

# Overview of triple negative breast cancer prognostic signatures in the context of data science-driven clinico-genomics research

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## Background

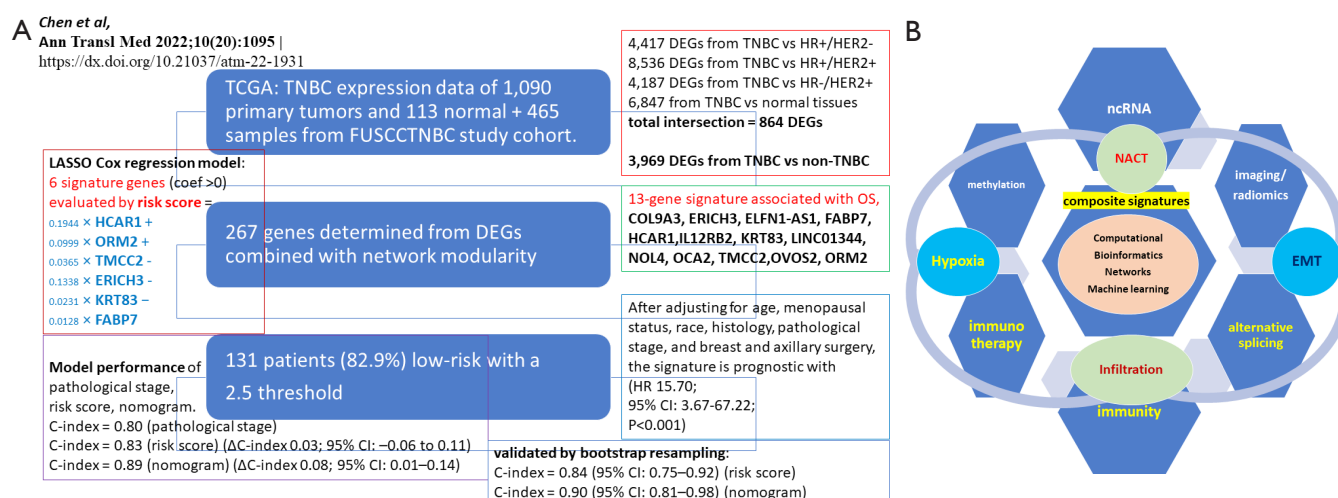
Triple-negative breast cancer (TNBC) is a highly aggressive tumor with inherent heterogeneity leading to a variety of outcomes, including early relapse, metastatic spread, and poor survival. Chen *et al.*'s (1) proposed a prognostic signature to improve risk stratification in TNBC patients obtained by combining expressions from The Cancer Genome Atlas (TCGA), with 1,090 primary tumors and 113 normal, and RNA-seq data FUSCCTNBC (Sequence Read Archive, SRP15). As the samples were classified according to subtypes (HR+/HER2-, HR+/HER2+, HR-/HER2+, TNBC, and normal samples with status of ER, PR, and HER2), the differentially expressed gene (DEG) profiles were computed from pairwise comparisons between TNBC and the other subtypes.

In general, signatures may inherently determine stratifications based on groups of patients sharing similar tumor characteristics and biological properties. Ideally, a signature should be linked to some novel mechanism, a pathway or a biological process or a drug mechanism. Algorithmic or semi-algorithmic identifications of (non-redundant) groups call for validation at both biological and clinical levels. For instance, prognostic validity may be established because of its association with overall survival. One usually relies on the 'big data evidence' that omics are expected to support. However, the reliability assigned to these and other types of signatures is a controversial matter. First, many cancer signatures are not fully justified. Signatures may fail to show biological value with reference

to the specific cancer they address. This is typically proven algorithmically, by enabling gene signature replacement and showing that surrogate genes have measurable prognostic prediction power not affecting the overall signature, but with opposite biological interpretation compared to the replaced genes with respect to mechanisms. Therefore, the problem becomes lack of provable causality regarding outcome prediction, depending on the usable surrogate gene space.

Second, reproducibility implies the ability to assess the robustness of a signature relatively to specific methods or platforms. A demonstration for association with outcomes should be generally valid across independent groups and replicate experiments, and with no role played by factors excluded from the model generating the signature, including noise. Gene expression signatures (GES) connecting phenotypes to mRNA suffer from quite limited reproducibility. Therefore, one possible strategy is to combine significantly enhanced GES across datasets and experiments covering various phenotypes. Another strategy is to correlate gene expressions with other measurements to increase the signature's prediction power, for instance through co-expression networks.

The strategy followed by Chen *et al.* [2022] to obtain the prognostic GES involves multiple steps (*Figure 1A*). Chen *et al.*' results are now contextualized (*Figures 1B,2*) with reference to a selected non-comprehensive list of recent signature results retrieved from PubMed (query: "TNBC prognostic signatures", 164 results).



**Figure 1** Overview of Chen *et al.*'s work and contextualization of their signature. (A) Basic sketch of the approach in Chen *et al.* The results have highlighted a multi-year (1-to-5) prognostic 6-gene signature integrated to a clinical scoring system utilizing pathological stage information through a stably performing nomogram and resulting in an improved concordance index. (B) Biomedical contexts selected from recent literature and defining TNBC prognostic signatures. Note that the highlighted composite signatures have data science as a driver. TNBC, triple-negative breast cancer.

## Contextualization

### NACT and EMT

TNBC patients respond better to neoadjuvant therapy compared to other BC subtypes, though some patients still have a poor prognosis (2). A study represented the transcriptional portrait associated with neoadjuvant chemotherapy (NACT) resistance and found candidate genes as prognostic markers (3). In Blaye *et al.*'s study (4), an immunologic transcriptomic signature was found using 115 post-NACT samples to predict relapse in TNBC patients by profiling 770 TME-related genes using the NanoString PanCancer IO360 panel. The 8 genes linked to immunity (*BLK*, *GZMM*, *CXCR6*, *LILRA1*, *SPIB*, *CCL4*, *CXCR4*, *SLAMF7*) were predictive of intra-cohort relapse (validated externally in KMplot and METABRIC), indicating that lack of immune activation after NACT is associated with a high risk of distant relapse. Another study (5) revealed a 6-epithelial-mesenchymal transition (EMT) gene signature (*LUM*, *SFRP4*, *COL6A3*, *MMP2*, *CXCL12*, and *HTRA1*) at diagnosis potentially predictive for metastasis in TNBC patients who did not achieve pathological complete response to NACT and in patients treated with surgery in combination with adjuvant therapy. The EMT pathway was significantly enriched in pre- and post-NACT patients with metastasis after NACT. The 6-EMT gene signature was validated in a GEO dataset (HR =0.36,

P=0.0008, 95% CI: 0.200–0.658) and in another GEO dataset of TNBC patients who received surgery followed by adjuvant chemotherapy (HR =0.46, 95% CI: 0.225–0.937). Other results (6) informing on cell plasticity relatively to EMT were centered on a signature of circulating tumor cells (CTCs) from 32 patients aimed to identify markers associated with TNBC outcome and aggressiveness, especially tissue inhibitor of metalloproteinases 1 (*TIMP1*) and androgen receptor (AR), whose blockade was further analyzed *in vitro* and *in vivo* in relation to proliferation and dissemination.

### Immunotherapy & hypoxia-related

Immunotherapy (ImT) research and TNBC prognosis present signature gaps. A hypoxia-immune-based cross-cohort classifier predictive of prognosis was developed and validated in (7) to potentially guide TNBC ImT. In study of Yang *et al.* (8), six gene cross-cohort prognostic hypoxia-immune related signatures were identified to stratify patients into high-/low-risk groups, with the former group showing poorer prognosis, and correlation with hypoxia status in tumor microenvironment (TME). Interestingly, there were relatively less activated immune cells and lower expression levels in immune checkpoint inhibitors (ICI). In study of Sun *et al.* (9), a robust predictive 3-gene signature (*PEKL*, *ALDOA*, *PGK1*) associated with the glycolytic process was

## Immunity Prognostic Signatures

| NACT & relapse            |  | Immune Infiltration |          | Immune Checkpoint Inhibitors               |              | TME            | Hypoxia        |
|---------------------------|--|---------------------|----------|--|--------------|----------------|----------------|
|                           |  | Network-driven      |          | Low risk                                   | OS favorable | Poor prognosis |                |
| BLK                       |  | RMND5A              | TOP2A    | PD-1                                       | PDCD1        | PPFIA4         | PEKL           |
| GZMM                      |  | ZNF829              | CCNA2    | PD-L1                                      | PDCD1LG2     | TNFRSF1B       | ALDOA          |
| CXCR6                     |  | KDM5B               | PCNA     | PD-L2                                      | KIR3DL2      | ARHGAP9        | PGK1           |
| LILRA1                    |  | NCBP2               | MSH2     | CTLA4                                      |              | ZNF831         |                |
| SPIB                      |  | GPI                 | CDK6     |  |              | CTLA4          |                |
| CCL4                      |  | BGN                 |          |  |              | BLK            |                |
| CXCR4                     |  | CCND2               |          |  |              | ANKRD22        |                |
| SLAMF7                    |  | PLBD1               |          |  |              | CLEC4E         |                |
|                           |  | ZYG11A              |          |  |              |                |                |
|                           |  | IL17                |          |  |              |                |                |
| Macrophage and Resistance |  | EMT                 |          | Paracancer                                 |              |                | Bioinfo-driven |
| Network-driven            |  |                     | With CTC |  |              |                | Poor OS        |
| CD79A                     |  | LUM                 | AR       | ABCC9 (cg06951626)                         |              |                | CENPW          |
| CXCL13                    |  | SFRP4               | TIMP1    | NKAPL (cg18675097, cg01031101, cg17384889) |              |                | HORMAD1        |
| IGLL5                     |  | COL6A3              |          | TMEM132C (cg03530754)                      |              |                | APOD           |
| LHFPL2                    |  | MMP2                |          |  |              |                | PIP            |
| PLEKHF1                   |  | CXCL12              |          |  |              |                | ZNF703         |
|                           |  | HTRA1               |          |  |              |                |                |
| DRFS                      |  | DFS                 |          | Poor, GEO, network-driven                  |              | PFI            |                |
| CD1B                      |  | IL18R1              | FAM83B   | BUB1                                       |              | FAM83B         |                |
| CD53                      |  | CD53                | KITLG    | CCNA2                                      |              | EXO1           |                |
| CT45A1                    |  | TRIM                | CFD      | PACC1                                      |              | S100B          |                |
| GTF3C1                    |  | Jaw1                | RBM24    |  |              | TYMS           |                |
| IL11RA                    |  | LTB                 |          |  |              | CFD            |                |
| IL1RN                     |  | PTPRCAP             |          |  |              |                |                |
| LRRN3                     |  |                     |          |  |              |                |                |
| MAPK1                     |  |                     |          |  |              |                |                |
| NEFL                      |  |                     |          |  |              |                |                |
| PRKCE                     |  |                     |          |  |              |                |                |
| PTPRC                     |  |                     |          |  |              |                |                |
| SPACA3                    |  |                     |          |  |              |                |                |
| TNFSF11                   |  |                     |          |  |              |                |                |

DFS= disease-free survival

PFI= progression-free interval

DRFS=Distant recurrence-free survival

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HCA1

ORM2

TMCC2

ERICH3

KRT83

FABP7

## Methylation Prognostic Signatures

| m6A modification-hypoxia | m6A       | m6A-related lncRNA |
|--------------------------|-----------|--------------------|
|                          | Favorable |                    |
| PIM2                     | METTL14   | SAMD12-AS1         |
| PET117                   | ALKBH5    | BVES-AS1           |
| SMARCA5                  |           | LINC00593          |
| TAF9                     |           | MIR205HG           |
| ABCB10                   |           | LINC00571          |
| MKP1                     |           | ANKRD10-IT1        |
|                          |           | CIRBP-AS1          |
|                          |           | SUCLG2-AS1         |
|                          |           | BLACAT1            |
|                          |           | HOXB-AS1           |

## ncRNA Prognostic Signatures

| miRNA              | OS             | Distant metastases-free | lncRNA immune-related |
|--------------------|----------------|-------------------------|-----------------------|
| Relapse predictive |                |                         |                       |
| miR-139-5p         | hsa-miR-342-3p | hsa-miR-29c             | RP11-890B15.3         |
| miR-10b-5p         | hsa-miR-195    | hsa-miR-342-3p          | RP11-1024P17.1        |
| miR-486-5p         | hsa-miR-155    | hsa-let-7c              | MFI2-AS1              |
| miR-455-3p         | hsa-miR-497    | hsa-let-7b              | RP11-180N14.1         |
| miR-107            |                | hsa-miR-497             |                       |
| miR-146b-5p        |                |                         |                       |
| miR-324-5p         |                |                         |                       |
| miR-20a-5p         |                |                         |                       |

Figure 2 Prognostic signatures associated to multiple domains and macro-categorized.

identified based on 48 hypoxia-related prognostic genes and a risk score model was set as a prognostic biomarker for TNBC. Future ImT developments will involve ICI toward TNBC treatment, as in Islam *et al.*'s study (10) (PI3K, MEK, PARP, EGFR, VEGF, and AR). Immune prognostic signatures were investigated (11) based on 66 prognostic genes, showing predictive power of the response to ICI (PD-1, PD-L1, PD-L2 and CTLA4 gene expressions higher in the low-risk group). ICI expression (and clinical) profiles of TNBC samples from TCGA and METABRIC were used to build 1-, 2-, 3-year OS-related GES (PDCD1, PDCD1LG2 and KIR3DL2, with receiver operating characteristic (ROC) curves showing AUC values of 0.925, 0.822 and 0.835, respectively) from potentially favorable OS predictors (*IDO1*, *CD274*, *PDCD1LG2*, *PDCD1*, *CTLA4*, *ICOS*, *KIR3DL2*, *HLA-B*, *HLA-F*, *LGA3*) (12).

Studies of underlying mechanisms will include the TME and the immune infiltration (ImI). In study of Qin *et al.* (13), the TNBC TME landscape was elucidated (GEO, N=107, training: METABRIC, N=299, validation cohorts). Two distinct TME clusters were identified, one characterized by low ImI with poor prognosis, the other with high ImI and better survival probability. These clusters showed differential expression (37 upregulated and 778 downregulated genes) delivering then a final 8-gene TME signature (*PPF1A4*, *TNFRSF1B*, *ARHGAP9*, *ZNF831*, *CTLA4*, *BLK*, *ANKRD22* and *CLEC4E*) in which *TNFRSF1B* and *BLK* were risk factors for poor prognosis in TNBC patients. In (14), from the study of 1,145 patients a tumor immune risk score (TIRS) was obtained with 8 biomarkers stratifying patients in high- and low-risk groups. Significant survival and ImI pattern differences were found at both transcriptome and protein levels indicating 4 different tumor immune microenvironment types (TIMT): one associated with the best prognosis and immune status, another with the opposite effect, the other two associated with highly unstable genomes and stem-cell-related characteristics along with high stromal scores.

### Bioinformatics composite signatures

TNBC prognostic signatures may involve hub genes and pathways (15), without establishing genes as drivers of network causality and directionality (16). For example, in (17) a 5-gene prognostic signature (*CD79A*, *CXCL13*, *IGLL5*, *LHFPL2*, and *PLEKHF1*) was found based on co-expressed hub genes associated with macrophages and related to gemcitabine resistance in TNBC patients. Then,

a survival-related 5-gene hub signature (*TOP2A*, *CCNA2*, *PCNA*, *MSH2*, *CDK6*) was combined with ImI analysis to show relevance for immune monitoring across GEO cancer data. Tumor-infiltrating lymphocytes (TILs) are a robust prognostic factor for improved patient survival, with T cells being predominant, but without resolving the uncertainty about the relationship with patient prognosis (18).

The real predictive power of cancer-related immune prognostic markers was investigated in (19) through a bioinformatic approach identifying a 6-gene signature (*IL18R1*, *CD53*, *TRIM*, *JAW1*, *LTB*, and *PTPRCAP*) positively correlated with disease-free survival (DFS) in TNBC patients and showing significant immune expression profiles in cancer cells and in intra- and peri-TILs. In Kim *et al.*'s study (20), it was identified the associations between gene expression and distant recurrence-free survival (DRFS) from a 13-gene expression profile (*CD1B*, *CD53*, *CT45A1*, *GTF3C1*, *IL11RA*, *IL1RN*, *LRRN3*, *MAPK1*, *NEFL*, *PRKCE*, *PTPRC*, *SPACA3* and *TNFSF11*) associated with patient prognosis ( $P < 0.05$ ) (AUC = 0.923). Combined with stage, it was predictive of distant recurrence of early TNBC, with 3 genetic risk groups classifying DRFS rate significantly even intra-stage ( $P < 0.001$ ) and informing on treatment with new possible ImT targets. Finally, cell differentiation together with grade- and tumor mutational burden-related DEGs were identified using integrated single-cell and bulk RNA-seq data analysis, delivering a composite 10-gene prognostic signature (*RMND5A*, *ZNF829*, *KDM5B*, *NCBP2*, *GPI*, *BGN*, *CCND2*, *PLBD1*, *ZYG11A*, *IL17RD*), closely related to tumor ImI (21).

### Epigenetics

Despite its relevance, aberrant DNA methylation is still underutilized in prognostic models for TNBC. For instance, the N6-methyladenosine ( $m^6A$ ) modification is relevant in cancer development, but not enough evidenced in TNBC. Based on the TCGA transcriptome profiles of the 13  $m^6A$  methylation regulators measured from 98 TNBC tumor and normal tissue samples, two expression levels were emphasized, specifically ALKBH5 (unfavorable prognostic factor, HR = 3.327,  $P = 0.006$ ) and METTL14 (favorable, HR = 0.425,  $P = 0.009$ ), both contributing to improve the accuracy of a prognostic model of risk prediction when combined with TNM stage (AUC = 0.791) (22). Aberrant DNA methylation was also investigated in study of Zhang *et al.* (23) from TCGA paracancer samples (PCS). Differential methylation-DEG correlation revealed

1,525 DEGs and 150 differentially methylated genes between TNBC and PCS, with a signature including prognostic markers such as ABCC9 (cg06951626), NKAPL (cg18675097, cg01031101, and cg17384889), and TMEM132C (cg03530754).

In Gao *et al.*'s study (24), methylation and expression data from TCGA were combined, and 743 differentially methylated sites (DMS) were identified corresponding to 332 genes, with 357 hypermethylated sites and 386 hypomethylated sites, of which 103 were prognosis related. A LASSO-driven 5-DMSs prognostic signature was found to classify TNBC patients with significant survival difference (log-rank  $P=4.97E-03$ ), then validated in GSE78754 (HR =2.42, 95% CI: 1.27–4.59, log-rank  $P=0.0055$ ) and verified for DFS in GSE141441 (HR =2.09, 95% CI: 1.28–3.44, log-rank  $P=0.0027$ ). Two DMS were related to high risk (cg21234506 and cg21580376; HR >1), and three DMS were protective (cg15724876, cg17887364, and cg19419246; HR <1). Interestingly, a recent study (25) on TME of TNBC associated m<sup>6</sup>A modification and hypoxia status, identifying 26 genes related to both regulation types and characterizing two clusters, one being with significantly worse prognosis. A 6-gene prognostic signature was identified (*PIM2*, *PET117*, *SMARCA5*, *TAF9*, *ABCB10*, *MKP1*) among the m<sup>6</sup>A modification-hypoxia genes to evaluate risk and predict ImT response of patients.

#### Network- and machine learning (ML)-driven signatures

Key DEGs (*CENPW*, *HORMAD1*, *APOD*, *PIP*, and *ZNF703*) were found as candidate prognostic markers associated with poor OS through integrated bioinformatics performed on GEO microarray TNBC data (26). Network analysis focused on 147 co-DEGs in TNBC vs non-cancerous tissue samples, identifying a 15-gene signature inclusive of *BUB1* and *CENPF* significantly correlated with OS, while *BUB1*, *CCNA2*, and *PACC1* showed significant poor DFS (27). In study of Liu *et al.* (28), 105 heterogeneous DEGs were identified between TNBC and other subtypes. Two prognostic signatures were significant: a 4-gene (*FAM83B*, *KITLG*, *CFD* and *RBM24*) in disease-free interval, and a 5-gene (*FAM83B*, *EXO1*, *S100B*, *TYMS* and *CFD*) in progression-free interval. The multivariate Cox regression models showed predictive performance by time-dependent ROC analysis and survival analysis of TNBC subtypes (mesenchymal stem-like and mesenchymal) emphasized the FAM83B prognostic role.

In study of Liu *et al.* (29), bioinformatics and network

analyses explored the role of alternative splicing events (ASE) events and their correlation with TNBC prognostic DEGs, delivering ASE profiles, prognostic interaction networks, and splice factor-AS interaction networks. Finally, the use of ML was central to (30) where first a Recursive Feature Elimination (RFE) algorithm identified signatures (20, 25, 30, 35, 40, 45, and 50 genes) differentiating TNBC from other subtypes, then XGBoost was found the best performer for 45 genes (mixed signatures), of which 34 genes differentially regulated, 4 specifically relevant for distant metastasis free survival, and 2 potentially prognostic (*POU2AF1* and *S100B*) associated with MAPK, PI3-AkT, Wnt, TGF- $\beta$ , and other signal transduction pathways involved in metastasis. In study of Gong *et al.* (31), 151 TNBC patients obtained from the TCGA SpliceSeq database showed relevance for the Exon Skip (ES) type of AS events, more robust in predicting performance in TNBC prognosis. The ES AS signature confers a strong oncogenic phenotypic enrichment in a high-risk group (cell cycle and SUMOylating pathways of tumorigenesis), and a low-risk group (programmed cell death and metabolism pathways).

#### Non-coding RNAs

Many ncRNAs are likely dysregulated in TNBC, some being promising biomarkers and prognostic too. For instance, an 8-miRNAs signature (miR-139-5p, miR-10b-5p, miR-486-5p, miR-455-3p, miR-107, miR-146b-5p, miR-324-5p and miR-20a-5p) supported an accurate predictive model of relapse in TNBC patients highly correlated with prognosis (AUC of 0.80), independently validated twice (AUCs of 0.89 and 0.90) (32). In study of Zaka *et al.* (33), Cox analysis was employed to identify 25 miRNAs associated with prognosis with both risky and protective outcomes. Some miRNAs were associated with OS (hsa-miR-342-3p, hsa-miR-195, hsa-miR-155, and hsa-miR-497) and others with distant metastases-free survival (hsa-miR-29c, hsa-miR-342-3p, hsa-let-7c, hsa-let-7b, and hsa-miR-497). Based on both TCGA and GEO, and supported by ImmPort relatively to immune-related mRNAs, of 62 identified immune-related lncRNAs a signature of 4 (RP11-890B15.3, RP11-1024P17.1, MFI2-AS1 and RP11-180N14.1) showed regulation power mediated by miRNAs and prognostic value stratifying TNBC patients into high- and low-risk groups. The high-risk group had unfavorable outcomes together with significant immune response to tumor cell and high infiltrating abundance of regulatory T cell (34). In study



of Wu *et al.* (35), a 10-m6A-related lncRNA signature predictive of the prognostic risk in TNBC (SAM12-AS1, BVES-AS1, LINC00593, MIR205HG, LINC00571, ANKRD10-IT1, CIRBP-AS1, SUCLG2-AS1, BLACAT1, and HOXB-AS1) was found, establishing a prognostic score risk model (AUC of 0.997 and 0.864 in TCGA and GSE76250 datasets, respectively) and showing patients with lower score associated to better OS.

### *Radiomics & integrated signatures*

A TNBC radiogenomic study (N=202) generated quantitative radiomic features (n=860) from contrast-enhanced magnetic resonance images (CE-MRI). Notably, a radiomic feature capturing peritumoral heterogeneity (PH) was an established prognostic factor for RFS (P=0.01) and OS (P=0.004). Combined with matching transcriptomic and metabolomic data, the PH was associated with immune suppression and upregulated fatty acid synthesis (36). A role for radiomics with reference images to achieve consistent assessments is linked to the assessment of stromal TILs to be incorporated into clinical practice scores based on the reproducibility of multiple factors central to the heterogeneity in lymphocyte distribution (slide-related issues; outside tumor boundary; minimal assessable stroma; inflammatory cells etc.) (37). In study of Wu *et al.* (38), 17 dynamic CE-MRI features and TILs were combined to characterize tumor and parenchyma in a TCGA cohort (n=126). A LASSO imaging signature for TILs was found and a prediction model built with imaging signature and molecular features, resulting in 4 imaging features significantly associated with TILs: tumor volume, cluster shade of signal enhancement ratio (SER), mean SER of tumor-surrounding background parenchymal enhancement (BPE), and proportion of BPE. The imaging signature during validation allowed the predicted TILs to separate TNBC patients into two groups with distinct RFS (log-rank P=0.042), with TNBC showing no/minimal TILs associated with a worse prognosis.

To conclude, TNBC research offers room for further improvement. Especially longitudinal studies based on patients with long or more prolonged survival life periods (5–10 years) and involving a multitude of data/marker types including imaging, will be valuable, bringing future challenges for integrative data science approaches.

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### **Footnote**

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*Ethical Statement:* The author is accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

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