa,2a,5 β ,5a β ,6 β ,8aa,9a,10aa)]-	430	6	1	5.048	122.754	1
2,4,6-Decatrienoic acid, 1a,2,5,5a,6,9,10,10a-octahydro-5,5a-dihydroxy-4-(hydroxymethyl)-1,1,7,9-tetramethyl-11-oxo-(2,3-Diphenylcyclopropyl)methyl phenyl sulfoxide, trans-2-Naphthalenemethanol, decahydro-a,a,4a-trimethyl-8-methylene-, [2R-(2a,4aa,8a β)]-benzene, 1,1',1''-[5-methyl-1-pentene-1,3,5-triyl]tris-	496	6	2	6.142	142.969	2
	332	1	0	4.417	94.863	0
	240	2	2	3.833	81.209	0
	312	0	0	5.123	98.352	1

infection, microbial infection, inflammation, antihistaminic, antiulcer, and local anesthetic [3]. Matcha, which is a green tea preparation in powder form [4], is known to have many benefits, including as a source of antioxidants and having antiviral activities [5].

The data described here include the binding free energy value (kcal/mol) of the phytoconstituents contained in betel leaf and matcha green tea which serve as ligands against various targets of SARS-CoV-2. Data on phytoconstituent from betel leaf were obtained from the results of Gas chromatography-mass spectrometry (GC-MS), while information about the phytoconstituent of matcha was obtained through literature searches. The data on the drug-likeness of the ligands based on Lipinski's rule of five are listed in **Table 1** for betel leaf and fruit, and **Table 2** for matcha green tea. The phytoconstituents of matcha green tea were obtained from the references listed in **Table 2**. The data on binding free energy resulted from the docking of betel leaf and matcha green tea is presented in **Tables 3** and **4**, respectively. **Tables 5** and **6** show the type of interaction and the interacting amino acids of the receptors with the ligands contained in

Table 2

Lipinski's rule of five value of the matcha green tea phytoconstituents.

Compound name	References	Molecular weight	No. H-bond acceptors	No. H-bond donors	$\log P$	Molar refractivity	No. of violations
(-)epicatechin 3,5-di-O-digallate (EC35G)	[7]	594	14	9	3.16	141.986	4
Rutin	[8]	610	16	10	-1.88	137.495	4
(-) epigallocatechin gallate (EGCG)	[9]	458	11	8	2.23	108.921	2
Apigenin glycoside	[10]	578	12	6	1.86	144.558	4
Flavonol 3-O- <i>b</i> -glucoside (FOG)	[11]	400	8	4	0.45	99.615	0
Myricetin 3-glucoside (M-G)	[12]	480	13	9	-1.01	107.939	1
(-) epigallocatechin (EGC)	[13]	306	7	6	1.25	74.288	1
Kaempferol	[14]	286	6	4	2.31	72.386	0
(-)epicatechin gallate (ECG)	[9]	442	10	7	2.53	107.256	1
(+)-catechin	[9]	290	6	5	1.55	72.623	1
(-)epicatechin (EC)	[9]	290	6	5	1.55	72.623	1
Myricetin	[14]	318	8	6	1.72	75.715	1
Kaempferol-3-O-glucoside	[10]	448	11	7	-0.44	104.609	2
Quercetin	[14]	302	7	5	2.01	74.050	1
(-)Epigallocatechin 3-(3-methyl-gallate) (3'Me-EGCG)	[15]	472	11	7	2.54	113.808	2
Caffeoylquinic acid (CQA)	[15]	354	9	6	-0.65	82.519	1
Chlorogenic acid	[8]	354	9	6	-0.65	82.519	1
Coumaric acid	[8]	164	3	2	1.49	44.776	0
Caffeic acid	[8]	180.16	4	3	1.20	46.441	0
Gallic Acid	[9]	170.12	5	4	0.50	38.395	1
Caffeine	[9]	194.19	5	0	0.06	49.100	0

betel plant and matcha green tea, respectively. The detail of interaction and visualization of the docking results of all phytoconstituents are provided in the supplementary data. The interaction visualization of the best 10 docking results of betel leaf and fruit phytoconstituents is provided in Fig. 1. The visualization of the interaction of matcha green tea with SARS-CoV-2 receptors is available from Supplementary data [6].

2. Experimental Design, Materials, and Methods

2.1. Receptors and ligands selection

The selection of receptors is based on the information contained in the literature. Five essentials enzyme and proteins of SARS-CoV-2 selected as receptors in this study were spike ectodomain structure (open state) (PDB code: 6VYB), receptor-binding domain (RDB) (PDB code: 6YLA), main protease (M^{pro}) (PDB code: 6LU7), RNA-dependent RNA polymerase (RdRp) (PDB code: 6M71), and papain-like protease (PL pro) (PDB code: 6WX4). The phytoconstituents of betel leaf which serve as ligands were based on GC-MS data [16]. The GC-MS procedure was carried out following the research conducted by Tumilaar et al. [17]. The phytoconstituents of matcha green tea were selected based on a literature survey as listed in Table 2.

2.2. Receptors and ligands preparation

The structures of the receptor (M^{pro}) were retrieved from Protein Data Bank (<http://www.rcsb.org>) and opened in BIOVIA Discovery Studio Visualizer 2020 [18]. After removing the water molecules and native ligands, the receptor was saved in a .pdb format. All the structures of the

Table 3Binding free energy of betel leaf phytoconstituents against SARS-CoV-2 M^{pro} (6LU7).

Ligands	Chemical formula	PubChem ID	Binding affinity (kcal/mol)
(5 β)Pregnane-3,20 β -diol, 14a,18a-[4-methyl-3-oxo-(1-oxa-4-azabutane-1,4-diyl)]-, diacetate	C ₂₈ H ₄₃ NO ₆	537,242	-11.5
N1-Benzyl-N2(bezylidenyl-benzylamino)-25-Norisopropyl-9,19-cyclolanostan-22-en-24-one, 3-acetoxyl-24-phenyl-4,4,14-trimethyl-	C ₂₈ H ₂₅ N ₃ C ₃₅ H ₄₈ O ₃	562,008 5,373,661	-8.5 -8.1
Milbemycin B, 6,28-anhydro-15-chloro-25-isopropyl-13-dehydro-5-O-demethyl-4-methyl-1H-2,8a-Methanocyclopental[a]cyclopropa[e]cycloden-11-one,	C ₃₃ H ₄₇ ClO ₇	5,367,225	-8
1a,2,5,5a,6,9,10,10a-octahydro-5,5a,6-trihydroxy-1,4-bis(hydroxymethyl)-1,7,9-trimethyl-, [1S-(1a,1aa,2a,5 β ,5a β ,6 β ,8aa,9a,10aa)]-	C ₂₀ H ₂₈ O ₆	119,057,278	-7.9
1H-2,8a-methanocyclopental[a]cyclopropa[e]cycloden-6-yl ester, [1aR-(1aa,2a,5 β ,5a β ,6 β ,8aa,9a,10aa)]-	C ₂₄ H ₃₄ O ₆	6,918,670	-7.9
2,4,6-Decatrienoic acid, 1a,2,5,5a,6,9,10,10a-octahydro-5,5a-dihydroxy-4-(hydroxymethyl)-1,1,7,9-tetramethyl-11-oxo-(2,3-Diphenylcyclopropyl)methyl phenyl sulfoxide, trans-2-Naphthalenemethanol, decahydro-a,a,4a-trimethyl-8-methylene-, [2R-(2a,4aa,8a β)]-	C ₃₀ H ₄₀ O ₆	5,367,323	-7.8
benzene, 1,1',1''-[5-methyl-1-pentene-1,3,5-triyl]tris-[3-[3-Bromophenyl]-7-chloro-3,4-dihydro-10-hydroxy-1,9(2H,10H)-acridinedione	C ₂₄ H ₂₄ C ₁₉ H ₁₃ BrClNO ₃	20,138,399 536,420	-7.6 -7.4
(22S)-21-Acetoxy-6a,11 β -dihydroxy-16a,17a-propylmethylenedioxypregna-1,4-diene-3,20-dione	C ₂₇ H ₃₆ O ₈	544,325	-7.4
2(1H)-Pyrimidinone, 5-chloro-4,6-diphenyl-Benz[e]azulene-3,8-dione, 5-[(acetyloxy)methyl]-3a,4,6a,7,9,10,10a,10b-octahydro-Alpha-phenyl-alpha-tropylacetaldehyde tosylhydrazone	C ₁₆ H ₁₁ ClN ₂ O C ₁₉ H ₂₄ O ₆	624,638 540,437	-7.3 -7.3
Pregnan-20-one, 3-(acetyloxy)-5,6:16,17-diepoxy-, (3 β ,5a,6a,16a)-Isoaromadendrene epoxide	C ₂₂ H ₂₂ N ₂ O ₂ S C ₂₃ H ₃₂ O ₅	9,602,323 265,665	-7.3 -7.2
2-[4-methyl-6-(2,6,6-trimethylcyclohex-1-enyl)hexa-1,3,5-trienyl]cyclohex-1-en-1-carboxaldehyde	C ₁₅ H ₂₄ O C ₂₃ H ₃₂ O	534,398 5,363,101	-7.1 -7.1
5a-Pregn-16-en-20-one, 3 β ,12a-dihydroxy	C ₂₅ H ₃₆ O ₅	1,756,337	-7.1
(22S)-6a,11 β ,21-Trihydroxy-16a,17a-propyl Neointermedeol	C ₂₅ H ₃₄ O ₇ C ₁₅ H ₂₆ O	11,877,394	-7
6-Chloro-3-(2-nitro-1-phenylethyl)-3,4-dihydro-1H-naphthalen-2-one	C ₁₈ H ₁₆ ClNO ₃	586,644	-7
Ethyl isoallocholate	C ₂₆ H ₄₄ O ₅	6,452,096	-7

ligands were retrieved from PubChem (<http://pubchem.ncbi.nlm.nih.gov>) in .sdf format. The files were converted into a .pdb format using Open Babel [19]. After adjusting the torque, the files were saved in .pdbqt format.

2.3. The docking process and visualization

Autodock Vina [20] was used in the docking analysis. The .pdbqt formats of ligands and receptors were copied into the Vina folder. Vina configuration was typed in notepad and saved as conf.txt. Vina program was performed in a command prompt mode. The most negative Gibbs' free energy of binding indicated the best pose. The visualization of the interacting amino acids of the receptors with the ligands was performed in Biovia Discovery Studio 2020 [18].

Table 4

Binding free energy of matcha phytoconstituents against several SARS-CoV-2 receptors.

Ligands	PubChem CID	SARS-CoV-2 Receptors PDB ID				
		6VYB	GYLA	6LU7	6MT1	6WX4
(-)epicatechin 3,5-di-O-digallate (EC35G)	467,299	-9.7	-10.0	-9.1	-9.2	-8.8
Rutin	5,280,805	-9.9	-10.1	-8.8	-8.8	-7.2
(-)epigallocatechin gallate (EGCG)	65,064	-9.2	-9.4	-8.2	-8.5	-7.6
Apigenin glycoside	44,257,854	-9.2	-10.1	-8.7	-9.1	-7.5
Flavonol 3-O-D-glucoside (FOG)	11,953,828	-9.0	-8.1	-7.8	-7.8	-6.7
Myricetin 3-glucoside (M-G)	44,259,426	-8.8	-9.2	-8.8	-8.0	-6.6
(-)Epigallocatechin (EGC)	72,277	-8.6	-8.4	-7.1	-7.5	-6.7
Kaempferol	5,280,863	-8.6	-7.9	-7.7	-7.1	-6.7
(-)epicatechin gallate (ECG)	107,905	-8.5	-9.0	-8.2	-8.3	-7.4
(+)-catechin	9064	-8.4	-8.2	-7.2	-6.8	-6.6
(-)epicatechin (EC)	72,276	-8.4	-7.8	-7.1	-7.0	-6.7
Myricetin	5,281,672	-8.4	-8.4	-7.3	-7.3	-7.1
Kaempferol-3-O-glucoside	5,282,102	-8.4	-8.9	-8.4	-7.9	-6.6
Quercetin	5,280,343	-8.3	-8.4	-7.4	-7.6	-7.0
(-)Epigallocatechin	9,804,842	-8.3	-8.1	-7.6	-8.6	-7.6
3-(3-methyl-gallate) (3'Me-EGCG)						
Caffeoyquinic acid (CQA)	10,155,076	-8.1	-8.0	-7.2	-7.0	-6.6
Chlorogenic acid	1,794,427	-7.9	-7.3	-7.6	-6.9	-7.0
Coumaric acid	9,840,292	-6.7	-7.0	-7.2	-5.3	-6.4
Caffeic acid	689,043	-6.7	-5.9	-5.7	-5.3	-5.3
Gallic acid	370	-6.3	-6.1	-5.5	-5.6	-5.2
Caffeine	2519	-6.1	-6.0	-5.2	-5.1	-5.2

Table 5

Interacting amino acids of the main protease (6LU7) with the 10 best ligands of betel leaf and fruit.

PubChem CID	Binding free energy (kcal/mol)	No. of bonds	Interacting residues and H-bond formation
537,242	-11.5	18	Van der Waals: ASN(A142), GLY(A143) CYS(A145), HIS(A164), ASP(A187), MET(A49), TYR(A54), ARG(A188), PRO(A168), LEU(A167), THR(A190), GLN(A189), GLU(A166), MET(A165); conventional H-bond: GLN(A192); unfavorable positive-positive: HIS(A41); attractive charge and pi-anion: HIS(A41); pi-sigma: HIS(A41).
562,008	-8.5	21	Van der Waals: GLU(A166), MET(A49), THR(A24), THR(A25), THR(A26), GLY(A143), ASN(A142), ARG(A188), ASP(A187), HIS(A164), LEU(A141), GLN(A189), HIS(A163), HIS(A172), PHE(A140); unfavorable positive-positive: HIS(A41); pi-cation: HIS(A41); pi-pi t-shaped: HIS(A41); pi-alkyl: CYS(A145), LEU(A27), MET(A165).
5,373,661	-8.1	19	Van der Waals: THR(A26), THR(A25), ASN(A142), GLY(A143), HIS(A41), CYS(A145), SER(A144), LEU(A141), GLU(A166), ARG(A188), THR(A190), GLN(A192), GLN(A189), HIS(A164), MET(A49), THR(A24); conventional H-bond: THR(A45), SER(A46); pi-alkyl: MET(A165).
5,367,225	-8	14	Van der Waals: THR(A25), LEU(A27), MET(A49), GLN(A189), CYS(A145), HIS(A41), SER(A144), MET(A165), PHE(A140), LEU(A141), GLU(A166); conventional H-bond: THR(A26), GLY(A143); pi-alkyl: HIS(A163).
119,057,278	-7.9	14	Van der Waals: MET(A165), GLN(A189), ASN(A142), SER(A144), GLY(A143), HIS(A172), PHE(A140); conventional H-bond: GLU(A166), 2HIS(A163), LEU(A141); unfavorable positive-positive: 2HIS(A41); alkyl: CYS(A145)
6,918,670	-7.9	16	Van der Waals: ASN(A142), GLN(A189), HIS(A164), ASP(A187), ARG(A188), MET(A165), LEU(A141), PHE(A140), LEU(A167), PRO(A168); conventional H-bond: 3GLU(A166), HIS(A172); unfavorable positive-positive: HIS(A163); pi-alkyl: HIS(A41).

(continued on next page)

Table 5 (continued)

PubChem CID	Binding free energy (kcal/mol)	No. of bonds	Interacting residues and H-bond formation
5,367,323	-7.8	13	Van der Waals: GLY(A143), HIS(A172), PHE(A140), ASN(A142), MET(A165), PRO(A168), LEU(A167); conventional H-bond: GLU(A166), HIS(A163); unfavorable positive-positive: GLN(A189); pi-alkyl: 3LEU(A141)
562,543	-7.8	15	Van der Waals: LEU(A141), PHE(A140), GLU(A166), HIS(A163), HIS(A172), HIS(A164), ASP(A187), TYR(A54), ARG(A188), GLN(A189), CYS(A145); conventional H-bond: HIS(A41); pi-cation: HIS(A41); pi-sulfur: MET(A49); pi-pi stacked & pi-alkyl: MET(A165).
165,258	-7.8	12	Van der Waals: ASP(A187), ARG(A188), GLN(A189), MET(A165), HIS(A164), MET(A49), LEU(A27), GLY(A143), ASN(A142), GLU(A166); conventional H-bond: HIS(A41); alkyl: CYS(A145)
20,138,399	-7.6	15	Van der Waals: LEU(A141), PHE(A140), HIS(A172), HIS(A163), HIS(A164), ASP(A187), ARG(A188), TYR(A54), THR(A190), GLN(A189), GLU(A166); pi-sulfur: MET(A165), CYS(A145); pi-pi stacked: HIS(A41); pi-alkyl: MET(A49).

Table 6

Hydrogen bond interaction of the amino acids of the receptors with phytoconstituents in matcha. The remaining interaction data are available from Tallei et al. [6].

Receptors	Ligands	Interacting amino acids			
		Conventional H-bond	Carbon H-bond	Pi-donor H-bond	Pi-carbon H-bond
6VYB	(-)epicatechin	ASN(C1108), LYS(C1038)			
	3,5-di-O-digallate (EC35G)		GLY(C910),		
	Rutin	ARG(B1039), ARG(C1039), ARG(A1039), 2ASN(C1023), ARG(A1019)	TYR(A904)		
	(-)epigallocatechin gallate (EGCG)	2SER(B94), ASN(B99), ARG(B190)			
	Apigenin glycoside	ARG(A1039), SER(B1030), THR(B1027), ASP(A1041)			
	Flavonol 3-O-D-glucoside (FOG)	THR(B998), TYB(B756), THR(A998), 2ASP(A994), THR(C998)			
	Myricetin 3-glucoside (M-G)	GLN(A954), GLN(A1010)	GLY(B769), 2GLN(A954)		
	(-)Epigallocatechin (EGC)	LEU(A861), LYS(A733), GLY(A1059)	PRO(A1057)		
	Kaempferol	2THR(A549), ASN(B978), MET(B740), TYR(B714), ARG(B1000)	GLY(A541)		
	(-)epicatechin gallate (ECG)	GLY(C744), TYR(C741), 3ARG(31,000), LEU(C977)			

(continued on next page)

Table 6 (continued)

Receptors	Ligands	Interacting amino acids			
		Conventional H-bond	Carbon H-bond	Pi-donor H-bond	Pi-carbon H-bond
6YLA	(-)Epigallocatechin 3-(3-methyl-gallate) (3''Me-EGCG)	2IHK(B1027), ARG(B1029)		IHK(C1027)	
	Caffeoylquinic acid (CQA)	GLN(A672), ARG(A675), ARG(C1014), ARG(A1019), GLU(A773), GLN(C954)			
	Chlorogenic acid	THR(A961), GLU(A1017), GLU(B773), GLN(A954)		GLY(B769)	
	Coumaric acid	SER(A514), TYR(B200), PHE(A515), THR(A430)			
	Caffeic acid	HIS(A1048), 2GLN(B1036)		GLY(B1035)	
	Gallic acid	GLN(A1005), GLN(B1002), THR(B1006)			
	Caffeine	GLU(A166), GLY(A143), SER(A144)			CYS(A142)
	(-)epicatechin 3,5-di-O-digallate (EC35G)	TYR(L:93), LYS(H:43), ALA(H:172), 2GLU(H:152), THR(H:116), GLY(L:47)			
	Rutin	GLY(H:112), THR(H:114), GLU(H:152), ALA(H:92), VAL(H:115)		GLY(L:47)	
	(-)epigallocatechin gallate (EGCG)	SER(C:62), MET(H:2), GLU(L:61), THR(H:0), THR(B:0), ASP(B:107), GLN(B:1)			
	Apigenin glycoside	SER(C:174), GLN(C:172), SER(H:75);		SER(H:75), SER(C:174)	
	Flavonol 3-O-D-glucoside (FOG)	2GLN(L:48)		GLN(H:39)	
	Myricetin 3-glucoside (M-G)	THR(E:385), THR(H:0), ASP(H:107), SER(L:62), THR(B:0), GLN(B:1), ASP(B107)			
	(-)Epigallocatechin (EGC)	GLN(H:39)			
	Kaempferol	LYS(L:45), GLN(L:44)			
	(-)epicatechin gallate (ECG)	LYS(C:213), 2ASN(C:216), 4GLU(C:219), GLY(C:218), LYS(B:218)		PRO(C:125), SER(B:132), PHE(C:122)	
	(+)-catechin	GLU(H:152), GLN(L:44), LYS(L:45), GLN(H:39)			
	(-)epicatechin (EC)	LYS(L:45), GLN(L:44)			
	Myricetin	ILE(H:93), GLN(H:39), GLN(L:44), LYS(L:45)			GLY(L:47)
	Kaempferol-3-O-glucoside	MET(H:2), LYS(A:386), GLN(H:3)			
	Quercetin	LYS(L:45), 2GLN(L:48), ILE(H:93)			
	(-)Epigallocatechin 3-(3-methyl-gallate) (3''Me-EGCG)	VAL(C:64), GLY(C:63), LYS(E:528), ASP(A:389), 2LYS(A:529), GLU(E:327)			
	Caffeoylquinic acid (CQA)	2GLN(L:48), GLY(H:42), 2VAL(H:115), ALA(H:92)		PRO(H:41), GLN(H:39)	

(continued on next page)

Table 6 (continued)

Receptors	Ligands	Interacting amino acids			
		Conventional H-bond	Carbon H-bond	Pi-donor H-bond	Pi-carbon H-bond
6LU7	Chlorogenic acid	GLY(H:28), 2ASN(H:77), ILE(H:30)			
	Coumaric acid	ASP(E:389), LYS(E:386), GLY(C:63), TYR(E:369), ASN(E:370), VAL(C:64), SER(E:366), 2ASP(C:66)			
	Caffeic acid	LYS(A:528), ASP(L:66), ASN(A:370)			
	Gallic acid	ASN(A:388), TYR(A:369), GLU(L:61), VAL(L:64), ASP(A:364)			
	Caffeine	ARG(H:59), TYR(L:31), SER(H:103)	PRO(E:412), TYR(L:98), TYR(L:31)		
	(-)epicatechin	THR(A24), THR(A26), THR(A46), HIS(A163), HIS(A164), MET(A165)	GLN(A189)		
	3,5-di-O-digallate (EC35G)				
	Rutin	THR(A26), PHE(A140), LEU(A141), ASN(A142), GLY(A143), HIS(A163), GLU(A166), THR(A190)			
	(-)epigallocatechin gallate (EGCG)	PHE(A140), HIS(A164), MET(A165)			
	Apigenin glycoside	LEU(A141), 2SER(A144), CYS(A145), HIS(A163), GLU(A166)	CYS(A145)		
	Flavonol 3-O- <i>p</i> -glucoside (FOG)	LEU(A141), GLY(A143), SER(A144), CYS(A145)	MET(A165), GLU(A166)		
	Myricetin 3-glucoside (M-G)	LEU(A141), ASN(A142), GLY(A143)			
	(-)Epigallocatechin (EGC)	HIS(A41)			
	Kaempferol	TYR(A54), ASP(A187); PHE(A140), HIS(164), MET(A165)	GLU(A166)		
	(-)epicatechin gallate (ECG)				
	(+)-catechin	GLU(A166), THR(A190); THR(A26), HIS(A41), GLN(A189)	GLN(A192)		
	(-)epicatechin (EC)				
	Myricetin	GLY(A143), SER(A144), ARG(A188)	GLU(A166)		
	Kaempferol-3-O-glucoside	THR(A24), THR(A26), THR(A46), HIS(A163), HIS(A164), MET(A165)	GLN(A189)		
GLN(A189)	Quercetin	TYR(A54), ASP(A187)	GLU(A166)		
	(-)Epigallocatechin 3-(3-methyl-gallate) (3''Me-EGCG)	LEU(A141), 2CYS(A145), THR(A190), ASN(A188)	GLU(A166),		
	Caffeoylquinic acid (CQA)	LEU(A141), GLY(A143), 2SER(A144), CYS(A145), HIS(A163), GLU(A166), 2THR(A190);	MET(A165)		
	Chlorogenic acid	ASN(A142), SER(A144), 2THR(A190)			

(continued on next page)

Table 6 (continued)

Receptors	Ligands	Interacting amino acids			
		Conventional H-bond	Carbon H-bond	Pi-donor H-bond	Pi-carbon H-bond
6M71	Coumaric acid	LEU(A141), GLY(A143), 2SER(A144), CYS(A145), THR(A190)		PRO(A168)	
	Caffeic acid	GLU(A166), GLY(A143), SER(A144)		CYS(A145).	
	Gallic acid	LEU(A141), GLY(A143), CYS(A145), GLU(A166), GLN(A189)			CYS(A145)
	Caffeine	GLY(A143), GLU(A166)	LEU(A141), 2CYS(A145), GLN(A189)		
	(-)epicatechin	THR(A:710), ASN(A:781)		LYS(A:780)	
	3,5-di-O-digallate (EC35G)	2SER(A:708), LYS(A:780), SER(A:784), HIS(A:133), ASN(A:138)			
	Rutin	LYS(A:47), 2TYR(A:129), SER(A:784), LYS(A:780), ASN(A:138)		ASP(A:135)	
	(-)epigallocatechin gallate (EGCG)	3ASN(A:781), 2SER(A:709), ALA(A:706), 2SER(A:784)			
	Apigenin glycoside	THR(A:394), ARG(A:349), LEU(A:245), LEU(A:251)	THR(A:319), CYS(A:395)		
	Flavonol 3-O- <i>D</i> -glucoside (FOG)	VAL(A:675)	ARG(A:457)		
6M71	Myricetin 3-glucoside (M-G)	LYS(A:47) THR(A:710), ASN(A:781), SER(A:709)		2GLY(A:774)	
	(-)Epigallocatechin (EGC)	LYS(A:47), TYR(A:129), 2SER(A:784)			
	Kaempferol	TYR(A:689), ILE(A:494), ARG(A:569), 2ASN(A:496)			
	(-)epicatechin gallate (ECG)	2SER(A:709), HIS(A:133), SER(A:784)			
	(+)-catechin	TYR(A:129), SER(A:784), PHE(A:134), LYS(A:780), SER(A:772)			
	(-)epicatechin (EC)	TYR(A:129), SER(A:709), GLN(A:778), 2THR(A:710), ASP(A:711), LYS(A:47)	TYR(A:129)		
	Myricetin	THR(A:710), 2SER(A:784)	GLY(A:774)		
	Kaempferol-3-O-glucoside	ASN(A:781), SER(A:709), ASN(A:138)		TYR(A:129), TYR(A:32)	
	Quercetin	ASN(A:628)			
	(-)Epigallocatechin 3-(3-methyl-gallate) (3'Me-EGCG)	LYS(A:780), SER(A:784), LYS(A:47), TYR(A:32)			
6M71	Caffeoylquinic acid (CQA)	ASP(A:623), CYS(A:622), LYS(A:621), PHE(A:793), SER(A:795), ASP(A:618)			
	Chlorogenic acid	THR(A:206), ASN(A:208)			

(continued on next page)

Table 6 (continued)

Receptors	Ligands	Interacting amino acids			
		Conventional H-bond	Carbon H-bond	Pi-donor H-bond	Pi-carbon H-bond
6WX4	Coumaric acid	LYS(A:47), SER(A:709), 2ASN(A:781), SER(A:784)			
	Caffeic acid	TYR(A:619), GLU(A:811), TRP(A:800)			
	Gallic acid	ASP(A:761), TRP(A:617)			
	Caffeine	TYR(B:135), GLY(B:144), TYR(B:138)		TYR(B:135), ASP(B:148)	
	Rutin	2GLU(D:252), TYR(D:252), SER(D:212), LEU(D:211)			
	(-)epicatechin	TYR(D:213), GLU(D:214), LYS(D:306)			
	3,5-di-O-digallate (EC35G)	164), TYR(D:273), 2GLY(D:163)		PRO(D:248)	
	(-)epigallocatechin gallate (EGCG)	HIS(D:175), THR(D:74), TYR(D:154)		GLN(D:174)	
	Apigenin glycoside	2SER(D:180), ASN(D:308), GLU(D:124)			
	Flavonol 3-O-D-glucoside (FOG)	LEU(D:58), ASN(D:60), ASP(D:61)		GLN(D:30), PHE(D:31)	
	Myricetin 3-glucoside (M-G)	LYS(D:306), GLU(D:307), 2GLU(D:214), TYR(D:305)			
	(-)Epigallocatechin (EGC)	ASP(D:164)			
	Kaempferol	2GLY(D:163), ASP(D:164), TYR(D:273), ARG(D:166)			
	(-)epicatechin gallate (ECG)	SER(D:212)			
	(+)-catechin	LYS(D:306), 2GLU(D:307), TYR(D:305), TYR(D:213), GLU(D:214)			
	(-)epicatechin (EC)	GLY(D:266), THR(D:301), ASP(D:164)			
	Myricetin	LYS(D:297), SER(D:212), THR(D:210)			
	Kaempferol-3-O-glucoside	TYR(D:273)			
	Quercetin	THR(D:301), ASP(D:164), TYR(D:273)			
	(-)Epigallocatechin 3-(3-methyl-gallate) (3''Me-EGCG)	2GLU(D:307), 2LYS(D:217), 2LYS(D:306), THR(D:257), TYR(D:251)			
	Caffeoylquinic acid (CQA)	THR(D:257), TYR(D:213), TYR(D:251), GLU(D:214)			
	Chlorogenic acid	GLU(D:252), LYS(D:217)		THR(D:257)	
	Coumaric acid	LYS(D:217), SER(D:212)			
	Caffeic acid	GLU(D:214)			
	Gallic acid	ARG(D:116), THR(D:301)			
	Caffeine				

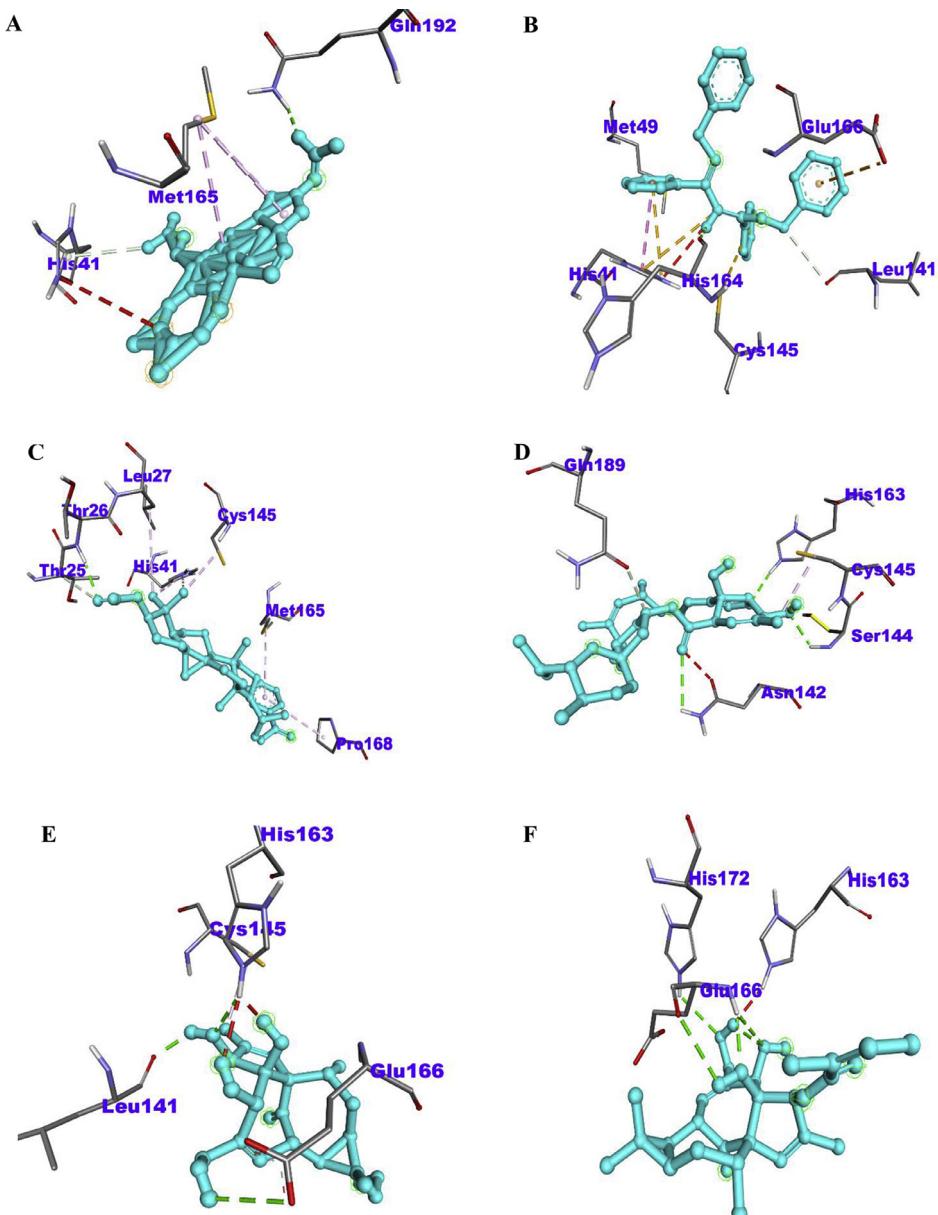


Fig. 1. The 2D diagram showing the types of amino acid residues involved in the bond between the phytoconstituents in betel plant and the M^{pro} receptor of Sars-CoV-2. (A) PubChem ID 537,242, (B) PubChem ID 562,008, (C) PubChem ID 5,373,661, (D) PubChem ID 5,367,225, (E) PubChem ID 119,057,278, (F) PubChem ID 6,918,670, (G) PubChem ID 5,367,323, (H) PubChem ID 562,543, (I) PubChem ID 165,258, (J) PubChem ID 20,138,399.

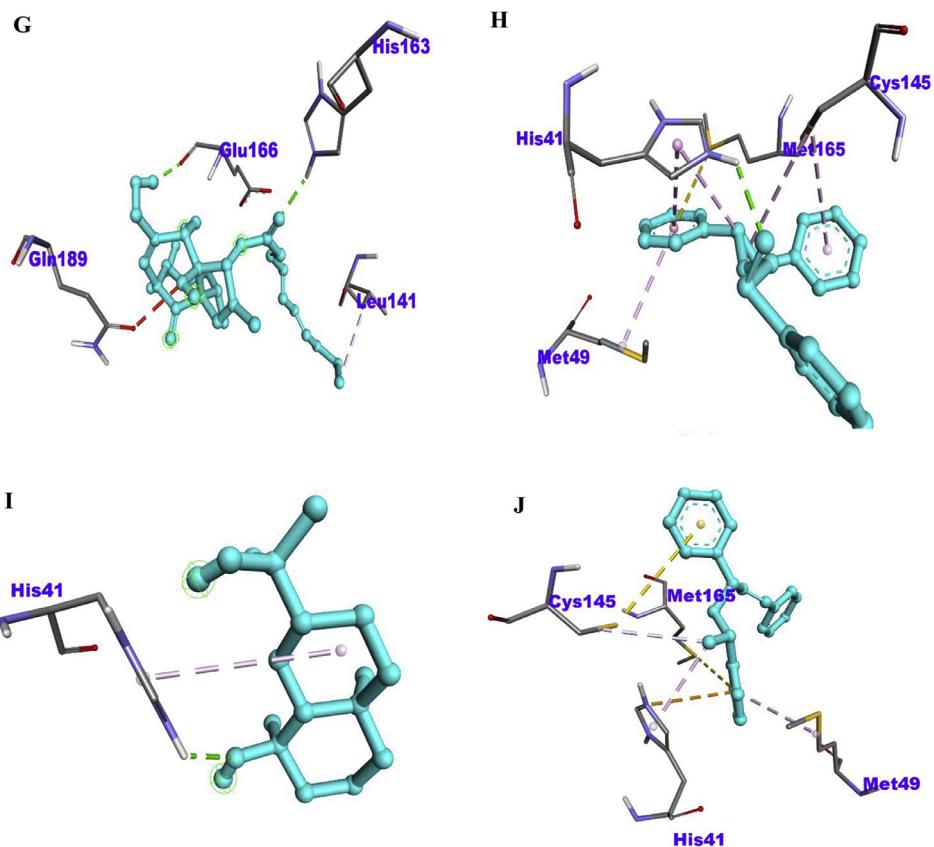


Fig. 1. Continued

Ethics Statement

The work did not involve the use of endangered species of wild flora.

Supplementary Data

Supplementary data to this article can be found at <http://dx.doi.org/10.17632/w8h74c6hsy.1> and <https://doi.org/10.17632/4dn4svm3jb.1>.

CRediT Author Statement

Fatimawali: Conceptualization, Methodology, Data curation, Writing – original draft, Writing – review & editing; **Rizky Ramadhan Maulana:** Software, Validation; **Axl Laurens Lukas Windah:** Software, Validation; **Irma Febrianti Wahongan:** Visualization, Investigation; **Sefren Geiner Tumilaar:** Visualization, Investigation; **Ahmad Akroman Adam:** Data curation, Writing – review & editing; **Billy Johnson Kepel:** Data curation, Writing – review & editing; **Widdhi Bodhi:** Visualization, Investigation; **Trina Ekawati Tallei:** Conceptualization, Methodology, Data curation, Writing – review & editing, Supervision, Writing – review & editing.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships which have or could be perceived to have influenced the work reported in this article.

Acknowledgments

This research was funded by the Directorate of Research and Community Service, Ministry of Research and Technology / National Research and Innovation Agency under the scheme of Excellent Applied Research in Higher Education (Contract No. [1179/UN12.13/LT/2020](#)) and the COVID-19 Refocusing Research scheme for the fiscal year 2020 (Contract No. [206/SP2H/AMD/LT/DRPM/2020](#)).

References

- [1] T.E. Tallei, S.G. Tumilaar, N.J. Niode, F. Fatimawali, B.J. Kepel, R. Idroes, Y. Effendi, S.A. Sakib, T.B. Emran, Potential of plant bioactive compounds as SARS-CoV-2 main protease (M^{pro}) and spike (S) glycoprotein inhibitors: a molecular docking study, *Scientifica* (Cairo) (2020) 2020Article ID 6307457, doi:[10.1155/2020/6307457](#).
- [2] A. Rakib, A. Paul, M. Chy, S.A. Sami, S.K. Baral, S.M. Majumder, A.M. Tareq, M.N. Amin, A. Shahriar, M.Z. Uddin, M. Dutta, T.E. Tallei, T.B. Emran, J. Simal-Gandara, Biochemical and computational approach of selected phyto-compounds from *Tinospora crispa* in the management of COVID-19, *Molecules* 25 (17) (2020) 3936, doi:[10.3390/molecules25173936](#).
- [3] R. Sengupta, J.K. Banik, A review on betel leaf (PAN), *Int. J. Pharm. Sci.* 4 (12) (2013) 4519–4524, doi:[10.13040/IJPSR.0975-8232.4\(12\).4519-24](#).
- [4] S. Farooq, A. Sehgal, Antioxidant activity of different forms of green tea: loose leaf, bagged and matcha, *Curr. Res. Nutr. Food. Sci.* J. 6 (2018) 35–40, doi:[10.12944/CRNFSJ.6.1.04](#).
- [5] K. Jakubczyk, J. Kochman, A. Kwiatkowska, J. Kałduńska, K. Dec, D. Kawczuga, K. Janda, Antioxidant properties and nutritional composition of matcha green tea, *Foods* 9 (4) (2020) 483, doi:[10.3390/foods9040483](#).
- [6] T.E. Tallei, Supplementary Dataset on the Docking of Phytoconstituents of Matcha Green Tea on SARS-CoV-2 . Mendeley Data, V1, 2020 </Dataset>, doi:[10.17632/4dn4svm3jb.1](#).
- [7] L. Yao, Y. Jiang, N. Datta, N. Datta, R. Singanayagam, X. Liu, J. Duan, K. Raymont, A. Lisle, Y. Xu, HPLC analyses of flavanols and phenolic acids in the fresh young shoots of tea (*Camellia sinensis*) grown in Australia, *Food Chem* 84 (2) (2004) 253–263, doi:[10.1016/s0308-8146\(03\)00209-7](#).

- [8] M. Jeszka-Skowron, A. Zgola-Grześkowiak, Analysis of antioxidant activity, chlorogenic acid, and rutin content of *Camellia sinensis* infusions using response surface methodology optimization, Food Anal. Meth. 7 (2014) 2033–2041, doi:[10.1007/s12161-014-9847-1](https://doi.org/10.1007/s12161-014-9847-1).
- [9] Y. Kim, K.G. Lee, M.K. Kim, M. K., Volatile and non-volatile compounds in green tea affected in harvesting time and their correlation to consumer preference, Int. J. Food Sci. Technol. 53 (10) (2016) 3735–3743, doi:[10.1007/s13197-016-2349-y](https://doi.org/10.1007/s13197-016-2349-y).
- [10] I.M. Savić, V.D. Nikolić, I.M. Savić, L.B. Nikolić, M.D. Jović, M.D. Jović, The qualitative analysis of the green tea extract using ESI-MS method, Savremene tehnologije 3 (1) (2014) 30–37, doi:[10.3724/sp.j.1123.2015.04028.10.5937/savteh1401030s](https://doi.org/10.3724/sp.j.1123.2015.04028.10.5937/savteh1401030s).
- [11] H. Jiang, U.H. Engelhardt, C. Thräne, Maiwald B, J. Stark, Determination of flavonol glycosides in green tea, oolong tea and black tea by UHPLC compared to HPLC, Food Chem 183 (2015) 30–35, doi:[10.1016/j.foodchem.2015.03.024](https://doi.org/10.1016/j.foodchem.2015.03.024).
- [12] Z. Wang, Y. Sha, X. Yu, Y. Liang, Determination of flavonol glycosides in tea samples by ultra-high performance liquid chromatography-photodiode array detection-tandem mass spectrometry, Se Pu 33 (9) (2015) 974–980 PMID: 26753286, doi:[10.3724/sp.j.1123.2015.04028](https://doi.org/10.3724/sp.j.1123.2015.04028).
- [13] W.C. Reyaert, Green tea catechins: their use in treating and preventing infectious diseases, BioMed. Res. Int. 9105261. (2018), doi:[10.1155/2018/9105261](https://doi.org/10.1155/2018/9105261).
- [14] B. Jegannathan, P.A. Punyasiri, J.D. Kottawa-Arachchi, M.A. Ranatunga, I.S. Abeysinghe, M.T. Gunasekare, B.M. Bandara, Genetic variation of flavonols, myricetin, and kaempferol in the Sri Lankan Tea (*Camellia sinensis* L.) and their health-promoting aspects, Int. J. Food Sci. 6057434 (2016), doi:[10.1155/2016/6057434](https://doi.org/10.1155/2016/6057434).
- [15] K. Kang, K.H. Cha, D. Song, S.M. Kim, E.H. Lee, H.J. Lee, C.W. Nho, C. Pan, Inhibitory effect of gastrointestinal lipolysis by green tea, coffee, and gomchui (*Ligularia fischeri*) tea during simulated digestion, FASEBJ 26 (2012) 1b360–1b360, doi:[10.1096/fasebj.26.1_supplement.1b360](https://doi.org/10.1096/fasebj.26.1_supplement.1b360).
- [16] F. Fatimawali, T.E. Tallei, Supplementary Dataset of Betel Plant GC-MS and Docking results, Mendeley Data V1 (2021), doi:[10.17632/s72rcpk82b.1](https://doi.org/10.17632/s72rcpk82b.1).
- [17] S.G. Tumilaar, F. Fatimawali, N.J. Niode, Y. Effendi, R. Idroes, A.A. Adam, A. Rakib, T.B. Emran, T.E. Tallei, The potential of leaf extract of *Pangium edule* Reinw as HIV-1 protease inhibitor: a computational biology approach, J. Appl. Pharm. Sci. 11 (01) (2021) 101–110, doi:[10.7324/JAPS.2021.110112](https://doi.org/10.7324/JAPS.2021.110112).
- [18] Systèmes D.B. Discovery Studio Modeling Environment, Release 2020. <https://discover.3ds.com/discovery-studio-visualizer-download>, 2020 (accessed 10 June 2020).
- [19] N.M. O'Boyle, M. Banck, C.A. James, C. Morley, T. Vandermeersch, G.R. Hutchison, Open Babel: an open chemical toolbox, J. Cheminform. 3 (2011) 33, doi:[10.1186/1758-2946-3-33](https://doi.org/10.1186/1758-2946-3-33).
- [20] O. Trott, A.J. Olson, AutoDock Vina: improving the speed and accuracy of docking with a new scoring function, efficient optimization and multithreading, J. Com. Chem. 31 (2010) 455–461, doi:[10.1002/jcc.21334](https://doi.org/10.1002/jcc.21334).