

Review

Genetics and Epigenetics of Atopic Dermatitis: An Updated Systematic Review

Maria J Martin ^{1,2,†}, Miguel Estravís ^{1,2,3,†}, Asunción García-Sánchez ^{1,2,3,*}, Ignacio Dávila ^{1,2,4}, María Isidoro-García ^{1,2,5,6} and Catalina Sanz ^{1,2,7}

- ¹ Institute for Biomedical Research of Salamanca (IBSAL), 37007 Salamanca, Spain; mjmartinma@saludcastillayleon.es (M.J.M.); estravis@usal.es (M.E.); idg@usal.es (I.D.); misidoro@usal.es (M.I-G.); catsof@usal.es (C.S.)
- ² Network for Cooperative Research in Health–RETICS ARADyAL, 37007 Salamanca, Spain
- ³ Department of Biomedical and Diagnostics Sciences, University of Salamanca, 37007 Salamanca, Spain
- ⁴ Department of Immunoallergy, Salamanca University Hospital, 37007 Salamanca, Spain
- ⁵ Department of Clinical Biochemistry, University Hospital of Salamanca, 37007 Salamanca, Spain
- ⁶ Department of Medicine, University of Salamanca, 37007 Salamanca, Spain
- ⁷ Department of Microbiology and Genetics, University of Salamanca, 37007 Salamanca, Spain
- * Correspondence: chonela@usal.es; Tel.: +3-492-329-1100
- + These authors equally contribute to the manuscript.

Received: 12 March 2020; Accepted: 15 April 2020; Published: 18 April 2020



Abstract: Background: Atopic dermatitis is a common inflammatory skin disorder that affects up to 15–20% of the population and is characterized by recurrent eczematous lesions with intense itching. As a heterogeneous disease, multiple factors have been suggested to explain the nature of atopic dermatitis (AD), and its high prevalence makes it necessary to periodically compile and update the new information available. In this systematic review, the focus is set at the genetic and epigenetic studies carried out in the last years. Methods: A systematic literature review was conducted in three scientific publication databases (PubMed, Cochrane Library, and Scopus). The search was restricted to publications indexed from July 2016 to December 2019, and keywords related to atopic dermatitis genetics and epigenetics were used. Results: A total of 73 original papers met the inclusion criteria established, including 9 epigenetic studies. A total of 62 genes and 5 intergenic regions were described as associated with AD. Conclusion: *Filaggrin (FLG)* polymorphisms are confirmed as key genetic determinants for AD development, but also epigenetic regulation and other genes with functions mainly related to the immune system and extracellular matrix, reinforcing the notion of skin homeostasis breakage in AD.

Keywords: atopic dermatitis; genetics; epigenetics; skin barrier; genetic association studies; DNA methylation; omics

1. Introduction

Atopic dermatitis (AD), also known as atopic eczema, is a common inflammatory skin disorder that affects up to 15–20% of children [1] and 7–10% of adults [2] in developed countries. AD typically develops during childhood and is characterized by recurrent eczematous lesions with intense itching [1]. It is considered the first step of the atopic march, associated with an increased risk of developing allergic rhinoconjunctivitis, asthma, or food allergy [3]. Worldwide, the prevalence of AD used to be higher in countries with higher incomes. However, due to the globalization process and a more westernized way of life, an increase of AD prevalence in low-income countries of Africa and East Asia has been reported, stressing the role of environment together with genetic and immunologic factors in the pathogenicity of AD [4].



AD is a heterogeneous disease. Thus, interactions between susceptibility genes, environmental factors, impaired barrier skin integrity, skin microbiota, and immune deregulation have been proposed to explain the nature of AD [5]. An urban way of life is one of the most clearly related environmental factors, as supported by a consistently higher incidence of eczema in urban versus rural areas [6]. Diet is also a risk factor, and a regular intake of fresh fruit and fish during pregnancy and childhood has shown some effectiveness in preventing AD [7–9]. Furthermore, it has been reported that a family history of asthma, hay fever, or eczema is associated with AD in the offspring, the risk being higher when both parents suffer from eczema [10].

AD is associated with atopic comorbidities such as asthma, allergic rhinoconjunctivitis, and food allergy, as well as with non-atopic entities such as inflammatory diseases and psychological disorders [11,12]. Genetic association studies have confirmed that atopic comorbidities share genetic susceptibility [13,14]. The heritability of AD in twin studies was estimated to be nearly 75%, and the association between asthma and AD was nearly 85% explained by genetic pleiotropy [15].

Two main reviews about the genetics of AD have been performed in the last decade. The first one was published in 2010 and compiled all the existent genetic studies related to AD [16], reporting variants in 81 genes, 46 of which had shown positive association with the disease. In 2016, Bin and Leung published an update on the topic, including genetic and epigenetic studies from 2009 to June 2016 [17]. The aim of this systematic review is to compile the most recent publications on the genetics of atopic dermatitis, also including epigenetic studies. Original articles about genetic variation and polymorphisms in patients with atopic dermatitis or atopic eczema were sought. Both adult and children studies were included. Comparison to healthy control was preferred, but not all studies performed it. Although literature reviews were excluded, we analyzed some meta-analyses due to the valuable information they contained.

2. Materials and Methods

This systematic review has been performed using the PRISMA guidelines for Systematic Reviews and Meta-Analysis 2009 checklist and GRADE recommendations [18].

Original articles and meta-analyses indexed from July 2016 to December 2019, describing genetic or epigenetic aspects of atopic dermatitis, were searched. We identified eligible studies using the following inclusion criteria: (1) primary study or meta-analysis, (2) written in English or Spanish, (3) human subjects, both children and adults, (4) patients suffering from atopic dermatitis or atopic eczema, and (5) studies describing mutations, single nucleotide polymorphisms (SNPs), or epigenetic modifications in association with disease onset, severity, or prevalence in the population. The exclusion criteria were: (1) animal, in vitro or in silico studies, (2) review articles, (3) proteomics or expression analysis without epigenetic/genotyping study, (4) articles focused in other diseases, such as psoriasis or ichthyosis vulgaris, in which AD was merely mentioned, and (5) articles whose full-text version was not available to us.

We performed the literature search between December 2019 and January 2020 in PubMed, Cochrane Library, and Scopus databases, using the following terms: "atopic dermatitis" OR "atopic eczema" AND "gene" OR "genetic" OR "mutation" OR "epigenetic" OR "DNA methylation" OR "sequencing" OR "microRNA" OR "polymorphism" OR "genome-wide association study" OR "microarray" OR "gene profiling".

Three authors individually reviewed the database search results, assessing titles, evaluating abstracts, and considering or not the study for full review. Any disagreements in either the title/abstract or the full manuscript review phases were resolved by consensus. All eligible studies were formally evaluated and included in this systematic review.

The authors independently evaluated the quality appraisal and graded the risk of bias of the included studies. The risk of bias was assessed by Rob2, the recommended tool to assess the risk of bias in randomized trials included in Cochrane Reviews [19], slightly adapted by the authors to fit the nature of the selected articles. Studies were classified as low, moderate, or high risk of bias.

Quality was assessed using the Newcastle–Ottawa scale [20]. Each study was awarded one point per positive item, according to the scale. Scores over 6 merited "high quality"; those below 4 were considered as "low quality"; the rest being classified as "moderate".

Epigenetic methodology has some peculiarities that prevent the application of bias and quality scoring by the commonly used scales. Some notes regarding this will be mention when discussing the selected epigenetic studies.

Gene pathway analysis of the found genes was performed using FunRich [21], Reactome [22], and STRING [23].

3. Results

3.1. Selection, Bias and Quality of Articles

The database search yielded 914 articles (Figure 1). Atopic eczema was used for the search engine as a synonymous term. After title and abstract review, 810 articles were rejected since they did not fulfill eligibility criteria, i.e., those describing animal or in vitro studies, literature reviews, analysis of protein or gene expression, and articles written in languages other than English or Spanish. Therefore, 104 articles qualified for full text review. Of those, we eliminated 13 studies that mentioned AD in comparison to other conditions but were not fully dedicated to it and 18 studies that did not perform any genetic association with the disease. As a result, 73 articles were evaluated. Out of 73 studies, 11 were related to epigenetics [13,24–33], 39 were candidate gene studies [34–73], 5 were genome-wide association studies (GWAS) [5,13,74–76], whole-exome sequencing (WES) was performed in 7 articles [77–83], and phenome-wide association sequencing was done in 1 article [84]. Four studies described results from next-generation sequencing (NGS) [85–88], and 2 showed analyses of copy number variations (CNV) [89,90]. Besides, 6 meta-analyses were also included [91–96].



Figure 1. The flow diagram depicts the flow of information through the different phases of the systematic review. It maps out the number of records identified, included and excluded, and the reasons for exclusions.

A description of the 64 selected non-epigenetic studies is presented in Table 1. Epigenetic articles are summarized in Table 2.

Reference	Study Type	Population/Country	Objective	Sample Size	Genes	SNP/Mutation	Results/Conclusion
	WES					c.3332delT,	Mutation present in hyperimmunoglobulin
[77]	(Whole-exome sequencing)	Iraq	To determine DOCK8 deficiency	1 child	DOCK8	Phe1113Leufs *2 (rs140392509)	 E syndrome (HIES) and non-Hodgkin lymphoma patient.
			To determine association of atopic dermatitis (AD) with ichthyosis vulgaris (IV) and actinic keratosis (AK)			1537C>T(R501X) (rs61816761)	
[65] Candidate	Candidate gene	Denmark		481 AK patients, 9112 Healthy controls (HC)	FLG	2318_2321del (2282del4)	FLG homozygous loss of function and AK (in 0.8% of AK studied vs. 0.2% in control population)
						7375C>T (R2447X; rs138726443)	r •r •
[66]	Candidate gene	Ethiopia	To elucidate SNVs associated with	184 patients of AD	SPINK5	rs2303063;	Significant association with AD.
[]	0	Ī	AD	and 186 HC	0111110	rs2303067	0
		Ethiopia	To elucidate SNVs associated with AD To establish the role of <i>CLDN1</i>	22 patients for		rs17501010	rs893051 is associated with development of AD in early life.
[35]	Candidate gene, WES			WES; 159 AD patients and 192	CLDN1	rs9290927	
			variants in Ethiopian AD patients	HC for genotyping		rs9290929	
						rs893051	_
[46]	Candidate gene	Jordan	To study the association between <i>resistin</i> gene polymorphisms and AD	162 AD patients, 161 HC	RETN	SNP +299 G>A (rs3745367)	rs3745367 associated with AD in a gender- and age-specific manner (male, less than 10 y)
_						SNP +157 C>T (rs3219177)	
[57]	Candidate gene	Iran	To identify association of SNPs in <i>IL-10</i> and <i>TGF-61</i> and AD in Iranian	89 children with	TGF-β1	cdn 10	cdn10/C allele, CC genotype associated with AD
			patients	11D, 100 11C		cdn 25	cdn 25/C allele associated with AD
[68]	Candidate gene	Chinese Han	To identify variants in Chinese Han population associated to AD	4619 AD patients and 10789 HC	CD207/VAX2	rs112111458 (allele G/A)	Association of rs112111458 and AD
[69]	Candidate gene	Turkish children	To evaluate if some <i>TLR2</i> gene	70 children with	TI R2	rs5743708 (R753Q)	None
[07]	Culturate gene	Turkish children	polymorphisms are associated with AD	AD, 69 HC	TERZ	rs4696480 (A-16934T)	
[70]	Candidata gene	Isle of Wight	Wight To study the association of <i>FLG</i> loss of function with atopic march	1150 participants	FLC	R501X (rs61816761)	<i>FLG</i> loss of function mutations are associated
[,]	2 gene			birth cohort	1 60	2282del	and was consistently associated with rhinitis
						S3247X (rs150597413)	from 4 years onwards

Table 1. Summary of findings from the selected genetic studies.

Reference	Study Type	Population/Country	Objective	Sample Size	Genes	SNP/Mutation	Results/Conclusion
						R501X (rs61816761)	
			To determine whether variations in <i>FLG</i> and <i>TSLP</i> genotype corresponded to differences in		FLG	2282del4	
	Candidate genes	USA (mixed				R2447X (rs138726443)	Variations in FLG and TSLP genotype were
[71]		population of children)		842 children with AD		S3247X (rs150597413)	associated with differences in self-reported skin clearance, TCI usage, and steroid usage
			inclupeute treatment use over time	_		rs1898671	
					TSLP	2282del4	
[78]	WES	Canada	To identify the genetic aberration in 4 related patients with combined immunodeficiency, early-onset asthma, eczema, and food allergies, as well as autoimmunity	4 related patients	CARD11	hg19:chr7:2987341:G>A NM_032415:exon3: c.C88T:p.R30W (rs145474800)	CARD11- R30W is associated with recurrent infections, autoimmunity, and severe atopy. The novel R30W mutations described abrogate the NF-κB pathway and lead to decreased IL-2 and IFN-γ secretion and lymphocyte proliferation
	Candidate gene		To investigate the importance of 4 common <i>FLG</i> null mutations in the susceptibility to eczema in Polish children population	50 children with AD, 37 children with non-atopic eczema and 71 HC		R501X (rs61816761)	
[72]		Poland			FLG	2282del4	<i>FLG</i> null mutations and AD are associated but explain only a part of AD cases (13.8%)
						R2447X (rs138726443)	but explaint only a part of The cases (10.076)
				children		S3247X (rs150597413)	
		Different ethnic	To study the association of known			2282del4	
[73]	Candidate gene	origins (Dutch, Cape Verdean, Dutch Antillean,	genetic factors and ethnic origin with the development of eczema	3096 children	FLG	R2447X (rs138726443)	Carrier frequencies of <i>FLG</i> mutations in children of non-Dutch origins were low.
		Moroccan,				R501X (rs61816761)	
		Surinamese-Creole, Surinamese-Hindusta Turkish children)	ani,			S3247X (rs150597413)	
	Meta-analysis					2282del4	Association between FLG loss of function
[91]		Canada (Caucasian)	Study the effect of <i>FLG</i> mutations on contact dermatitits (CD)	165 patients with CD, 891 HC	FLG	R2447X (rs138726443)	mutations and contact polysensitivity,
		Cuucuoluity				S3247X (rs150597413)	especially in K501X polymorphism.
						R501X (rs61816761)	

Reference	Study Type	Population/Country	Objective	Sample Size	Genes	SNP/Mutation	Results/Conclusion
					FLG	R501X (rs61816761)	This SNP is a stronger risk factor for eczema than for hay fever or asthma.
[13]				- 180 129 cases with	RPTN-[]-HRNR	rs12123821	This SNP is a stronger risk factor for eczema than for hay fever or asthma.
	GWAS (Genome-wide association	European ancestry	To identify shared risk variants of a broad allergic disease phenotype that	asthma and/or hay fever and/or	IL1R2-[]-IL18R1	rs12470864	This SNP is a stronger risk factor for eczema than for hay fever or asthma.
	study)		fever and eczema	eczema, and 180,709 HC	WDR36-[]-CAMK4	rs6594499	This SNP is a stronger risk factor for hay fever than for eczema or asthma.
				_	IL2RA	rs61839660	This SNP is a stronger risk factor for eczema than for hay fever or asthma.
				-	GSDMB	rs921650	This SNP is a stronger risk factor for eczema than for hay fever or asthma.
[36]	Candidate gene	Russia	To explore the frequency <i>FLG</i> mutations and CNVs in AD patients and control subjects of Russian and Tatar ethnic origin living in Volga-Ural region of Russia	177 Russian, 126 Tatar AD patients; and 152 Russian, 109 Tatar HC	FLG	2282del4 R501X (rs61816761) R2447X (rs138726443)	Significant differences in 2282del4 frequency were found between Tatar AD patients and HC. The allelic frequency of the R501X mutation in AD patients was 0.85% and in HC -0.47%. The allelic frequency of R2447X was 1.75% in patients, and 1.33% in HC.
						S2889X (rs782477344)	Mutations in S2889X constituted 96.4% of all FLG mutations.
			To investigate the personal			2282del4	No carrier of R501X and Q2417X mutations
[37]	Candidate gene	India	consequences of having atopic dermatitis and/or hand eczema and	163 patients and 86 HC	FLG	R501X (rs61816761)	was identified. FLG mutations are associated with irritant contact dermatitis with or without atopy
			FLG mutations			Q2417X (rs528722713)	allergic contact dermatitis without atopy,
						2282del4	and idiopathic subtypes. <i>FLG</i> mutations
						R2447X (rs138726443)	eczema.

Reference	Study Type	Population/Country	Objective	Sample Size	Genes	SNP/Mutation	Results/Conclusion
[83]	WES	Korea	To identify family-specific candidate	3 families (2 affected AD and 2 unaffected	COL646	rs16830494	<i>COL6A6</i> variants may be risk factors for AD because the minor allele (AA) in rs16830494 and
	WLO	Rorea	early-onset AD in Koreans.	individuals) for WES.		rs59021909	the rs59021909 (TT) allele and the rs200963433
				112 AD and 61 HC for validation studies.		rs200963433	 heterozygous (CT) frequency were all higher in AD cases compared to controls, but no significant association was reached.
					TLR1-TLR6	rs2101521	
					WDR36-CAMK4	rs1438673	
			To study in the Chinese Ham		PTGER4	rs7720838	_
[38]	Candidate gene	Chinese Han	population the association AD with	3013 AD patients, 5483 HC	NFATC2	rs6021270	SNPs rs2158177 and rs1837253 are associated with
			previously reported SNPs		IL1RL1-IL18R1	rs3771175	
					THSD7B	rs1469621	
					RAD50/IL13	rs2158177	_
					TSLP	rs1837253	_
	Candidate gene	Sweden	To explore the longitudinal relation between preschool eczema, <i>FLG</i> mutation, or both and IgE sensitization in childhood.	1890 children		2282del4	Preschool eczema is associated with IgE
[39]					FLG	R501X (rs61816761)	aeroallergens up to 16 years of age. FLG mutation is associated with IgE sensitization to peanut but
						R2447X (rs138726443)	not to other allergens. Sensitized children with preceding PSE are more often polysensitized.
						rs9784600	
						rs9784675	
						rs11740584	
						rs7737031	
			To elucidate the associations between	7000 children and 1020 HC.		rs17691077	<i>KIF3A</i> is associated with asthma + eczema. The
[40]	Candidate gene	USA	KIF3A SNPs and asthma, eczema,	Results were replicated in 762 children with atopy	KIF3A	rs2299007	genetic association of <i>KIF3A</i> with asthma or even
			and AR, alone and in combination.	702 children with atopy.	_	rs3798130	with asthma+ eczema.
						rs12186803	
						rs1468216	
					_	rs2023822	_
						rs2237059	_
						rs2023823	

Reference	Study Type	Population/Country	Objective	Sample Size	Genes	SNP/Mutation	Results/Conclusion
						rs2787094	
						rs543749	-
						rs2280090	rs2280090 was associated with reduced
						rs2280091	flow after 240 s of hypertonic saline
					ADAM33	rs3918396	inhalation with respect to the age- and
						rs6127096	an increased risk of allergic bronchitis;
	Phenome-WAS (Phenome-wide association study)					rs511898	rs3918396 was associated with wheezing and
						rs2280090	- eczema comorbidity.
			To dissect the role of immunogenetics in allergy and asthma.	- 974 children - -		rs3918396	-
[84]		Turkey			IL4	rs2243250	rs2243250 is associated with increased
						rs2070874	240 s of hypertonic saline inhalation).
					CD14	rs2569190	Associated with asthma.
					ADRB2	rs1042713	No association
						rs1042714	
					IL13	rs1800925	No association
						rs1295686	
						rs20541	
					II 4R	rs1805015	No association
					1211	rs1801275	
					MS4A2	rs1441586	No association
						rs569108	
					SERPINE1	rs1799768	No association
					TNF	rs1800629	No association
[42]	Candidate gene	Korean	To study <i>MIF</i> promoter polymorphisms and total plasma IgE in AD Korean patients	178 AD patients, 80 HC	MIF	rs755622 (–173 G to C)	MIF promoter polymorphisms in the –173 C allele and the MIF C/5-CATT and MIF C/7-CATT haplotypes were significantly associated with an increased risk for AD.

Reference	Study Type	Population/Country	Objective	Sample Size	Genes	SNP/Mutation	Results/Conclusion
			To identify FLG SNP variations and			rs71626704	rs71626704 and rs76413899 were significantly
[41]	Candidate gene	Korea	evaluated its association with clinical phenotypes, including AD and other parameters.	81 patients	FLG	rs76413899	 associated with a history of asthma and cheilitis.
						rs62623409	rs62623409 and rs71625199 were associated
						rs71625199	— with sensitization to environmental allergens.
						rs2289276	rs3806932, rs2289276, and rs2289278 are
[40]	Condition		lo investigate the association between four possible TSLP	-	741 D	rs2289278	 associated with susceptibility of AD. rs3806932, rs3806933, and rs2289276 form
[43]	Candidate gene	Korea	polymorphisms and atopic disease in		TSLP	rs3806932	one linkage disequilibrium block. The GTT
			a Korean population.			rs3806933	 haplotype strongly contributes to atopic march
						2282del4	
[44]	Candidate gene	UK		224 patients and 40 HC	EL C	R501X (rs61816761)	 Subjects with FLG-null mutations have more mature Langerbans cells in non-lesional skin
[44]					TLG	S3247X (rs150597413)	irrespective of whether they have AD.
						R2447X	
						(rs138726443)	
		Chinasa Han/				3321delA	_
						K4022X (rs146466242)	c.3321delA was found in all populations.
		Singapore,				6950del8	Some mutations showed south-to-north (or north-to-south) distribution gradient:
[45]	Candidate cone	Chinese Han/ Shanghai, Chinese	To assess the significance of <i>FLG</i>	1384 patients and	FLC	Q2417X (rs528722713)	p.K4022X, the most prevalent <i>FLG</i> mutation in northern China and Korea, declined in
	Candidate gene	Korean, Japanese/ Kyushu and	East Asian populations.	1031 HC	FLG	E2422X (rs374588791)	frequency moving southward; in contrast, c.6950del8 (e.g., p.Q2417X, p.E2422X)
		Japanese/ mainland				S2554X (rs121909626)	 showed the reverse. p.S2554X/p.S2889X/p.S3296X/Q1701X mutations were Japanese-specific.
						S2889X (rs782477344)	
						S3296 (rs760426769)	_
						Q1701X (rs4547271)	_

Table 1. Cont.

Reference	Study Type	Population/Country	Objective	Sample Size	Genes	SNP/Mutation	Results/Conclusion
[92]	Meta-analysis	China, Taiwan, Japan and Saudi Arabia (Asian population) and Poland, Czech R., Macedonia, Egypt (Caucasian)	To study the association between IL-4-590C/T polymorphism and AD susceptibility.	923 patients and 1215 HC	IL-4	-590C/T	The <i>IL-4</i> -590C/T polymorphism may contribute to AD susceptibility in the overall population and children, especially for Asian children.
[89]	CNV analysis	UK	To assess the contribution of <i>LILR</i> and <i>LILRA3</i> genes CNV to AD	1482 patients from 378 families	LILR, LILRA3		The transmission of one copy of <i>LILRA6</i> within families was potentially related to the development of AD.
						R501X (rs61816761)	2822del4 was significantly associated with early-onset AD and asthma.
		Finland	To test the association of the 4 most prevalent European <i>FLG</i> null mutations, the 2 Finnish enriched <i>FLG</i> null mutations, the <i>FLG</i> 12-repeat allele, and 50 additional	501 patients with AD and 1710 HC	- FLG	R2447X (rs138726443)	R501X was associated with early-onset and suggestively with keratosis pilaris.
					_	S3247X (rs150597413)	 R2447X showed suggestive association wit early-onset AD. Baseline IgE values were higher in patients with FLG null mutation
					_	S1020X (rs200360684)	but the association was not significant; <i>FLG</i> null mutations were not associated with
[47]	Candidate gene				501 patients with	_	V603M (rs2306942)
	0		epidermal barrier gene variants, with		=	rs12730241	—
			features, risk of other atopic diseases,		CLDN1		
			age of onset, and treatment response.		CLDN4		
					CLDN20		
					CLDN23		No significant association with AD
					OCLN		
					IVL		
					FLG2		
					LOR		
					JAM-1		
					TJP1		

Table 1. Cont.

Reference	Study Type	Population/Country	Objective	Sample Size	Genes	SNP/Mutation	Results/Conclusion
			To study CARD11 mutations in four families with recalcitrant, severe			L194P	The study describes rare hypomorphic
[79]	WES	USA		8 patients	CARD11	R975W (rs1064795307)	dominant negative mutations in <i>CARD11</i> in 4 unrelated families, which lead to
			atopic disease.			E57D	dominantly inherited, severe atopy, with
					-	dup183_196	
			To test whether genetically lowered	33996 children – –	DHCR7	rs12785878	No association
[5]	GWAS	UK	with risk of asthma, atopic dermatitis,		CYP2R1	rs10741657	
			or elevated serum IgE levels.		CYP24A1	rs6013897	
						rs79808464	
						rs116222149	
					-	rs11584340	
					-	rs113136594	
					-	rs145828067	
			To evaluate the role of <i>FLG</i>		-	rs374910442	
			polymorphisms expression and risk	100 children with	-	rs747005144	<i>FLG</i> mutations are associated with early
[48]	Candidate gene	Italy	of developing a concomitant Molluscum contagiosum sustained skin	AD and 97 healthy	FLG	rs145627745	disease, and a significantly increased risk of
			infection in the pediatric population	children	-	rs144209313	M. contagiosum sustained skin infection
			with AD.		-	rs74129443	
					-	rs192455877	_
					-	rs150957860	_
						rs138055273	
					-	rs147472105	
					-	rs183942200	—

rs558269137

Reference	Study Type	Population/Country	Objective	Sample Size	Genes	SNP/Mutation	Results/Conclusion
			To identific discussion is then in the			A407T	
						R518W (rs142940671)	_
[86]	NGS (Next-generation	Cormon	locus 11q13.5 using combination of	31 AD patients	I DDC22	R312	Association of low-frequency and rare
[00]	sequencing)	German	sequencing and functional annotation.	51 MD putients	LKKC52	S411R (rs201424816)	with AD.
						R414W (rs201431152)	_
						R652C	
	Meta-analysis	analysis French, French- Canadian and UK	To detect new interacting genes involved in eczema	388 French families. Replication in 253	COL5A3	rs2287807	Identified significant interaction between two new genes, COL5A3 and MMP9, which
[93]				French-Canadian and 207 UK family datasets.	MMP9	rs17576	may be accounted for by a degradation of <i>COL5A3</i> by <i>MMP9</i> influencing eczema susceptibility.
						R501X (rs61816761)	
						S3316X (rs149484917)	_
						R826X (rs115746363)	Rare FLG LoF variants in African American
[85]	NGS	USA	To evaluate <i>FLG</i> LoF variation in children of African ancestry and the	262 African American children	FLG	R2447X (rs138726443)	— children are associated with AD and more persistent AD. In contrast to Europeans, no FLG LoF variants predominate in African
			persistence.	and 133 Caucasians		Q570X (rs192402912)	American children. The most common variants were R501X,
						R3409X (rs201356558)	— 53316X, and K826X.
						S3247X (rs150597413)	_
						Q3818X (rs148606936)	_
						H440fs	

Reference	Study Type	Population/Country	Objective	Sample Size	Genes	SNP/Mutation	Results/Conclusion
[90]						R501X (rs61816761)	
	CNV (Copy number variations) analysis	African American (USA)	To study <i>FLG</i> LoF and CNV in African American population	39 children with AD	FLG	R826X (rs115746363)	rs149484917 is a population-specific <i>FLG</i> LoF unique to several populations of African
						S3316X (rs149484917)	Ancestry. Two new <i>FLG</i> LoF were identified (488delG and S3101X)
						488delG	
						S3101X	_
						Q267R (rs6892205)	
						A335V (rs34482796)	Only S368N frequency differed between
[49]	Candidate gene	Ianan	To study polymorphisms of SPINK5	57 patients 50 HC	SDINK5	S368N (rs230306)	Genetic Variation Database.
	Canalate gene	Jupan	gene in Japanese AD patients	57 patients, 50 HC	51 IINK5	D386N (rs2303064)	Algorithms predicting functional effects of
						R711Q (rs3777134)	scores for R654H
						E825D (rs2303070)	

Reference	Study Type	Population/Country	Objective	Sample Size	Genes	SNP/Mutation	Results/Conclusion
						R501X (rs61816761)	
						3321delA	_
[50]						Y1767X (rs1222103354)	_
						S1695X (rs772851618)	
				70 patients	FLG	Q1701X (rs145738429)	_
	Candidate gene	Korea	To examine the spectrum of null-mutations and compare with			Q1745X (rs1209640261)	Only 11 AD patients had <i>FLG</i> mutations. This frequency was lower than that
	-		other Asian countries			Q1790X (rs200622741)	 described for other Asian populations (Chinese, Japanese, Singaporean)
						S2554X (rs121909626)	
						S2889X	
						S3296X (rs761212672)	
						K4022X (rs146466242)	
						3222del4	_
						S1515X (rs180768115)	
						Q2417X (rs528722713)	_
[51]	Candidate gene	Korea	To investigate the genetic polymorphisms of <i>FLG</i> in Korean AD patients	9 ichtyosis vulgaris patients 50 AD patients 55 HC	FLG	K4022X (rs146466242)	This loss-of-function mutation was only found in AD patients. 62 new SNPs were identified
[87] NGS	NGS	Korea	To investigate clinical characteristics of AD patients with <i>FLG</i> mutations. To determine differences between patients with and without <i>FLG</i> mutations	1110 patients, 68 with mutations in <i>FLG</i> gene	FLG	K4022X (rs146466242)	Null alleles were associated with early onset of AD and higher risk of developing the disease by age 2 years.
						3321delA	EASI score was also higher in these patients

Reference	Study Type	Population/Country	Objective	Sample Size	Genes	SNP/Mutation	Results/Conclusion
					INPP5D	rs1057258-c	
				-	PRR5L	rs12295535-t	_
					STAT3	rs17881320-t	_
					PPP2R3D	rs2143950-t	Six classes based on temporal trajectories of
					ACTL9	rs2164983-a	early-onset/late resolving,
			To investigate longitudinal	individuals	IL6R	rs2228145-c	early-onset/early-resolving,
[76] GWAS	GWAS	UK, Netherlands	phenotypes of AD in two	NL: 3652	KIF3A	rs2897442-c	<i>FLG</i> null mutations were strongly associated
			independent conorts	individuals	OVOL1	rs479844-g	with early-onset and late-onset of AD
				(PIAMA) -	C11orf30	rs7927894-t	(p < 0.00001; ALSPAC) and early-onset/late-resolving ($p = 0.0006;$
					IL22	rs2227482-t	PIAMA)
					IL21	rs17389644-a	_
				-	IL2RA	rs6602364-g	
[80]	[80] WES	WES USA (Hispanic, To identify rare DNA variants Caucasian, conferring significant risk for AD African-American)	To identify rare DNA variants	3 patients	CARD14	c.1778T>C, I593T	Downregulation of CARD14 led to severe – AD and reduced skin protection against
					c.2206A>C,N737H (rs535171797)	infection and dysregulated cutaneous inflammation pathways	
				-	SCAND3		
					TCHHL1	-	
					ADCY10	-	
					MTF1	-	
	WES, rare		To analyze the genetic architecture of		MCM10	-	Some rare sequence variations of candidate
[81]	enrichment	Bangladeshi	patients with AD from a Bangladeshi	from 70 families	ORM2	-	loss-of-function variations were carried by
	anarysis		continuinty in London, OK	-	CUX2	-	almost 50% of AD-affected individuals.
				-	MAST2	-	
				-	PHLDB1	-	
				-	FLG	-	
			To explore the role of different SNPs			rs7927894	The haplotype TATG in these SNPs fully
[52]	Candidate gene	Poland	at 11q13.5 in predisposing to allergic	270 AD patients, 540 HC		rs2513517	explained the association with AD ($p = 0.00021$)The TG haplotype in the last two
			phenotypes	340 NC		rs7930794	SNPs was also related to allergic rhinitis
						rs7125552	

Table	1. (Cont.
-------	------	-------

Reference	Study Type	Population/Country	Objective	Sample Size	Genes	SNP/Mutation	Results/Conclusion
			To assess the genetic relationship	16 case-control		<i>IL-10</i> -1082a/G	These polymorphisms showed a weak
[94]	Meta-analysis	Asian, Caucasian	between II-10 polymorphisms and susceptibility to AD	studies	IL-10	IL-10 -819T/C	association with AD susceptibility
			susceptionity to TiD.			IL-10 -592a/C	_
[53] Candidate gene					R501X (rs61816761)		
		Netherlands	To investigate whether <i>FLG</i> mutations influence the outcome of immunosuppressive therapy	42 patients with severe AD: 3 Asians, 1 Afro-Caribbean, 38		2282del4	 FLG mutation group showed a trend towards less improvement in the course of 24 weeks of treatment with methotrexate and azothiopine
	Candidate gene	(Caucasians, Asians, Afro- Caribbean)			Asians, 1 FLG Afro-Caribbean, 38 Caucasians	R2447X (rs138726443)	
				Caucasians		S3247X (rs150597413)	
						5321delA	_
						Q1790X+S3296X	
						Q1790X+S2889X	 The most severe AD was associated with c.3321delA+S2889X bi-allelic combination. By contrast, individuals with S2880X +S206X did not devide AD
[54]	Candidate gene	To andidate gene Japan FLG	To elucidate the effect of bi-allelic	6 individuals from FLG 3 families	EL C	S2889X+S3296X	
	Cancillate gelle		severity		I LG	Q1701X+S2889X	
						3321delA+S2889X	

Reference	Study Type	Population/Country	Objective	Sample Size	Genes	SNP/Mutation	Results/Conclusion
						rs612709	
					ADAM	rs528557	-
						rs44707	_
					-	rs2787094	_
					ALOX5	rs4986832	_
	To study the eff genotype-phen genetically hom living in tw					rs892690	_
					rs2115819		
			To study the effect of environment on genotype-phenotype association in a genetically homogeneous population, living in two separate areas	615 Greenlandic Inuit individuals 643 Danish Inuit individuals		rs2844484	 LTα rs2844484 was associated with AD in Greenland patients (p = 0.035) The risk of AD was related to the genotype distribution of this SNP, with a significant interaction with the place of residence
[67]		Inuit E			LT-α	rs909253	
						rs1041981	
					LTC4S	rs730012	
					NOS I	rs7977109	
					ORMDL3	rs4065275	
						rs12603332	
					TBXA2R	rs4523	
						rs1799964	
					TNF-α	rs1800630	_
						rs1800629	
					АРОВ	rs145862664	_
					CYP27A1	rs199691576	_
		-	To investigate rare genetic variants	469 AD patients.	C3orf15	rs193146105	Gene polymorphism in CYP27A1, a gene
[82]	WES	Japan	associated with AD	469 AD patients, 935 HC	GAK	rs142107211	involved in vitamin D3 metabolism, was
					VNN2	rs200230703	
					USP35	rs200193128	_
					ZNF749	rs76428401	

Reference	Study Type	Population/Country	Objective	Sample Size	Genes	SNP/Mutation	Results/Conclusion
						g.480G>A	
						g.4576A>T	_
						g.5070C>T	_
						rs200698932	_
						rs769935596	- SNIPs a 1320 a 1331 a 1313 a 1311 a 5363
			To investigate the potential role of	(EAD motion to		rs760937092	were present both in patients and controls.
[55] Candida	Candidate gene	China	SHARPIN in the pathogenesis of AD	65 AD patients, 100 HC	SHARPIN	rs1280524235	The mutations in <i>SHARPIN</i> were only
						g.4320	expression in AD lesions.
						g.4334	_ •
						g.4343	_
						g.4344	
						g.5363	_
						R501X	_
						(rs61816761)	
						S2554X (rs121909626)	
				Japan: 26 IV		S2889X	Mutation S3296X only appeared in Japanese
[56]	Candidate gene	Japanese, Korean	To determine prevalence of <i>FLG</i> mutation in AD and IV patients in	patients, 91 AD patients	FLG	G1109EfsX (rs133912394)	AD patients. R501X and R826X only appeared in IV
			Japan and South Korea	AD patients		K4022X (rs146466242)	 patientsThe rest of the mutations were found in both Korean and Japanese patients. –
						S1695X (rs772851618)	
						Q1701X (rs145738429)	
						R826X (rs115746363)	-

Reference	Study Type	Population/Country	Objective	Sample Size	Genes	SNP/Mutation	Results/Conclusion
			To examine the association between	1		R501X (rs61816761)	Within the dizygotic twin population, 11 pairs were discordant for <i>FLG</i> mutation. The
[58]	Candidate gene	Denmark	loss-of-function mutations in <i>FLG</i> and AD and asthma in adult twins	575 adults twins with asthma	FLG	2282del4	risk of AD increased in the twin with <i>FLG</i>
						R2447X (rs138726443)	No significant association was found with FLG mutations and asthma
				417 children: 186		R501X (rs61816761)	
[59] Candidate gene			patients (40 with	FLG	2282del4	- 29% of patients with persistent AD had FLC	
	Candidate gene	Denmark	To explore heritable, environmental, and clinical factors related to persistent AD	persistent AD), 231 HC. Follow-up study from birth to age 13 y		R2447X (rs138726443)	The higher AD genetic risk score, the higher risk of persistent AD.
						S3247X (rs150597413)	
				-	FLG2		_
				-	SPRR3	-	
						rs284544	In FLG WT patients, RPTN rs3001978CC was
					RPTN	rs28441202	significantly associated with AD early age
						rs3001978	onset ($p = 0.033$), pruritus ($p = 0.021$), severity of AD ($p = 0.045$) and concomitant
[60]	Candidate gene	Poland	To identify new potential markers of	159 AD patients,		rs12117644	asthma ($p = 0.041$)
		-	AD	10811C -	CRNN	rs941934	rs 941934 allele A was more frequent in AD
				FLG	FLG	R2447X (rs138726443)	only appeared in AD patients ($p = 0.019$). The association depended on <i>FLG</i> mutations.
						S3247X (rs150597413)	_

Reference	Study Type	Population/Country	Objective	Sample Size	Genes	SNP/Mutation	Results/Conclusion
[61]	Candidate gene	Russia	To determine the relationship of <i>TLRs</i> polymorphisms with AD	25 AD patients, 25 AD and rhinitis/asthma patients, 100 HC	TLR2	rs55743708 (G>A)	Increased levels of IL-4 and IL-10. Dysfunction of cell activation.
			I J I I		TLR4	rs4986790(A>G)	Increased levels of IL-4 and IL-1. Weaker cell response to microbial antigens.
		gene USA	USA To examine the effect of <i>FLG</i> mutations and <i>TSLP</i> (thymic stromal lymphopoietin) polymorphisms on the age of AD onset			R501X (rs61816761)	
	Candidate gene				FLG	2282del4	 FLG null mutations were associated with early onset of AD. Number of mutations was associated with timing of onset. No association was found between TSLP polymorphism and timing of onset.
[62]				822 children (age 2–17 y)		R2447X (rs138726443)	
						S3247X (rs150597413)	
				-	TSLP	rs1898671	-
[63]	Candidate gene	Taiwan	To investigate the association between gene–environmental interaction and childhood AD	839 mother–child pairs	GST	GST-T1/M1 mutants	GST null genotypes in association with high levels of perfluoroalkyl substances in blood increased the risk of developing AD

Reference	Study Type	Population/Country	Objective	Sample Size	Genes	SNP/Mutation	Results/Conclusion
						S1515X (rs180768115)	
						E2422X (rs374588791)	_
						S406X (rs189114758)	-
						c.6950-6957del8	_
						c.1640delG	_
				334 patients with FLC AD and/or IV FLC		Q368X (rs746899204)	-
					FLG	3321delA	_
		Singapore population: Chinese, Indian, Malay	To sequence the entire <i>FLG</i> coding region in Singaporeans from different ethnicities			7945delA	_
	NGS					Q2417X (rs528722713)	 A new technology that improved accuracy and cost-effectiveness is described.
[88]						2952delC	
						9040-9058dup19	New mutations have been identified
						Q1790X (rs200622741)	-
						S1302X (rs754812742)	
						S1515X (rs180768115)	
						4004del2	
						2282del4	
						R2447X (rs138726443)	
						477insA	_
						678delA	-
						S378X (rs755134998)	
						3036delT	
						10866delA	_
						rs10067777	

Reference	Study Type	Population/Country	Objective	Sample Size	Genes	SNP/Mutation	Results/Conclusion
						rs7701890	
					-	rs13360927	
				3031 cases, 5075 HC	-	rs13361382	
[74] GW.			To identify AD susceptibility genes in		-	rs5870408	rs11357450 had the strongest association
	GWAS	China	5q22.1 and observe expression in AD		TMEM232	rs1400764268	with risk of AD (OR = 1.20 ; $p = 0.04$)
			ussues		-	rs35639206	
					-	rs137936676	_
					-	rs10617471	
					-	rs11357450	
				_	KLK7	KLK7 MT	More frequent in AD patients ($p = 0.04$). No differences between AD groups
					FLG	3321delA	More frequent in moderate/sever AD patients ($p = 0.038$)
						K4022X (rs14646429)	More frequent in patients but not statistically different
						1156	More frequent in AD vs HC ($v < 0.001$). No
					SPINK5	1188	differences between AD groups
						2475	
[64]	Candidate gene	Korea	barrier- or immune-related genes	279 AD patients, 224 HC	DEFB1	rs5743399	More frequent in AD vs HC ($p < 0.002$). No differences between AD groups
					KDR	rs2305948	More frequent in AD vs HC ($p = 0.03$). No differences between AD groups
					IL5RA	rs334809	More frequent in AD vs HC ($p < 0.001$). More frequent in mild AD vs moderate/severe
					IL9	rs31563	More frequent in AD vs HC (<i>p</i> < 0.001). No differences between mild AD and HC; and between moderate AD vs. HC
					II 120 B1	rs393548	More frequent in AD vs HC ($p = 0.02$).
					ILIZNDI -	rs436857	More frequent in AD vs HC ($p = 0.01$). More frequent in mild AD vs. HC. No differences between moderate AD and HC
					IL13	rs20541	Heterozygous less frequent in patients vs. HC

Reference	Study Type	Population/Country	Objective	Sample Size	Genes	SNP/Mutation	Results/Conclusion
[95]	Meta-analysis	Germany, Turkey, Italy, Finland,	any, Turkey, To assess whether <i>TLRs</i> y, Finland, polymorphisms are associate with	<i>TLR2</i> : 9 studies; 733 cases, 807 HC	TLR2	rs5743708	Increased risk of AD for GA heterozygous
	Ukraine, Russia	Ukraine, Russia	risk of AD	<i>TLR4</i> : 6 studies; 646 cases, 601 HC	TLR4	rs4986790	AG showed some correlation with risk of AD, but was non-conclusive.
		Saudi Arabia, Iran, India, Netherlands,	— • • • • • •			IL-10-1092 G/A	
[96]	Meta-analysis	Italy, Poland, I Meta-analysis Macedonia, Czech Republic, China, P Korea, Taiwan, Germany, UK	on association between <i>IL-10</i>	15 case-control studies, 1647	IL-10 IL-10-592 A/C IL-10-819 G/A	IL-10-592 A/C	IL-10-819 G/A mutation in Caucasian subjects and with <i>IL-10-</i> 1092 G/A in Asian
			AD	patients, 2031 HC		patients	
[75]	GWAS	UK	To assess whether <i>FLG</i> expression in umbilical cord blood associates with	94 infants	FLG	S3247X (rs150597413)	FLG expression in cord blood correlated with AD risk.
			and predicts AD			2282del4	2.94-increased risk for mutated <i>FLG</i> variants

Cell/Ttissue Types	Epigenetic Assay	Significant Findings	Reference
Primary adult human keratocytes	miFinder miRNA PCR Array	Broad dysregulation of miRNAs upon IL-4 treatment	[27]
Whole blood samples	DNA methylation profiling	Identifying CpG methylation sites in IL4 and IL13 associated with AD phenotype	[28]
Serum	Real-time PCR for miR-146a	miR-146a levels are unaltered in AD patients	[29]
Serum	RNA sequencing	miR-151a and miR-409 overexpressed in Chinese AD patients	[30]
AD lesioned skin	Microarray expression data from GSE32924	Regulatory network comprising 182 miRNAS	[24]
Umbilical cord serum	Exiqon Serum/Plasma Focus micro- RNA PCR Panel (179 miRNAs)	miR-144 levels are higher in the umbilical cord of AD children	[26]
Whole blood samples	DNA methylation profiling	Association between smoking and the methylation state of <i>PITPNM2</i>	[13]
Monocytes and neutrophils	VSTM1 methylation	Polymorphism dependent VSTM1 methylation	[31]
AD lesioned skin	Microarray miRNA expression (GSE31408)	Hsa-let-7a-5p, has-miR-26a-5p and has-miR-143-3p differentially expressed in lesioned tissues	[32]
Whole blood samples	NLRP2 methylation	NLRP2 methylation is associated both to the environment and SNP	[33]
AD lesioned skin	Real-time PCR assay	miR-124 downregulated in AD lesional tissue	[25]

Table 2. Summary of findings from the selected epigenetic studies.

As mentioned above, we followed the Cochrane guidelines to assess the risk of bias of the selected non-epigenetic studies, using the current version of the Rob2 tool [19]. As this tool has been developed for randomized trials, the authors decided to make some assumptions in order to adapt it to the specific nature of the genetic analysis. Taking into account that our main concern with respect to bias referred to the lack of appropriate controls or non-adequate genetic or epigenetic techniques for achieving the intended aim, we responded to questions about intervention or randomization consequently. Therefore, a study was classified as high risk when healthy controls were missing or the methodology to analyze the samples was not clearly explained in the text.

Under these conditions, 20.3% of the non-epigenetic studies were considered at a high risk of bias. Healthy controls were not included in 14 studies, some of them referring to very few patients. One of the studies had no reference to the sample size, and another one did not describe the used methodology. We found some concerns in 4 studies, mainly referring to the selection of subjects. The rest of the selected studies (71.9%) accomplished our criteria for low risk of bias (Figure 2, Table S1).

Correspondingly, 71.2% of the articles merited high quality after running the NOS questionnaire (Table S2). Overall, the representativeness of the cases was the better-scored category. Thirteen articles were considered low-quality studies, mainly due to failed selection and definition of relevant controls.

3.2. Genetic Studies

A total of 62 genes and 5 intergenic regions were described as associated with AD in the selected studies; 32 of them were related to the disease for the first time during this period. Among them, filaggrin was the most widely reported gene, being over 90% of the other genes cited in only one article.

The interaction analysis performed by STRING showed some connectivity enrichment of the listed proteins (*p*-value < 1e-16). The network was clustered using the k-means method. Clustering results are shown in Figure 3 (non-linked proteins were removed from the graph). Three main clusters were found. The most populated included cytokines related to STAT3, with connections with ORM2 and RETN, and TNF.



Figure 2. Risk of bias (upper panel) and quality assessment (bottom panel) of the selected genetic studies, as a percentage of the total.



Figure 3. Functional association protein networks in STRING software (STRING consortium: Swiss Institute of Bioinformatics, Lausanne, Switzerland; Novo Nordisk Foundation CPR, Copenhagen, Denmark; EMBL Heidelberg, Germany) established from the genes reported in the selected studies. The diverse clusters are colored differently. Protein–protein interactions are drawn in blue, when obtained from curated databases, or purple if experimentally determined. Inter-cluster edges are represented by dashed-lines.

With respect to the pathway analysis, a genome-wide overview from Reactome is shown as a Reacfoam in Figure 4 (for a higher resolution, a zoomable pdf version is available as Figure S1). Reacfoam shows a high-level pathway overview visualization based on Voronoi tessellation. Darker functions correspond to those that are over-represented in the list of genes identified in the selected studies, i.e., immune system, developmental biology, signal transduction, and extracellular matrix (ECM). Immune system functions stood up among the high hierarchy pathways (False Discovery Rate (FDR) 2.21e-6; *p*-value 2.22e-8).



Figure 4. Reacfoam shows a high-level pathway overview of the genes reported in the selected studies. Significantly enriched pathways are shown in dark shade. The three main pathways have been zoomed in as (**A**) cytokine signaling in immune system, (**B**) extracellular matrix organization, and (**C**) signaling by receptor tyrosine kinases.

The most significant pathways were related to signaling by interleukins (FDR 1.7e-11; *p*-value!4.17e-11) or different variants, i.e., IL-4/IL-13 (FDR 7.02e-10; *p*-value 5.2e-10), IL-2 (FDR 3.89e-5; *p*-value 4.8e-7) or IL-12 signaling (FDR 1.42e-4; *p*-value 2.11e-6). Identifiers found in the former pathway were *IL6R*, *IL10*, *TGFB1*, *TNF*, *STAT3*, *ADAM33*, *IL4*, *IL13*, and *MMP9*. *IL5RA*, *STAT3*, *IL2RA*, *INPP5D*, *IL9*, and *IL21* were associated with the IL-2 signaling pathway, while *MIF*, *STAT3*, *IL10*, and *IL12RB1* were the IL-12 pathway entities found in the analysis. *IL22* and *INNP5D* had not been related to AD before 2016. Moreover, enrichment was also found in transcriptional regulation of granulopoiesis (FDR 0.074; *p*-value 0.009), the process leading to the production of neutrophils, eosinophils, and basophils. Signaling cascades by *MAPK1* (FDR 0.04; *p*-value 0.002) and *FGFR* (FDR 0.04; *p*-value 0.003) were also significantly enriched. The related entities found in the analysis were *STAT3*, *FLG*, *IL2RA*, *IL5RA*, *IL6R*, and *MCM10*.

Seven out of the 62 genes described in the period covered by this review have been curated with functions in pathways related to extracellular matrix organization (FDR 4.58e-2; *p*-value 3.81e-3). Interestingly, 6 of these genes (*COL5A3*, *COL6A6*, *KDR*, *MCM10*, *MMP9*, and *STS*) had not been associated with AD yet, and only TGF- β 1, involved in a broad spectrum of pathways, had been previously associated [97].

We also performed an analysis of disease-related genes using the FunRich software. The results are shown in Figure 5. The appearance of DOCK8 in all these biological processes stands out, taking into account that it has been described as related to AD only in one study reporting a single case [77]. COSMIC analysis located 75.4% of genes in the skin (*ACTL9, ADAM33, ADCY10, C11orf30, CARD11, CARD14, CLDN1, COL5A3, CRNN, CUX2, CYP27A1, DEFB1, DOCK8, FLG, GSDMB, IL12RB1, IL22, IL2RA, IL4, IL5RA, IL6R, IL9, KDR, LILRA6, LRRC32, MAST2, MCM10, MMP9, MTF1, NLRP2, ORM2, PANX3, PHLDB1, PRR5L, RPTN, RTEL1, SPINK5, STS, TCHHL1, TGFB1, TLR2, TLR4, and TNF).*



Biological process

Figure 5. Most relevant biological processes involving the reported genes. Percentages indicate the number of genes with respect to total that is included in each process.

Additionally, 17 of the new AD identified genes could not be ascribed to significant biological functions, i.e., *CARD14*, *CRNN*, *TCHHL1*, *RPTN*, *PANX3*, *PHLDB1*, *LILRA6*, *NLRP2*, *MTF1*, *LTA*, *MAST2*, *DOCK8*, *CUX2*, *ADCY10*, *VSTM1*, and *RTEL1*.

3.3. Filaggrin

During the period of this revision, we have identified 33 studies on the *filaggrin* (*FLG*) gene association with AD. It is remarkable that 16 novel mutations have been reported [56,81,85,88].

3.3.1. Filaggrin Mutations and Other Allergic Diseases

Eleven studies analyzed the association between *FLG* mutations and allergic sensitization, showing that *FLG* alleles conferred an increased risk, mainly in children with eczema [13,39,41,44,47,50,51,58,70,72,76]. In Polish children, Debinska et al. showed that several *FLG* mutations predisposed patients to eczema plus asthma, increasing more than 6-fold the risk of this complex phenotype (*p*-value 0.043) [72]. By contrast, such increased risk of asthma in *FLG* mutation was not confirmed in adult twins, although the risk of having AD was increased in those individuals with asthma, compared to individuals without asthma (27.6% vs. 18.5%; OR 1.68, 95% CI 1.12–2.52; *p*-value 0.012) [58]. Chan et al. showed a significant effect of *FLG* loss-of-function (LoF) mutations on both asthma and rhinitis at ages 1, 2, 4, 10. and 18 years, particularly at the age of 10 years (RR 1.96; 95% CI 1.70–2.26; *p*-value 0.003), early eczema being a requisite to suffer asthma at all ages [70].

Ferreira et al. carried out GWAS on individuals suffering from asthma, hay fever, and eczema to identify shared risk variants. The rs6181676[A] FLG variant was 1.32-fold more common in individuals suffering only from eczema when compared to individuals suffering only from hay fever (p-value 7.2e-8), and 1.26-fold comparing with asthma-only cases [13]. Two FLG single nucleotide polymorphisms (SNP), rs71626704 and rs76413899, were significantly associated with a history of asthma and cheilitis (p-value 0.002 and p-value 0.003, respectively) and rs62623409 and rs71625199 SNPs were associated with sensitization to environmental allergens (*p*-value 0.038 and *p*-value 0.008, respectively). Rs11584340 was associated with an increase of eosinophil-derived neurotoxin serum levels in allergic rhinitis patients and eosinophilic cationic protein serum levels in asthmatic patients [41]. Park et al. reported an association between FLG LoF mutation and early onset of asthma and AD [87]. The association between an FLG mutation and IgE sensitization to peanut at age 4 years (OD, 1.88; 95%) CI 103–3.44), but not to other allergens was reported by Johansson et al. [39]. Equally FLG mutations were significantly associated with elevated IgE in a population of Korean patients with AD (>200KIU/L and/or MAST-CLA>+, p-value 0.005), palmar hyperlinearity (p < 0.001), and a family history of allergic disease (p-value 0.021) [50]. However, there was no significant difference in IgE levels between AD patients with non-mutated FLG and those carrying FLG LoF mutations (p-value 0.062) [44].

3.3.2. Filaggrin Mutations and Early Onset of AD

Patients who carried *FLG* mutation alleles are associated with early-onset AD [62,72]. Wan et al. found a dose-dependent association between the number of common *FLG* mutations and early onset [62]. In a Finish population, the combination of *FLG* mutations was shown to be significantly associated with early-onset of AD (<2 years) (OR 4.15, *p*-value 1.82e-10) and asthma (OR 2.76, *p*-value 1,57e-6) [47]. In two independent cohorts, *FLG* LoF mutations were associated with subphenotypes of AD. Thus, in a cohort study of 14,701 children from Avon (UK), the strongest association was detected with early-onset-persistent AD (OR 4.31; 95% CI 3.29–5.63; *p*-value 2e-26) and in a Dutch cohort of 3963 children, only the group of children with early-onset-late-resolving AD was associated with *FLG* LoF mutations (OR 5.63; 95% CI 2.65–11.95; *p*-value 7e-6) [76].

Additionally, it has been demonstrated that *FLG* expression in umbilical cord blood was associated with eczema development in infancy, being significantly lower in children with *FLG* variants when compared to children with wild-type *FLG* genotype (*p*-value 0.007) [75].

3.3.3. Filaggrin Mutations and other Skin Diseases

Andersen et al. studied the prevalence of *FLG* null mutations in adult patients with actinic keratosis (AK), premalignant intra-epidermal skin lesions that can progress into squamous cell carcinomas (SCCs). In their study, 7.5% AK patients had an *FLG* LoF mutation, of whom only the homozygous mutation carriers (0.8%), but not heterozygous, showed an increased risk of AK compared with wild-types (*p*-value 0.0017) [65]. Elhaji et al. found a significant association between the R501X mutation with polysensitivity in contact dermatitis when three or more positive patch test reaction occurred (8.5% patients vs. 4% controls; *p*-value 0.008) [91].

FLG mutations were significantly associated with palmar hyperlinearity in a population of Korean patients with AD (p < 0.001) [50], and also in a Finish population (OR 4.67, *p*-value 1.46e-5) [47].

The specific variant rs558269137 was exclusively detected in Italian children with AD and *Molluscum contagiosum* virus (MCV) infection, while rs374910442, rs138055273, rs113136594, and rs11584340 variants were found both in AD children and AD plus MCV-infected children [48].

3.3.4. Filaggrin Mutations and Eczema Severity

Seven studies analyzed the association between *FLG* mutations and eczema severity [48,50,53,71, 87,88,90]. Chang et al. found that *FLG* LoF homozygotes and heterozygotes were less likely to report periods of skin clearance (OR 0.20; 95% CI 0.07–0.55) and more likely to report frequent steroid use (OR 3.18; 95% CI 1.22–8.30) [71].

Specific gene variants of *FLG* have been associated with moderate to severe SCORAD (Scoring Atopic Dermatitis) indexes. Rs145627745, rs79808464, rs150957860, and rs145828067 variants were entirely associated with moderate disease severity (SCORAD 25-50), rs747005144 variant was associated with severe disease (SCORAD >50), whereas rs374910442, rs138055273, rs183942200, rs1584340, and rs113136594 variants were associated with both moderate and severe disease [48].

In a population of African American (AA) children with AD and *FLG* LoF, 77% exhibited severe AD (SCORAD >50) [90]. A negative effect on the success of the immunosuppressive treatment was reported in *FLG* mutated patients when compared with those without *FLG* mutations [53]. When exploring the correlation between Eczema Area and Severity Index (EASI) and *FLG*-related AD in Korea, contradictory results were reported [50,87]. In addition, Wong et al. did not find any significant association between *FLG* LoF and the severity of AD [88].

3.3.5. Filaggrin Mutations and Ethnicity Risk Factors

Fifteen studies were carried out on European ancestor populations [12,13,36,39,44,47,48,53,58,70, 72,75,76,91,98], 11 studies in Asia populations [37,45,50,51,54,56,64,67,81,87,88], 2 studies in African ancestor populations [85,90], and 3 studies in mixed populations [62,71,73]. The most prevalent LoF in patients with European ancestors were R501X, 2282del4, S3247X and R2447X, analyzed in 27 studies [12,13,36,37,39,44,47,48,50,53,54,56,58,60,62,64,70–73,75,76,81,85,88,90,91]. Other ethnic ancestors and rare variants were analyzed in 15 studies [37,41,45,47,50,51,53,54,56,64,81,85,87,88,90].

Elbert et al. carried out a study to analyze the association of ethnic origin with *FLG* mutations and environmental risk factors in children from multiethnic origins but living in the Netherlands, showing that minority ethnicity children had a higher risk of eczema than Dutch children [73]. Gimalova et al. studied LoF variants in Russians and Tartars AD patients, reporting that c.2282del4 was the most prevalent mutation in both populations, whereas R501X and R2447X mutations were rare [36]. In India, a study of the association between *FLG* mutations and hand eczema showed that mutations in S2889X constituted 96.4% of all *FLG* mutations, while European mutations were not found [37].

An overview of the genetic map and geographic distribution of *FLG* mutations across East Asia found that 3321delA is a pan-Asian mutation [45]. K4022X, the most prevalent *FLG* mutation in Korea and northern China, showed a south-to-north distribution gradient. In contrast, c.6950del8 showed the reverse effect. On the other hand, S2554X, S2889x, S3296X, and Q1701X mutations were Japanese-specific. These *FLG* mutations were associated with an increased risk of AD but did not confer a risk of asthma [45].

On et al. carried on a study of *FLG* mutations previously detected in Korean, Japanese, and Chinese patients on seventy Korean patients with AD. Four LoF mutations (3321delA, K4022X, S3296X, and S2889X) were identified in 15.7% patients [50]. The most commonly detected variants in Korean patients with AD were 3321delA (9.1%) and Y1767X (1.6%), K4022X (4.3–4.5%) [50,51,87]. Interestingly, Y1767X was only found in AD patients, whereas K4022X was found in both patients and controls [51].

Pigors et al. analyzed the genetic scheme of AD patients from the South Asian Bangladeshi community using WES combined with rare variant enrichment analysis [81], showing that *FLG* carried the highest number of enriched dominant (OR 12.1; *p*-value <0.0001) and recessive (OR 43.4; *p*-value <0.0001) LoF mutations. Three of the LoF mutations were previously unreported (S923Ffs*2, T1545Qfs*163, and S2352X). Furthermore, these genetic data revealed intrafamilial heterogeneity with multiple *FLG* variants often segregating within the Bangladeshi families with AD [81].

The common European *FLG* LoF R501X and 2282del4 were significantly associated with the risk of developing AD (OR 11.29, *p*-value 0.00022 and OR 2.66, *p*-value 0.00016, respectively) in Finnish patients [47]. In addition, having two 12-repeat alleles (rs12730241) was found to be significantly associated with a higher risk of AD (OR 1.96, 95% CI 1.36–2.81, *p*-value 0.00056) [47].

Using massively parallel sequencing, Margolis et al. identified nine *FLG* LoF variants in AA children, including 6 newly reported and 3 previously described, suggesting multiple and rare *FLG* LoF variants. Those children with *FLG* LoF variants had more persistent AD than wild-type children for

FLG LoF [85]. These findings were supported by Mathyer et al., who identified five *FLG* LoF variants in 9 heterozygous AA AD patients (488delG, R501X, R826X, S3101X, and S3316X) [90].

New mutations were also found in Japanese (R826X) and Korean (S2889X) ichthyosis vulgaris (IV) patients [56]. Although the R826X mutation has not been detected in Japanese and Korean AD patients to date, it had been previously reported in Chinese and AA populations [95,99], suggesting that it is not a population-specific *FLG* mutation. Japanese and Korean patients shared 4 *FLG* mutations, Gly1109Glufs*13, Ser2889X, Ser3296X, and Lys4022X, being the latter more frequent in Korean than in Japanese AD or IV patients [56].

A robust and cost-effective high-throughput PCR-based method using microfluidics technology and NGS was applied to study the *FLG* coding region in cohorts of Chinese, Malaysian, and Indian AD patients living in Singapore. Thirty-three *FLG* LoF variants were identified in Chinese subjects, being 5 of them novel mutations. Unreported *FLG* LoF variants in Indian and Malaysian patients confirmed the diversity depending on the ethnic group [88].

3.4. Other Genes

Significant associations with AD have been reported for most of the analyzed genes and variants. Thus, *ACTL9*, *C11orf30*, *IL6R*, *IL21*, *IL22*, *INPP5D*, *KIF3A*, *OVOL1*, *PRR5L*, *PPP2R3D*, and *STAT3* were investigated in two cohorts, ALSPAC and PIAMA [76]. *ADCY10*, *CUX2*, *MAST2*, *MCM10*, *MTF1*, *ORM2*, *PHLDB1*, and *TCHHL1* were analyzed in Bangladeshi patients [81]. A *CLDN1* polymorphism was positively associated with early onset of AD in Ethiopian patients [35], but no association was found in the Finnish variants [47]. *CARD11*-R30W has been associated with recurrent infections, autoimmunity, and severe atopy [78], and other dominant, negative mutations in *CARD11*, leading to dominantly inherited, severe atopy have been described in 4 unrelated USA families [79]. Within the same protein family, downregulation of *CARD14* was reported to lead to severe AD and reduced skin protection against infection as well as dysregulated cutaneous inflammation pathways [80]. Rs199691576, a polymorphism of *CP27A1*, was also related to AD in Japan [82].

Mutations in barrier and immune-related genes, i.e., *KLK7*, *SPINK5*, *DEFB1*, *KDR*, *IL5RA*, *IL9*, *IL12RB1*, and *IL13*, were found more frequently in Korean AD patients than in healthy controls [64]. *SPINK5* (serine peptidase inhibitor kazal type 5), a protein involved in epidermal cell differentiation, was also associated with AD in Ethiopian [66] and Japanese patients [49].

Other genes involved in immune functions, such as *IL2RA* [13,76], *IL4* and *ADAM33* [84], *TGFB1* [57], and *MIF* promoter [42] have also been significantly associated to increased risk of AD. The relationship of TLRs polymorphisms with AD, i.e., *TLR2* rs55743708(G>A) and *TLR4* rs4986790(A>G) were reported to increase levels of IL-4 and IL-10 in Russian AD patients [61], while other authors found no association of *TLR2* polymorphisms, i.e., rs5743708 and rs4696480, in Turkish children with AD [69]. Also, significant SNPs in *TSLP*, a lymphopoietin, have been reported in Chinese Han [38], Korean [43], and American [62,71] patients. Lymphotoxin α , a protein involved in IL-2 and IL-4 signaling events, more specifically, *LTA* rs2844484, was associated with AD in Greenland patients [67]. In a study performed in Jordan investigating the relationship between *RETN* gene polymorphisms and AD, rs3745367 was found significantly associated with AD in a gender- and age-specific manner [46].

Variants in extracellular matrix genes such as *COL5A3* rs2287807 and *MMP9* rs17575 were found as significantly related to AD in a meta-analysis performed in French, Canadian, and UK families [93]. *COL6A6* minor allele (*AA*) in rs16830494 and the rs59021909 (*TT*) allele and the rs200963433 heterozygous (*CT*) showed higher frequency in patients than in controls, although no statistical significance was reached [83]. *TMEM232* rs11357450 had the strongest association with the risk of AD among all the variants analyzed by Wu et al. [74]. Mutations in *SHARPIN*, a protein involved in epidermis development, that were exclusively present in patients, decreased its expression in AD lesions [55]. *GSDMB* rs921650 was characterized as a strong risk factor for eczema [13].

3.5. Epigenetic Studies

31 of 46

Over the period included in this revision, the studies of epigenetic modifications on atopic dermatitis were focused on DNA methylation and microRNAs. These epigenetic mechanisms have been shown to be crucial regulators in different allergic conditions [100,101], although histone modifications have also been studied in the context of allergic diseases and have been shown to play a role in their development [102].

With respect to DNA methylation, two articles studied such modification at a whole-genome level in blood samples. In the first one, Ferreira et al. analyzed the association of DNA methylation with different allergic risk factors. In this manner, they detected 36 genes with DNA methylation sites nearby that were associated with differences in gene expression between allergic patients and healthy controls. Additionally, they found an association between smoking and the methylation state of *PITPNM2* [13], potentially involved in neutrophil function [103,104]. The second study found 490 CpGs differentially methylated between AD patients with eczema herpeticum and healthy controls and 6 CpGs differentially methylated when comparing AD patients without eczema herpeticum and healthy controls. Among these sites, they identified CpG methylation sites in IL4 and IL13, which suggested that there was a significant association between these methylations and the phenotype observed in the patients [28]. Another two studies were centered on the methylation state and its effect on gene expression of *NLRP2* [33] and *SIRL-1* [31]. These studies showed the influence of single nucleotide polymorphisms, a correlation of the gene expression level, and the presence or absence of AD condition.

Regarding the miRNA research, the different analyzed studies can be grouped by two main approaches to the function of miRNAs in the development of AD. On the one hand, the studies that assessed for the upregulation or downregulation of miRNAs in lesioned tissue of AD patients. In this way, several differentially expressed miRNAs were described in AD lesions. When comparing the lesional tissue of AD patients with normal skin samples of healthy controls, Yang et al. described miR-124 downregulation in AD lesional tissue [25] and in an in silico interaction analysis of differentially expressed miRNAs, Li et al. [32] postulated that downregulation of hsa-let-7a-5p would potentially upregulate CCR7, a chemokine receptor involved in the activation of T cells [105]. They also found a differential expression of miR-143, whose potential target is DENND1B, which is involved in the proliferation of T-cells [34]. In addition, the authors suggest that the downregulation of miR-26 would regulate hyaluronan synthase 3 (HAS3), which is upregulated in AD skin [32,106]. After comparing lesional skins samples with non-lesional skin samples in AD patients, Ding et al. proposed a regulatory network of differentially expressed genes that included 182 miRNAs, and among them, hsa-miR-148b, hsa-miR-152, and hsa-miR-324 [24]. Finally, using primary adult human keratocytes, several out of the 372 most common miRNAs were dysregulated when exposed to IL-4, which plays a key role in the development of AD [27].

On the other hand, there are some studies that look for miRNAs differentially expressed in sera from AD patients compared with healthy controls. Thus, the levels of miR-144 detected in umbilical cord serum were higher in those Japanese children that would develop AD at one year of age [26], levels of miR-151a and miR-409 were found to be in higher in sera from Chinese AD patients compared with healthy controls [30], and finally, miR-146a showed no difference in serum levels between patients with AD and healthy individuals [29], despite previously demonstrating a role in the regulation of the immune system and inflammatory responses pathways [107,108]. This second method of retrieving candidate miRNAs may serve as a feasible and less invasive way of obtaining new AD biomarkers, whereas the former procedure in lesioned tissue focused more on the mechanistic of action of such RNAs.

4. Discussion

Herein, we have systematically reviewed the literature related to genetics and epigenetics of AD published between June 2016 and December 2019. We have found 58 original articles and

6 meta-analyses and also 9 epigenetic studies. A total of 62 genes have been analyzed in the selected publications, 31 of which had not been reported as potentially associated with AD before June 2016.

One remarkable feature about allergic diseases is the diversity in the potential phenotypes sharing the same genotype, indicating that there appear to be additional components that increase the complexity of the regulation and development of such conditions, or at least shape its evolution over time [109,110]. Epigenetic regulation has emerged as a key factor that was missing to completely understand the molecular basis of allergic disease [111].

4.1. Filaggrin

Up to the revision date of this review, *FLG* LoF mutations were the most significantly associated genetic variants for AD. Filaggrin is a key protein in the differentiation of the epidermis and the formation of the skin barrier, which is necessary to prevent water loss through the epidermis and to avoid the entry of allergens, toxins, and pathogens [112]. Its precursor profilaggrin is encoded by the *FLG* gene, which is located on chromosome 1q21.3 [113] within a region known as the epidermal differentiation complex (EDC) comprising over 50 genes encoding proteins involved in terminal differentiation and cornification of keratinocytes [114]. LoF mutations in the exon 3 completely hinder FLG protein expression, increasing the risk of AD [113,115–119]. A meta-analysis of 24 studies on *FLG* untations determined a 3-fold increased risk of AD in those individuals carrying one or more *FLG* LoF, singling out the influence of one gene in such a heterogeneous disease [120]. More than 300 *FLG* LoF variants have been identified in the *gnomAD* browser, an international database of exome and genome sequencing data (https://gnomad.broadinstitute.org), more than 20 of them associated with susceptibility to AD [85].

The results showing that *FLG* null mutations conferred risk for allergic sensitization and susceptibility to ezcema-associated asthma are well aligned with previous studies [121–127]. All these findings support the idea that *FLG* mutations lead to functional epidermal barrier defects, increasing skin permeability and subsequent allergic sensitization, promoting the Th2 inflammatory response, and eventually leading to asthma [122]. The "outside–inside" theory of AD pathogenesis proposes that epidermal APCs in AD patients are overexposed to danger signals because of their impaired skin barrier, leading to APC maturation and T-cell-mediated inflammatory skin disease [128].

AD has been divided into early-onset and late-onset forms. Early-onset AD would be especially driven by genetic factors, whereas late-onset AD might depend on environmental exposures [129]. Common *FLG* null mutations associated with early-onset AD are described in different populations [126, 130–133]. *FLG* LoF mutations have also associated with moderate-or-severe AD cases [117,122,134–138].

The most frequent *FLG* LoF mutations (R501X, 2282del4, S3247X, and R2447X) are present in 7–10% of Europeans [115,120] while these mutations are rare in Asian patients, who carry specific mutations [45,50,139,140]. Thus, K4022X has been reported as the most prevalent variant in Korean AD patients [50,51,87]. Interestingly, *FLG* mutations in Korean AD patients seem to be less frequent than in other East Asian countries, most likely due to genetic and environmental factors or mutations in other barrier genes [50,87]. Also, the analysis of *FLG* mutations in East Asia showed a geographic distribution in agreement with the history of human migrations [45].

On the other hand, *FLG* LoF mutations could be less common in patients with African descent than in those with European or Asian descent [139,140], although other studies have shown that AA children had an increased risk of AD compared with European children [140]. The prevalence of AD in the US was reported as the highest among AA patients, but this population remains largely understudied [141,142]. Recently, two studies using current sequencing methods were able to identify rare *FLG* LoF variants in AA children associated with more persistent AD [85,90]. The prevalence of common *FLG* variants in children of African ancestry is less frequent than those of European or Asian ancestry [85]. Moreover, hygiene habits, vitamin D level, sun exposure, microbiota, genetics, and skin barrier characteristics could also influence the association of ethnicity with AD [1,143].

The implementation of new technologies like NGS to analyze cohorts of understudied populations, as well as newer bioinformatic tools, will allow the identification of new *FLG* LoF mutations and confirm the variation among the different ethnicities.

4.2. Other Genes

Regarding research contributions in the period of this review, new components of the extracellular matrix have been described to be associated with AD [64,81,83,93,144]. These new associations highlight the importance of such structure in the development of AD and in the integrity of the skin barrier. Thus, COL5A3, COL6A6, and MMP9 are important for the collagen formation [145–147]; KDR, one of the two receptors of the VEGF, has been associated with integrin cell surface interactions with extracellular matrix [148]; mutations in STS (steroid sulfatase) have been associated to X-linked ichthyosis [148]; rare variants of MCM10, a key component of the pre-replication complex, have been described for the first time associated to AD [81]. Regarding TGF- β 1 [57], besides its roles in other pathways, different works have shown its role in extracellular matrix assembly and disassembly [149,150].

Some genes associated to AD have been related to innate immune system pathways, providing solid evidences of the relationship of the innate immune system with the disease and its progression. On its behalf, most of the genes of the review associated to innate immune pathways (ADAM33, MIF, MMP9, ORM2, RETN, and TLR2) are related to neutrophil degranulation that contributes to the inflammation of the tissue in the AD [151,152]. In addition, a substantial number of publications [61,69,95,153] emphasize the importance of Toll-like receptor cascades on the development of AD and its link with other allergic diseases [154,155]. The TLR-2 rs4696480 polymorphism has also been associated with AD severity in adult patients in two different populations, and supported with functional studies [156,157]. However, one of the studies included in this review reported a lack of association of this polymorphism in Turkish children with AD [69], which could be due to the fact that differences in genetics and environmental factors appear to be relevant in the development of allergy. New pieces of evidence have been added to the previously reported association of genes such as ADAM33, CARD11, and DEFB1 with AD [64,67,78,79,84]. In this line, ADAM33 has also been associated to other allergic diseases like asthma [158] and allergic rhinitis [159]. CARD11 is required for B- and T-cell receptor signal transduction and activation of NF-KB transcription factor [160]. DEFB1 is an antimicrobial peptide implicated in the resistance of epithelial surfaces to microbial colonization. It is a member of the family of defensins, peptides made by neutrophils, and it has been proposed as a link between the innate and the adaptive immune systems [161,162].

It is noteworthy that several of the new genes listed in this revision do not fall into any of these functional groups. This may be due to different causes, as because of little knowledge about some genes or because they are not properly curated yet. This might be the cases of CARD14, with similar functions to CARD11 [163]; VSTM1, which behaves as a cytokine [164]; LILRA6, a member of the leukocyte immunoglobulin-like receptor family [165]; mutations in DOCK8 are responsible for an immunodeficiency syndrome [166]; NLRP2 is involved in inflammatory processes [167,168]; RTEL1, a helicase involved in telomere maintenance, has also been found associated to severe dyskeratosis congenita [169]; LT- α (also known as TNF- β), which is a cytokine produced by lymphocytes [170]. Additionally, ADCY10, CUX2, MAST2, MTF1, PANX3, PHLDB1, and SCAND3 have been associated to AD in a single study of exome sequencing [81]. The S100 fused type protein (SFTP) family includes genes which are mainly expressed in stratified epithelia and play a role in epithelial homeostasis [114]. SFTPs contain two calcium-binding domain EF-hand motifs and are associated with cytoplasmic intermediate filaments as well as minor components of the cornified envelope [114]. This family of proteins include 7 members, FLG being its most studied member and certainly showing an association with AD. Besides FLG, only FLG2 and HRNR were previously associated with AD [171–173]. Interestingly, over the last 5 years, other 3 members of the family have been associated with AD. The three members now associated with AD, CRNN, RPTN, and TCHHL1 were known to be involved in different epithelial

disorders [174–176]. Taken together, SFTP family proteins pose as pivotal players in the proper skin cornification and in the development of AD.

4.3. AD Epigenetics

In recent years, the search for risk factors that help to understand how allergic diseases develop has become one of the main objectives of the research. The plasticity observed in the different phenotypes associated to an underlying genotype suggests that additional components may provide complexity to the processes that lead to the development of the disease, or, at least, influence its evolution [109,177]. In the last years, the focus has been set on epigenetic modifications, which can lead to the development of allergic diseases. Epigenetic regulation has emerged as a pivotal key in the comprehension of the molecular basis of allergic conditions [111,178].

Most of the research on AD epigenetic regulation has focused on the posttranscriptional regulation mediated by miRNAS. miRNAs constitute a class of small non-coding RNAs, with a size ranging from 17 to 25 nucleotides, and a sequence that allows them to bind to specific mRNAs. This key feature permits the posttranscriptional modulation of targeted genes by triggering mRNA degradation and/or inhibition of translation [179]. According to several functional studies, miRNAs are involved in virtually every cellular process [180]. miRNAs have also been related to immune system regulation, miR-21, miR-146a, and mIR-155 being the most extensively studied. A role in the regulation of the immune response and tissue inflammation in allergic diseases has been shown [101].

The analysis of lesional tissues has provided new miRNA molecules that could regulate different mechanisms and signalling pathways that are altered in AD lesions. As a general feature, a decrease of miRNAs involved in the regulation of the immune response and an increase of miRNAs involved in epidermis development is observed in these studies. Thus, downregulation of miR-124 in AD lesions has been shown to control NF-κB-dependent inflammatory responses in keratinocytes and chronic skin inflammation in atopic eczema [25]. Bioinformatics analyses from two studies suggest that miRNAs can influence the transcriptional regulation of signalling pathways related to the synthesis of extracellular matrix components, such as arachidonic acid and hyaluronic acid, as well as participate in processes such angiogenesis, lymphangiogenesis and apoptosis, all of which are involved in AD progression [24,27].

In addition, the use of miRNAs as biomarkers in allergic diseases is increasingly described [181,182]. In this sense, miR-151a and hsa-mir-144-3p have been proposed as potential biomarkers in AD, as they have been shown to be differentially expressed in serum and umbilical cord serum, respectively [26,30]. miR-151a would reduce IL12RB2 levels in T-cells, favouring the increase of Th2 cells, which are central in the pathogenesis of AD [30,183]. Also, an increased expression of miR-144 would reduce *ABCA1* mRNA and protein levels and induce a proinflammatory response via NF- $\kappa\beta$ [26]. Nevertheless, the differences in the expression of these small non coding RNAs observed in the lesional skin does not necessarily translate to changes in serum levels, which impairs their use as biomarkers. This is the case of miR-146a, which has been shown to be upregulated in AD lesional skin when compared to healthy controls [184] but showed no differences in serum levels [29].

Another extensively studied epigenetic mechanisms is DNA cytosine methylation [185]. This modification occurs in CpG dinucleotides which are grouped in the so called CpG islands, frequently located in intergenic regions, as well as in promoter region of genes [186]. CpG islands methylation of gene promoters is related to the repression of the transcription, i.e., CpG islands of genes that are being actively transcribed do not usually present methylation, while non-transcribed genes present CpG islands with high degrees of methylation [187]. Cytosine methylation suppresses gene transcription, as it causes chromatin condensation and prevents transcription factors from binding to their target sequences in promoters [185]. Different EWAS have shown differential methylation patterns associated with some pathologies [188,189] or even with the exposure to different agents [190–192]. Two studies showed gene expression modulation due to exposure to tobacco smoke in AD [13,33]. Thürmann et al. showed that *NLRP2* was differentially methylated, due to both the effect of polymorphisms and tobacco

smoke [33], and Ferreira et al. found differences in the methylation of *PITPNM2*, which were partly associated with environmental tobacco smoke [13]. Interestingly, both genes are involved in innate immune responses, with *NLRP2* involved in inflammatory processes related to macrophages [193,194] and *PITPNM2* related to neutrophils [103,104]. Other two studies found differences in DNA methylation patterns in AD patients. The first one found differences in methylation of the promotor region of the VSTM1 gene locus [31], which encodes the protein SIRL-1 that has been proposed to inhibit crucial pro-inflammatory functions in human myeloid cells [195,196]. The second one was a genome wide methylation study that found different patterns of methylation in over 490 sites in AD patients with eczema herpeticum, and where Boorgula et al. identified a significant association between *IL4* and *IL13* methylation and the AD phenotype, as well as with serum IgE levels [28]. However, this methylation patterns where shown to be highly influenced by the eosinophilic count [28].

5. Final Remarks

In the present article, we have evaluated the last 5 years of AD-related literature using systematic review methodology. We have focused on genetics and epigenetics aspects of the disease, monitoring the different polymorphisms and gene variations associated to the onset or severity of AD, comparing studies performed in different locations and including several ethnicities, therefore showing an up-to-date picture of current knowledge. Some of the retrieved articles used state-of-the-art technology when assessing their findings, including genome-wide sequencing of representative samples of patients. An exhaustive analysis of risk of bias and quality of the 64 selected articles have allowed us to ponder the validity of the reported associations. Another strong point is the inclusion of epigenetic studies.

Regarding limitations to the present review, it has to be point out that we have restricted our analysis to those genes included in the articles published in the last 5 years. Although the main genes related to disease onset and development, i.e. filaggrin, have been included, we are aware that other important genes, already reported elsewhere, may be missing here. Since our goal is to update the topic with new results, we highly recommend the interested reader to consult the previous reviews for more information [16,17].

In addition, we should remark that most genes have been described only once and for a limited number of patients. For instance, *DOCK8* has been identified in a single case report. Larger clinical trials would be required to unambiguously link these genes to AD. The universalization of the whole genome techniques will allow the discovery of new mutations or confirm the already known ones in different populations.

New developments in genetics and epigenetics technology offer opportunities to improve the diagnosis of AD patients, ascribing them to specific genetic groups and allowing the tailoring of therapy with the best response to ensure the most convenient patient care.

Supplementary Materials: The following are available online at http://www.mdpi.com/2073-4425/11/4/442/s1, Figure S1: Reactfoam of pathways overview of the genes reported in the selected studies; Table S1: Analysis of the risk of bias for the selected genetic studies; Table S2: Quality assessment of the selected genetic studies.

Author Contributions: M.J.M., M.E., and A.G.-S. designed and performed the systematic review, analyzed and validate the data, and co-wrote the original draft. M.J.M. and M.E. performed bioinformatic analyses. I.D., M.I.-G., and C.S. supervised the review, revising it critically, and gave the final approval of the manuscript. All authors have read and agreed to the published version of the manuscript.

Acknowledgments: This work was supported by the Instituto de Salud Carlos III (ISCIII), cofounded by the European Regional Development Fund (Grant PI17/00818, PI: Ignacio Dávila), the Network for Cooperative Research in Health, RETICS ARADYAL (RD16/0006/0019, PI: Ignacio Dávila), Junta de Castilla y León Health Council (GRS1596/A/17, PI: María Isidoro), Junta de Castilla y León Education Council (CAS086P17, PI: Ignacio Dávila).

Conflicts of Interest: The authors declare no conflict of interest.

References

- Weidinger, S.; Beck, L.A.; Bieber, T.; Kabashima, K.; Irvine, A.D. Atopic dermatitis. *Nat. Rev. Dis. Prim.* 2018, 4, 1. [CrossRef] [PubMed]
- 2. Silverberg, J.I. Atopic Dermatitis in Adults. Med. Clin. N. Am. 2020, 104, 157–176. [CrossRef] [PubMed]
- 3. Dharmage, S.C.; Lowe, A.J.; Matheson, M.C.; Burgess, J.A.; Allen, K.J.; Abramson, M.J. Atopic dermatitis and the atopic march revisited. *Allergy* **2014**, *69*, 17–27. [CrossRef] [PubMed]
- 4. Bonamonte, D.; Filoni, A.; Vestita, M.; Romita, P.; Foti, C.; Angelini, G. The Role of the Environmental Risk Factors in the Pathogenesis and Clinical Outcome of Atopic Dermatitis. *Biomed. Res. Int.* **2019**, 2019, 2450605. [CrossRef]
- Manousaki, D.; Paternoster, L.; Standl, M.; Moffatt, M.F.; Farrall, M.; Bouzigon, E.; Strachan, D.P.; Demenais, F.; Lathrop, M.; Cookson, W.O.C.M.; et al. Vitamin D levels and susceptibility to asthma, elevated immunoglobulin E levels, and atopic dermatitis: A Mendelian randomization study. *PLoS Med.* 2017, 14, e1002294. [CrossRef] [PubMed]
- 6. Schram, M.E.; Tedja, A.M.; Spijker, R.; Bos, J.D.; Williams, H.C.; Spuls, P.I. Is there a rural/urban gradient in the prevalence of eczema? A systematic review. *Br. J. Dermatol.* **2010**, *162*, 964–973. [CrossRef]
- 7. Romieu, I.; Torrent, M.; Garcia-Esteban, R.; Ferrer, C.; Ribas-Fitó, N.; Antó, J.M.; Sunyer, J. Maternal fish intake during pregnancy and atopy and asthma in infancy. *Clin. Exp. Allergy* **2007**, *37*, 518–525. [CrossRef]
- Leermakers, E.T.M.; Sonnenschein-van der Voort, A.M.M.; Heppe, D.H.M.; de Jongste, J.C.; Moll, H.A.; Franco, O.H.; Hofman, A.; Jaddoe, V.W.V.; Duijts, L. Maternal fish consumption during pregnancy and risks of wheezing and eczema in childhood: The Generation R Study. *Eur. J. Clin. Nutr.* 2013, 67, 353–359. [CrossRef]
- 9. Willers, S.M.; Devereux, G.; Craig, L.C.A.; McNeill, G.; Wijga, A.H.; Abou El-Magd, W.; Turner, S.W.; Helms, P.J.; Seaton, A. Maternal food consumption during pregnancy and asthma, respiratory and atopic symptoms in 5-year-old children. *Thorax* **2007**, *62*, 773–779. [CrossRef]
- 10. Apfelbacher, C.J.; Diepgen, T.L.; Schmitt, J. Determinants of eczema: Population-based cross-sectional study in Germany. *Allergy* **2011**, *66*, 206–213. [CrossRef]
- Dalgard, F.J.; Gieler, U.; Tomas-Aragones, L.; Lien, L.; Poot, F.; Jemec, G.B.E.; Misery, L.; Szabo, C.; Linder, D.; Sampogna, F.; et al. The Psychological Burden of Skin Diseases: A Cross-Sectional Multicenter Study among Dermatological Out-Patients in 13 European Countries. *J. Investig. Dermatol.* 2015, 135, 984–991. [CrossRef] [PubMed]
- 12. Andersen, Y.M.F.; Egeberg, A.; Skov, L.; Thyssen, J.P. Comorbidities of Atopic Dermatitis: Beyond Rhinitis and Asthma. *Curr. Dermatol. Rep.* **2017**, *6*, 35–41. [CrossRef] [PubMed]
- Ferreira, M.A.; Vonk, J.M.; Baurecht, H.; Marenholz, I.; Tian, C.; Hoffman, J.D.; Helmer, Q.; Tillander, A.; Ullemar, V.; van Dongen, J.; et al. Shared genetic origin of asthma, hay fever and eczema elucidates allergic disease biology. *Nat. Genet.* 2017, *49*, 1752–1757. [CrossRef] [PubMed]
- 14. Ferreira, M.A.R.R.; Vonk, J.M.; Baurecht, H.; Marenholz, I.; Tian, C.; Hoffman, J.D.; Helmer, Q.; Tillander, A.; Ullemar, V.; Lu, Y.; et al. Eleven loci with new reproducible genetic associations with allergic disease risk. *J. Allergy Clin. Immunol.* **2019**, *143*, 691–699. [CrossRef]
- 15. Elmose, C.; Thomsen, S.F. Twin Studies of Atopic Dermatitis: Interpretations and Applications in the Filaggrin Era. *J. Allergy* **2015**, *2015*, 902359. [CrossRef]
- 16. Barnes, K.C. An update on the genetics of atopic dermatitis: Scratching the surface in 2009. *J. Allergy Clin. Immunol.* **2010**, *125*, 16–29. [CrossRef]
- Bin, L.; Leung, D.Y.M. Genetic and epigenetic studies of atopic dermatitis. *Allergy Asthma Clin. Immunol.* 2016, 12, 52. [CrossRef]
- Guyatt, G.; Oxman, A.D.; Akl, E.A.; Kunz, R.; Vist, G.; Brozek, J.; Norris, S.; Falck-Ytter, Y.; Glasziou, P.; Debeer, H.; et al. GRADE guidelines: 1. Introduction—GRADE evidence profiles and summary of findings tables. J. Clin. Epidemiol. 2011, 64, 383–394. [CrossRef]
- Sterne, J.A.C.; Savović, J.; Page, M.J.; Elbers, R.G.; Blencowe, N.S.; Boutron, I.; Cates, C.J.; Cheng, H.-Y.; Corbett, M.S.; Eldridge, S.M.; et al. RoB 2: A revised Cochrane risk-of-bias tool for randomized trials. *BMJ* 2019, 366, 148981.
- 20. Stang, A. Critical evaluation of the Newcastle-Ottawa scale for the assessment of the quality of nonrandomized studies in meta-analyses. *Eur. J. Epidemiol.* **2010**, *25*, 603–605. [CrossRef]

- Pathan, M.; Keerthikumar, S.; Ang, C.; Gangoda, L.; Quek, C.Y.J.; Williamson, N.A.; Mouradov, D.; Sieber, O.M.; Simpson, R.J.; Salim, A. FunRich: An open access standalone functional enrichment and interaction network analysis tool. *Proteomics* 2015, *15*, 2597–2601. [CrossRef] [PubMed]
- 22. Jassal, B.; Matthews, L.; Viteri, G.; Gong, C.; Lorente, P.; Fabregat, A.; Sidiropoulos, K.; Cook, J.; Gillespie, M.; Haw, R.; et al. The reactome pathway knowledgebase. *Nucleic Acids Res.* **2020**, *48*, D498–D503. [CrossRef] [PubMed]
- Szklarczyk, D.; Gable, A.L.; Lyon, D.; Junge, A.; Wyder, S.; Huerta-Cepas, J.; Simonovic, M.; Doncheva, N.T.; Morris, J.H.; Bork, P.; et al. STRING v11: Protein-protein association networks with increased coverage, supporting functional discovery in genome-wide experimental datasets. *Nucleic Acids Res.* 2019, 47, D607–D613. [CrossRef] [PubMed]
- 24. Ding, Y.; Shao, X.; Li, X.; Zhai, Y.; Zhang, Y.; Wang, S.; Fang, H. Identification of candidate genes in atopic dermatitis based on bioinformatic methods. *Int. J. Dermatol.* **2016**, *55*, 791–800. [CrossRef]
- Yang, Z.; Zeng, B.; Wang, C.; Wang, H.; Huang, P.; Pan, Y. MicroRNA-124 alleviates chronic skin inflammation in atopic eczema via suppressing innate immune responses in keratinocytes. *Cell. Immunol.* 2017, 319, 53–60. [CrossRef]
- 26. Dissanayake, E.; Inoue, Y.; Ochiai, S.; Eguchi, A.; Nakano, T.; Yamaide, F.; Hasegawa, S.; Kojima, H.; Suzuki, H.; Mori, C.; et al. Hsa-mir-144-3p expression is increased in umbilical cord serum of infants with atopic dermatitis. *J. Allergy Clin. Immunol.* **2019**, *143*, 447–450. [CrossRef]
- 27. Bao, L.; Chau, C.; Bao, J.; Tsoukas, M.M.; Chan, L.S. IL-4 dysregulates microRNAs involved in inflammation, angiogenesis and apoptosis in epidermal keratinocytes. *Microbiol. Immunol.* **2018**, *62*, 732–736. [CrossRef]
- 28. Boorgula, M.P.; Taub, M.A.; Rafaels, N.; Daya, M.; Campbell, M.; Chavan, S.; Shetty, A.; Cheadle, C.; Barkataki, S.; Fan, J.; et al. Replicated methylation changes associated with eczema herpeticum and allergic response. *Clin. Epigenetics* **2019**, *11*, 122. [CrossRef]
- 29. Carreras-Badosa, G.; Runnel, T.; Plaas, M.; Karner, J.; Ruckert, B.; Lattekivi, F.; Koks, S.; Akdis, C.A.; Kingo, K.; Rebane, A. microRNA-146a is linked to the production of IgE in mice but not in atopic dermatitis patients. *Allergy* **2018**, *73*, 2400–2403. [CrossRef]
- 30. Chen, X.-F.; Zhang, L.-J.; Zhang, J.; Dou, X.; Shao, Y.; Jia, X.-J.; Zhang, W.; Yu, B. MiR-151a is involved in the pathogenesis of atopic dermatitis by regulating interleukin-12 receptor beta2. *Exp. Dermatol.* **2018**, 27, 427–432. [CrossRef]
- Kumar, D.; Puan, K.J.; Andiappan, A.K.; Lee, B.; Westerlaken, G.H.A.; Haase, D.; Melchiotti, R.; Li, Z.; Yusof, N.; Lum, J.; et al. A functional SNP associated with atopic dermatitis controls cell type-specific methylation of the VSTM1 gene locus. *Genome Med.* 2017, *9*, 18. [CrossRef]
- 32. Li, H.M.; Xiao, Y.J.; Min, Z.S.; Tan, C. Identification and interaction analysis of key genes and microRNAs in atopic dermatitis by bioinformatics analysis. *Clin. Exp. Dermatol.* **2019**, *44*, 257–264. [CrossRef]
- Thürmann, L.; Grützmann, K.; Klös, M.; Bieg, M.; Winter, M.; Polte, T.; Bauer, T.; Schick, M.; Bewerunge-Hudler, M.; Röder, S.; et al. Early-onset childhood atopic dermatitis is related to NLRP2 repression. J. Allergy Clin. Immunol. 2018, 141, 1482–1485. [CrossRef] [PubMed]
- 34. Yang, C.-W.; Hojer, C.D.; Zhou, M.; Wu, X.; Wuster, A.; Lee, W.P.; Yaspan, B.L.; Chan, A.C. Regulation of T Cell Receptor Signaling by DENND1B in T H 2 Cells and Allergic Disease. *Cell* **2016**, *164*, 141–155. [CrossRef]
- 35. Asad, S.; Winge, M.C.G.; Wahlgren, C.-F.; Bilcha, K.D.; Nordenskjöld, M.; Taylan, F.; Bradley, M. The tight junction gene Claudin-1 is associated with atopic dermatitis among Ethiopians. *J. Eur. Acad. Dermatol. Venereol.* **2016**, *30*, 1939–1941. [CrossRef] [PubMed]
- Gimalova, G.F.; Karunas, A.S.; Fedorova, Y.Y.; Khusnutdinova, E.K. The study of filaggrin gene mutations and copy number variation in atopic dermatitis patients from Volga-Ural region of Russia. *Gene* 2016, 592, 85–89. [CrossRef] [PubMed]
- 37. Handa, S.; Khullar, G.; Pal, A.; Kamboj, P.; De, D. Filaggrin gene mutations in hand eczema patients in the Indian subcontinent: A prospective case-control study. *Contact Dermat.* **2019**, *80*, 359–364. [CrossRef]
- 38. Jiang, X.-Y.; Zhao, J.-H.; Yu, C.-X.; Fang, L.; Zheng, X.-D.; Yin, X.-Y.; Wu, Y.-Y.; Tang, X.-F.; Zhou, F.-S.; Zhang, X.-J.; et al. Association analyses identify two susceptibility loci 5q31 and 5q22.1 for atopic dermatitis in Chinese Han population. *Asian Pac. J. Allergy Immunol.* **2018**, *35*, 196–202.
- Johansson, E.K.; Bergström, A.; Kull, I.; Lind, T.; Söderhäll, C.; van Hage, M.; Wickman, M.; Ballardini, N.; Wahlgren, C.-F. IgE sensitization in relation to preschool eczema and filaggrin mutation. *J. Allergy Clin. Immunol.* 2017, 140, 1572–1579. [CrossRef]

- Johansson, E.; Biagini Myers, J.M.; Martin, L.J.; He, H.; Pilipenko, V.; Mersha, T.; Weirauch, M.; Salomonis, N.; Ryan, P.; LeMasters, G.K.; et al. KIF3A genetic variation is associated with pediatric asthma in the presence of eczema independent of allergic rhinitis. *J. Allergy Clin. Immunol.* 2017, 140, 595–598. [CrossRef]
- 41. Kim, M.; Yoo, J.; Kim, J.; Park, J.; Han, E.; Jang, W.; Chae, H.; Lee, J.H.; Park, Y.M.; Kim, Y. Association of FLG single nucleotide variations with clinical phenotypes of atopic dermatitis. *PLoS ONE* **2017**, *12*, e0190077. [CrossRef]
- 42. Kim, J.S.; Choi, J.; Hahn, H.-J.; Lee, Y.-B.; Yu, D.-S.; Kim, J.-W. Association of Macrophage Migration Inhibitory Factor Polymorphisms with Total Plasma IgE Levels in Patients with Atopic Dermatitis in Korea. *PLoS ONE* **2016**, *11*, e0162477. [CrossRef]
- 43. Ko, E.J.; Heo, W.I.; Park, K.Y.; Lee, M.-K.; Seo, S.J. Genetic polymorphism of thymic stromal lymphopoietin in Korean patients with atopic dermatitis and allergic march. *J. Eur. Acad. Dermatol. Venereol.* **2018**, *32*, e468–e470. [CrossRef] [PubMed]
- 44. Leitch, C.S.; Natafji, E.; Yu, C.; Abdul-Ghaffar, S.; Madarasingha, N.; Venables, Z.C.; Chu, R.; Fitch, P.M.; Muinonen-Martin, A.J.; Campbell, L.E.; et al. Filaggrin-null mutations are associated with increased maturation markers on Langerhans cells. *J. Allergy Clin. Immunol.* **2016**, *138*, 482–490. [CrossRef] [PubMed]
- 45. Li, K.; Oh, W.J.; Park, K.Y.; Kim, K.-H.; Seo, S.J. FLG mutations in the East Asian atopic dermatitis patients: Genetic and clinical implication. *Exp. Dermatol.* **2016**, *25*, 816–818. [CrossRef] [PubMed]
- 46. Banihani, S.A.; Abu-Alia, K.F.; Khabour, O.F.; Alzoubi, K.H. Association between Resistin Gene Polymorphisms and Atopic Dermatitis. *Biomolecules* **2018**, *8*, 17. [CrossRef]
- Luukkonen, T.M.; Kiiski, V.; Ahola, M.; Mandelin, J.; Virtanen, H.; Poyhonen, M.; Kivirikko, S.; Surakka, I.; Reitamo, S.; Palotie, A.; et al. The Value of FLG Null Mutations in Predicting Treatment Response in Atopic Dermatitis: An Observational Study in Finnish Patients. *Acta Derm. Venereol.* 2017, *97*, 456–463. [CrossRef]
- 48. Manti, S.; Amorini, M.; Cuppari, C.; Salpietro, A.; Porcino, F.; Leonardi, S.; Del Giudice, M.M.; Marseglia, G.; Caimmi, D.P.; Salpietro, C. Filaggrin mutations and Molluscum contagiosum skin infection in patients with atopic dermatitis. *Ann. Allergy. Asthma Immunol.* **2017**, *119*, 446–451. [CrossRef]
- 49. Morizane, S.; Ouchida, M.; Sunagawa, K.; Sugimoto, S.; Kobashi, M.; Sugihara, S.; Nomura, H.; Tsuji, K.; Sato, A.; Miura, Y.; et al. Analysis of All 34 Exons of the SPINK5 Gene in Japanese Atopic Dermatitis Patients. *Acta Med. Okayama* **2018**, *72*, 275–282.
- 50. On, H.R.; Lee, S.E.; Kim, S.C.S.E.; Hong, W.J.; Kim, H.J.; Nomura, T.; Suzuki, S.; Shimizu, H.; Kim, S.C.S.E. Filaggrin Mutation in Korean Patients with Atopic Dermatitis. *Yonsei Med. J.* **2017**, *58*, 395–400. [CrossRef]
- 51. Park, K.Y.; Li, K.; Seok, J.; Seo, S.J. An Analysis of the Filaggrin Gene Polymorphism in Korean Atopic Dermatitis Patients. *J. Korean Med. Sci.* **2016**, *31*, 1136–1142. [CrossRef]
- 52. Ponińska, J.K.; Samoliński, B.; Tomaszewska, A.; Raciborski, F.; Samel-Kowalik, P.; Walkiewicz, A.; Lipiec, A.; Piekarska, B.; Krzych-Fałta, E.; Namysłowski, A.; et al. Haplotype dependent association of rs7927894 (11q13.5) with atopic dermatitis and chronic allergic rhinitis: A study in ECAP cohort. *PLoS ONE* 2017, 12, e0183922. [CrossRef] [PubMed]
- 53. Roekevisch, E.; Leeflang, M.M.G.; Schram, M.E.; Campbell, L.E.; Irwin McLean, W.H.; Kezic, S.; Bos, J.D.; Spuls, P.I.; Middelkamp-Hup, M.A. Patients with atopic dermatitis with filaggrin loss-of-function mutations show good but lower responses to immunosuppressive treatment. *Br. J. Dermatol.* **2017**, 177, 1745–1746. [CrossRef]
- 54. Sekiya, A.; Kono, M.; Tsujiuchi, H.; Kobayashi, T.; Nomura, T.; Kitakawa, M.; Suzuki, N.; Yamanaka, K.; Sueki, H.; McLean, W.H.I.; et al. Compound heterozygotes for filaggrin gene mutations do not always show severe atopic dermatitis. *J. Eur. Acad. Dermatol. Venereol.* **2017**, *31*, 158–162. [CrossRef]
- Tang, L.; Wang, J.; Zhu, J.; Liang, Y. Down-regulated SHARPIN may accelerate the development of atopic dermatitis through activating interleukin-33/ST2 signalling. *Exp. Dermatol.* 2018, 27, 1328–1335. [CrossRef] [PubMed]
- 56. Teye, K.; Numata, S.; Krol, R.P.; Ishii, N.; Matsuda, M.; Lee, J.-B.; Hamada, T.; Hashimoto, T. Prevalence of filaggrin gene mutations in patients with atopic dermatitis and ichthyosis vulgaris in Kyushu area of Japan and South Korea. *J. Dermatol. Sci.* **2017**, *86*, 174–177. [CrossRef]
- 57. Behniafard, N.; Amirzargar, A.A.; Gharagozlou, M.; Delavari, F.; Hosseinverdi, S.; Sotoudeh, S.; Farhadi, E.; Mahmoudi, M.; Khaledi, M.; Moghaddam, Z.G.; et al. Single nucleotide polymorphisms of the genes encoding IL-10 and TGF-β1 in Iranian children with atopic dermatitis. *Allergol. Immunopathol.* **2018**, 46, 155–159. [CrossRef] [PubMed]

- Thomsen, S.F.; Elmose, C.; Szecsi, P.B.; Stender, S.; Kyvik, K.O.; Backer, V.; Thyssen, J.P. Filaggrin gene loss-of-function mutations explain discordance of atopic dermatitis within dizygotic twin pairs. *Int. J. Dermatol.* 2016, 55, 1341–1344. [CrossRef]
- 59. Thorsteinsdottir, S.; Stokholm, J.; Thyssen, J.P.; Norgaard, S.; Thorsen, J.; Chawes, B.L.; Bonnelykke, K.; Waage, J.; Bisgaard, H. Genetic, Clinical, and Environmental Factors Associated with Persistent Atopic Dermatitis in Childhood. *JAMA Dermatol.* **2019**, *155*, 50–57. [CrossRef] [PubMed]
- 60. Trzeciak, M.; Sakowicz-Burkiewicz, M.; Wesserling, M.; Glen, J.; Dobaczewska, D.; Bandurski, T.; Nowicki, R.; Pawelczyk, T. Altered Expression of Genes Encoding Cornulin and Repetin in Atopic Dermatitis. *Int. Arch. Allergy Immunol.* **2017**, *172*, 11–19. [CrossRef] [PubMed]
- Tyurin, Y.A.; Shamsutdinov, A.F.; Kalinin, N.N.; Sharifullina, A.A.; Reshetnikova, I.D. Association of Toll-Like Cell Receptors TLR2 (p.Arg753GLN) and TLR4 (p.Asp299GLY) Polymorphisms with Indicators of General and Local Immunity in Patients with Atopic Dermatitis. *J. Immunol. Res.* 2017, 2017, 8493545. [CrossRef] [PubMed]
- 62. Wan, J.; Mitra, N.; Hoffstad, O.J.; Margolis, D.J. Influence of FLG mutations and TSLP polymorphisms on atopic dermatitis onset age. *Ann. Allergy Asthma Immunol.* **2017**, *118*, 737–738. [CrossRef] [PubMed]
- 63. Wen, H.-J.; Wang, S.-L.; Chen, P.-C.; Guo, Y.L. Prenatal perfluorooctanoic acid exposure and glutathione s-transferase T1/M1 genotypes and their association with atopic dermatitis at 2 years of age. *PLoS ONE* **2019**, *14*, e0210708. [CrossRef] [PubMed]
- 64. Yoon, N.Y.; Wang, H.Y.; Jun, M.; Jung, M.; Kim, D.H.; Lee, N.R.; Hong, K.-W.; Seo, S.J.; Choi, E.H.; Lee, J.; et al. Simultaneous detection of barrier- and immune-related gene variations in patients with atopic dermatitis by reverse blot hybridization assay. *Clin. Exp. Dermatol.* **2018**, *43*, 430–436. [CrossRef] [PubMed]
- Andersen, Y.M.F.; Egeberg, A.; Balslev, E.; Jørgensen, C.L.T.; Szecsi, P.B.; Stender, S.; Kaae, J.; Linneberg, A.; Gislason, G.; Skov, L.; et al. Filaggrin loss-of-function mutations, atopic dermatitis and risk of actinic keratosis: Results from two cross-sectional studies. *J. Eur. Acad. Dermatol. Venereol.* 2017, *31*, 1038–1043. [CrossRef]
- Asad, S.; Tapia-Páez, I.; Montano Montes, A.; Wahlgren, C.-F.; Bilcha, K.D.; Nordenskjöld, M.; Bradley, M. Evaluation of Single Nucleotide Variants in Ethiopian Patients with Atopic Dermatitis. *Acta Derm. Venereol.* 2019, 99, 101–102. [CrossRef]
- Song, Y.; Schwager, M.J.; Backer, V.; Guo, J.; Porsbjerg, C.; Khoo, S.-K.; Laing, I.A.; Moses, E.K.; LeSouëf, P.; Zhang, G. Environment Changes Genetic Effects on Respiratory Conditions and Allergic Phenotypes. *Sci. Rep.* 2017, 7, 6342. [CrossRef]
- Cai, X.-Y.; Zheng, X.-D.; Fang, L.; Zhou, F.-S.; Sheng, Y.-J.; Wu, Y.-Y.; Yu, C.-X.; Zhu, J.; Xiao, F.-L. A variant on chromosome 2p13.3 is associated with atopic dermatitis in Chinese Han population. *Gene* 2017, 628, 281–285. [CrossRef]
- Can, C.; Yazicioglu, M.; Gurkan, H.; Tozkir, H.; Gorgulu, A.; Sut, N.H. Lack of Association between Toll-like Receptor 2 Polymorphisms (R753Q and A-16934T) and Atopic Dermatitis in Children from Thrace Region of Turkey. *Balk. Med. J.* 2017, 34, 232–238. [CrossRef]
- 70. Chan, A.; Terry, W.; Zhang, H.; Karmaus, W.; Ewart, S.; Holloway, J.W.; Roberts, G.; Kurukulaaratchy, R.; Arshad, S.H. Filaggrin mutations increase allergic airway disease in childhood and adolescence through interactions with eczema and aeroallergen sensitization. *Clin. Exp. Allergy* **2018**, *48*, 147–155. [CrossRef]
- Chang, J.; Mitra, N.; Hoffstad, O.; Margolis, D.J. Association of Filaggrin Loss of Function and Thymic Stromal Lymphopoietin Variation with Treatment Use in Pediatric Atopic Dermatitis. *JAMA Dermatol.* 2017, 153, 275–281. [CrossRef] [PubMed]
- 72. Debinska, A.; Danielewicz, H.; Drabik-Chamerska, A.; Kalita, D.; Boznanski, A. Filaggrin loss-of-function mutations as a predictor for atopic eczema, allergic sensitization and eczema-associated asthma in Polish children population. *Adv. Clin. Exp. Med.* **2017**, *26*, 991–998. [CrossRef] [PubMed]
- 73. Elbert, N.J.; Duijts, L.; den Dekker, H.T.; Jaddoe, V.W.V.; Sonnenschein-van der Voort, A.M.M.; de Jongste, J.C.; Pasmans, S.G.M.A. Role of environmental exposures and filaggrin mutations on associations of ethnic origin with risk of childhood eczema. The Generation R Study. *Pediatr. Allergy Immunol.* 2016, 27, 627–635. [CrossRef] [PubMed]
- 74. Wu, Y.-Y.; Tang, J.-P.; Liu, Q.; Zheng, X.-D.; Fang, L.; Yin, X.-Y.; Jiang, X.-Y.; Zhou, F.-S.; Zhu, F.; Liang, B.; et al. Scanning indels in the 5q22.1 region and identification of the TMEM232 susceptibility gene that is associated with atopic dermatitis in the Chinese Han population. *Gene* **2017**, *617*, 17–23. [CrossRef] [PubMed]

- Ziyab, A.H.; Ewart, S.; Lockett, G.A.; Zhang, H.; Arshad, H.; Holloway, J.W.; Karmaus, W. Expression of the filaggrin gene in umbilical cord blood predicts eczema risk in infancy: A birth cohort study. *Clin. Exp. Allergy* 2017, 47, 1185–1192. [CrossRef] [PubMed]
- 76. Paternoster, L.; Savenije, O.E.M.; Heron, J.; Evans, D.M.; Vonk, J.M.; Brunekreef, B.; Wijga, A.H.; Henderson, A.J.; Koppelman, G.H.; Brown, S.J. Identification of atopic dermatitis subgroups in children from 2 longitudinal birth cohorts. *J. Allergy Clin. Immunol.* 2018, 141, 964–971. [CrossRef]
- 77. Al-Kzayer, L.F.Y.; Al-Aradi, H.M.H.; Shigemura, T.; Sano, K.; Tanaka, M.; Hamada, M.; Ali, K.H.; Aldaghir, O.M.; Nakazawa, Y.; Okuno, Y. DOCK8 mutation diagnosed using whole-exome sequencing of the dried blood spot-derived DNA: A case report of an Iraqi girl diagnosed in Japan. *BMC Med. Genet.* 2019, 20, 114. [CrossRef]
- 78. Dadi, H.; Jones, T.A.; Merico, D.; Sharfe, N.; Ovadia, A.; Schejter, Y.; Reid, B.; Sun, M.; Vong, L.; Atkinson, A.; et al. Combined immunodeficiency and atopy caused by a dominant negative mutation in caspase activation and recruitment domain family member 11 (CARD11). *J. Allergy Clin. Immunol.* 2018, 141, 1818–1830. [CrossRef]
- Ma, C.A.; Stinson, J.R.; Zhang, Y.; Abbott, J.K.; Weinreich, M.A.; Hauk, P.J.; Reynolds, P.R.; Lyons, J.J.; Nelson, C.G.; Ruffo, E.; et al. Germline hypomorphic CARD11 mutations in severe atopic disease. *Nat. Genet.* 2017, 49, 1192–1201. [CrossRef]
- Peled, A.; Sarig, O.; Sun, G.; Samuelov, L.; Ma, C.A.; Zhang, Y.; Dimaggio, T.; Nelson, C.G.; Stone, K.D.; Freeman, A.F.; et al. Loss-of-function mutations in caspase recruitment domain-containing protein 14 (CARD14) are associated with a severe variant of atopic dermatitis. *J. Allergy Clin. Immunol.* 2019, 143, 173–181. [CrossRef]
- Pigors, M.; Common, J.E.A.; Wong, X.F.C.C.; Malik, S.; Scott, C.A.; Tabarra, N.; Liany, H.; Liu, J.; Limviphuvadh, V.; Maurer-Stroh, S.; et al. Exome Sequencing and Rare Variant Analysis Reveals Multiple Filaggrin Mutations in Bangladeshi Families with Atopic Eczema and Additional Risk Genes. *J. Investig. Dermatol.* 2018, 138, 2674–2677. [CrossRef] [PubMed]
- Suzuki, H.; Makino, Y.; Nagata, M.; Furuta, J.; Enomoto, H.; Hirota, T.; Tamari, M.; Noguchi, E. A rare variant in CYP27A1 and its association with atopic dermatitis with high serum total IgE. *Allergy* 2016, *71*, 1486–1489. [CrossRef]
- Heo, W.I.L.; Park, K.Y.; Jin, T.; Lee, M.-K.; Kim, M.; Choi, E.H.; Kim, H.-S.; Bae, J.M.; Moon, N.J.; Seo, S.J. Identification of novel candidate variants including COL6A6 polymorphisms in early-onset atopic dermatitis using whole-exome sequencing. *BMC Med. Genet.* 2017, *18*, 8. [CrossRef] [PubMed]
- Karaca, S.; Civelek, E.; Karaca, M.; Sahiner, U.M.; Ozgul, R.K.; Kocabas, C.N.; Polimanti, R.; Sekerel, B.E. Allergy-specific Phenome-Wide Association Study for Immunogenes in Turkish Children. *Sci. Rep.* 2016, *6*, 33152. [CrossRef] [PubMed]
- 85. Margolis, D.J.; Mitra, N.; Gochnauer, H.; Wubbenhorst, B.; D'Andrea, K.; Kraya, A.; Hoffstad, O.; Gupta, J.; Kim, B.; Yan, A.; et al. Uncommon Filaggrin Variants Are Associated with Persistent Atopic Dermatitis in African Americans. *J. Investig. Dermatol.* **2018**, *138*, 1501–1506. [CrossRef]
- 86. Manz, J.; Rodriguez, E.; ElSharawy, A.; Oesau, E.-M.; Petersen, B.-S.; Baurecht, H.; Mayr, G.; Weber, S.; Harder, J.; Reischl, E.; et al. Targeted Resequencing and Functional Testing Identifies Low-Frequency Missense Variants in the Gene Encoding GARP as Significant Contributors to Atopic Dermatitis Risk. J. Investig. Dermatol. 2016, 136, 2380–2386. [CrossRef]
- 87. Park, K.Y.; Park, M.K.; Seok, J.; Li, K.; Seo, S.J. Clinical characteristics of Korean patients with filaggrin-related atopic dermatitis. *Clin. Exp. Dermatol.* **2016**, *41*, 595–600. [CrossRef]
- Wong, X.F.C.C.C.; Denil, S.L.I.J.I.J.; Foo, J.N.; Chen, H.; Tay, A.S.L.; Haines, R.L.; Tang, M.B.Y.Y.; McLean, W.H.I.I.; Sandilands, A.; Smith, F.J.D.D.; et al. Array-based sequencing of filaggrin gene for comprehensive detection of disease-associated variants. *J. Allergy Clin. Immunol.* 2018, 141, 814–816. [CrossRef]
- 89. Lopez-Alvarez, M.R.; Jiang, W.; Jones, D.C.; Jayaraman, J.; Johnson, C.; Cookson, W.O.; Moffatt, M.F.; Trowsdale, J.; Traherne, J.A. LILRA6 copy number variation correlates with susceptibility to atopic dermatitis. *Immunogenetics* **2016**, *68*, 743–747. [CrossRef]
- 90. Mathyer, M.E.; Quiggle, A.M.; Wong, X.F.C.C.; Denil, S.L.I.J.; Kumar, M.G.; Ciliberto, H.M.; Bayliss, S.J.; Common, J.E.; de Guzman Strong, C. Tiled array-based sequencing identifies enrichment of loss-of-function variants in the highly homologous filaggrin gene in African-American children with severe atopic dermatitis. *Exp. Dermatol.* 2018, 27, 989–992. [CrossRef]

- Elhaji, Y.; Sasseville, D.; Pratt, M.; Asai, Y.; Matheson, K.; McLean, W.H.I.; Hull, P.R. Filaggrin gene loss-of-function mutations constitute a factor in patients with multiple contact allergies. *Contact Dermat.* 2019, *80*, 354–358. [CrossRef] [PubMed]
- Liang, J.; Liu, Y.; Xue, R.; Chen, L.; Chen, H.; Shao, L.; Wang, J.; Zhang, X. Interleukin 4 -590C/T (rs2243250) Polymorphism Is Associated with Increased Risk of Atopic Dermatitis: Meta-Analysis of Case-Control Studies. *Dermat. Contact Atopic Occup. Drug* 2017, 28, 144–151. [CrossRef] [PubMed]
- Margaritte-Jeannin, P.; Babron, M.-C.; Laprise, C.; Lavielle, N.; Sarnowski, C.; Brossard, M.; Moffatt, M.; Gagne-Ouellet, V.; Etcheto, A.; Lathrop, M.; et al. The COL5A3 and MMP9 genes interact in eczema susceptibility. *Clin. Exp. Allergy* 2018, *48*, 297–305. [CrossRef] [PubMed]
- 94. Qi, Y.; Kong, J.; He, J. Genetic relationship between IL-10 gene polymorphisms and the risk of clinical atopic dermatitis. *BMC Med. Genet.* **2019**, *20*, 83. [CrossRef] [PubMed]
- 95. Zhang, Y.; Wang, H.-C.; Feng, C.; Yan, M. Analysis of the Association of Polymorphisms rs5743708 in TLR2 and rs4986790 in TLR4 with Atopic Dermatitis Risk. *Immunol. Investig.* **2019**, *48*, 169–180. [CrossRef] [PubMed]
- Zhao, J.; Chen, Z.-Y.; Li, L.-F. Association between the IL-10-1082G/A, IL-10-592A/C, and IL-10-819G/A Polymorphisms and Atopic Dermatitis Susceptibility: A Meta-Analysis. *Genet. Test. Mol. Biomark.* 2019, 23, 332–341. [CrossRef]
- 97. Arkwright, P.D.; Chase, J.M.; Babbage, S.; Pravica, V.; David, T.J.; Hutchinson, I.V. Atopic dermatitis is associated with a low-producer transforming growth factor beta(1) cytokine genotype. *J. Allergy Clin. Immunol.* **2001**, *108*, 281–284. [CrossRef]
- Trzeciak, M.; Sakowicz-Burkiewicz, M.; Wesserling, M.; Dobaczewska, D.; Glen, J.; Nowicki, R.; Pawelczyk, T. Expression of Cornified Envelope Proteins in Skin and Its Relationship with Atopic Dermatitis Phenotype. *Acta Derm. Venereol.* 2017, 97, 36–41. [CrossRef]
- 99. Polcari, I.; Becker, L.; Stein, S.L.; Smith, M.S.; Paller, A.S. Filaggrin gene mutations in African Americans with both ichthyosis vulgaris and atopic dermatitis. *Pediatr. Dermatol.* **2014**, *31*, 489–492. [CrossRef] [PubMed]
- Rebane, A. microRNA and Allergy. In *Advances in Experimental Medicine and Biology*; Springer: Cham, Switzerland, 2015; Volume 888, pp. 331–352. ISBN 9783319226705.
- Dissanayake, E.; Inoue, Y. MicroRNAs in Allergic Disease. Curr. Allergy Asthma Rep. 2016, 16, 67. [CrossRef]
 [PubMed]
- 102. Alaskhar Alhamwe, B.; Khalaila, R.; Wolf, J.; von Bülow, V.; Harb, H.; Alhamdan, F.; Hii, C.S.; Prescott, S.L.; Ferrante, A.; Renz, H.; et al. Histone modifications and their role in epigenetics of atopy and allergic diseases. *Allergy Asthma Clin. Immunol.* 2018, 14, 39. [CrossRef] [PubMed]
- Kamen, L.A.; Schlessinger, J.; Lowell, C.A. Pyk2 Is Required for Neutrophil Degranulation and Host Defense Responses to Bacterial Infection. *J. Immunol.* 2011, 186, 1656–1665. [CrossRef] [PubMed]
- 104. Yan, S.R.; Novak, M.J. Beta2 integrin-dependent phosphorylation of protein-tyrosine kinase Pyk2 stimulated by tumor necrosis factor alpha and fMLP in human neutrophils adherent to fibrinogen. *FEBS Lett.* **1999**, 451, 33–38. [CrossRef]
- 105. Calabresi, P.A.; Allie, R.; Mullen, K.M.; Yun, S.H.; Georgantas, R.W.; Whartenby, K.A. Kinetics of CCR7 expression differ between primary activation and effector memory states of T(H)1 and T(H)2 cells. *J. Neuroimmunol.* 2003, 139, 58–65. [CrossRef]
- 106. Malaisse, J.; Bourguignon, V.; De Vuyst, E.; Lambert de Rouvroit, C.; Nikkels, A.F.; Flamion, B.; Poumay, Y. Hyaluronan metabolism in human keratinocytes and atopic dermatitis skin is driven by a balance of hyaluronan synthases 1 and 3. *J. Investig. Dermatol.* 2014, 134, 2174–2182. [CrossRef] [PubMed]
- Quinn, S.R.; O'Neill, L.A. A trio of microRNAs that control Toll-like receptor signalling. *Int. Immunol.* 2011, 23, 421–425. [CrossRef]
- 108. Sonkoly, E.; Ståhle, M.; Pivarcsi, A. MicroRNAs and immunity: Novel players in the regulation of normal immune function and inflammation. *Semin. Cancer Biol.* **2008**, *18*, 131–140. [CrossRef]
- 109. Renz, H.; Autenrieth, I.B.; Brandtzæg, P.; Cookson, W.O.; Holgate, S.; von Mutius, E.; Valenta, R.; Haller, D. Gene-environment interaction in chronic disease: A European Science Foundation Forward Look. J. Allergy Clin. Immunol. 2011, 128, S27–S49. [CrossRef]
- 110. Von Mutius, E. Gene-environment interactions in asthma. J. Allergy Clin. Immunol. 2009, 123, 3–11. [CrossRef]
- 111. Potaczek, D.P.; Harb, H.; Michel, S.; Alhamwe, B.A.; Renz, H.; Tost, J. Epigenetics and allergy: From basic mechanisms to clinical applications. *Epigenomics* **2017**, *9*, 539–571. [CrossRef]

- Candi, E.; Schmidt, R.; Melino, G. The cornified envelope: A model of cell death in the skin. *Nat. Rev. Mol. Cell Biol.* 2005, *6*, 328–340. [CrossRef] [PubMed]
- 113. Irvine, A.D.; McLean, W.H.H.I.; Leung, D.Y.M.M. Filaggrin mutations associated with skin and allergic diseases. *N. Engl. J. Med.* **2011**, *365*, 1315–1327. [CrossRef] [PubMed]
- 114. Kypriotou, M.; Huber, M.; Hohl, D. The human epidermal differentiation complex: Cornified envelope precursors, S100 proteins and the "fused genes" family. *Exp. Dermatol.* 2012, 21, 643–649. [CrossRef] [PubMed]
- Brown, S.J.; McLean, W.H.I. One remarkable molecule: Filaggrin. J. Investig. Dermatol. 2012, 132, 751–762.
 [CrossRef]
- 116. Margolis, D.J.; Apter, A.J.; Gupta, J.; Hoffstad, O.; Papadopoulos, M.; Campbell, L.E.; Sandilands, A.; McLean, W.H.I.; Rebbeck, T.R.; Mitra, N. The persistence of atopic dermatitis and filaggrin (FLG) mutations in a US longitudinal cohort. *J. Allergy Clin. Immunol.* **2012**, *130*, 912–917. [CrossRef]
- 117. Palmer, C.N.A.; Irvine, A.D.; Terron-Kwiatkowski, A.; Zhao, Y.; Liao, H.; Lee, S.P.; Goudie, D.R.; Sandilands, A.; Campbell, L.E.; Smith, F.J.D.; et al. Common loss-of-function variants of the epidermal barrier protein filaggrin are a major predisposing factor for atopic dermatitis. *Nat. Genet.* 2006, *38*, 441–446. [CrossRef]
- 118. Rogers, A.J.; Celedón, J.C.; Lasky-Su, J.A.; Weiss, S.T.; Raby, B.A. Filaggrin mutations confer susceptibility to atopic dermatitis but not to asthma. *J. Allergy Clin. Immunol.* **2007**, *120*, 1332–1337. [CrossRef]
- 119. Smith, F.J.D.; Irvine, A.D.; Terron-Kwiatkowski, A.; Sandilands, A.; Campbell, L.E.; Zhao, Y.; Liao, H.; Evans, A.T.; Goudie, D.R.; Lewis-Jones, S.; et al. Loss-of-function mutations in the gene encoding filaggrin cause ichthyosis vulgaris. *Nat. Genet.* **2006**, *38*, 337–342. [CrossRef]
- Rodríguez, E.; Baurecht, H.; Herberich, E.; Wagenpfeil, S.; Brown, S.J.; Cordell, H.J.; Irvine, A.D.; Weidinger, S. Meta-analysis of filaggrin polymorphisms in eczema and asthma: Robust risk factors in atopic disease. *J. Allergy Clin. Immunol.* 2009, 123, 1361–1370. [CrossRef]
- 121. Van Den Oord, R.A.H.M.; Sheikh, A. Filaggrin gene defects and risk of developing allergic sensitisation and allergic disorders: Systematic review and meta-analysis. *BMJ* **2009**, *339*, 86–88. [CrossRef]
- 122. Marenholz, I.; Nickel, R.; Rüschendorf, F.; Schulz, F.; Esparza-Gordillo, J.; Kerscher, T.; Grüber, C.; Lau, S.; Worm, M.; Keil, T.; et al. Filaggrin loss-of-function mutations predispose to phenotypes involved in the atopic march. *J. Allergy Clin. Immunol.* **2006**, *118*, 866–871. [CrossRef] [PubMed]
- 123. Bønnelykke, K.; Pipper, C.B.; Tavendale, R.; Palmer, C.N.A.; Bisgaard, H. Filaggrin gene variants and atopic diseases in early childhood assessed longitudinally from birth. *Pediatr. Allergy Immunol.* 2010, 21, 954–961. [CrossRef]
- 124. Weidinger, S.; O'Sullivan, M.; Illig, T.; Baurecht, H.; Depner, M.; Rodriguez, E.; Ruether, A.; Klopp, N.; Vogelberg, C.; Weiland, S.K.; et al. Filaggrin mutations, atopic eczema, hay fever, and asthma in children. J. Allergy Clin. Immunol. 2008, 121, 1203–1210. [CrossRef] [PubMed]
- Weidinger, S.; Rodríguez, E.; Stahl, C.; Wagenpfeil, S.; Klopp, N.; Illig, T.; Novak, N. Filaggrin mutations strongly predispose to early-onset and extrinsic atopic dermatitis. *J. Investig. Dermatol.* 2007, 127, 724–726. [CrossRef] [PubMed]
- 126. Henderson, J.; Northstone, K.; Lee, S.P.; Liao, H.; Zhao, Y.; Pembrey, M.; Mukhopadhyay, S.; Smith, G.D.; Palmer, C.N.A.; McLean, W.H.I.; et al. The burden of disease associated with filaggrin mutations: A population-based, longitudinal birth cohort study. *J. Allergy Clin. Immunol.* 2008, 121, 872–877. [CrossRef] [PubMed]
- 127. Marenholz, I.; Kerscher, T.; Bauerfeind, A.; Esparza-Gordillo, J.; Nickel, R.; Keil, T.; Lau, S.; Rohde, K.; Wahn, U.; Lee, Y.A. An interaction between filaggrin mutations and early food sensitization improves the prediction of childhood asthma. *J. Allergy Clin. Immunol.* 2009, *123*, 911–916. [CrossRef]
- Elias, P.M. Therapeutic Implications of a Barrier-based Pathogenesis of Atopic Dermatitis. *Ann. Dermatol.* 2010, 22, 245–254. [CrossRef]
- 129. Loo, E.X.L.; Shek, L.P.; Goh, A.; Teoh, O.H.; Chan, Y.H.; Soh, S.E.; Saw, S.M.; Kwek, K.; Gluckman, P.D.; Godfrey, K.M.; et al. Atopic Dermatitis in Early Life: Evidence for at Least Three Phenotypes? Results from the GUSTO Study. *Int. Arch. Allergy Immunol.* 2015, *166*, 273–279. [CrossRef]
- 130. Greisenegger, E.; Novak, N.; Maintz, L.; Bieber, T.; Zimprich, F.; Haubenberger, D.; Gleiss, A.; Stingl, G.; Kopp, T.; Zimprich, A. Analysis of four prevalent filaggrin mutations (R501X, 2282del4, R2447X and S3247X) in Austrian and German patients with atopic dermatitis. *J. Eur. Acad. Dermatol. Venereol.* 2010, 24, 607–610. [CrossRef]

- 131. Rupnik, H.; Rijavec, M.; Korošec, P. Filaggrin loss-of-function mutations are not associated with atopic dermatitis that develops in late childhood or adulthood. *Br. J. Dermatol.* **2015**, *172*, 455–461. [CrossRef]
- 132. Flohr, C.; Johansson, S.G.O.; Wahlgren, C.-F.; Williams, H. How atopic is atopic dermatitis? *J. Allergy Clin. Immunol.* **2004**, *114*, 150–158. [CrossRef] [PubMed]
- 133. Brown, S.J.; Sandilands, A.; Zhao, Y.; Liao, H.; Relton, C.L.; Meggitt, S.J.; Trembath, R.C.; Barker, J.N.W.N.; Reynolds, N.J.; Cordell, H.J.; et al. Prevalent and Low-Frequency Null Mutations in the Filaggrin Gene Are Associated with Early-Onset and Persistent Atopic Eczema. *J. Investig. Dermatol.* 2008, 128, 1591–1594. [CrossRef] [PubMed]
- 134. Sandilands, A.; Terron-Kwiatkowski, A.; Hull, P.R.; O'Regan, G.M.; Clayton, T.H.; Watson, R.M.; Carrick, T.; Evans, A.T.; Liao, H.; Zhao, Y.; et al. Comprehensive analysis of the gene encoding filaggrin uncovers prevalent and rare mutations in ichthyosis vulgaris and atopic eczema. *Nat. Genet.* 2007, *39*, 650–654. [CrossRef] [PubMed]
- 135. Barker, J.N.W.N.; Palmer, C.N.A.; Zhao, Y.; Liao, H.; Hull, P.R.; Lee, S.P.; Allen, M.H.; Meggitt, S.J.; Reynolds, N.J.; Trembath, R.C.; et al. Null mutations in the filaggrin gene (FLG) determine major susceptibility to early-onset atopic dermatitis that persists into adulthood. *J. Investig. Dermatol.* 2007, 127, 564–567. [CrossRef] [PubMed]
- Stemmler, S.; Parwez, Q.; Petrasch-Parwez, E.; Epplen, J.T.; Hoffjan, S. Two common loss-of-function mutations within the filaggrin gene predispose for early onset of atopic dermatitis. *J. Investig. Dermatol.* 2007, 127, 722–724. [CrossRef] [PubMed]
- 137. Ercan, H.; Ispir, T.; Kirac, D.; Baris, S.; Ozen, A.; Oztezcan, S.; Cengizlier, M.R. Predictors of atopic dermatitis phenotypes and severity: Roles of serum immunoglobulins and filaggrin gene mutation R501X. *Allergol. Immunopathol.* (*Madr*) 2013, 41, 86–93. [CrossRef]
- Weidinger, S.; Illig, T.; Baurecht, H.; Irvine, A.D.; Rodriguez, E.; Diaz-Lacava, A.; Klopp, N.; Wagenpfeil, S.; Zhao, Y.; Liao, H.; et al. Loss-of-function variations within the filaggrin gene predispose for atopic dermatitis with allergic sensitizations. J. Allergy Clin. Immunol. 2006, 118, 214–219. [CrossRef]
- Brunner, P.M.; Guttman-Yassky, E. Racial differences in atopic dermatitis. *Ann. Allergy Asthma Immunol.* 2019, 122, 449–455. [CrossRef]
- Kaufman, B.P.; Guttman-Yassky, E.; Alexis, A.F. Atopic dermatitis in diverse racial and ethnic groups-Variations in epidemiology, genetics, clinical presentation and treatment. *Exp. Dermatol.* 2018, 27, 340–357. [CrossRef]
- 141. Margolis, D.J.; Gupta, J.; Apter, A.J.; Hoffstad, O.; Papadopoulos, M.; Rebbeck, T.R.; Wubbenhorst, B.; Mitra, N. Exome sequencing of filaggrin and related genes in African-American children with atopic dermatitis. *J. Investig. Dermatol.* 2014, 134, 2272–2274. [CrossRef]
- 142. Shaw, T.E.; Currie, G.P.; Koudelka, C.W.; Simpson, E.L. Eczema Prevalence in the United States: Data from the 2003 National Survey of Children's Health. *J. Investig. Dermatol.* **2011**, *131*, 67–73. [CrossRef] [PubMed]
- 143. Stefanovic, N.; Flohr, C.; Irvine, A.D. The exposome in atopic dermatitis. *Allergy* **2020**, *75*, 63–74. [CrossRef] [PubMed]
- 144. Zhang, Q.; Si, N.; Liu, Y.; Zhang, D.; Wang, R.; Zhang, Y.; Wang, S.; Liu, X.; Deng, X.; Ma, Y.; et al. Steroid sulfatase and filaggrin mutations in a boy with severe ichthyosis, elevated serum IgE level and moyamoya syndrome. *Gene* **2017**, *628*, 103–108. [CrossRef] [PubMed]
- 145. Imamura, Y.; Scott, I.C.; Greenspan, D.S. The pro-alpha3(V) collagen chain. Complete primary structure, expression domains in adult and developing tissues, and comparison to the structures and expression domains of the other types V and XI procollagen chains. *J. Biol. Chem.* 2000, 275, 8749–8759. [CrossRef] [PubMed]
- 146. Fitzgerald, J.; Holden, P.; Hansen, U. The expanded collagen VI family: New chains and new questions. *Connect. Tissue Res.* **2013**, *54*, 345–350. [CrossRef] [PubMed]
- 147. LeBert, D.C.; Squirrell, J.M.; Rindy, J.; Broadbridge, E.; Lui, Y.; Zakrzewska, A.; Eliceiri, K.W.; Meijer, A.H.; Huttenlocher, A. Matrix metalloproteinase 9 modulates collagen matrices and wound repair. *Development* 2015, 142, 2136–2146. [CrossRef]
- 148. Soldi, R.; Mitola, S.; Strasly, M.; Defilippi, P.; Tarone, G.; Bussolino, F. Role of alphavbeta3 integrin in the activation of vascular endothelial growth factor receptor-2. *EMBO J.* **1999**, *18*, 882–892. [CrossRef]

- Maegdefessel, L.; Azuma, J.; Toh, R.; Merk, D.R.; Deng, A.; Chin, J.T.; Raaz, U.; Schoelmerich, A.M.; Raiesdana, A.; Leeper, N.J.; et al. Inhibition of microRNA-29b reduces murine abdominal aortic aneurysm development. J. Clin. Investig. 2012, 122, 497–506. [CrossRef]
- 150. Salazar, K.D.; Lankford, S.M.; Brody, A.R. Mesenchymal stem cells produce Wnt isoforms and TGF-beta1 that mediate proliferation and procollagen expression by lung fibroblasts. *Am. J. Physiol. Lung Cell. Mol. Physiol.* **2009**, 297, L1002–L1011. [CrossRef]
- 151. Rørvig, S.; Østergaard, O.; Heegaard, N.H.H.; Borregaard, N. Proteome profiling of human neutrophil granule subsets, secretory vesicles, and cell membrane: Correlation with transcriptome profiling of neutrophil precursors. *J. Leukoc. Biol.* **2013**, *94*, 711–721. [CrossRef]
- 152. Foley, S.C.; Mogas, A.K.; Olivenstein, R.; Fiset, P.O.; Chakir, J.; Bourbeau, J.; Ernst, P.; Lemière, C.; Martin, J.G.; Hamid, Q. Increased expression of ADAM33 and ADAM8 with disease progression in asthma. *J. Allergy Clin. Immunol.* 2007, 119, 863–871. [CrossRef] [PubMed]
- 153. Huls, A.; Klumper, C.; MacIntyre, E.A.; Brauer, M.; Melen, E.; Bauer, M.; Berdel, D.; Bergstrom, A.; Brunekreef, B.; Chan-Yeung, M.; et al. Atopic dermatitis: Interaction between genetic variants of GSTP1, TNF, TLR2, and TLR4 and air pollution in early life. *Pediatr. Allergy Immunol.* 2018, 29, 596–605. [CrossRef] [PubMed]
- 154. Nakashima, K.; Hirota, T.; Obara, K.; Shimizu, M.; Jodo, A.; Kameda, M.; Doi, S.; Fujita, K.; Shirakawa, T.; Enomoto, T.; et al. An association study of asthma and related phenotypes with polymorphisms in negative regulator molecules of the TLR signaling pathway. *J. Hum. Genet.* 2006, *51*, 284–291. [CrossRef] [PubMed]
- 155. Kormann, M.S.D.; Depner, M.; Hartl, D.; Klopp, N.; Illig, T.; Adamski, J.; Vogelberg, C.; Weiland, S.K.; von Mutius, E.; Kabesch, M. Toll-like receptor heterodimer variants protect from childhood asthma. *J. Allergy Clin. Immunol.* **2008**, *122*, 86–92. [CrossRef] [PubMed]
- 156. Oh, D.-Y.; Schumann, R.R.; Hamann, L.; Neumann, K.; Worm, M.; Heine, G. Association of the toll-like receptor 2 A-16934T promoter polymorphism with severe atopic dermatitis. *Allergy* 2009, 64, 1608–1615. [CrossRef]
- 157. Potaczek, D.; Nastalek, M.; Okumura, K.; Wojas-Pelc, A.; Undas, A.; Nishiyama, C. An association of TLR2-16934A > T polymorphism and severity/phenotype of atopic dermatitis. *J. Eur. Acad. Dermatol. Venereol.* 2011, 25, 715–721. [CrossRef]
- 158. Li, H.-F.; Yan, L.-P.; Wang, K.; Li, X.-T.; Liu, H.-X.; Tan, W. Association between ADAM33 polymorphisms and asthma risk: A systematic review and meta-analysis. *Respir. Res.* **2019**, *20*, 38. [CrossRef]
- 159. Li, Z.; Yan, F.; Yang, Z.; Zhou, J.; Chen, Y.; Ding, Z. Association between ADAM33 S2 and V4 polymorphisms and susceptibility to allergic rhinitis: A meta-analysis. *Allergol. Immunopathol.* **2016**, *44*, 170–176. [CrossRef]
- Bedsaul, J.R.; Carter, N.M.; Deibel, K.E.; Hutcherson, S.M.; Jones, T.A.; Wang, Z.; Yang, C.; Yang, Y.-K.; Pomerantz, J.L. Mechanisms of Regulated and Dysregulated CARD11 Signaling in Adaptive Immunity and Disease. *Front. Immunol.* 2018, *9*, 2105. [CrossRef]
- 161. Yang, D.; Chertov, O.; Bykovskaia, S.N.; Chen, Q.; Buffo, M.J.; Shogan, J.; Anderson, M.; Schröder, J.M.; Wang, J.M.; Howard, O.M.; et al. Beta-defensins: Linking innate and adaptive immunity through dendritic and T cell CCR6. *Science* 1999, 286, 525–528. [CrossRef]
- 162. Jurevic, R.J.; Bai, M.; Chadwick, R.B.; White, T.C.; Dale, B.A. Single-nucleotide polymorphisms (SNPs) in human beta-defensin 1: High-throughput SNP assays and association with Candida carriage in type I diabetics and nondiabetic controls. *J. Clin. Microbiol.* 2003, 41, 90–96. [CrossRef] [PubMed]
- 163. Bertin, J.; Wang, L.; Guo, Y.; Jacobson, M.D.; Poyet, J.L.; Srinivasula, S.M.; Merriam, S.; DiStefano, P.S.; Alnemri, E.S. CARD11 and CARD14 are novel caspase recruitment domain (CARD)/membrane-associated guanylate kinase (MAGUK) family members that interact with BCL10 and activate NF-kappa B. *J. Biol. Chem.* 2001, 276, 11877–11882. [CrossRef]
- 164. Guo, X.; Zhang, Y.; Wang, P.; Li, T.; Fu, W.; Mo, X.; Shi, T.; Zhang, Z.; Chen, Y.; Ma, D.; et al. VSTM1-v2, a novel soluble glycoprotein, promotes the differentiation and activation of Th17 cells. *Cell. Immunol.* 2012, 278, 136–142. [CrossRef] [PubMed]
- Hirayasu, K.; Arase, H. Leukocyte Immunoglobulin-Like Receptor (LILR). In *Encyclopedia of Signaling Molecules*; Springer International Publishing: Cham, Switzerland, 2018; pp. 2854–2861.
- 166. Su, H.C.; Jing, H.; Angelus, P.; Freeman, A.F. Insights into immunity from clinical and basic science studies of DOCK8 immunodeficiency syndrome. *Immunol. Rev.* **2019**, *287*, 9–19. [CrossRef] [PubMed]

- 167. Minkiewicz, J.; de Rivero Vaccari, J.P.; Keane, R.W. Human astrocytes express a novel NLRP2 inflammasome. *Glia* **2013**, *61*, 1113–1121. [CrossRef]
- 168. Tschopp, J.; Martinon, F.; Burns, K. NALPs: A novel protein family involved in inflammation. *Nat. Rev. Mol. Cell Biol.* 2003, *4*, 95–104. [CrossRef]
- 169. Walne, A.J.; Vulliamy, T.; Kirwan, M.; Plagnol, V.; Dokal, I. Constitutional mutations in RTEL1 cause severe dyskeratosis congenita. *Am. J. Hum. Genet.* **2013**, *92*, 448–453. [CrossRef]
- 170. Gray, P.W.; Aggarwal, B.B.; Benton, C.V.; Bringman, T.S.; Henzel, W.J.; Jarrett, J.A.; Leung, D.W.; Moffat, B.; Ng, P.; Svedersky, L.P. Cloning and expression of cDNA for human lymphotoxin, a lymphokine with tumour necrosis activity. *Nature* **1984**, *312*, 721–724. [CrossRef]
- 171. Trzeciak, M.; Wesserling, M.; Bandurski, T.; Glen, J.; Nowicki, R.; Pawelczyk, T. Association of a Single Nucleotide Polymorphism in a Late Cornified Envelope-like Proline-rich 1 Gene (LELP1) with Atopic Dermatitis. *Acta Derm. Venereol.* **2016**, *96*, 459–463. [CrossRef]
- 172. Knüppel, S.; Esparza-Gordillo, J.; Marenholz, I.; Holzhütter, H.-G.; Bauerfeind, A.; Ruether, A.; Weidinger, S.; Lee, Y.-A.; Rohde, K. Multi-locus stepwise regression: A haplotype-based algorithm for finding genetic associations applied to atopic dermatitis. *BMC Med. Genet.* **2012**, *13*, 8. [CrossRef]
- 173. Margolis, D.J.; Gupta, J.; Apter, A.J.; Ganguly, T.; Hoffstad, O.; Papadopoulos, M.; Rebbeck, T.R.; Mitra, N. Filaggrin-2 variation is associated with more persistent atopic dermatitis in African American subjects. *J. Allergy Clin. Immunol.* **2014**, 133, 784–789. [CrossRef] [PubMed]
- 174. Li, C.; Xiao, L.; Jia, J.; Li, F.; Wang, X.; Duan, Q.; Jing, H.; Yang, P.; Chen, C.; Wang, Q.; et al. Cornulin Is Induced in Psoriasis Lesions and Promotes Keratinocyte Proliferation via Phosphoinositide 3-Kinase/Akt Pathways. J. Investig. Dermatol. 2019, 139, 71–80. [CrossRef] [PubMed]
- Turksen, K.; Troy, T.-C. Permeability barrier dysfunction in transgenic mice overexpressing claudin 6. Development 2002, 129, 1775–1784. [PubMed]
- 176. Yamakoshi, T.; Makino, T.; Ur Rehman, M.; Yoshihisa, Y.; Sugimori, M.; Shimizu, T. Trichohyalin-like 1 protein, a member of fused S100 proteins, is expressed in normal and pathologic human skin. *Biochem. Biophys. Res. Commun.* **2013**, 432, 66–72. [CrossRef] [PubMed]
- 177. Kantor, R.; Silverberg, J.I. Environmental risk factors and their role in the management of atopic dermatitis. *Expert Rev. Clin. Immunol.* **2017**, *13*, 15–26. [CrossRef]
- Isidoro-García, M.; Dávila-González, I.; Pascual de Pedro, M.; Sanz-Lozano, C.; Lorente-Toledano, F. Interactions between genes and the environment. Epigenetics in allergy. *Allergol. Immunopathol.* 2007, 35, 254–258. [CrossRef]
- Makeyev, E.V.; Maniatis, T. Multilevel Regulation of Gene Expression by MicroRNAs. Science 2008, 319, 1789–1790. [CrossRef] [PubMed]
- Krol, J.; Loedige, I.; Filipowicz, W. The widespread regulation of microRNA biogenesis, function and decay. *Nat. Rev. Genet.* 2010, *11*, 597–610. [CrossRef] [PubMed]
- 181. Maes, T.; Cobos, F.A.; Schleich, F.; Sorbello, V.; Henket, M.; De Preter, K.; Bracke, K.R.; Conickx, G.; Mesnil, C.; Vandesompele, J.; et al. Asthma inflammatory phenotypes show differential microRNA expression in sputum. J. Allergy Clin. Immunol. 2016, 137, 1433–1446. [CrossRef]
- 182. Sinha, A.; Yadav, A.K.; Chakraborty, S.; Kabra, S.K.; Lodha, R.; Kumar, M.; Kulshreshtha, A.; Sethi, T.; Pandey, R.; Malik, G.; et al. Exosome-enclosed microRNAs in exhaled breath hold potential for biomarker discovery in patients with pulmonary diseases. *J. Allergy Clin. Immunol.* 2013, 132, 219–222. [CrossRef]
- Bauer, S.M. Atopic Eczema: Genetic Associations and Potential Links to Developmental Exposures. *Int. J. Toxicol.* 2017, 36, 187–198. [CrossRef] [PubMed]
- 184. Rebane, A.; Runnel, T.; Aab, A.; Maslovskaja, J.; Rückert, B.; Zimmermann, M.; Plaas, M.; Kärner, J.; Treis, A.; Pihlap, M.; et al. MicroRNA-146a alleviates chronic skin inflammation in atopic dermatitis through suppression of innate immune responses in keratinocytes. *J. Allergy Clin. Immunol.* 2014, 134, 836–847. [CrossRef] [PubMed]
- Deaton, A.M.; Bird, A. CpG islands and the regulation of transcription. *Genes Dev.* 2011, 25, 1010–1022. [CrossRef]
- Takai, D.; Jones, P.A. Comprehensive analysis of CpG islands in human chromosomes 21 and 22. *Proc. Natl. Acad. Sci. USA* 2002, 99, 3740–3745. [CrossRef] [PubMed]
- 187. Schübeler, D. Function and information content of DNA methylation. Nature 2015, 517, 321–326. [CrossRef]

- 188. Nestor, C.E.; Barrenäs, F.; Wang, H.; Lentini, A.; Zhang, H.; Bruhn, S.; Jörnsten, R.; Langston, M.A.; Rogers, G.; Gustafsson, M.; et al. DNA Methylation Changes Separate Allergic Patients from Healthy Controls and May Reflect Altered CD4+ T-Cell Population Structure. *PLoS Genet.* 2014, 10, e1004059. [CrossRef]
- 189. Pascual, M.; Suzuki, M.; Isidoro-Garcia, M.; Padrón, J.; Turner, T.; Lorente, F.; Dávila, I.; Greally, J.M. Epigenetic changes in B lymphocytes associated with house dust mite allergic asthma. *Epigenetics* 2011, 6, 1131–1137. [CrossRef]
- 190. Joubert, B.R.; Felix, J.F.; Yousefi, P.; Bakulski, K.M.; Just, A.C.; Breton, C.; Reese, S.E.; Markunas, C.A.; Richmond, R.C.; Xu, C.-J.J.; et al. DNA Methylation in Newborns and Maternal Smoking in Pregnancy: Genome-wide Consortium Meta-analysis. *Am. J. Hum. Genet.* 2016, *98*, 680–696. [CrossRef]
- 191. Joehanes, R.; Just, A.C.; Marioni, R.E.; Pilling, L.C.; Reynolds, L.M.; Mandaviya, P.R.; Guan, W.; Xu, T.; Elks, C.E.; Aslibekyan, S.; et al. Epigenetic Signatures of Cigarette Smoking. *Circ. Cardiovasc. Genet.* 2016, 9, 436–447. [CrossRef]
- 192. Liang, Y.; Chang, C.; Lu, Q. The Genetics and Epigenetics of Atopic Dermatitis-Filaggrin and Other Polymorphisms. *Clin. Rev. Allergy Immunol.* **2016**, *51*, 315–328. [CrossRef]
- 193. Fontalba, A.; Gutierrez, O.; Fernandez-Luna, J.L. NLRP2, an inhibitor of the NF-kappaB pathway, is transcriptionally activated by NF-kappaB and exhibits a nonfunctional allelic variant. *J. Immunol.* **2007**, *179*, 8519–8524. [CrossRef] [PubMed]
- 194. Bruey, J.M.; Bruey-Sedano, N.; Newman, R.; Chandler, S.; Stehlik, C.; Reed, J.C. PAN1/NALP2/PYPAF2, an inducible inflammatory mediator that regulates NF-kappaB and caspase-1 activation in macrophages. *J. Biol.Chem.* 2004, 279, 51897–51907. [CrossRef] [PubMed]
- 195. Steevels, T.A.M.; van Avondt, K.; Westerlaken, G.H.A.; Stalpers, F.; Walk, J.; Bont, L.; Coffer, P.J.; Meyaard, L. Signal inhibitory receptor on leukocytes-1 (SIRL-1) negatively regulates the oxidative burst in human phagocytes. *Eur. J. Immunol.* 2013, 43, 1297–1308. [CrossRef] [PubMed]
- Steevels, T.A.M.; Lebbink, R.J.; Westerlaken, G.H.A.; Coffer, P.J.; Meyaard, L. Signal inhibitory receptor on leukocytes-1 is a novel functional inhibitory immune receptor expressed on human phagocytes. *J. Immunol.* 2010, 184, 4741–4748. [CrossRef]



© 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).