

# Use of Multiple Sequencing Technologies To Produce a High-Quality Genome of the Fungus *Pseudogymnoascus destructans*, the Causative Agent of Bat White-Nose Syndrome

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**White-nose syndrome has recently emerged as one of the most devastating wildlife diseases recorded, causing widespread mortality in numerous bat species throughout eastern North America. Here, we present an improved reference genome of the fungal pathogen *Pseudogymnoascus destructans* for use in comparative genomic studies.**

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Since emerging in 2006, white-nose syndrome has rapidly spread across eastern North America from an index site in New York, causing the mortality of millions of bats of numerous species (1). The pathogen was identified as *Pseudogymnoascus destructans* (2, 3), a psychrophilic fungus that infects bats during hibernation, when bat body temperatures decrease to the ambient temperature of their hibernacula (4). The growth of this fungus on bats and subsequent invasion of epidermal tissues can cause a cascade of deleterious physiological changes resulting in high mortality rates (5, 6). The fungus is widespread in Eurasia (7–9), and initial genetic and experimental results suggest that it was recently introduced to North America (10, 11). Critically missing, however, are detailed genomic analyses of *P. destructans* to shed light on the origins, evolution, global dispersal, and pathogenicity of this fungus.

Initial genome sequencing efforts for *P. destructans* used 454 and Illumina platforms; however, unresolved repeat regions in the genome resulted in fragmented draft assemblies (12). Therefore, to generate a high-quality contiguous assembly of the *P. destructans* genome, we utilized a combination of data from PacBio SMRT reads, Illumina MiSeq 250-bp paired-end reads (SRR1952982), 8-kb 454 “jumping” libraries (SRP001346), and end sequences from an unbiased random shear bacterial artificial chromosome (BAC) library with an average insert size of 100 kb (13).

The PacBio reads were assembled with the PacBio SMRT version 2.2 analysis pipeline (14) and yielded 153 contigs. Using Illumina 250-bp paired-end MiSeq reads quality-trimmed to Q30 with ea-utils version 1.1.2 (15) and error-corrected with Hammer (16), 454 8-kb jumping reads, and >100-kb BAC library end sequences, the contigs were extended and scaffolded with SSPACE version 3.0 (17); gaps were removed with GapFiller version 1.10

(18). Twenty iterations of SSPACE/GapFiller were followed by Pilon version 1.8 (19) to correct local misalignments and indels from the assembly, resulting in 96 scaffolds. A self-query of the improved scaffolds with the NUCmer application of MUMmer version 3.23 (20) indicated that the smaller scaffolds (<42,633 bp) and a few larger scaffolds were complete or nearly complete duplications of the sequence in other scaffolds. Thirteen scaffolds, which had a >95% identity over 95% of their length with a larger scaffold in the assembly, were removed. The final assembly contains 83 scaffolds, is 35.818201 Mb in size, has an  $N_{50}$  value of 1.168637 Mb, and contains only 7,812 bp of gaps. This is a significant improvement compared to previous draft assemblies of *P. destructans* (AEFC01: 1,847 scaffolds, 30.6849 Mb,  $N_{50}$  105.158 kb, 2,328,879 bp Ns; AYK01: 5,304 scaffolds, 30.2827 Mb,  $N_{50}$  17.914 kb, 0 Ns).

The new *P. destructans* genome contains 38.17% repetitive sequence elements (RepeatModeler version 1.08, RepeatMasker version 4.05, <http://www.repeatmasker.org>), which required a combination of long-read and large-insert sequencing technologies to scaffold. The nearly complete assembly presented here should serve as a valuable resource to facilitate fungal disease research.

**Nucleotide sequence accession numbers.** This whole-genome shotgun project has been deposited in DDBJ/ ENA/GenBank under the accession number LAJJ00000000. The version described in this paper is the first version, LAJJ01000000.

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## REFERENCES

- Turner GG, Reeder DM, Coleman JH. 2011. A five-year assessment of mortality and geographic spread of white-nose syndrome in North American bats and a look to the future. *Bat Res News* 52:13–27.
- Lorch JM, Meteyer CU, Behr MJ, Boyles JG, Cryan PM, Hicks AC, Ballmann AE, Coleman JTH, Redell DN, Reeder DM, Blehert DS. 2011. Experimental infection of bats with *Geomyces destructans* causes white-nose syndrome. *Nature* 480:376–378. <http://dx.doi.org/10.1038/nature10590>.
- Blehert DS, Hicks AC, Behr M, Meteyer CU, Berlowski-Zier BM, Buckles EL, Coleman JTH, Darling SR, Gargas A, Niver R, Okoniewski JC, Rudd RJ, Stone WB. 2009. Bat white-nose syndrome: an emerging fungal pathogen? *Science* 323:227. <http://dx.doi.org/10.1126/science.1163874>.
- Verant ML, Boyles JG, Waldrep W, Wibbelt G, Blehert DS. 2012. Temperature-dependent growth of *Geomyces destructans*, the fungus that causes bat white-nose syndrome. *PLoS One* 7:e46280. <http://dx.doi.org/10.1371/journal.pone.0046280>.
- Cryan PM, Meteyer CU, Boyles JG, Blehert DS. 2010. Wing pathology of white-nose syndrome in bats suggests life-threatening disruption of physiology. *BMC Biol* 8:135. <http://dx.doi.org/10.1186/1741-7007-8-135>.
- Verant ML, Meteyer CU, Speakman JR, Cryan PM, Lorch JM, Blehert DS. 2014. White-nose syndrome initiates a cascade of physiologic disturbances in the hibernating bat host. *BMC Physiol* 14:10. <http://dx.doi.org/10.1186/s12899-014-0010-4>.
- Hoyt JR, Sun K, Parise KL, Lu G, Langwig KE, Jiang T, Yang S, Frick WF, Kilpatrick AM, Foster JT, Feng J. 2015. Widespread bat white-nose syndrome fungus, northeastern China. *Emerg Infect Dis* 22:140–142. <http://dx.doi.org/10.3201/eid2201.151314>.
- Puechmaille SJ, Frick WF, Kunz TH, Racey PA, Voigt CC, Wibbelt G, Teeling EC. 2011. White-nose syndrome: is this emerging disease a threat to European bats? *Trends Ecol Evol* 26:570–576. <http://dx.doi.org/10.1016/j.tree.2011.06.013>.
- Zukal J, Bandouchova H, Brichta J, Cmokova A, Jaron KS, Kolarik M, Kovacova V, Kubátová A, Nováková A, Orlov O, Pikula J, Presetnik P, Šuba J, Zahradníková A, Jr, Martínková N. 2016. White-nose syndrome without borders: *Pseudogymnoascus destructans* infection tolerated in Europe and palearctic Asia but not in North America. *Sci Rep* 6:19829. <http://dx.doi.org/10.1038/srep19829>.
- Leopardi S, Blake D, Puechmaille SJ. 2015. White-nose syndrome fungus introduced from Europe to North America. *Curr Biol* 25:R217–R219. <http://dx.doi.org/10.1016/j.cub.2015.01.047>.
- Warnecke L, Turner JM, Bollinger TK, Lorch JM, Misra V, Cryan PM, Wibbelt G, Blehert DS, Willis CKR. 2012. Inoculation of bats with European *Geomyces destructans* supports the novel pathogen hypothesis for the origin of white-nose syndrome. *Proc Natl Acad Sci USA* 109:6999–7003. <http://dx.doi.org/10.1073/pnas.1200374109>.
- Chibucos MC, Crabtree J, Nagaraj S, Chaturvedi S, Chaturvedi V. 2013. Draft genome sequences of human pathogenic fungus *Geomyces pannorum sensu lato* and bat white nose syndrome pathogen *Geomyces (Pseudogymnoascus) destructans*. *Genome Announc* 1(6):e01045-13. <http://dx.doi.org/10.1128/genomeA.01045-13>.
- Bok JW, Ye R, Clevenger KD, Mead D, Wagner M, Krowicz A, Albright JC, Goering AW, Thomas PM, Kelleher NL, Keller NP, Wu CC. 2015. Fungal artificial chromosomes for mining of the fungal secondary metabolome. *BMC Genomics* 16:343. <http://dx.doi.org/10.1186/s12864-015-1561-x>.
- Chin CS, Alexander DH, Marks P, Klammer AA, Drake J, Heiner C, Clum A, Copeland A, Huddleston J, Eichler EE, Turner SW, Korlach J. 2013. Nonhybrid, finished microbial genome assemblies from long-read SMRT sequencing data. *Nat Methods* 10:563–569. <http://dx.doi.org/10.1038/nmeth.2474>.
- Aronesty E. 2013. Comparison of sequencing utility programs. *Open Bioinforma J* 7:1–8. <http://dx.doi.org/10.2174/1875036201307010001>.
- Medvedev P, Scott E, Kakaradov B, Pevzner P. 2011. Error correction of high-throughput sequencing datasets with non-uniform coverage. *Bioinformatics* 27:i137–i141. <http://dx.doi.org/10.1093/bioinformatics/btr208>.
- Boetzer M, Henkel CV, Jansen HJ, Butler D, Pirovano W. 2011. Scaffolding pre-assembled contigs using SSPACE. *Bioinformatics* 27:578–579. <http://dx.doi.org/10.1093/bioinformatics/btq683>.
- Nadalin F, Vezzi F, Policriti A. 2012. GapFiller: a de novo assembly approach to fill the gap within paired reads. *BMC Bioinformatics* 13(suppl 1):S8. <http://dx.doi.org/10.1186/1471-2105-13-S14-S8>.
- Walker BJ, Abeel T, Shea T, Priest M, Abouelliel A, Sakthikumar S, Cuomo CA, Zeng Q, Wortman J, Young SK, Earl AM. 2014. Pilon: an integrated tool for comprehensive microbial variant detection and genome assembly improvement. *PLoS One* 9:e112963. <http://dx.doi.org/10.1371/journal.pone.0112963>.
- Kurtz S, Phillippy A, Delcher AL, Smoot M, Shumway M, Antonescu C, Salzberg SL. 2004. Versatile and open software for comparing large genomes. *Genome Biol* 5:R12. <http://dx.doi.org/10.1186/gb-2004-5-2-r12>.