





## Draft Genome Sequence of *Halobacillus* sp. Strain KGW1, a Moderately Halophilic and Alkaline Protease-Producing Bacterium Isolated from the Rhizospheric Region of *Phragmites karka* from Chilika Lake, Odisha, India

Ananta Narayan Panda,<sup>a</sup> Samir R. Mishra,<sup>a</sup> Lopamudra Ray,<sup>a</sup> Neha Sahu,<sup>a</sup> Ankita Acharya,<sup>c</sup> Sudhir Jadhao,<sup>c</sup> Mrutyunjay Suar,<sup>a</sup> Tapan Kumar Adhya,<sup>a</sup> Gurdeep Rastogi,<sup>b</sup> Ajit Kumar Pattnaik,<sup>b</sup> Vishakha Raina<sup>a</sup>

School of Biotechnology, KIIT University, Bhubaneswar, Odisha, India<sup>a</sup>; Wetland Research and Training Center, Chilika Development Authority, Department of Forest and Environment, Bhubaneswar, Odisha, India<sup>b</sup>; Bionivid Technology Private Limited, Kasturi Nagar, Bangalore, India<sup>c</sup>

Halobacillus sp. strain KGW1 is a moderately halophilic, rod shaped, Gram-positive, yellow pigmented, alkaline protease-producing bacterium isolated from a water sample from Chilika Lake, Odisha, India. Sequencing of bacterial DNA assembled a 3.68-Mb draft genome. The genome annotation analysis showed various gene clusters for tolerance to stress, such as elevated pH, salt concentration, and toxic metals.

Received 18 March 2016 Accepted 21 March 2016 Published 30 June 2016

Citation Panda AN, Mishra SR, Ray L, Sahu N, Acharya A, Jadhao S, Suar M, Adhya TK, Rastogi G, Pattnaik AK, Raina V. 2016. Draft genome sequence of *Halobacillus* sp. strain KGW1, a moderately halophilic and alkaline protease-producing bacterium isolated from the rhizospheric region of *Phragmites karka* from Chilika Lake, Odisha, India. Genome Announc 4(3):e00361-16. doi:10.1128/genomeA.00361-16.

**Copyright** © 2016 Panda et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license. Address correspondence to Vishakha Raina, vishakha raina@qmail.com.

having low G+C content, as identified by Spring et al. (1). Including the other type species *Halobacillus halophilus* (formerly *Sprorosarcina halophile*), *Halobacillus littoralis* (DSM 10405<sup>T</sup>) and *Halobacillus truperi* (DSM 10404 <sup>T</sup>) (2), the *Halobacillus* genus is composed of 21 species that are validly published. This genus features pigment-producing (3) moderate halophiles that are capable of producing a variety of enzymes at a wide range of salinity and pH (4). Thus, halophiles are suitable species for screening of enzyme and carotenoid production useful for various industrial applications. Chilika Lake is situated between 19°28′ and 19°54′N latitude and 85°05′ and 85°38′E longitude. *Phragmites karka* (a common reed) forms monoculture dense patches in lagoon Chilika and is considered to be a highly invasive weed in Chilika Lake (5, 6).

The microbial communities associated with the rhizospheric region of these macrophytes contribute toward overall biogeochemical cycling and play an important role in nutrient cycling at the ecosystem level (7). *Halobacillus* sp. strain KGW1<sup>T</sup> was isolated from rhizospheric region water sample of macrophytedominated areas at Kalupara Ghat, Chilika Lake (19.84699'N and 85.40778'E) by dilution plating technique at 30°C on Zobell's marine broth (ZMB; HiMedia, India). It is a Gram-positive, alkaline protease-producing bacterium that can grow from a temperature 15°C to 45°C and can tolerate alkaline pH (pH 5.5 to 11.5) and salt (0 to 25% [wt/vol] NaCl).

Genomic DNA was extracted using the Gnome kit (MP Biomedicals, Santa Ana, CA). The genome sequence of strain KGW1<sup>T</sup> was sequenced using an Illumina MiSeq sequencing platform. The data generated were assembled using the Velvet (version 1.2.10) assembler (8), resulting in 13 contigs, out of a total of

3,683,719 bp, and an  $N_{50}$  contig size of 1,191,089 bp (1.19 Mb). The estimated complete genome size was 3.68 Mb, with a G+C content of 46.98%. Genome annotation was performed using the Rapid Annotations using Subsystems Technology (RAST) (9, 10), which predicted a total of 3,900 protein-coding sequences, 74 pseudogenes, 67 tRNAs, and 7 rRNA clusters. The taxonomy identification was performed using EzTaxon and MEGA6, which identified H. trueperi as the putative species per 16S rRNA gene sequence homology. PHAST analysis (11) revealed a putative intact phage integrated in the genome, with a length of 54.3 kb, 52 protein-coding sequences, and a G+C content of 42.36%.

The RAST annotations identified various major gene clusters for stress regulation, resistance to toxic compounds and heavy metals, protein degradation, carbohydrate degradation, degradation of multiple aromatic compounds, lipid metabolism, 1-aminocyclopropane-1-carboxylate deaminase activity, auxin biosynthesis, nitrogen, metabolism, siderophore production, phosphorous solubilization, and sulfur metabolism.

**Nucleotide sequence accession numbers.** This whole-genome shotgun project has been deposited in DDBJ/EMBL/GenBank under the accession no. LSOC00000000. The version described in this paper is the first version, LSOC01000000.

## **ACKNOWLEDGMENTS**

We are grateful to the Chilika Development Authority (CDA), Bhubaneswar. The staff of the CDA are acknowledged for their help and providing necessary facilities in collection of water and sediment samples.

## **FUNDING INFORMATION**

This research received no specific grant from any funding agency in the public, commercial, or not-for-profit sectors.

## **REFERENCES**

- 1. Spring S, Ludwig W, Marquez MC, Ventosa A, Schleifer KH. 1996. *Halobacillus* gen. nov., with descriptions of *Halobacillus litoralis* sp. nov., and *Halobacillus trueperi* sp. nov., and transfer of *Sporosarcina halophila* to *Halobacillus halophilus* comb. Int J Syst Bacteriol 46:492–496. http://dx.doi.org/10.1099/00207713-46-2-492.
- Kushner DJ. 1986. Molecular adaptation of enzymes, metabolic systems and transport systems in halophilic bacteria. FEMS Microbiol Rev 39: 121–127. http://dx.doi.org/10.1111/j.1574-6968.1986.tb01852.x.
- 3. Köcher S, Breitenbach J, Müller V, Sandmann G. 2009. Structure, function and biosynthesis of carotenoids in the moderately halophilic bacterium *Halobacillus halophilus*. Arch Microbiol 191:95–104. http://dx.doi.org/10.1007/s00203-008-0431-1.
- Ventosa A, Nieto JJ, Oren A. 1998. Biology of moderately halophilic bacteria. Microbiol Mol Biol Rev 62:504–544.
- Kumar R, Pattnaik AK. 2012. Chilika—an integrated management planning framework for conservation and wise use. Wetlands International, South Asia, New Delhi, India, and Chilika Development Authority, Bhubaneswar, Odisha, India.
- Chilika Development Authority. 2012. Annual report: 2010–11. Third Eye Communications, Bhubaneswar, India. http://www.chilika.com/pdf /CDA%20Annual%20Report%202012.pdf.
- 7. Borsodi AK, Rusznyák A, Molnár P, Vladár P, Reskóné MN, Tóth EM,

- Sipos R, Gedeon G, Márialigeti K. 2007. Metabolic activity and phylogenetic diversity of reed (*Phragmites australis*) periphyton bacterial communities in a Hungarian shallow soda lake. Microb Ecol 53:612–620. http://dx.doi.org/10.1007/s00248-006-9133-x.
- Zerbino DR, Birney E. 2008. Velvet: algorithms for *de novo* short read assembly using de Bruijn graphs. Genome Res 18:821–829. http:// dx.doi.org/10.1101/gr.074492.107.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. BMC Genomics 9:75. http://dx.doi.org/10.1186/1471-2164-9-75.
- Overbeek R, Olson R, Pusch GD, Olsen GJ, Davis JJ, Disz T, Edwards RA, Gerdes S, Parrello B, Shukla M, Vonstein V, Wattam AR, Xia F, Stevens R. 2014. The SEED and the rapid annotation of microbial genomes using subsystems technology (RAST). Nucleic Acids Res 42: D206-D214. http://dx.doi.org/10.1093/nar/gkt1226.
- Zhou Y, Liang Y, Lynch KH, Dennis JJ, Wishart DS. 2011. PHAST: a fast phage search tool. Nucleic Acids Res 39:W347–W352. http://dx.doi.org/ 10.1093/nar/gkr485.