

## Editorial

# Advances in Confocal Microscopy of the Eye

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Received 6 April 2016; Accepted 6 April 2016

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This special issue focuses on the recent advances on *in vivo* confocal microscopy (IVCM), a technique used to investigate eye structures at the cellular level without tissue damage.

In 1985 Lemp and coworkers were the first to study *ex vivo* corneas by confocal microscopy and to suggest a possible *in vivo* use [1]. In the early 1990s, the groups directed by Cavanagh and Kaufman published the first IVCM in humans; thereafter the scientific interest in ocular IVCM rapidly increased. In the last decade, IVCM progressively gained a relevant role in the clinical setting, being of help in the diagnosis and management of a number of conditions such as toxicity induced by preservatives [2, 3] and different eye treatments [4, 5], iatrogenic damage [6], infections [7], and dystrophies [8], pathology of the conjunctiva [9–11] and limbus [12], ocular surface tumors [13], and corneal deposits [14, 15].

IVCM is a valuable tool for enhancing our understanding of anterior segment physiology and pathology, as pointed out by Dr. V. Fasanella et al. in their paper investigating meibomian gland changes occurring with aging and a number of ocular surface diseases.

Examples of the usefulness of IVCM in detecting early corneal changes can be found in the paper by Dr. D. Wang et al., who highlighted that a higher-than-normal immune activity may be observed in a subgroup of patients with clear corneal grafts in the absence of any other clinical signs. Dr. P. Song et al. studied the confocal and histopathological features

of the stromal scar in keratoconus and suggested that IVCM is capable of detecting subtle keratocyte activation associated with fibrosis.

In recent years, IVCM has become increasingly useful in evaluating corneal innervation (which is of key importance in regulating ocular surface homeostasis) [16] and corneal immune and inflammatory responses [17]. In this special issue, three papers explored the changes occurring in corneal innervation in both local and systemic diseases. The paper by Dr. E. F. Wang et al. reviewed the impact of various systemic diseases on corneal innervation and discussed the potential use of IVCM as a noninvasive marker of peripheral neuropathy. The reports by Dr. R. Shetty et al. and Dr. N. K. Pahuja et al. investigated nerve changes occurring, respectively, in dry eye disease and keratoconus.

Finally, IVCM plays a role in studying the therapeutic effects of topical eye treatments, as shown in this issue by the paper of Dr. A. M. Fea et al.

This special issue provides a useful update on the advances in the rapidly evolving field of IVCM and highlights the contribution of this technology to our understanding of the anterior segment in health and disease.

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## References

- [1] M. A. Lemp, P. N. Dilly, and A. Boyde, "Tandem-scanning (confocal) microscopy of the full-thickness cornea," *Cornea*, vol. 4, no. 4, pp. 205–209, 1985.
- [2] M. Iester, F. Oddone, P. Fogagnolo, P. Frezzotti, and M. Figus, "Changes in the morphological and functional patterns of the ocular surface in patients treated with prostaglandin analogues after the use of TSP 0.5%® preservative-free eyedrops: a prospective, multicenter study," *Ophthalmic Research*, vol. 51, no. 3, pp. 146–152, 2014.
- [3] M. Iester, S. Telani, P. Frezzotti et al., "Ocular surface changes in glaucomatous patients treated with and without preservatives beta-blockers," *Journal of Ocular Pharmacology and Therapeutics*, vol. 30, no. 6, pp. 476–481, 2014.
- [4] P. Fogagnolo, A. Dipinto, E. Vanzulli et al., "A 1-year randomized study of the clinical and confocal effects of tafluprost and latanoprost in newly diagnosed glaucoma patients," *Advances in Therapy*, vol. 32, no. 4, pp. 356–369, 2015.
- [5] P. Fogagnolo, M. Sacchi, G. Ceresara et al., "The effects of topical coenzyme Q10 and vitamin e d- $\alpha$ -tocopheryl polyethylene glycol 1000 succinate after cataract surgery: a clinical and in vivo confocal study," *Ophthalmologica*, vol. 229, no. 1, pp. 26–31, 2013.
- [6] S. De Cillà, P. Fogagnolo, M. Sacchi et al., "Corneal involvement in uneventful cataract surgery: an in vivo confocal microscopy study," *Ophthalmologica*, vol. 231, no. 2, pp. 103–110, 2014.
- [7] M. Randon, H. Liang, M. El Hamdaoui et al., "In vivo confocal microscopy as a novel and reliable tool for the diagnosis of Demodex eyelid infestation," *British Journal of Ophthalmology*, vol. 99, no. 3, pp. 336–341, 2015.
- [8] D. V. Patel, C. N. Grupcheva, and C. N. J. McGhee, "Imaging the microstructural abnormalities of Meesmann corneal dystrophy by in vivo confocal microscopy," *Cornea*, vol. 24, no. 6, pp. 669–673, 2005.
- [9] P. Frezzotti, P. Fogagnolo, G. Haka et al., "In vivo confocal microscopy of conjunctiva in preservative-free timolol 0.1% gel formulation therapy for glaucoma," *Acta Ophthalmologica*, vol. 92, no. 2, pp. e133–e140, 2014.
- [10] A. Labbé, B. Dupas, P. Hamard, and C. Baudouin, "In vivo confocal microscopy study of blebs after filtering surgery," *Ophthalmology*, vol. 112, no. 11, article 1979, 2005.
- [11] A. Labbé, L. Gheck, V. Iordanidou, C. Mehanna, F. Brignole-Baudouin, and C. Baudouin, "An in vivo confocal microscopy and impression cytology evaluation of pterygium activity," *Cornea*, vol. 29, no. 4, pp. 392–399, 2010.
- [12] D. V. Patel, T. Sherwin, and C. N. J. McGhee, "Laser scanning in vivo confocal microscopy of the normal human corneoscleral limbus," *Investigative Ophthalmology and Visual Science*, vol. 47, no. 7, pp. 2823–2827, 2006.
- [13] E. E. Gabison, A. Labbé, F. Brignole-Baudouin et al., "Confocal biomicroscopy of corneal intraepithelial neoplasia regression following interferon alpha 2b treatment," *British Journal of Ophthalmology*, vol. 94, no. 1, pp. 134–135, 2010.
- [14] G. Ceresara, P. Fogagnolo, S. De Cillà et al., "Corneal involvement in crohn's disease: an in vivo confocal microscopy study," *Cornea*, vol. 30, no. 2, pp. 136–142, 2011.
- [15] G. Ceresara, P. Fogagnolo, M. Zuin, S. Zatelli, J. Bovet, and L. Rossetti, "Study of corneal copper deposits in wilson's disease by in vivo confocal microscopy," *Ophthalmologica*, vol. 231, no. 3, pp. 147–152, 2014.
- [16] S. Allgeier, A. Zhivov, F. Eberle et al., "Image reconstruction of the subbasal nerve plexus with in vivo confocal microscopy," *Investigative Ophthalmology & Visual Science*, vol. 52, no. 9, pp. 5022–5028, 2011.
- [17] W. J. Mayer, M. J. Mackert, N. Kranebitter et al., "Distribution of antigen presenting cells in the human cornea: correlation of in vivo confocal microscopy and immunohistochemistry in different pathologic entities," *Current Eye Research*, vol. 37, no. 11, pp. 1012–1018, 2012.