

# Preliminary analysis of the protein profile in saliva during physiological term and preterm delivery

MACIEJ ŁOPUCKI<sup>1\*</sup>, JACEK WAWRZYKOWSKI<sup>2\*</sup>, TOMASZ GĘCA<sup>3\*</sup>, ANDRZEJ MITURSKI<sup>3\*</sup>,  
MONIKA FRAN CZYK<sup>2\*</sup> and MARTA KANKOFER<sup>2\*</sup>

<sup>1</sup>Department of Gynaecological Oncology and Gynaecology, Medical University of Lublin, 20-081 Lublin;

<sup>2</sup>Department of Biochemistry, Faculty of Veterinary Medicine, University of Life Sciences in Lublin, 20-033 Lublin;

<sup>3</sup>Department of Obstetrics and Pathology of Pregnancy, Medical University of Lublin, 20-081 Lublin, Poland

Received October 18, 2017; Accepted March 1, 2018

DOI: 10.3892/mmr.2018.8909

**Abstract.** From a clinical point of view, easily obtainable and useful markers of a particular pathological status are required for appropriate diagnosis and treatment. Analysis of the proteomic profile of saliva may allow for the selection of potential marker of preterm delivery in humans. Saliva samples were collected from 12 patients diagnosed with threatened preterm delivery and 10 controls with uncomplicated pregnancies at the same gestational age. Samples were analysed using 2D electrophoresis. Based on statistical analysis, spots of interest were selected and collected from gels. Subsequently, spots were decoloured and proteins were identified by mass spectrometry using the matrix assisted laser desorption ionization-time of flight technique. The results of identification were compared with the Swiss-Prot database. A total of 1,393 spots were detected in the present study with 59 significantly different between control and preterm samples. Increased intensity of staining of 32 spots was observed in the premature delivery group compared with control patients and 27 spots were stained more intensely in the control group compared with the premature delivery group. A total of nine spots, which were significantly different between examined samples were identified and three of them exhibited increased intensity of staining in premature delivery group compared with controls, including dedicator of cytokinesis protein 1, metallothionein-2, guanylyl cyclase-activating protein 1. The six remaining spots included, epithelial-stromal interaction protein 1, serum albumin, tyrosine-tRNA ligase, cytoplasmic, protein chibby homolog 3, leukemia inhibitory factor receptor

and adenosylhomocysteinase 3, and exhibited increased intensity of staining in healthy controls compared with premature delivery group. Further studies with an increased number of patients and identification of the complete protein profile are required to confirm the results of the present study and applicability of saliva as a source of disease biomarkers.

## Introduction

Saliva is an easily obtainable biological fluid, which can reflect body metabolism (1). Certain components of saliva are secreted by salivary glands, while other components originate from plasma. Of human saliva proteins, 30% are of blood origin and the majority of them exhibit antimicrobial activities, indicating that composition of saliva is dependent on the composition of plasma (2). Analysis of human saliva can be used as a diagnostic tool to detect markers of diseases and physiological states (3).

Certain reports have stated that several parameters in human saliva resemble those in plasma. Öztürk *et al* (4) examined lipid peroxidation intensity during human gestation and postpartum, and identified an association between the concentration of examined parameters in plasma and saliva. Salivary glands express oestrogen receptors and therefore, pregnancy may affect the composition of saliva (5).

Preterm delivery is defined as parturition before the 37th week of gestation and is the most common cause of perinatal morbidity and mortality. A general hormonal profile of patients in the periparturient period has already been described, however detailed mechanisms underlying the initiation of delivery remain unclear (6). Elucidation and annotation of protein profile in plasma and in saliva may be useful for understanding the physiological processes underlying delivery and potential pathologies (7). There are several reports describing plasma and placental protein profile during human gestation (8-10); however, data regarding salivary proteins are scarce.

Menon *et al* (11), reviewed several co-regulated processes associated with delivery, including the study of foetal membranes, maternal signals, maturation of foetal organs and hormonal profile. The study emphasized the role of proteins in the aforementioned processes associated with delivery (11).

---

*Correspondence to:* Dr Andrzej Miturski, Department of Obstetrics and Pathology of Pregnancy, Medical University of Lublin, Staszica 16, 20-081 Lublin, Poland  
E-mail: a.miturski@gmail.com

\*Contributed equally

**Key words:** premature delivery, saliva, proteins, preterm delivery, proteomic profile

From a clinical point of view, it is necessary to develop a rapid and accurate method for predicting the time of delivery, and to identify women at risk for spontaneous preterm birth or other alterations during pregnancy. The aim of the present study was to evaluate protein profile in saliva from patients with uncomplicated pregnancy and delivery at term, and to compare the profile with that from premature delivery.

## Materials and methods

**Patients.** The present study included 22 pregnant women receiving treatment and delivering between February 2015 and November 2016 at the Department of Obstetrics and Pathology of Pregnancy at the Independent Clinical Hospital No. 1 in Lublin, Medical University of Lublin (Lublin, Poland). The study was approved by The Ethics Committee of the Medical University of Lublin (KE-0254/77/2014).

The study group [partus praematurus imminens (PPI)] included 12 patients with preterm uterine contractions and diagnosis of threatened preterm delivery. Iatrogenic preterm birth occurred in 1 patient as a result of foetal distress and 3 patients delivered at term despite symptoms of threatened preterm delivery. These 4 patients were excluded from further analysis. Saliva was collected at the time of admission to the Independent Clinical Hospital No 1 in Lublin, Poland.

The control group (C) included 10 patients with uncomplicated pregnancies. During routine control visits between 27-32 weeks of gestation in the outpatients clinic, saliva was collected from each patient. One patient from the control group declined to continue participation in the study and another delivered in a different hospital; thus, these patients were excluded from the study. Therefore, data from 16 women were further analysed, including 8 C group patients and 8 PPI patients. Inclusion criteria was as follows: Maternal age, >18; delivery prior to 35 weeks of gestation; intact membranes; and regular uterine contractions with cervical effacement  $\geq 50\%$ . Exclusion criteria were as follows: Multiple pregnancy; no confirmation of gestational age by ultrasound examination performed in the first trimester; iatrogenic preterm birth; and smoking during pregnancy. Written informed consent was obtained from all participants at the time of enrolment. Participation in the study was voluntary. The characteristics of study participants are presented in Table I. Mean gestational age at the time of sample collection in both groups was  $31.5 \pm 2.8$  weeks. Saliva was collected in the morning prior to eating.

**Preparation of saliva.** Saliva samples were collected in Salivette tubes (Sarstedt, Inc., Newton, NC, USA). Samples were centrifuged for 15 min at  $5,000 \times g$  in  $4^\circ\text{C}$ . Protease inhibitor cocktail (10  $\mu\text{l}$ ; cat. no. I3911; Sigma-Aldrich; Merck KGaA, Darmstadt, Germany) was added to 1,000  $\mu\text{l}$  of supernatant. The samples were stored at  $-20^\circ\text{C}$  prior to further analysis.

Then thawed saliva (700  $\mu\text{l}$ ) was concentrated to  $\sim 50 \mu\text{l}$  using Vivaspin columns for ultrafiltration (polyethersulfone membrane, molecular weight cutoff 3 kDa; Sartorius AG, Göttingen, Germany), washed twice with 100  $\mu\text{l}$  7.0 pH 20 mM PBS and suspended in 330  $\mu\text{l}$  rehydration buffer

containing 8 M urea, 2% CHAPS, 50 mM dithiothreitol (DTT), 0.2% Bio-Lyte 3/10 ampholyte and 0.001% bromophenol blue.

**Electrophoresis.** Isoelectric focusing was performed using strips with linear gradient of pH 3-10 and length of 17 cm. Each saliva sample was loaded onto one strip. Preparation of SDS-PAGE (16x20 cm; 1.0 mm; T=11%, C=2.6%) was performed as previously described by Laemmli (12). The second dimension was performed using a Protean II XL cell (Bio-Rad Laboratories, Inc., Hercules, CA, USA) according to the manufacturer's protocol. Gels were stained with silver nitrate according to MS compatible protocol (13).

**Identification of spots.** Selected spots of interest (27 spots) were removed from the gels manually (in accordance to significance and intensity of staining), cut into small pieces and transferred to 0.5 ml Eppendorf tubes. The gel pieces were subsequently washed three times with 100  $\mu\text{l}$  100 mM  $\text{NH}_4\text{HCO}_3$  buffer (pH 8.5; Sigma-Aldrich; Merck KGaA) for 5 min. Gel pieces were dehydrated by adding 100  $\mu\text{l}$  acetonitrile (ACN) and dried for 15 min at room temperature in vacuum concentrator CentriVap with 220 x g (Labconco Corporation, Kansas City, MO, USA). Samples were resuspended in 100  $\mu\text{l}$  of 10 mM DTT in 50 mM  $\text{NH}_4\text{HCO}_3$  buffer in order to perform reduction at  $56^\circ\text{C}$  for 60 min. Following cooling to room temperature, the solution was replaced by 100  $\mu\text{l}$  of 50 mM iodoacetamide in 50 mM  $\text{NH}_4\text{HCO}_3$  buffer and samples were incubated in the dark for 45 min at room temperature. Trypsin digestion of proteins was carried out on ice by gradual addition of 10  $\mu\text{l}$  12.5 ng/ml enzyme solution (Trypsin Gold; mass spectrometry grade; Promega Corporation, Madison, WI, USA) in 50 mM  $\text{NH}_4\text{HCO}_3$  buffer until proteins were completely rehydrated. Finally, 30  $\mu\text{l}$  50 mM  $\text{NH}_4\text{HCO}_3$  buffer was added to each sample and digestion reaction was allowed to proceed at  $37^\circ\text{C}$  overnight. Following digestion, the supernatant was collected and peptides were extracted three times with 50  $\mu\text{l}$  70% ACN and 1.5% trifluoroacetic acid (TFA) by sonication for 15 min at room temperature in an ultrasonic water bath (Ultron U-507; Ultron, Dywity, Poland). The supernatant was collected and dried in the CentriVap (Labconco Corporation) for 45 min at  $40^\circ\text{C}$  and 220 x g.

The obtained pellet of peptides was resuspended in 10  $\mu\text{l}$  0.1% TFA and purified with  $\mu\text{C}18$  ZipTip columns (Eppendorf, Hamburg, Germany) in accordance with the manufacturer's protocol. After purification, 1  $\mu\text{l}$  peptide solution (1.5% TFA, 30% ACN) was applied to  $\alpha$ -cyano-4-hydroxycinnamic acid-Pre spotted AnchorChip frame (Bruker Corporation, Billerica, MA, USA) and left to dry at room temperature. Mass spectra were obtained using Ultraflex III matrix-assisted laser desorption/ionization time-of-flight/time-of-flight spectrometer (MALDI; Bruker Corporation). Acquisition was performed in positive ion reflector mode with a 25 kV acceleration voltage. External calibrations were prepared using the peptide calibration standard (Bruker Corporation). FlexAnalysis 3.0 software (Bruker Corporation) was used for selection of monoisotopic peptide masses.

Identification of peptides and proteins from mass spectrometry data was performed using Mascot algorithm

Table I. Characteristics of the patients and controls.

Characteristic	Experimental group (n=8)	Control group (n=8)
Mean maternal age (years)	27.8±3.6	28.4±5.8
Mean gestational age at delivery (weeks)	31.5±2.8	39.2±1.3
Number primiparous	5 (62%)	3 (37%)
Mean birth weight (g)	1952±980	3608±505
Maternal BMI (kg/m <sup>2</sup> )	28.4±7.2	29.6±6.8

Values are presented as the mean ± standard deviation. BMI, body mass index.

(Mascot 2.2 software; Matrix Science, Ltd., London, UK) and Swiss-Prot database (UniProt release 2016\_07; uniprot.org/statistics/Swiss-Prot) restricted to the human taxonomy. Search parameters were set in the following mode: Enzyme-trypsin, up to 1 missed cleavages, fixed modification-carboamidomethylation cysteine, variable modification-oxidation of methionine, error of 50 ppm. Analyses and comparisons with the database were made by the BioTools 3.2 software (Bruker Corporation).

**Statistical analysis.** Delta2D software (version 4.6; Decodon GmbH, Greifswald, Germany) was used to differentiate the presence of particular spots and intensity of staining within examined saliva samples using one-way analysis of variance, as previously described (14). As a result, a list of most common and differential spots between groups C and PPI ( $P < 0.05$ ) was created and spots for further analysis were selected. A principal components analysis scatterplot (PCA 3D) of all spots was constructed using Delta2D software (15).

Spot selection for identification was based on the presence of spots in one or both of the examined groups as well as their intensity of staining that reflects the amount of protein necessary for further identification analysis (highly visible spots with amount of protein more than 5-10 ng). The number of analysed saliva samples followed proteomic requirements (16,17). The specificity and sensitivity of used laboratory methods (16,17) allowed for reliable statistical analysis. Data is presented as the mean ± standard deviation.  $P < 0.05$  was considered to indicate a statistically significant difference.

The protein identification of spots was based on the comparison with UniProt databases and confirmed by Mascot scores ([http://www.matrixscience.com/help/scoring\\_help.html](http://www.matrixscience.com/help/scoring_help.html)). Scores  $> 67$  were considered to indicate a statistical significance.

## Results

Out of 1,393 detected spots, 59 were significantly different between the C and PPI groups. Statistical analysis demonstrated that 32 spots exhibited increased intensity of staining in PPI group samples compared with the C group, while 27

had the opposite trend (Table II). Out of 59 spots that differed significantly between examined groups 9 were identified and are shown in Table III. Three spots exhibited increased intensity of staining in the PPI group compared with the C group: Dedicator of cytokinesis protein 1 (DOCK1), metallothionein-2 (MT2A) and guanylyl cyclase-activating protein 1 (GUCA1A). Six spots exhibited an opposite pattern in PPI and C groups: Epithelial-stromal interaction protein 1 (EPSTI1), serum albumin, tyrosine--tRNA ligase, cytoplasmic (YARS), protein chibby homolog 3, leukemia inhibitory factor receptor (LIFR) and adenosyl-homocysteinase 3. DOCK1 is a haemostasis-associated protein, and GUCA1A and EPSTI1 are proteins associated with programmed cell death. YARS is associated with gene expression and LIFR has a role in the immune system. MT2A involved in antioxidative defence.

In addition, 18 spots that did not differ in intensity of staining between groups were identified. The identified proteins included, carboxypeptidase E (Cpe), fucose-1-phosphate guanylyltransferase (FKGP), E3 ubiquitin-protein ligase TRIM56 (TRIM56), myosin light chain kinase 2, skeletal/cardiac muscle (Myk2), CMP-N-acetylneuraminic- $\beta$ -galactosamide- $\alpha$ -2,3-sialyltransferase 4 (ST8SIA4), methyltransferase-like protein 13 (METTL13) and pancreatic lipase-related protein 3 (PNLIPRP3). The above proteins have roles in cell metabolism, but are not associated with pregnancy and were not considered as clinically significant.

Fig. 1 presents fused image of spots from both examined groups, which were overlapped by use of Delta2D software. Orange spots are from C group, blue from PPI group while common spots for both groups are marked with black. The figure demonstrates the profile and location of the spots distributed in accordance to the isoelectric point and molecular weight of proteins, which were in examined the biological fluids.

Fig. 2 presents PCA scatterplot from C group marked with green area and PPI group marked with red area. The location of areas represents the association between the obtained spots and indicates that the majority of proteins are similar in examined samples as they overlay.

Fig. 3 represents histograms of all spots that differ significantly in accordance to intensity of staining what indirectly reflects the amount of protein. All 59 pairs are significantly different between each other at ( $P < 0.05$ ). Y-axis reflects the mean spot volume (intensity of staining x spot area). X axis-experimental groups.

## Discussion

The present study compared the proteomic pattern of saliva from patients that delivered at term, with those that delivered prematurely. Statistical analysis enabled identification of the spots that differed significantly in intensity of staining between the examined groups of patients. As the intensity of staining reflects the amount of protein in spot, this analysis indicated quantitative differences in saliva protein profile between examined spots. The majority of detected spots expressed similar intensity. Proteins corresponding to certain spots were identified.

Table II. Statistical analysis of significantly different (P&lt;0.05) spots between examined groups of patients, C and PPI.

Spot label	PPI mean	Control mean	C/PPI ratio	P-value
1	0.053	0.001	0.027	0.024
2	0.045	0.002	0.035	0.021
3	0.011	0.002	0.181	0.015
4	0.024	0.005	0.225	0.024
5	0.015	0.002	0.118	0.034
6	0.145	0.219	1.509	0.048
7	0.307	0.597	1.948	0.031
8	0.107	0.283	2.639	0.045
9	0.025	0.005	0.189	0.004
10	0.015	0.003	0.197	0.011
11	0.064	0.229	3.574	0.011
12	0.207	0.364	1.759	0.048
13	1.233	2.411	1.956	0.023
14	0.068	0.235	3.434	0.016
15	0.009	0.002	0.184	0.044
16	0.01	0.002	0.174	0.03
17	0.032	0.003	0.105	0.015
18	0.1	0.037	0.368	0.024
19	0.05	0.139	2.793	0.028
20	0.028	0.008	0.273	0.033
21	0.014	0.062	4.396	0.019
22	0.037	0.011	0.307	0.014
23	0.017	0.053	3.071	0.028
24	0.015	0.002	0.14	0.021
25	0.008	0.059	7.127	0.012
26	0.096	0.025	0.262	0.016
27	0.818	0.311	0.38	0.003
28	0.335	0.042	0.125	0.036
29	1.116	0.542	0.485	0.026
30	0.023	0.002	0.099	0.044
31	0.118	0.318	2.681	0.017
32	0.009	0.002	0.238	0.041
33	0.011	0.001	0.119	0.042
34	0.027	0.003	0.13	0.0005
35	0.021	0.005	0.215	0.044
36	0.084	0.254	3.03	0.006
37	0.347	1.062	3.06	0.002
38	0.018	0.074	4.14	0.024
39	0.135	0.035	0.26	0.047
40	0.013	0.003	0.218	0.043
41	0.009	0.025	2.606	0.03
42	0.31	0.975	3.144	0.009
43	0.017	0.003	0.18	0.022
44	0.021	0.005	0.254	0.039
45	0.02	0.086	4.383	0.006
46	0.058	0.138	2.386	0.007
47	0.07	0.862	12.254	0.01
48	0.475	0.069	0.144	0.016
49	0.103	0.357	3.473	0.0005
50	0.045	0.006	0.133	0.049

Table II. Continued.

Spot label	PPI mean	Control mean	C/PPI ratio	P-value
51	0.046	0.011	0.244	0.026
52	0.776	1.185	1.526	0.045
53	0.143	0.363	2.539	0.005
54	0.049	0.019	0.384	0.041
55	0.056	0.175	3.13	0.006
56	0.04	0.127	3.144	0.013
57	0.207	0.692	3.338	0.026
58	0.029	0.134	4.617	0.025
59	0.105	0.024	0.228	0.001

Ratio >1 indicates increased abundance of a spot in the PPI group compared with the C group. C, control; PPI, partus praematurus imminens. Mean values are obtained from densitometric analysis of gels from all C and PPI patients in duplicate-mean spot volume (intensity of staining x spot area). Table corresponds with Figure 3.

A preliminary report featuring a plasma protein profile in humans during pregnancy was published in 1978 by Joseph *et al* (8). Term 2D protein profile of human placenta was described by Mushahary *et al* (9). Pregnancy-associated proteins, which have a role in maintenance of human placenta include proteins detected in maternal serum, foetal proteins and soluble, solubilized or membrane-associated placental proteins (18).

Yuan *et al* (10) described protein profile of human placental blood plasma obtained from patients that underwent elective caesarean section (not in labour), spontaneous vaginal delivery, induction of labour vaginal delivery, spontaneous delivery, emergency caesarean section and induction of delivery. Apolipoproteins A-IV, E and C-III were considered to be associated with the onset of delivery while hepatocyte nuclear factor 1 homeobox A was common for human parturition regardless of type of delivery (10).

Preterm delivery may cause perinatal mortality and morbidity, and is defined as birth prior to 37 weeks of gestation (19). Although several factors are known to be involved in spontaneous preterm termination of pregnancy, biochemical markers of preterm delivery remain to be elucidated. Previous studies indicated that intrauterine inflammation may be one of the initiating factors of preterm delivery (20,21). There are several other possible factors associated with this pathological condition, including uteroplacental ischemia, decidual haemorrhage, failure in the maternal immunological tolerance of the foetus, allergies, abnormal uterine size, cervical incompetence, maternal and foetal stress and hormonal irregularities (22).

Numerous studies associated with preterm birth have demonstrated that alterations in the expression of particular molecules are associated with certain disturbances but none of them were considered diagnostic tools and markers (19,21-24). These molecules include membrane and soluble proteins present or absent in the placental tissue of patients with preterm birth (23). Proteins that are only

Table III. The list of identified spots in saliva of the examined groups of patients.

Spot label	Protein	UniProt entry	Mascot Score	Protein sequence coverage (%)	Molecular weight (kDa)	pI
6	Epithelial-stromal interaction protein 1	Q96J88 (ESIP1_HUMAN)	58	16	36.80	9.90
12	Serum albumin	P02768 (ALBU_HUMAN)	61	21	69.37	5.92
18	Dedicator of cytokinesis protein 1	Q14185 (DOCK1_HUMAN)	87	8	21.53	7.29
19	Tyrosine-tRNA ligase, cytoplasmic	P54577 (SYYC_HUMAN)	80	18	59.14	6.61
21	Protein chibby homolog 3	A6NI87 (CBY3_HUMAN)	59	33	27.34	10.65
22	Metallothionein-2	P02795 (MT2_HUMAN)	65	54	6.04	8.23
23	Leukemia inhibitory factor receptor	P42702 (LIFR_HUMAN)	133	16	123.74	5.50
25	Adenosylhomocysteinase 3	Q96HN2 (SAHH3_HUMAN)	160	19	66.72	7.13
29	Guanylyl cyclase-activating protein 1	P43080 (GUC1A_HUMAN)	62	57	22.92	4.34

pI, isoelectric point. Mascot Scores greater than 67 were considered significant at ( $P < 0.05$ ).

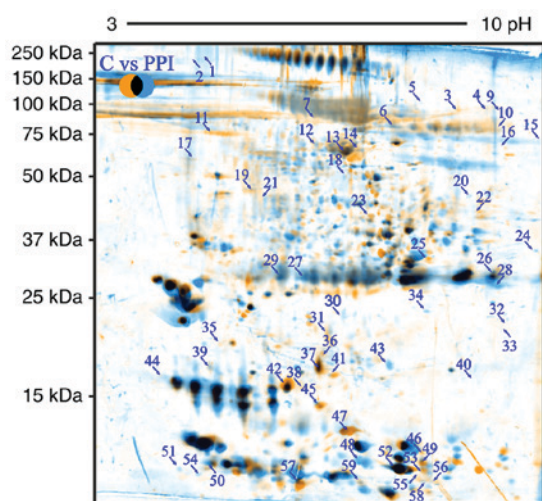


Figure 1. Merged images of spots from healthy individuals (orange) and patients with PPI (blue) and spots identified in both groups (black). Spots which were statistically significant and identified (based on intensity of staining) are marked with arrows and correspond with numbers in Tables II and III. Picture represents overlapped samples from all C and PPI patients in duplicate. Gel was stained with silver nitrate but Delta2D software allowed to distinguish between spots which are common for both examined groups and individual for C group or PPI group and mark them in colour separately. PPI, partus praematurus imminens.

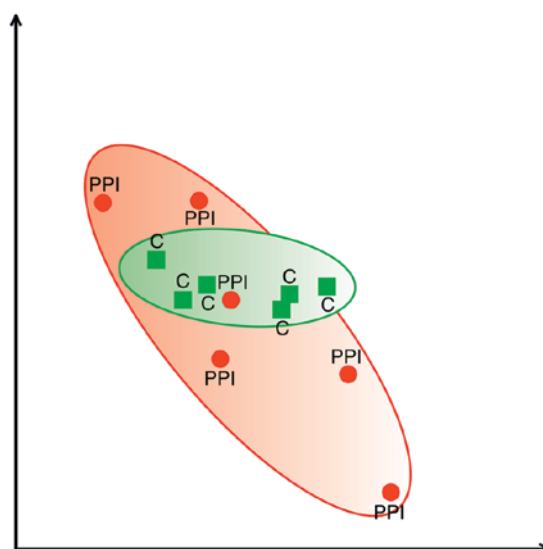


Figure 2. PCA scatterplot presenting analysis of spots from PPI group (red area) and C group (green area). Statistical analysis of association between spots obtained from examined groups of patients (PCA 3D) (15). The range of overlapping areas represent the differences between examined groups of patients. The picture was created by Delta2D software based on statistical analysis (15). C, control; PPI, partus praematurus imminens; PCA, principal components analysis.

detectable in preterm placenta compared with placenta from full-term births include, vimentin,  $\beta$ -actin and  $\gamma$ -actin as representatives of structural proteins, and soluble proteins, including lactoylglutathione lyase, transgelin and 78 kDa glucose-regulated protein. Preterm labour is also characterized by lack of certain placental proteins, which are present in normal term placenta. These include membrane proteins annexin A4 and type I keratin, and proteins present in the soluble fraction, including endoplasmic, and  $\beta$ - and  $\gamma$ -actin (24).

Elevated levels of MT2A in saliva of PPI patients compared with C patients detected in the present study may indicate oxidative stress (25). Oxidative stress alters protein molecules via peroxidative damage and complicate preterm births (26).

These results are supported by the present study where, LIFR, which has a role in ovulation, implantation and fertility, was downregulated in the PPI group compared with the C group, suggesting potential alterations in protein metabolism.

Identification of proteins that exhibit similar patterns of expression in the examined saliva may aid in elucidating the sequence of events that leads to premature delivery. Molecules identified in the present study belong to different functional groups, including enzymes, and may indicate the presence of reactions that take place regardless of pregnancy-associated alterations and appear to be necessary both in physiological and pathological conditions. Among identified proteins were enzymes, including, Cpe, FKBP, TRIM56, Mylk2, ST8SIA4, METTL13 and PNLIPRP3.

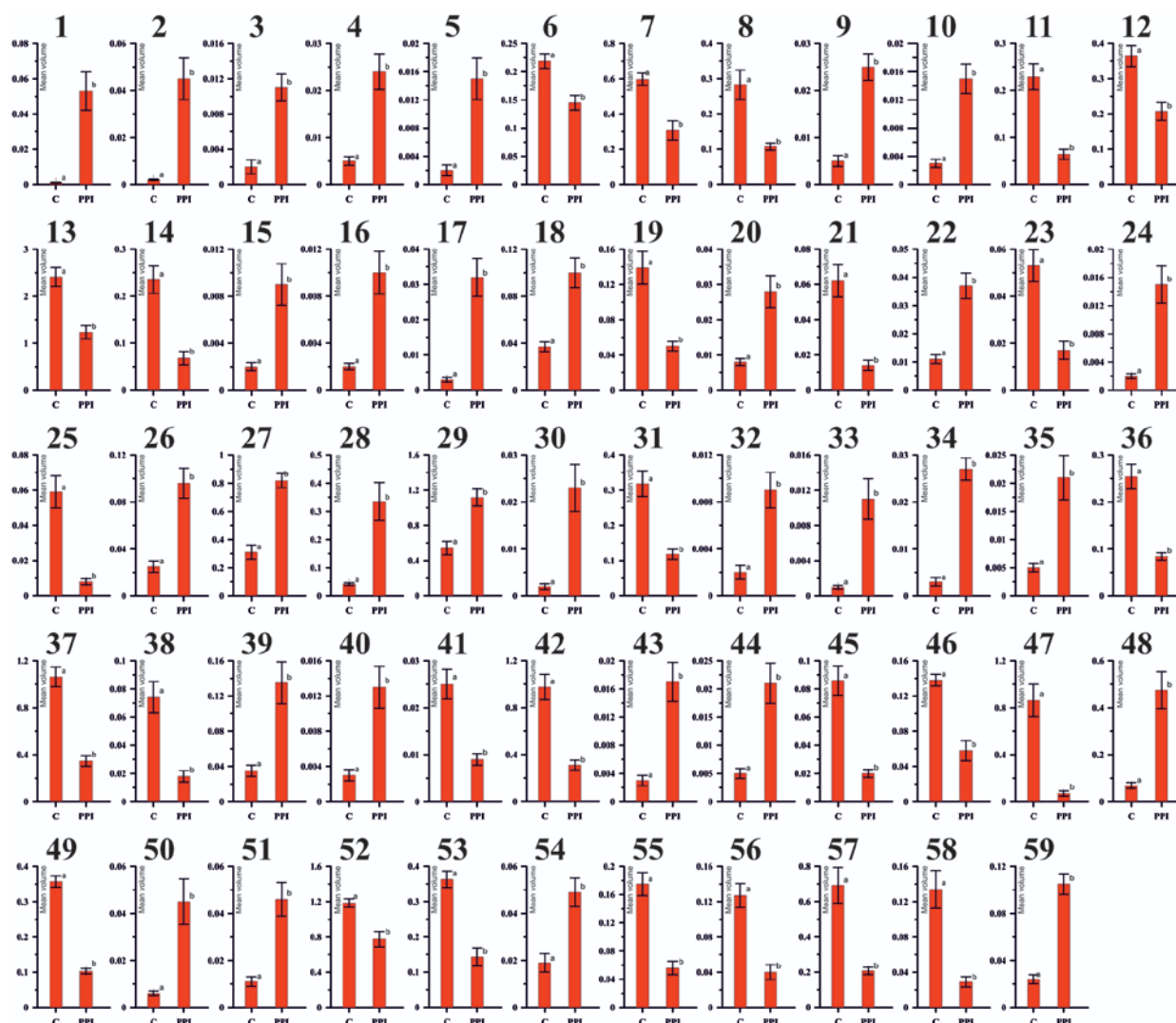


Figure 3. Histograms of proteins that differ in intensity of staining between C and PPI groups. The intensity of staining was analysed densitometrically during the analysis of gels by Delta2D software. Y-axis represents the mean spot volume (intensity of staining x spot area) while X-axis experimental groups. All pairs differ significantly at  $^aP < 0.05$  vs. PPI,  $^bP < 0.05$  vs. C. Detailed statistical analysis is in Table II. C, control; PPI, partus praematurus imminens.

As described in the review by Menon *et al* (11), the onset of parturition requires a complex and simultaneous transduction of several signals from the foetus, foetal membranes and the mother via local and general systems. Certain proteins may have roles in modulation of reactions during delivery in physiological and pathological conditions. Previous studies indicated that saliva may be a source of parameters of diagnostic use in diseases of the oral cavity, diabetes or breast cancer; however, the use of saliva samples for diagnostics in obstetrics requires further investigation (27-29). The present investigated saliva in the context of threatened preterm delivery.

In conclusion, although saliva is an easy obtainable biological fluid, its use in diagnostic procedures is currently limited, due to the lack of information regarding its content. The results of the present study suggested that MT2A in saliva, which has a role in oxidative stress, may be a potential marker indicating risk of premature delivery. Further studies with an increased number of patients are necessary for verification and validation of the results. The present study suggests a potential for

targeted proteomic analysis of saliva to identify biomarkers that may predict premature delivery and other alterations associated with the periparturient period.

#### Acknowledgements

Not applicable.

#### Funding

The present study was supported by Statutory Activity of University of Life Sciences in Lublin (WKB-DS-4) and Statutory Activity of Medical University of Lublin (DS/122/2015-2017).

#### Availability of data and materials

All data generated or analyzed during this study are included in this published article.

### Authors' contributions

MŁ and MK conceived the presented idea. TG and AM developed the theory and performed the computations. MŁ and MK verified the analytical methods. JW and MF expressed and purified all proteins. TG and AM analysed the data. JW designed the figures. MŁ, TG and AM supervised the findings of this work. All authors discussed the results and contributed to the final manuscript.

### Ethics approval and consent to participate

Written informed consent was obtained from all participants at the time of enrolment. The study was approved by the Ethics Committee of the Medical University of Lublin (KE-0254/77/2014).

### Consent for publication

Informed patient consent was obtained from all participants at the time of enrolment.

### Competing interests

The authors declare that they have no competing interests.

### References

- Lima DP, Diniz DG, Moimaz SA, Sumida DH and Okamoto AC: Saliva: Reflection of the body. *Int J Infect Dis* 14: e184-e188, 2010.
- Schulz BL, Cooper-White J and Punyadeera CK: Saliva proteome research: Current status and future outlook. *Crit Rev Biotechnol* 33: 246-259, 2013.
- Xiao H and Wong DT: Proteomics and its applications for biomarker discovery in human saliva. *Bioinformatics* 5: 294-296, 2011.
- Öztürk LK, Akyüz S, Yarat A, Koç S, Gül N and Doğan BN: Salivary lipid peroxidation and total sialic acid levels during healthy gestation and postpartum: A longitudinal study. *Clin Biochem* 43: 430-434, 2010.
- Valimaa H, Savolainen S, Soukka T, Silvoniemi P, Mäkelä S, Kujari H, Gustafsson JA and Laine M: Estrogen receptor-beta is the predominant estrogen receptor subtype in human oral epithelium and salivary glands. *J Endocrinol* 180: 55-62, 2004.
- Catalano RD, Lannagan TR, Gorowiec M, Denison FC, Norman JE and Jabbour HN: Prokineticins: Novel mediators of inflammatory and contractile pathways at parturition? *Mol Hum Reprod* 16: 311-319, 2010.
- Shankar R, Gude N, Cullinane F, Brennecke S, Purcell AW and Moses EK: An emerging role for comprehensive proteome analysis in human pregnancy research. *Reproduction* 129: 685-696, 2005.
- Joseph JC, Baker C, Sprang ML and Bermes EW: Changes in plasma proteins during pregnancy. *Ann Clin Lab Sci* 8: 130-141, 1978.
- Mushahary D, Gautam P, Sundaram CS and Sirdeshmukh R: Expanded protein expression profile of human placenta using two-dimensional gel electrophoresis. *Placenta* 34: 193-196, 2013.
- Yuan W, Heesom K, Phillips R, Chen L, Trinder J and López Bernal A: Low abundance plasma proteins in labour. *Reproduction* 144: 505-518, 2012.
- Menon R, Bonney EA, Condon J, Mesiano S and Taylor RN: Novel concepts on pregnancy clocks and alarms: Redundancy and synergy in human parturition. *Hum Reprod Update* 22: 535-560, 2016.
- Laemmli UK: Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature* 227: 680-685, 1970.
- Shevchenko A, Wilm M, Vorm O and Mann M: Mass spectrometric sequencing of proteins silver-stained polyacrylamide gels. *Anal Chem* 68: 850-858, 1996.
- Jerrold HZ: *Biostatistical Analysis*. 4th edition. Prentice Hall, New Jersey, 1999.
- Jolliffe IT: *Principal Component Analysis*. 2nd edition. Springer, 2002.
- Smit S, Hoefsloot HC and Smilde AK: Statistical data processing in clinical proteomics. *J Chromatogr B Analyt Technol Biomed Life Sci* 866: 77-88, 2008.
- Chich JF, David O, Villers F, Schaeffer B, Lutowski D and Huet S: Statistics for proteomics: Experimental design and 2-DE differential analysis. *J Chromatogr B Analyt Technol Biomed Life Sci* 849: 261-272, 2007.
- Mine K, Katayama A, Matsumura T, Nishino T, Kuwabara Y, Ishikawa G, Murata T, Sawa R, Otsubo Y, Shin S and Takeshita T: Proteome analysis of human placenta: Pre-eclampsia versus normal pregnancy. *Placenta* 28: 676-687, 2007.
- Parry S, Zhang H, Biggio J, Bukowski R, Varner M, Xu Y, Andrews WW, Saade GR, Esplin MS, Leite R, *et al*: Maternal serum serpin B7 is associated with early spontaneous preterm birth. *Am J Obstet Gynecol* 211: 678.e1-e12, 2014.
- Nelson DM, Sadovsky Y, Robinson JM, Croy BA, Rice G and Kniss DA: Advanced techniques in placental biology-workshop report. *Placenta* 27 (Suppl A): S87-S90, 2006.
- Kim MA, Lee YS and Seo K: Assessment of predictive markers for placental inflammatory response in preterm births. *PLoS One* 9: e107880, 2014.
- Pařízek A, Koucký M and Dušková M: Progesterone, inflammation and preterm labor. *J Steroid Biochem Mol Biol* 139: 159-165, 2014.
- Romero R, Espinoza J, Gotsch F, Kusanovic JP, Friel LA, Erez O, Mazaki-Tovi S, Than NG, Hassan S and Tromp G: The use of high-dimensional biology (genomics, transcriptomics, proteomics, and metabolomics) to understand the preterm parturition syndrome. *BJOG* 113: 118-135, 2006.
- Butt RH, Lee MW, Pirshahid SA, Backlund PS, Wood S and Coorsen JR: An initial proteomic analysis of human preterm labor: Placental membranes. *J Proteome Res* 5: 3161-3172, 2006.
- Vašák M and Meloni G: Chemistry and biology of mammalian metallothioneins. *J Biol Inorg Chem* 16: 1067-1078, 2011.
- Menon R: Oxidative stress damage as a detrimental factor in preterm birth pathology. *Front Immunol* 5: 567, 2014.
- Zhang A, Sun H, Wang P and Wang X: Salivary proteomics in biomedical research. *Clin Chim Acta* 415: 261-265, 2013.
- Al Kawas S, Rahim ZH and Ferguson DB: Potential uses of human salivary protein and peptide analysis in the diagnosis of disease. *Arch Oral Biol* 57: 1-9, 2012.
- Al-Tarawneh SK, Border MB, Dibble CF and Bencharit S: Defining salivary biomarkers using mass spectrometry-based proteomics: A systematic review. *OMICS* 15: 353-361, 2011.



This work is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International (CC BY-NC-ND 4.0) License.