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Modifying the tumour microenvironment and reverting tumour cells: New strategies for treating malignant tumours

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Abstract

The tumour microenvironment (TME) plays a pivotal role in tumour fate determination. The TME acts together with the genetic material of tumour cells to determine their initiation, metastasis and drug resistance. Stromal cells in the TME promote the growth and metastasis of tumour cells by secreting soluble molecules or exosomes. The abnormal microenvironment reduces immune surveillance and tumour killing. The TME causes low anti-tumour drug penetration and reactivity and high drug resistance. Tumour angiogenesis and microenvironmental hypoxia limit the drug concentration within the TME and enhance the stemness of tumour cells. Therefore, modifying the TME to effectively attack tumour cells could represent a comprehensive and effective anti-tumour strategy. Normal cells, such as stem cells and immune cells, can penetrate and disrupt the abnormal TME. Reconstruction of the TME with healthy cells is an exciting new direction for tumour treatment. We will elaborate on the mechanism of the TME to support tumours and the current cell therapies for targeting tumours and the TME-such as immune cell therapies, haematopoietic stem cell (HSC) transplantation therapies, mesenchymal stem cell (MSC) transfer and embryonic stem cell-based microenvironment therapies-to provide novel ideas for producing breakthroughs in tumour therapy strategies.

1 | INTRODUCTION

Tumour incidence and mortality are increasing yearly, with particularly rising trends in younger populations.¹ In 2018, 18.1 million new tumour cases were reported worldwide, and 9.6 million people died from tumours, making them one of the greatest threats to human health.²

The generation and development of tumours were previously believed to depend on only tumour suppressor or oncogene mutations, the basis of the "tumour-centric" view.³ Therapies derived from this theory, whether drugs, surgeries or radiation therapies, are all based on killing tumour cells with inevitable secondary damage

and increasing treatment resistance. Researchers have found that the tumour microenvironment (TME) plays a pivotal role in the generation, progression and metastasis of tumours. A century ago, Stephen Paget found that breast cancer metastasis displayed organ (tissue) preference, which related to the cell environment of the targeted organ (tissue). He boldly assumed that tumour progression is controlled by the interaction of tumour cells and the external environment, and first proposed the concept of the TME.⁴ Various components of the TME constitute an intricate network that precisely regulates tumour fate and the interactions of tumour cells with other components. This enables tumour cells to steadily proliferate, resist apoptosis, escape from immune elimination, maintain stemness and

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metastasize to distant sites. The TME theory superseded the theory that the fate of tumour cells is determined only by their genetic material and provided a new perspective for comprehensively understanding tumour metastasis and drug resistance mechanisms.

Traditional anti-tumour chemoradiotherapy is strongly cytotoxic because it denatures nucleic acids and proteins in tumour cells; however, this also results in damage to normal cells and causes serious adverse reactions, even secondary tumour formation.⁵⁻⁸ Tumour cells escape apoptosis by constantly generating new gene mutations that mediate tumour drug resistance. To solve the problem of the poor specificity of chemoradiotherapy, targeted therapies and immune therapies have been developed.⁹ Although immune therapies, such as anti-programmed death 1(PD-1)/PD-L1 treatment, show considerable efficacy in several tumours, they still have individual specificity. Meanwhile, the high incidence of severe autoimmune adverse reactions after immune therapy poses a new threat to patients' lives.¹⁰⁻¹³

With the gradual deepening of understanding of TME, targeting TME compounds to undermine protecting hotbed of tumours have become an effective means of cancer treatment. Large amount of pre-clinic and clinic study proved the quietly success in targeting angiogenesis, extracellular matrix (ECM) and cells components within TME.¹⁴ In recent years, cell therapies are fast rising and have been proven to have powerful functions and ensured safety. Compared with the single role of drug, cells may act on TME from multi-angle and through many ways at one time due to its better plasticity. It is manifested that cell therapies can inhibit or reverse tumours for which there is currently no effective therapy. We suggest that utilizing a therapeutic cell's own microenvironment to regulate and modify the TME, thereby destroying the tumour nests that tumour cells depend on for survival, constitutes a new direction for tumour treatment. We will elaborate on the current therapies, especially cell therapies, for targeting tumours and the TME-such as immune cell therapies, stem cell replacement therapies (mainly used for bone marrow-derived tumours), mesenchymal stem cell (MSC) transfer and embryonic stem cell-based microenvironment therapies-to provide novel ideas for the optimization of tumour therapy strategies.

2 | COMPOSITION AND FUNCTION OF THE TME

The components surrounding the tumour cells constitute the functional TME in which tumour cells initiate and grow and from which they invade and metastasize. The TME is a sophisticated network that includes various tumour-associated cells, such as cancer-associated fibroblasts (CAFs), cancer stem cells (CSCs), MSCs, tumourassociated immune and inflammatory cells, pluripotent stromal cells, cancer-associated adipocytes, pericytes and endothelial cells (ECs). The tumour-associated cells secrete tumour-associated exosomes (TEXs) and soluble molecules, including cytokines, kinases, protein, transcription factors, growth factors, hormones and free radicals. They regulate and respond to each other, construct the scaffold of the tumour ECM, and are nourished by tumour angiogenesis, manifesting as a special niche for innutrition, acidity, hypoxia and ischaemia.^{15,16} The TME has a degree of tumour-derived organ and tissue specificity, reviews of Schumacher et al¹⁷ also summarized that human tumours vary substantially in the composition of their microenvironment, and this is likely to regulate cancer cell morphology and influence the ability of the individual T-cell immunotherapy.

2.1 | The ECM forms the 3-dimensional tumour protective nest

The tumour ECM, comprising collagen, fibronectin, laminin, vitronectin and tenascin, is secreted by cells in the TME. Via autocrine and paracrine signalling, tumour cells and microenvironment cells, especially CAFs, transform the ECM to an advantageous phenotype for tumour progression.¹⁸ Collagen and fibronectin in the ECM provide physical support for tumour cells, and proteoglycans act as binding factors for growth factors and cytokines.¹⁶ Thus, the ECM not only provides a 3-dimensional structure for tumour growth, but, more importantly, it provides precise biomechanical and biochemical regulation for tumour cells.¹⁹

The alteration of the orderly isotropic arrangement of the collagen, called matrix remodelling, results in tumour-associated collagen signatures (TACS).²⁰ Different TACS stages correspond to different degrees of tumour progression. Over activated and transformed CAFs induce type I collagen aberrations, transforming type I TACS to type III, which manifests as fibre thickening, cross-linking, deposition, distribution perpendicular to the tumour boundary, developing tensile stresses and formation of a tumour invasion track.²¹ This growing fibrosis and stiffness of ECM results in tissue "desmoplasia," which stimulate tumorigenesis, metastasis and immune surveillance.^{22,23} Furthermore, the ECM stiffness is obstructive for effective permeation of drugs to the intratumoral area and promotes transformation of cancer cells to CSCs, and these two aspects can induce resistance to anti-cancer therapies.²⁴ Whatcott et al²⁵ demonstrated that high desmoplasia is related to low survival rates of pancreatic ductal adenocarcinoma patient, indicating the reprogramming in the tumour stroma is most effective for targeting desmoplastic tumours, such as pancreas.²⁶

2.2 | Angiogenesis causes hypoxia and low drug penetration while promoting tumour immune escape and metastasis

The infinite proliferation and malignant metastasis of solid tumours are inseparable from angiogenesis in the TME. Tumour angiogenesis is the formation of abnormal vascular tissue with an immature structure, abnormal function and high permeability; it contributes to abnormal haemodynamics, poor nutrient, and oxygen supply, and TME hypoxia and hypoperfusion. Hypoxia is a common condition in many solid tumours. The hypoxic TME harbours cells with a burst of proliferation activity, effective for overgrowing a mass of tumour.²⁷ It has been identified a bidirectional cross-linking between hypoxia and angiogenesis so that the activity of either one potentiating another.^{28,29} Tumour angiogenesis also results in low intratumoral drug concentrations and inhibits immune cell infiltration of the TME, thereby laying the foundation for drug resistance, immune escape and metastasis.³⁰ Vascular endothelial growth factor (VEGF) is believed to be the most important factor for angiogenesis, a process which depends on the activation, germination and tube formation of ECs.^{31,32} Most cells in the TME, including tumour cells, can activate vascular ECs by secreting VEGF. This prompts the vascular ECs to accelerate proliferation and stretch out more pseudopodia to enhance their signal capture and invasion capabilities, which initiates angiogenesis. In addition to VEGF, matrix metalloproteinase (MMP) 2/9, transforming growth factor α/β (TGF α/β), fibroblast growth factor (FGF) and platelet-derived growth factor (PDGF) in the TME can also promote tumour angiogenesis.³³

2.3 | Cells in the TME are core links in the signal communication network

The cells in the TME are collectively referred to as tumour-associated stromal cells; they regulate tumour fate through complex signal communication with tumour cells. Tumour-associated stromal cells include CAFs, tumour-associated ECs, pericytes, CSCs, MSCs and immune cells, and other rarer stromal cell populations, such as neurons, fibrocytes, adipocytes and follicular dendritic cells, have also been observed in some tumours.

2.3.1 | CAFs

Normal fibroblasts and adipose-derived stem cells in the TME are transformed into CAFs that overexpress α -smooth muscle actin (α -SMA) and fibroblast activation protein.³⁴ CAFs occupies approximately 80% of pancreatic and breast tumour volume, and from cellular view, CAFs take about 40%-50% of the whole population of cells within the tumour microenvironment.²⁶ The secretory capacity of CAFs is much greater than that of normal stromal cells, and CAFs are the main source of tumour cell ECM.³⁵ CAFs and tumour cells reciprocally regulate each other through a feedback mechanism. CAFs represent metabolic symbiosis with cancer cells, which is important therapeutically.^{36,37} TGFβ1, PDGF, FGFβ and connective tissue growth factor secreted by tumour cells promote the transformation of normal fibroblasts into CAFs. TGF^β, FGF and hepatocyte growth factor (HGF) secreted by CAFs act on tumour cells, inducing epithelial-mesenchymal transition (EMT) and a pre-metastatic state.³⁸⁻⁴⁰ The matrix proteins secreted by CAFs differ in composition and content from those produced by normal tissues, forming the TME-specific ECM stiffness. However, CAFs also play an important role in matrix degradation by secreting matrix enzymes such as MMPs, which engages cancer cells to reshape their morphology and breakthrough basement membrane. Both deposition and degradation of ECM induced by CAFs are cancer promoting and CAFs could be essential target for cancer therapies.⁴¹

In addition, CAFs interplay with other stromal cells to regulate TME ecology. CAFs' secretomes reprogramme cancer cells, immune cells and ECs to facilitate cancer cell invasion and metastasis. Increased glycolysis of CAFs creates acidity in TME to increase generation of myeloid-derived suppressor cells (MDSCs), inhibit maturation of tumour-associated dendritic cells (TADCs) and impair the activity of NK and effector T cells.¹⁵ As the most population of stomal cells, CAFs accumulation in the TME is often correlated with poor prognosis in many tumours.⁴²

2.3.2 | Immune cells

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Proliferation

Both innate and adaptive immune cells in the TME show low tumourrecognition and killing ability, which is a key factor contributing to tumorigenesis and progression (Figure 1).

The TME affects innate immune cells in various ways. (a) Tumourassociated macrophages (TAMs) are mainly generated from bone marrow-derived monocytes or tissue-resident macrophages and are tumour-promoting by participating in tumour angiogenesis and metastasis. TAMs are highly heterogeneity and recently identified as a spectrum concept for macrophages activation based on the stimuli and secreted cytokines and chemokines. M1 macrophages, stimulated by LPS and interferon gamma, are identified by their production of high levels of pro-inflammatory cytokines and promotion of the Th1 response in the adaptive immune response. M2 macrophages are activated by IL-4 or IL-13 ligands and notably polarized into M2a (anti-inflammatory, tissue remodelling), M2b (Th2 activation, immunoregulation), M2c (phagocytosis of apoptotic cells), M2d (angiogenesis, tumour progression) subtypes, which are response for tumour progression and the main population of macrophages in TME.⁴³ IL-10, IL-4, IL-13, CCL2, CCL3, CCL4, CCL5 and MFG-E8 in TME promote M2 macrophages generation. TAMs participate in the tumour angiogenesis by secreting pro-angiogenic factors, including VEGF-A, EGF, PIGF, TGF-β, TNF-α, IL-1β, IL-8, CCL2, CXCL8 and CXCL12, and enable tumour metastasis by up-regulated N-cadherin and Snail, whereas down-regulated E-cadherin.⁴⁴ Furthermore, M2 subtype TAMs regulate the emergence of regulatory immune-inhibiting cells by secreting abundant anti-inflammatory cytokines.⁴⁵ (b) Signalling molecules in the TME, such as MHC class I chain-related protein A and B, drive low expression of activating receptors like NKG2D, NKp30, NKp46 and NKG2C in natural killer (NK) cells, thereby inhibiting their anti-tumour effects.^{46,47} (c) Dendritic cells (DCs) present antigen to T cells through MHC molecules and recruit other immune cells by secreting pro-inflammatory cytokines and chemokines. Antiinflammatory factors in the TME disrupt the normal differentiation of monocytes into DCs and enhance transformation of immature regulatory DCs (RegDCs). RegDCs can inhibit T-cell function by secreting TGF β and prostaglandin E₂.⁴⁸ (d) MDSCs generated by abnormal differentiation of monocytes. It secretes S100A8/A9, which



FIGURE 1 Abnormal and inactivated immune cell populations in the TME. Monocytes in the TME are prompted to differentiate into tumour-supporting M2 macrophages and MDSCs, whereas their differentiation into M1 macrophages and DCs is impaired. The presence of fewer DCs and more MDSCs results in the inhibition of effector T-cell responses through the down-regulation of the TCR and IL-2R. NK cells in the TME are inhibited through the down-regulation of their expression of NK activating receptors, such as NKG2D, NKp30, NKp46 and NKG2C. Traditional effector T cells transform into Tregs that have tumour protective effects. DC, dendritic cell; IL-2R, interleukin-2 receptor; MDSC, myeloid-derived suppressor cell; NFκB, nuclear factor kappa-B; NK, natural killer; TCR, T-cell receptor; TME, tumour microenvironment; Tregs, regulatory T cells

enhance tumour cell survival, and can inhibit T-cell responses by secreting TGF β , raising intracellular reactive oxygen species (ROS) and L-arginine level, and activating the IL-10 pathway.⁴⁹ MDCSs act together with TAMs to remodelling ECM, promoting angiogenesis and metastasis.50

Adaptive immune cells in the TME are altered to promote tumour immune escape. The expression of the T-cell receptor (TCR) and interleukin-2 receptor (IL-2R) are down-regulated in otherwise immunocompetent CD8+ T cells, resulting in immune suppression. Stomal cells, such as BECs, pericytes, CAFs and MSCs, express PD-L1, T-cell immunoglobulin and mucin-containing domain molecule 3 (TIM3) and CD95L (also known as FASL) on their surface, and they secrete indoleamine 2,3-dioxygenase (IDO), interleukin-6 (IL-6), prostaglandin E2 (PGE2) and transforming growth factor- β (TGF β) to induce exhausting and anergy of effector T cells.¹⁶ Meanwhile, expression of members of the FAS/FASL and PD-1/PDL-1 pathways is up-regulated in the anomalous T cells, whereas the PI3K/AKT pathway is down-regulated, which inhibits T-cell proliferation and induces apoptosis.^{51,52} In addition, regulatory T cells (Tregs), which are CD4⁺CD25^{high}FOXP3⁺, are induced from traditional T cells trough TGFβ1 and IL-10 signalling by TAMs, RegDCs and other non-immune cells. NK cells and cytotoxic T Lymphocytes (CTLs) play a key role for elimination of malignant tumour. The two cell types show metabolic interdependencies with tumour cells, which causing metabolic competition, nutrient restriction and immunosuppression.53 M2 macrophages, CAFs and Tregs make immunologic restrains against CD8+ T cells, thus causing CTLs dysfunction and exhaustion. Therefore, the T cells lose their normal functions and even mediate some of the maior tumour protective effects in the TME.⁵⁴ Recalling immunity

cycle in tumour through priming and activation of effector T cells is the key to exert durable and fruitful effects against tumour.⁵⁵

2.3.3 | Tumour-associated ECs

Tumour-associated ECs include blood endothelial cells (BECs) and lymphatic endothelial cells (LECs). VEGFA in the microenvironment promotes the formation of new blood vessels from BECs, and VEGFC and VEGFD activate LECs to form new lymphatic vessels in the TME.⁵⁶ Low expression of E-selectin, intercellular adhesion molecule (ICAM) 1/2 and vascular adhesion molecule 1 (VCAM1) in tumour-associated ECs results in the loss of tight connections in the tumour vascular endothelium. This leads to vascular structural abnormalities and hemodynamic disorders, inhibits the recruitment of immune cells and induces CD8⁺ T-cell immune tolerance by up-regulating the expression of programmed deathligand 1/2 and the CD28-CTLA4 family receptor ligands B7-H3 and B7-H4.⁵⁷ The leaky vasculature also causes precipitation and accumulation of waste products, and promotes TME acidity that influences tumour metabolism.⁵⁸ The acidified TME drives tumour local invasion.59

2.3.4 | Pericytes

Pericytes differentiate from mesenchymal precursors and are recruited to tumours by platelet-derived growth factor- β (PDGF β) gradients.⁶⁰ Pericytes are heterogenous in their function and are coming into focus these years. Nestin+/NG2+ "Type-2" pericytes contribute to tumour angiogenesis by promoting ECs survival through their secretomes including VEGFA, ANGPT1 and ECM components. They also express neural cell adhesion molecule 1 (NCAM1) and the NG2 proteoglycan, which contribute to vascular maturation by increasing pericyte recruitment.⁶¹ Furthermore, pericytes are involved in CSCs maintenance, tumour metastasis and immune microenvironment. Upregulation of PD-L1, CD90, PDGFR β , CD248 and Rgs5 which inhibit CD4+ and CD8+ cytotoxic T-cell activity was reported in pericytes derived from within tumour microenvironments, which facilitates immunosuppression and eventual immune evasion of tumour cells.⁶²

2.3.5 | MSCs

During the progression of carcinogenesis, MSCs are recruited to the TME by growth factors, cytokines and chemokines, such as IL-6, IL-1 β , TGF- β 1, EGF, PDGF, TNF- α and SDF-1 α , secreted by cancer cells and stomal cells.⁶³ Upon entry into the tumour microenvironment niche, MSCs promote or inhibit tumour progression by various mechanisms. The anti-tumour effect of MSCs is largely dependent on their immune-activating function: MSCs could enhance the phagocytic ability of co-cultured macrophages, and TLR-3 activated MSCs preserves viable and functional neutrophils by amplifying the antiapoptotic effects. MSCs have the ability to stimulate resting T cells to become activated and to proliferate, and can behave as conditional DCs in syngeneic immune responses by secreting proinflammatory cytokines. Many researches also state the pro-tumour effect of MSCs: once homing to TME, MSCs could differentiate into several stomal cells, such as CAFs, pericytes, adipocytes and endothelial-like cells to participate in TME construction and evolution.^{64,65} MSCs promotes tumour growth by increasing the number of cancer stem cells through bone morphogenetic protein signalling and WNT, TGF- β , IL-6/JAK2/STAT3 signalling pathways.⁶⁶

2.4 | TEXs are TME messengers

Exosomes are secreted vesicle-like membrane structures with a diameter between 30 and 100 nm. They carry a variety of proteins, lipids and nucleic acids, and contribute to intercellular communication.⁶⁷ TEXs, as the microcosms of tumour cells, carry a large amount of tumour-derived materials with source-cell specificity; TEXs are one of the main methods of TME signal interaction. TEXs participate in almost all tumour processes, including angiogenesis, matrix remodelling, tumour metastasis and immune evasion.

TEXs regulate tumour angiogenesis by activating ECs.⁶⁸ Pancreatic cancer-derived CD106⁺CD49d⁺ TEXs are recruited, recognized and internalized by tumour-associated ECs. They induce the expression of VEGF and other angiogenic proteins, such as CXCL5, MIF, and CCR1, by ECs and promote angiogenesis in the TME.⁶⁹ In addition, many non-coding RNAs in TEXs have been shown to Cell Proliferation

play a key role in angiogenesis. Non-coding RNAs (including miR-9, miR-21 and miR-210) in TEXs can promote angiogenesis in lung cancer by activating STAT3, ephrin A3 and MMP2/9.⁷⁰⁻⁷³ Conigliaro et al found that exosomes secreted by CD90⁺ liver cancer cells are rich in long non-coding RNA (lncRNA) H19, which can up-regulate the expression of VEGF, VEGF receptor and ICAM in vascular ECs, thereby promoting tumour angiogenesis and tumour cell adhesion and migration to the site of neovascularization. This finding suggests that lncRNA H19 may also be related to haematological metastasis of tumours.⁷⁴ Lang et al^{75,76} confirmed that lncRNA CCAT2 and lncRNA POU3F3 in glioma-derived exosomes are related to angiogenesis in vivo. Meanwhile, TEXs derived from various tumours—such as melanoma,⁷⁷ chronic myeloid leukaemia,⁷⁸ glioma,⁷⁹ and breast,^{80,81} colon,⁸² and ovarian cancer⁸³ have been shown to promote angiogenesis in the TME.

Matrix enzymes in TEXs degrade normal ECM, promote matrix remodelling and create a pre-metastasis microenvironment.^{84,85} For example, melanoma-derived exosomes can promote glycolysis in skin dermal cells, inhibit oxidative phosphorylation and acidify the ECM through their delivery of miR-155 and miR-210 to prepare for tumour metastasis.⁸⁶

TEXs disrupt the immune function of tumour patients and promote immune escape of tumour cells. They inhibit the anti-tumour activities of monocytes, DCs, T cells, macrophages, and NK cells, and drive the development of MDSCs and Tregs to promote tumour immune tolerance.^{52,87,88}

Manipulation of extracellular vesicles (EVs) to carry a desired cargo is a novel strategy for tumour therapy. EVs can be modified with specific receptors so as to target the cell/s of interest; this will pursue a long-term content storage, virtue by no phenotypical alteration inside the TME. This approach is impressive and covers the current limitation for application of stem cells for cancer therapy due to encountering phenotypical alterations in the TME. Another impressive feature with EVs therapy is their stability and their capacity to cross biological barriers efficiently.⁸⁹

3 | TME-MEDIATED TUMOUR PROTECTION AND PROMOTION MECHANISMS

Various parts of the TME act as tumour hotbeds to promote their malignancy (Figure 2). To aid the search for effective anti-tumour targets, we will consider the role of the TME in maintaining the tumour reserve, promoting tumour metastasis and resisting killing from 2 angles: (a) promotion of CSCs generation and (b) initiation of EMT.

3.1 | The TME protects CSCs

CSCs are a relatively quiescent tumour cell subpopulation that undergoes active DNA repair, similar to normal adult stem cells. As the "foundation" of tumours, CSCs mediate therapy resistance and metastasis of tumours; they make a tumour "the endless weed under wildfire." $^{\rm 90}$

Oxygen depletion by highly proliferative cancer cells and dysfunctional tumour angiogenesis leads to microenvironmental hypoxia. CSCs are concentrated in the hypoxic niche in which hypoxia and hypoxia-inducible factors (HIFs), mediators of hypoxia, induce their resistance by promoting metabolic reprogramming, stem cell maintenance and tumorigenesis. Hypoxia and HIFs could maintain stemness of CSCs by induction of EMT, activation of stemness-related signalling pathways, activation of stemness-related genes (Sox2, Oct4, Nanog, BMI1, MYC and KLF4), suppression of differentiation-related genes and so on. CSCs in hypoxic TME uptake more glucose and reprogramme their metabolic mechanism from oxidative phosphorylation to glycolysis, which is responsible for resistance to radiation and chemotherapy and promoting DNA repair.⁹¹ 31734836 One of the key features of CSCs is the evolving of drug efflux pumps which are ATP-binding cassette (ABC) transporters. These are unidirectional cellular pumps that cause their resistance. The efflux pumps reduce accumulation of drugs within the stem cells and disable the activity of the chemotherapeutic drugs. The activity of efflux system is potentiated by hypoxia.^{28,92,93} Furthermore, hypoxia and HIFs induce dedifferentiation and enhances telomerase activity of CSCs to endow them resistance.⁹³ Hypoxia-induced stem phenotypes have been confirmed in many types of tumours. Hypoxia can increase the expression of the stem cell markers c-MYC, NANOG, SOX2 and OCT4 in chronic myeloid leukaemia (CML) tumour cells. Disrupting the HIF-mediated hypoxic environment and correcting hypoxic adaptive cell metabolism can reduce the malignancy of tumour cells and enhance drug responsiveness.94

The cellular components in the TME also play an important role in protecting CSCs. CAFs maintain the stemness of CSCs by secreting factors such as HGF and promoting EMT.⁹⁵ In addition, the high expression of the ECM receptor integrin on the membrane of CSCs can promote the formation of tight junctions with the ECM, which contributes to physical protection by the ECM and the maintenance of stemness. TACS III shelters CSCs by reducing drug exposure and forming a protective track.

However, according to the important role of tumour stem cells in tumour progression and drug resistance, targeting CSCs in TME could become a promising treatment of cancers.

3.2 | The TME promotes EMT to initiate metastasis

Metastasis is a sign of tumour deterioration and progression, and a leading cause of cancer death. Tumour metastasis is multistep process regulated by many factors, but EMT is considered the initial step.

EMT occurs when epithelial cells lose their unique polarity, adhesion ability, and surface expression of E-cadherin and β -catenin, then assume the morphology of mesenchymal cells and express mesenchymal cell markers such as α -SMA, vimentin, N-cadherin and SNAIL.⁹⁶ EMT is involved in normal physiological processes such as embryonic development and wound repair, but also plays an important role in tumour metastasis and CSCs generation. Tumour cells that undergo EMT dysregulate the expression of the myofilaments, microtubules and intermediate filaments that comprise the cytoskeleton, thereby promoting their degradation. These changes allow the cells to lose tight connection with the ECM, obtain higher deformability and motility capacity, and easily break through the tumour basement membrane to pass through gaps between the vascular ECs, and successfully implant in target metastasis organs.⁹⁷ Tumour cells also experience an increase in stemness during EMT.

The TME plays an important role in promoting EMT. TGF β , EGF, VEGF, HGF and MMP family members in the TME comprise a complex network that promotes EMT. Among those factors, TGF β is the key EMT-promoting molecule; it regulates EMT through SMAD-dependent or independent pathways. In the SMAD-dependent pathway, TGF β binds to receptors in the tumour cell membrane; they phosphorylate SMAD family members and promote SMAD2/3 dimer formation. The dimers are transported into the nucleus with the help of activated SMAD4, where they activate a series of EMT transcription factors such as SNAIL, TWIST, vimentin, fibronectin and ZEB.⁹⁸ In the SMAD-independent pathways in tumour cells, thereby promoting EMT-related gene expression.⁹⁹

Cells in the TME, especially CAFs, are the main sources of EMTpromoting molecules. CAFs participate in EMT and tumour metastasis by: (a) secreting paracrine EMT-related factors,¹⁰⁰ (b) making direct contact with tumours to promote EMT and (c) mediating matrix remodelling and secreting abnormal ECM collagen and fibronectin. In addition to CAFs, other cells in the TME contribute to EMT. M2 macrophages promote EMT by secreting the immune regulatory factors IL-6 and IL-8. Tregs and neutrophils also affect the tumour cell phenotype by up-regulating the expression of members of the TGF β pathway.¹⁰¹ Various macromolecular proteins in the tumour ECM, such as collagen fibres, are also important for EMT. Breast cancer cells cultured with type I collagen fibres can acquire mesenchymal cell characteristics and undergo EMT. Tumour cells cultured with tumour ECM fibronectin also readily undergo EMT.¹⁰² High expression of HIF1 α in the hypoxic TME inhibits caveolin-1 expression

FIGURE 2 Composition of the TME and its effects on tumour development. The TME is a complex network of tumour-associated cells (such as CAFs, tumour-associated immune cells, and ECs), TEXs, soluble molecules (such as cytokines, growth factors and hormones) and tumour-specific ECM, which is nourished by tumour angiogenesis. The components act as a tumour nest that maintains CSCs, promotes EMT, enables escape from immune surveillance and provides resistance to therapies through specific signal pathways. CAF, cancer-associated fibroblast; CSCs, cancer stem cells; ECM, extracellular matrix; ECs, endothelial cells; EMT, epithelial-mesenchymal transition; TEX, tumour-associated exosome; TME, tumour microenvironment



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in tumour cells; a negative feedback mechanism up-regulates the expression of the caveolin-1-related protein epidermal growth factor receptor, thereby activating the STAT3 signalling pathway and reducing the expression of epithelium-specific markers.¹⁰³ At the same time, mesenchymal transition of tumour-associated vascular ECs reduces tight junctions, thereby weakening the barrier effects of blood vessels.

However, the current understanding of cellular phenotypical modification during cancer metastasis is acquiring a partial EMT, namely a hybrid E/M phenotype. This indicates that tumours cells can represent mesenchymal phenotype while simultaneously exhibiting epithelial potentials. This hybrid phenotype can be seen in collective invasion of tumour cells, and it accounts for the greatest capacity to pursue tumour metastasis.¹⁰⁴ These all indicate that targeting EMT activity and weakening the EMT-promoting link can play a key role in inhibiting tumour metastasis.

4 | TARGETING THE TME TO BREAK THROUGH THE FORTRESS OF THE TUMOUR

Isolation of tumour cells by the TME results in low treatment efficiencies, low clinical response rates and drug resistance.^{13,105} It has been suggested that highly specific therapeutic strategies that break the barrier of the TME and disrupt its strong protective effects may be safer and more effective.¹⁰

4.1 | Targeting drugs attack the TME separately

Understanding the major events occurring in the TME that support primary tumour growth and how these events impact the modulation of the environment is of utmost relevance to assist the definition of efficient therapy strategies. A good deal of current strategies was used to target TME components.

For targeting tumour-associated ECM, the inhibition of the TGF-β signalling pathway mediated by Losartan and its analogs results in a reduced secretion of collagen I and consequently reduced ECM stiffness. However, drugs targeting MMPs, include JNJ0966, highly selective towards MMP-9, and the antibody Fab 3369 that targets MMP-14, have been developed. Among them, incyclinide went through several clinical trials for advanced carcinomas (Clinical trials NCT00004147, NCT00003721, NCT00001683 and NCT00020683).¹⁰⁶ Drugs targeted against CAFs have come into clinic therapy. Nindetanib or BIBF1120 having tyrosine kinase inhibitor activity towards receptors of VEGF, FGF and PDGF greatly reduce the protective effect of CAFs on tumour. Several antiangiogenic drugs have been tested in clinical trials. Monoclonal antibodies of VEGF/VEGFR, such as bevacizumab and pazopanib, increased overall survival or progression-free survival of patients when used alone or as adjuvants in a cocktail chemotherapeutic treatment.¹⁰⁷ In addition, targeting immune system in TME also

play an important role in tumour treatment. The inhibition of colony-stimulating factor-1 (CSF-1) signalling by anti-CSF-1R neutralizing antibodies has been shown to impair TAMs-recruiting and MDSCs mediated tumour proliferation. 28117416 Activation of the anti-tumour functions of the immune cells by GM-CSF, which stimulates the antibody-dependent cellular cytotoxicity of anti-cancer antibodies, and targeting of immune checkpoint inhibitors (CTLA-4, PD-1, PD-L1) has been widely exploited to prevent tumour progression.¹⁰⁸

4.2 | Cell therapy to modify the TME is a new strategy for tumour treatment

Even though targeting molecules inhibit tumour to a great extent, there is still some insurmountable difficulties. For example, agents that degrade and/or deconstruct ECM must be used carefully, since they may induce metastasis instead of avoiding tumour progression. We boldly assume that cell therapies may solve some of the problems of targeting drugs attributing to cell's natural advantages. (a) The cell itself has good plasticity, which provides the possibility of acting on multiple TME components at the same time to fully degrade TME, which can effectively prevent the compensatory regeneration and protection of other components when simply targeting a certain part; (b) the cell has homing properties. The "localization system" of the treatment cells has the tendency of tumour tissue and homing under the action of cytokines and chemokines, which greatly avoids the systemic side effects brought by non-selective drugs; (c) cell has high permeability. Due to tumour stiffness and abnormal angiogenesis, the drug's permeability to TME is poor, but cells can effectively enter TME through deformation movements, secretion of matrix-regulating enzymes, etc; (d) many cells have the ability to regulate normal tissue regeneration and repair, which can correctly repair tissue damage caused by chemotherapy drugs and avoid the adverse impact of bystander effects on normal cells (Figure 3).

4.2.1 | Immune cell therapy for precisely targeting tumour cells

The TME regulates the differentiation and activation of immune cells to reduce their ability to recognize and eliminate tumour cells. Immune cell therapies that more accurately target tumour cells while restoring normal immune cell populations within the TME, enhancing normal immune cell function and disrupting the protective effects of the abnormal immune cell networks have become a new anti-tumour option.¹⁰⁹ As early as 1998, studies demonstrated that transplanting immune-activated splenocytes into mice with breast cancer caused tumour shrinkage and conferred long-term tumour-free survival, suggesting that adoptive immune cells can be effective anti-tumour therapeutics.¹¹⁰ In recent years, immune cells such as NK cells, cytokine-induced killer cells, DCs, tumour-infiltrating lymphocytes and

FIGURE 3 Cell therapy to modify the TME. Traditional radiotherapy, chemotherapy and targeted drugs all rely of tumour cell killing as their main mechanism of action, with the attendant serious adverse reactions and rapid resistance. Current cell therapies for targeting tumours and the TME–modified immune cell therapies, haematopoietic stem cell transplantation, mesenchymal stem cell transfer, and embryonic stem cell-based microenvironment therapies–provide novel ideas for exploring breakthrough of tumour therapy strategies



chimeric antigen receptor (CAR)-modified immune cells have shown marked anti-tumour potential.¹¹¹ Among them, CAR-modified cells have the best effects and have gradually made the transition from scientific research products to clinical treatment option.

A CAR is a biological element that is transgenically expressed on the surface of an immune cell membrane where it recognizes specific antigens. The extracellular structure of a CAR is a variable single-chain fragment that recognizes a specific antigen. The intra-membrane structure comprises a signal transduction domain; the earliest CARs used a single CD3⁽ domain, but successive generations have included different numbers and types of co-stimulatory domains to improve the cell proliferation, cytokine secretion and killing capacities of CAR-equipped immune cells.¹¹² CAR-modified immune cells are purified and expanded from autologous or allogeneic immune cells or stem cell-derived cells, and then engineered with a CAR and transferred into patients. They have a lower risk of mediating off-target graft-versus-host responses while killing tumours. Upon recognition of tumour cells that express their target antigens, CAR-modified cells activate pathways to initiate their proliferation, release of inflammatory factors and phagocytosis to kill the targeted cells accurately and specifically. At present, the most commonly CAR-modified immune cells are CAR-T and CAR-NK cells, which have achieved good responses in the clinical treatment of haematological tumours.¹¹³ Many groups have explored the feasibility of applying these tools to solid tumours. As of 2018, 57% of CAR-T-cell clinical trials focused on the treatment of haematological malignancies, 11% on gastrointestinal tumours, 8% on neurological tumours and 5% on breast cancers.¹¹⁴

CAR-T cells are currently the most mature anti-tumour immune cells. Among these, CAR-T cells targeting CD19 have been the most

successful in the clinic. A clinical trial at Memorial Sloan-Kettering Cancer Center in 2016 (NCT01044069) showed that 91% of 32 B cell-derived acute lymphoblastic leukaemia (B-ALL) patients treated with CD19-targeted CAR-T cells achieved complete remission. Another clinical trial (NCT01626495) demonstrated that 30 patients with B-ALL had a complete response rate of 90% after CD19 CAR-Tcell therapy. In a National Institutes of Health clinical trial in 21 children and adults with ALL (NCT01593696), a complete response rate of up to 70% has been reported. CD19-specific CAR-T cells also show limited therapeutic effects on other haematologic malignancies such as chronic lymphocytic leukaemia, lymphoma and multiple myeloma (NCT02135406); however, the response rates of these diseases are lower than that of B-ALL.^{115,116} CAR-T cells targeting other molecules. such as CD20, CD22, CD30 and ROR1, have been used successfully to target different tumours (NCT02315612 and NCT00621452).¹¹⁷ In addition to blood tumours, many studies have tried to target solid tumours using CAR-T cells, such as GD2 CAR-T for neuroblastoma and GPC3-targeted CAR-T cells for hepatocellular carcinoma, which have prolonged patient survival.^{118,119} However, CAR-T is highly effective for malignancies of haematologic system; for solid tumours, the application of this strategy is elusive mainly because of their suppressive TME, weak TIL trafficking,^{120,121} and has a high probability of side effects. In clinical trials of CD19-specific CAR-T-cell treatment, all subjects without exception showed cytokine release syndrome. In addition, complications such as neurotoxicity, B-cell depletion and off-target effects also seriously threaten patients' lives.¹²²

In addition to CAR-modified T cells, CAR-NK cells have achieved a high response rate in the treatment of myelodysplastic syndrome and acute myeloid leukaemia (AML). CAR-NK cells for the treatment of B-cell lymphomas (NCT01974479 and NCT03056339) and

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metastatic solid tumours (NCT03415100) are in clinical trials; however, they face problems such as low CAR transfection efficiency and short in vivo survival time.¹²³ To resolve the issues of limited sources of NK cells and short survival time, NK cells have been generated from induced pluripotent stem cells (iPSCs) and cord blood, and their therapeutic effects on recurrent or refractory blood and solid tumours have been assessed (NCT01729091, NCT03019640, NCT02280525 and NCT03539406). However, the low induction and differentiation efficiencies of iPSCs and the relatively high risk of tumour formation still limit their application.

Exosomes secreted by CAR cells have been confirmed to carry the CAR structure. They maintain the original targeting characteristics of the parent cells in the context of lower off-target cytotoxicity, with high efficacy and safety in preclinical experiments.¹²⁴ CAR exosomes may be used to optimize or even replace existing cell-based CAR immunotherapies.

4.2.2 | Haematopoietic stem cell transplantation to reconstruct haematopoietic function

The origins of many tumours are associated with the dysplasia, depletion, and dysregulated proliferation and differentiation of normal stem cells. The existence of a normal stem cell microenvironment has become a prognostic indicator for tumour treatment.¹²⁵ Eliminating malignant CSCs and supplementing with normal stem cells has become a reliable treatment for many tumours, especially haematologic tumours.¹²⁶

Haematopoietic stem cell transplantation (HSCT) is to completely remove the abnormal bone marrow haematopoietic stem cell (HSC) microenvironment by means of radiotherapy and chemotherapy, and then rebuild the haematopoietic microenvironment and restore the normal haematopoietic function by transplanting normal stem cells. As the only curative treatment for malignant and non-malignant diseases of the haematopoietic system, HSCT has developed rapidly in the past 25 years.¹²⁷⁻¹³⁰ The first step of HSCT is the pretreatment of patients with radio-chemotherapy to destroy the dysfunctional haematopoietic and immune systems and to ensure low immune responsiveness by the patients, thus laying the foundation for long-term survival of the transplanted cells. Then, HSCs-mobilized from the donor by granulocyte-colony-stimulating factor-are purified, expanded in vitro, and transplanted into the recipient, where they typically home to the bone marrow and gradually reconstitute the recipient's haematopoietic system. HSCT brings graft-versus-leukaemia (GVL) effect in recipients, which helps eradicate tumours and is considered to be multifactorial.¹³¹ The mechanism of GVL, though poorly understood, might be similar to graft-versus-host disease (GVHD) phenomenon that is mediated by donor's immune compounds. The principal cytotoxicity in GVL is mediated by donor T cells and NK cells with ancillary roles played by dendritic cells, B cells and minor histocompatibility antigens.¹³² Although HSCT can fundamentally eradicate tumours, several side effects, including acute infection, graft rejection, chronic GVHD and secondary tumour development after transplantation, still impair the long-term

survival of patients.^{133,134} Approximately 1.2%-1.6% of patients have secondary tumours 5 years after HSCT, and the cumulative incidence of secondary tumours increases to 2.2%-6.1% after 10 years, and 3.8%-14.9% after 15 years.¹³⁵ To reduce the side effects and improve the survival rates of patients, HSCT programmes have continuously innovated. Peripheral blood- and umbilical cord blood-derived HSCs are gradually replacing bone marrow HSCs for transplantation, which solves the problem of insufficient cells.¹³⁶ HSCT performed under non-myelosuppressive conditions by reducing the doses of chemotherapy and radiation, or even without pretreatment, reduces the incidence of the GVHD response and improves long-term transplantation efficacy.¹³⁷ It is clear that maximizing the GVL effect while minimizing GVHD is the holy grail of transplant immunology. Given that GVHD and GVL have similar mechanisms, prophylaxis for GVHD might affect GVL intensity, which should be taken into account when formulating a transplantation strategy.

HSCT is becoming increasingly prominent in the treatment of some solid tumours, especially refractory tumours.¹³⁸ As early as 1997, researchers found that breast tumours in mice shrank after allogeneic HSCT.¹³⁹ In recent years, HSCT has been studied for the treatment of relapsed and refractory glioma, and as a consolidation therapy for the remission stage, resulting in a 1-year tumour-free survival rate of up to 90%–93% and a 3-year survival rate of 63.7%.^{140,141} After receiving allogeneic HSCT, especially non-myelosuppressive transplantation, patients with metastatic renal cell carcinoma showed a relatively high response rate, likely due to the restoration of T cells and other immune cells capable of disrupting the immunosuppressive TME.^{142,143}

4.2.3 | MSC therapy

MSCs are non-haematopoietic stem cells primarily found in the bone marrow. They can differentiate into adipose tissue, bone and cartilage under the appropriate conditions.¹⁴⁴ In addition to the bone marrow, MSCs also exist in cord blood, peripheral blood and adipose tissue.¹⁴⁵ Due to their relative abundance, shared features with tumours and homing characteristics, MSCs can effectively enter the TME, thereby overcoming low drug delivery efficiency and limitations on TME penetration.¹⁴⁶ Transfer of pre-edited and modified MSCs has become a new strategy for tumour cell therapy.¹⁴⁷

The reported effects of natural MSCs on tumours have been contradictory. Some studies have shown that MSCs have anti-tumour effects, such as inhibiting tumour cell proliferation and causing cell cycle arrest in leukaemia, bladder cancer, renal cell carcinoma, breast cancer and other tumours.¹⁴⁸⁻¹⁵¹ However, other studies have found that MSCs promote tumour progression. MSC transplantation can boost tumour invasiveness. This is due to that the cells highly express CXCR4, upon injection, these CXCR4 + cells are attracted into the areas enriched in CSCs or cancer stem niches. In these places, MSCs can alter their phonotypical characteristics into attaining a CSCs like feature, thus enriching the pool of cancer stem niches for the subsequent progression and metastasis of tumours

like glioblastoma.¹⁵² Relevant researches show that osteosarcoma cells have enhanced proliferation and migration ability after co-culture with MSCs in vitro¹⁵³; MSCs enhance the thermal resistance of ovarian cancer cells through CXCL12-dependent pathways and promote tumour development and angiogenesis in mice with colon cancer^{154,155}; With support from MSCs, breast cancer cells acquire high metastasis ability through the CCL5/CCR5 feedback axis.^{156,157} MSCs may secrete pro-tumour chemokines and differentiate into CAFs in the TME. Studies demonstrated that effect of MSCs is opposite within different injection site in the same animal model. When injected locally with breast cells, MSCs promoted the migration and invasion abilities of tumour cells, and drove the angiogenesis progress. To the contrary, distant injection of MSCs inhibited tumour progression and promoted altered immune cell populations in the TME. Treg and MDSC were decreased significantly, and CD8 T cells and DCs were increased. Immune-activating cytokines TNF, IFN γ , TLR3 and IL-12 were up-regulated. These results imply we should pay attention to the administration methods when apply MSCs to tumour patients.^{158,159} We hypothesize that MSCs could concurrently behave pro-tumour effects by differentiating into CAFs, pro-angiogenesis, and inhibit T cells to enhance TME compounds, while tend to suppress tumours by activating innate and adaptive immune, the final effect depends on the order and strength of the two sides and tumour context.

MSC exosomes, engineered MSC nanoparticles (nanoghosts), and other microvesicles have been used as substitutes for MSCs due to their similar compositions and better safety profiles. Exosomes derived from MSCs can inhibit the proliferation of various tumour cells.¹⁶⁰⁻¹⁶² However, there also have been reports that MSC exosomes promote tumours.¹⁶³ The best method to make use of the advantages of MSCs while avoiding their tumour-promoting activity has become the focus of MSC therapy research.

Researchers are increasingly modifying MSCs to optimize their anti-tumour function in 2 ways. First, they are enhancing the synthesis and release of endogenous and exogenous anti-tumour factors from MSCs. Second, they are strengthening their homing to the TME and prolonging their activity there to achieve more efficient TME penetration and anti-tumour effects.

In order to ensure the tumour-killing activity of MSCs and prevent them from promoting tumours, many researchers have enhanced MSC synthesis and release of IFN- γ , which inhibits tumour cell proliferation, and tumour necrosis factor-related apoptosis-inducing ligand, which promotes apoptosis.^{164,165} These approaches have enhanced MSC anti-tumour effectiveness in preclinical and clinical experiments. MSCs transfected with suicide genes and carrying biologically active anti-tumour substances reach the TME via recruitment by chemotactic signals such VEGF and TGF β 1 secreted by tumour cells and CAFs. Once in the TME, they initiate the suicide programme to release the drugs with greater specificity and in greater concentration in the TME than achievable by conventional methods.¹⁶⁶⁻¹⁶⁸ These findings suggest the feasibility of using modified MSCs as efficient drug carriers. MSCs transfected with only a CCL5 promoter-driven or ganciclovir-induced suicide gene can also cause a degree of tumour cytotoxicity in animal models and clinical trials of hepatocellular carcinoma, pancreatic cancer, and breast cancer.^{169,170}

In order to enhance the in vivo activity of MSCs, researchers have encapsulated MSCs with synthetic biodegradable ECM and implanted them in the brains of mice with glioblastoma. This approach resulted in dramatic shrinkage of the tumour and ensured the long-term biological activity of the MSCs.¹⁷¹ Other studies have enhanced the homing of MSCs through gene editing. HIF1 α induced by the hypoxic TME up-regulates downstream SDF-1 α to promote tumour proliferation and metastasis. Overexpression of the SDF-1 α receptor CXCR4 in MSCs enhances the penetration of MSCs into the hypoxic glioma microenvironment.¹⁷²

However, MSCs remain a double-edged sword. The stronger the effects of MSCs as drug carriers that penetrate the TME to kill tumours, the more serious the side effects caused by the drugs' non-selective destruction of normal cells. The current drawbacks associated with MSC therapies have prompted researchers to seek safer, more accurate anti-tumour treatment methods.

4.2.4 | Embryonic stem cell microenvironmental therapy

The early embryonic microenvironment has the powerful ability to repair erroneous genetic material and inhibit oncogene expression; thus, embryonic cells have innate tumour immunity characteristics. An increasing amount of research is being devoted to applying this tumour-hostile microenvironment to tumour therapy.^{173,174}

The embryonic microenvironment can reverse tumour fate. Lee et al¹⁷⁵ implanted human skin melanoma cells into zebrafish blastocysts and, surprisingly, found that the implanted cells did not form tumours in the embryo. Kasemeier-Kulesa et al¹⁷⁶ found that malignant melanoma cells implanted into chicken embryos did not produce tumours, and even reverted to expressing the normal melanocyte-specific marker MART-1. Durr et al did not observe tumour formation after they implanted human KG-1 myeloid leukaemia cells and primary human AML cells into 3.5-day (mulberry phase) mouse blastocysts. The tumour cells implanted into the embryonic microenvironments reverted to the expression of erythroid-specific haemoglobin and glycophorin A, and resumed proper erythroid homing to the aorta-gonad-mesonephros region (embryonic haematopoietic tissue), the yolk sac, and the peripheral blood.¹⁷⁷ The ability of the embryonic microenvironment to revert tumour cells gradually weakens as the embryo develops and differentiates.¹⁷⁸ (Table 1).

In order to apply the powerful tumour-reverting effects of the in situ embryonic microenvironment to the practical treatment of tumours, researchers have cultured embryonic stem cells (ESCs) in vitro to establish an ESC microenvironment (ESCM) that mimics that found in embryos with remarkable results. Zhou et al injected TK suicide gene-transfected ESCs into mice with CML via the tail vein and found that the cells restored normal blood cell counts in the mice and prolonged their survival. By timely activation of the

Mimic embryonic microenvironment	Target diseases	In vivo administration route	Mechanisms	Reference
hESC-conditioned medium/ hESC- exosomes	Colorectal adenocarcinoma, breast cancer	Subcutaneous injection	Rebalance ectopic expression of OCT4, SOX2, KLF and c-MYC	173
mESC-co-culture	Uveal melanoma	Subcutaneous injection	Down-regulation of PI3K/AKT pathway	180
mESC-intravenous injection	Chronic myeloid leukaemia	Intravenous injection	NA	179
mESC-conditioned medium	Breast cancer	Mammary fat pads injection	Down-regulation of STAT3 pathway	183
hESC-conditioned medium	Colorectal carcinoma	Subcutaneous injection	Suppress Notch1 pathway	184
3D decellularized matrix after hESCs culture	Cutaneous melanoma, breast cancer	Mammary fat pads injection	Down-regulation of nodal pathway	178

Abbreviations: hESC, human embryonic stem cell; mESC, mouse embryonic stem cell; TCTP, translationally controlled tumour protein.

suicide gene, the differentiated ESCs could be eliminated to avoid teratoma formation and guarantee the safety of the recipients.¹⁷⁹ Liu and Wang et al co-cultured embryonic stem cells with uveal melanoma or cutaneous melanoma cells. They found that the ESCM can inhibit malignant phenotype development in tumour cells. Upon co-culture of ESCs, healthy cells and tumour cells, the ESCM surprisingly down-regulated the PI3K pathway in the tumours, but up-regulated the PI3K pathway in the tumour, but up-regulated the PI3K pathway in normal cells. Thus, the ESCM played a bidirectional regulation role by reversing tumour-related signalling and enhancing signalling in healthy cells. This finding proved that killing tumours does not inevitably result in off-target killing of normal cells.^{180,181}

Other research has confirmed the tumour-reverting effects of the ESCM. For example, decellularized matrix components generated by 3-dimensional ESC culture can reverse metastatic breast and prostate cancers.¹⁷⁶ Furthermore, human ESC-conditioned medium and exosomes can suppress the malignant phenotypes of colon cancer and breast cancer cells. The researchers speculated that the effects of the ESCM on tumour reversion may result from the exchange of materials between cells, thereby rebalancing the expression of the stemness factors OCT4, SOX2, KLF and c-MYC in tumour cells.¹⁷³ Zebrafish embryo extracts can inhibit breast cancer by down-regulating the expression of translationally controlled tumour protein and promoting E-cadherin/ β -catenin redistribution to reshape the cytoskeleton.¹⁸² Mouse ESC-conditioned medium was found to inhibit breast cancer cell proliferation, promote apoptosis and inhibit malignant behaviour.¹⁸³

The mechanisms by which the ESCM reverts tumours are still not completely clear. Current studies suggest that tumour reversion may be related to the PI3K/AKT, STAT3, Notch1, and other pathways, and that exosomes derived from ESCs may play an important role in their signal transduction.^{178,183,184} In addition, transmembrane communication and cell contact-mediated signal transduction may play key roles in ESCM regulation of tumour cell fate. The totipotency of ESCs enables them to regulate the TME in many ways, not only by affecting tumour cells, but also by reshaping other TME components, enhancing normal cell functions, and supporting the regeneration and repair of the body. ESC treatment raises new hope for radically eliminating the side effects caused by killing normal cells during tumour treatment.

5 | SUMMARY AND PROSPECTS

The TME can mediate tumour cell immune escape, promote CSCs formation and enhance tumour metastasis ability, thereby promoting tumorigenesis and development. The distinctive features of the TME, including the abnormal haemodynamics due to neovascularization, the ECM that is difficult to penetrate, and the hypoxic state that confers resistance to oxidative damage, play important roles in the drug resistance of tumour cells. Cell therapies utilizing engineered immune cells have shown good anti-tumour effects, but poor curative effects for solid tumours. Furthermore, the main aim of this type of cell therapy is still to kill tumour cells. Thus, the body can develop resistance to the treatment, and it is impossible to avoid the complications caused by cell killing that are also associated with traditional therapy. Malignant tumour cells are derived from normal cells and develop in the TME. Suitable microenvironments such as the early embryonic environment can revert tumour cells into normal cells. HSCT, MSC transplantation, and ESC and other stem cell therapies can reach the interior of the TME due to their strong targeting and penetrating power. By reshaping the cellular components, ECM structure and soluble factor composition to comprehensively rebuild the TME, these stem cell therapies can reverse the malignant tumour phenotype while avoid cell killing and reducing treatment-related harm to patients. Stem therapies could, therefore, represent the most promising new direction for tumour treatment. New breakthroughs in tumour treatment await in the near future based on combining approaches to complement the strengths of individual cell therapies,

optimizing the favourable characteristics of cells with engineered modifications to compensate for their shortcomings, or replacing source cells with cell-specific microvesicles.

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CONFLICT OF INTEREST

The authors declare no conflict of interest.

AUTHOR CONTRIBUTIONS

CYQ, SBW and JHL wrote the first draft of the manuscript. JL, TL, CYL, YRS and YRL wrote sections of the manuscript. CYQ, XS, QW participated in the discussion of the manuscript. CL, LY and ZCW provided critical revisions. All authors approved the final version of the manuscript for submission.

DATA AVAILABILITY STATEMENT

Data available on request.

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REFERENCES

- Weinberg BA, Marshall JL, Salem ME. The growing challenge of young adults with colorectal cancer. Oncology. 2017;31(5):381-389.
- Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. 2018;68(6):394-424.
- Vogelstein B, Kinzler KW. The multistep nature of cancer. Trends Genetics. 1993;9(4):138-141.
- Paget S. The distribution of secondary growths in cancer of the breast. 1889. *Cancer Metastasis Rev.* 1989;8(2):98-101.
- Fu B, Wang N, Tan HY, Li S, Cheung F, Feng Y. Multi-component herbal products in the prevention and treatment of chemotherapy-associated toxicity and side effects: a review on experimental and clinical evidences. *Front Pharmacol.* 2018;9:1394.
- Accordino MK, Neugut AI, Hershman DL. Cardiac effects of anticancer therapy in the elderly. J Clin Oncol. 2014;32(24):2654-2661.
- Xiao Q, Zhu W, Feng W, et al. A review of resveratrol as a potent chemoprotective and synergistic agent in cancer chemotherapy. *Frontiers Pharmacol.* 2018;9:1534.
- Lorenz E, Scholz-Kreisel P, Baaken D, Pokora R, Blettner M. Radiotherapy for childhood cancer and subsequent thyroid cancer risk: a systematic review. *Eur J Epidemiol*. 2018;33(12):1139-1162.
- Yarchoan M, Johnson BA 3rd, Lutz ER, Laheru DA, Jaffee EM. Targeting neoantigens to augment antitumour immunity. *Nat Rev Cancer.* 2017;17(4):209-222.
- Villanueva N, Bazhenova L. New strategies in immunotherapy for lung cancer: beyond PD-1/PD-L1. Ther Adv Respir Dis. 2018;12:1753466618794133.
- Drilon A, Hu ZI, Lai GGY, Tan DSW. Targeting RET-driven cancers: lessons from evolving preclinical and clinical landscapes. *Nature Rev Clin Oncol.* 2018;15(3):151-167.

12. Seebacher NA, Stacy AE, Porter GM, Merlot AM. Clinical development of targeted and immune based anti-cancer therapies. *J Exp Clin Cancer Res.* 2019;38(1):156.

Proliferation

- 13. Wykosky J, Fenton T, Furnari F, Cavenee WK. Therapeutic targeting of epidermal growth factor receptor in human cancer: successes and limitations. *Chin J Cancer*. 2011;30(1):5-12.
- Mortezaee K, Parwaie W, Motevaseli E, et al. Targets for improving tumor response to radiotherapy. *Int Immunopharmacol.* 2019;76:105847.
- 15. Najafi M, Goradel NH, Farhood B, et al. Tumor microenvironment: Interactions and therapy. J Cell Physiol. 2019;234(5):5700-5721.
- 16. Wu T, Dai Y. Tumor microenvironment and therapeutic response. *Cancer Lett.* 2017;387:61-68.
- Schumacher TN, Schreiber RD. Neoantigens in cancer immunotherapy. Science. 2015;348(6230):69-74.
- Spugnini EP, Logozzi M, Di Raimo R, Mizzoni D, Fais S. A role of tumor-released exosomes in paracrine dissemination and metastasis. *Int J Mol Sci.* 2018;19(12):3968.
- 19. Pickup MW, Mouw JK, Weaver VM. The extracellular matrix modulates the hallmarks of cancer. *EMBO Rep.* 2014;15(12):1243-1253.
- Malik R, Lelkes PI, Cukierman E. Biomechanical and biochemical remodeling of stromal extracellular matrix in cancer. *Trends Biotechnol.* 2015;33(4):230-236.
- Gaggioli C, Hooper S, Hidalgo-Carcedo C, et al. Fibroblastled collective invasion of carcinoma cells with differing roles for RhoGTPases in leading and following cells. *Nat Cell Biol.* 2007;9(12):1392-1400.
- Liu M, Xu J, Deng H. Tangled fibroblasts in tumor-stroma interactions. Int J Cancer. 2011;129(8):1795-1805.
- Yamauchi M, Barker TH, Gibbons DL, Kurie JM. The fibrotic tumor stroma. J Clin Investig. 2018;128(1):16-25.
- Najafi M, Farhood B, Mortezaee K. Extracellular matrix (ECM) stiffness and degradation as cancer drivers. J Cell Biochem. 2019;120(3):2782-2790.
- Whatcott CJ, Diep CH, Jiang P, et al. Desmoplasia in primary tumors and metastatic lesions of pancreatic cancer. *Clin Cancer Res.* 2015;21(15):3561-3568.
- Najafi M, Mortezaee K, Majidpoor J. Stromal reprogramming: a target for tumor therapy. *Life Sci.* 2019;239:117049.
- Mennerich D, Kubaichuk K, Kietzmann T. DUBs, hypoxia, and cancer. Trends Cancer. 2019;5(10):632-653.
- Najafi M, Farhood B, Mortezaee K, Kharazinejad E, Majidpoor J, Ahadi R. Hypoxia in solid tumors: a key promoter of cancer stem cell (CSC) resistance. J Cancer Res Clin Oncol. 2020;146(1):19-31.
- Mortezaee K. Hypoxia induces core-to-edge transition of progressive tumoral cells: a critical review on differential yet corroborative roles for HIF-1alpha and HIF-2alpha. *Life Sci.* 2020;242:117145.
- Viallard C, Larrivee B. Tumor angiogenesis and vascular normalization: alternative therapeutic targets. *Angiogenesis*. 2017;20(4):409-426.
- Gianni-Barrera R, Butschkau A, Uccelli A, et al. PDGF-BB regulates splitting angiogenesis in skeletal muscle by limiting VEGF-induced endothelial proliferation. *Angiogenesis*. 2018;21(4):883-900.
- Ribeiro MF, Zhu H, Millard RW, Fan GC. Exosomes function in proand anti-angiogenesis. *Current Angiogenesis*. 2013;2(1):54-59.
- Fukumura D, Xavier R, Sugiura T, et al. Tumor induction of VEGF promoter activity in stromal cells. *Cell*. 1998;94(6):715-725.
- O'Sullivan J, Lysaght J, Donohoe CL, Reynolds JV. Obesity and gastrointestinal cancer: the interrelationship of adipose and tumour microenvironments. *Nat Rev Gastroenterol Hepatol.* 2018;15(11):699-714.
- Xing F, Saidou J, Watabe K. Cancer associated fibroblasts (CAFs) in tumor microenvironment. *Front Biosci.* 2010;15:166-179.

_EY-Proliferation

- Domingo-Vidal M, Whitaker-Menezes D, Martos-Rus C, et al. Cigarette smoke induces metabolic reprogramming of the tumor stroma in head and neck squamous cell carcinoma. *Mol Cancer Res.* 2019;17(9):1893-1909.
- Lisanti MP, Martinez-Outschoorn UE, Sotgia F. Oncogenes induce the cancer-associated fibroblast phenotype: metabolic symbiosis and "fibroblast addiction" are new therapeutic targets for drug discovery. *Cell Cycle*. 2013;12(17):2723-2732.
- Yu Y, Xiao CH, Tan LD, Wang QS, Li XQ, Feng YM. Cancerassociated fibroblasts induce epithelial-mesenchymal transition of breast cancer cells through paracrine TGF-beta signalling. Br J Cancer. 2014;110(3):724-732.
- Ao M, Franco OE, Park D, Raman D, Williams K, Hayward SW. Cross-talk between paracrine-acting cytokine and chemokine pathways promotes malignancy in benign human prostatic epithelium. *Can Res.* 2007;67(9):4244-4253.
- Fuyuhiro Y, Yashiro M, Noda S, et al. Cancer-associated orthotopic myofibroblasts stimulates the motility of gastric carcinoma cells. *Cancer Sci.* 2012;103(4):797-805.
- Farhood B, Najafi M, Mortezaee K. Cancer-associated fibroblasts: secretions, interactions, and therapy. J Cell Biochem. 2019;120(3):2791-2800.
- Tang H, Fu YX. Immune evasion in tumor's own sweet way. Cell Metab. 2018;27(5):945-946.
- 43. Messex JK, Byrd CJ, Liou GY. Signaling of macrophages that contours the tumor microenvironment for promoting cancer development. *Cells*. 2020;9:919.
- Fu LQ, Du WL, Cai MH, Yao JY, Zhao YY, Mou XZ. The roles of tumor-associated macrophages in tumor angiogenesis and metastasis. *Cell Immunol.* 2020;353:104119.
- Ostuni R, Kratochvill F, Murray PJ, Natoli G. Macrophages and cancer: from mechanisms to therapeutic implications. *Trends Immunol*. 2015;36(4):229-239.
- Ashiru O, Boutet P, Fernandez-Messina L, et al. Natural killer cell cytotoxicity is suppressed by exposure to the human NKG2D ligand MICA*008 that is shed by tumor cells in exosomes. *Can Res.* 2010;70(2):481-489.
- Bauernhofer T, Kuss I, Henderson B, Baum AS, Whiteside TL. Preferential apoptosis of CD56dim natural killer cell subset in patients with cancer. *Eur J Immunol.* 2003;33(1):119-124.
- Valenti R, Huber V, Filipazzi P, et al. Human tumor-released microvesicles promote the differentiation of myeloid cells with transforming growth factor-beta-mediated suppressive activity on T lymphocytes. *Can Res.* 2006;66(18):9290-9298.
- Nagaraj S, Gabrilovich DI. Myeloid-derived suppressor cells in human cancer. *Cancer J.* 2010;16(4):348-353.
- Hirata E, Sahai E. Tumor microenvironment and differential responses to therapy. Cold Spring Harb Perspect Med. 2017;7(7):a026781.
- Hoffmann TK, Dworacki G, Tsukihiro T, et al. Spontaneous apoptosis of circulating T lymphocytes in patients with head and neck cancer and its clinical importance. *Clin Cancer Res.* 2002;8(8):2553-2562.
- 52. Whiteside TL. Immune modulation of T-cell and NK (natural killer) cell activities by TEXs (tumour-derived exosomes). *Biochem Soc Trans*. 2013;41(1):245-251.
- Renner K, Singer K, Koehl GE, et al. Metabolic hallmarks of tumor and immune cells in the tumor microenvironment. *Front Immunol*. 2017;8:248.
- Wieckowski EU, Visus C, Szajnik M, Szczepanski MJ, Storkus WJ, Whiteside TL. Tumor-derived microvesicles promote regulatory T cell expansion and induce apoptosis in tumor-reactive activated CD8+ T lymphocytes. J Immunol. 2009;183(6):3720-3730.
- Farhood B, Najafi M, Mortezaee K. CD8(+) cytotoxic T lymphocytes in cancer immunotherapy: a review. J Cell Physiol. 2019;234(6):8509-8521.

- 56. Achen MG, McColl BK, Stacker SA. Focus on lymphangiogenesis in tumor metastasis. *Cancer Cell*. 2005;7(2):121-127.
- Zang X, Sullivan PS, Soslow RA, et al. Tumor associated endothelial expression of B7-H3 predicts survival in ovarian carcinomas. *Mod Pathol.* 2010;23(8):1104-1112.
- Kozlova N, Grossman JE, Iwanicki MP, Muranen T. The interplay of the extracellular matrix and stromal cells as a drug target in stroma-rich cancers. *Trends Pharmacol Sci.* 2020;41(3):183-198.
- 59. Estrella V, Chen T, Lloyd M, et al. Acidity generated by the tumor microenvironment drives local invasion. *Can Res.* 2013;73(5):1524-1535.
- Turley SJ, Cremasco V, Astarita JL. Immunological hallmarks of stromal cells in the tumour microenvironment. *Nat Rev Immunol*. 2015;15(11):669-682.
- Garza Trevino EN, Gonzalez PD, Valencia Salgado CI, Martinez GA. Effects of pericytes and colon cancer stem cells in the tumor microenvironment. *Cancer Cell Int.* 2019;19:173.
- 62. Sattiraju A, Mintz A. Pericytes in glioblastomas: multifaceted role within tumor microenvironments and potential for therapeutic interventions. *Adv Exp Med Biol.* 2019;1147:65-91.
- 63. Atiya H, Frisbie L, Pressimone C, Coffman L. Mesenchymal stem cells in the tumor microenvironment. *Adv Exp Med Biol.* 2020;1234:31-42.
- 64. Liu R, Wei S, Chen J, Xu S. Mesenchymal stem cells in lung cancer tumor microenvironment: their biological properties, influence on tumor growth and therapeutic implications. *Cancer Lett*. 2014;353(2):145-152.
- Liu T, Han C, Wang S, et al. Cancer-associated fibroblasts: an emerging target of anti-cancer immunotherapy. J Hematol Oncol. 2019;12(1):86.
- Yang X, Hou J, Han Z, et al. One cell, multiple roles: contribution of mesenchymal stem cells to tumor development in tumor microenvironment. *Cell Biosci.* 2013;3(1):5.
- Trams EG, Lauter CJ, Salem N Jr, Heine U. Exfoliation of membrane ecto-enzymes in the form of micro-vesicles. *Biochem Biophys Acta*. 1981;645(1):63-70.
- Zhou LI, Lv T, Zhang Q, et al. The biology, function and clinical implications of exosomes in lung cancer. *Cancer Lett.* 2017;407:84-92.
- Nazarenko I, Rana S, Baumann A, et al. Cell surface tetraspanin Tspan8 contributes to molecular pathways of exosome-induced endothelial cell activation. *Can Res.* 2010;70(4):1668-1678.
- Liu YI, Luo F, Wang B, et al. STAT3-regulated exosomal miR-21 promotes angiogenesis and is involved in neoplastic processes of transformed human bronchial epithelial cells. *Cancer Lett.* 2016;370(1):125-135.
- Zhuang G, Wu X, Jiang Z, et al. Tumour-secreted miR-9 promotes endothelial cell migration and angiogenesis by activating the JAK-STAT pathway. EMBO J. 2012;31(17):3513-3523.
- Cui H, Seubert B, Stahl E, et al. Tissue inhibitor of metalloproteinases-1 induces a pro-tumourigenic increase of miR-210 in lung adenocarcinoma cells and their exosomes. *Oncogene*. 2015;34(28):3640-3650.
- Grange C, Tapparo M, Collino F, et al. Microvesicles released from human renal cancer stem cells stimulate angiogenesis and formation of lung premetastatic niche. *Can Res.* 2011;71(15):5346-5356.
- Conigliaro A, Costa V, Lo Dico A, et al. CD90+ liver cancer cells modulate endothelial cell phenotype through the release of exosomes containing H19 IncRNA. *Mol Cancer.* 2015;14:155.
- Lang H-L, Hu G-W, Zhang BO, et al. Glioma cells enhance angiogenesis and inhibit endothelial cell apoptosis through the release of exosomes that contain long non-coding RNA CCAT2. Oncol Rep. 2017;38(2):785-798.
- Lang HL, Hu GW, Chen Y, et al. Glioma cells promote angiogenesis through the release of exosomes containing long non-coding RNA POU3F3. Eur Rev Med Pharmacol Sci. 2017;21(5):959-972.

- Ekström EJ, Bergenfelz C, von Bülow V, et al. WNT5A induces release of exosomes containing pro-angiogenic and immunosuppressive factors from malignant melanoma cells. *Mol Cancer*. 2014;13:88.
- Mineo M, Garfield SH, Taverna S, et al. Exosomes released by K562 chronic myeloid leukemia cells promote angiogenesis in a Src-dependent fashion. *Angiogenesis*. 2012;15(1):33-45.
- Kucharzewska P, Christianson HC, Welch JE, et al. Exosomes reflect the hypoxic status of glioma cells and mediate hypoxia-dependent activation of vascular cells during tumor development. *Proc Natl Acad Sci USA*. 2013;110(18):7312-7317.
- Maji S, Chaudhary P, Akopova I, et al. Exosomal Annexin II promotes angiogenesis and breast cancer metastasis. *Mol Cancer Res.* 2017;15(1):93-105.
- Kosaka N, Iguchi H, Hagiwara K, Yoshioka Y, Takeshita F, Ochiya T. Neutral sphingomyelinase 2 (nSMase2)-dependent exosomal transfer of angiogenic microRNAs regulate cancer cell metastasis. *J Biol Chem.* 2013;288(15):10849-10859.
- 82. Huang Z, Feng Y. Exosomes derived from hypoxic colorectal cancer cells promote angiogenesis through Wnt4-induced beta-catenin signaling in endothelial cells. *Oncol Res.* 2017;25(5):651-661.
- Millimaggi D, Mari M, D'Ascenzo S, et al. Tumor vesicle-associated CD147 modulates the angiogenic capability of endothelial cells. *Neoplasia*. 2007;9(4):349-357.
- Dolo V, Ginestra A, Cassara D, Ghersi G, Nagase H, Vittorelli ML. Shed membrane vesicles and selective localization of gelatinases and MMP-9/TIMP-1 complexes. *Ann N Y Acad Sci.* 1999;878:497-499.
- Dolo V, D'Ascenzo S, Violini S, et al. Matrix-degrading proteinases are shed in membrane vesicles by ovarian cancer cells in vivo and in vitro. *Clin Exp Metas.* 1999;17(2):131-140.
- Shu SL, Yang Y, Allen CL, et al. Metabolic reprogramming of stromal fibroblasts by melanoma exosome microRNA favours a pre-metastatic microenvironment. *Sci Rep.* 2018;8(1):12905.
- Gabrilovich DI, Nagaraj S. Myeloid-derived suppressor cells as regulators of the immune system. *Nat Rev Immunol*. 2009;9(3):162-174.
- Chen W, Jiang J, Xia W, Huang J. Tumor-related exosomes contribute to tumor-promoting microenvironment: an immunological perspective. *J Immunol Res.* 2017;2017:1073947.
- Veerman RE, Gucluler Akpinar G, Eldh M, Gabrielsson S. Immune cell-derived extracellular vesicles - functions and therapeutic applications. *Trends Mol Med.* 2019;25(5):382-394.
- Fuchs D, Daniel V, Sadeghi M, Opelz G, Naujokat C. Salinomycin overcomes ABC transporter-mediated multidrug and apoptosis resistance in human leukemia stem cell-like KG-1a cells. *Biochem Biophys Res Comm.* 2010;394(4):1098-1104.
- Majmundar AJ, Wong WJ, Simon MC. Hypoxia-inducible factors and the response to hypoxic stress. *Mol Cell*. 2010;40(2):294-309.
- 92. Najafi M, Farhood B, Mortezaee K. Cancer stem cells (CSCs) in cancer progression and therapy. *J Cell Physiol*. 2019;234(6):8381-8395.
- Najafi M, Mortezaee K, Majidpoor J. Cancer stem cell (CSC) resistance drivers. *Life Sci.* 2019;234:116781.
- Cheloni G, Tanturli M, Tusa I, et al. Targeting chronic myeloid leukemia stem cells with the hypoxia-inducible factor inhibitor acriflavine. *Blood*. 2017;130(5):655-665.
- 95. Vermeulen L, De Sousa E Melo F, van der Heijden M, et al. Wnt activity defines colon cancer stem cells and is regulated by the microenvironment. *Nat Cell Biol.* 2010;12(5):468-476.
- Liu Y, Liu B, Zhang GQ, Zou JF, Zou ML, Cheng ZS. Calpain inhibition attenuates bleomycin-induced pulmonary fibrosis via switching the development of epithelial-mesenchymal transition. *Naunyn Schmiedebergs Arch Pharmacol.* 2018;391(7):695-704.
- Catalano V, Turdo A, DiFranco S, Dieli F, Todaro M, Stassi G. Tumor and its microenvironment: a synergistic interplay. *Semin Cancer Biol.* 2013;23(6 Pt B):522-532.

98. Guo S, Deng CX. Effect of stromal cells in tumor microenvironment on metastasis initiation. *Int J Biol Sci*. 2018;14(14):2083-2093.

^Droliferation

- 99. Jing Y, Han Z, Zhang S, Liu Y, Wei L. Epithelial-Mesenchymal Transition in tumor microenvironment. *Cell Biosci*. 2011;1:29.
- 100. Goulet CR, Champagne A, Bernard G, et al. Cancer-associated fibroblasts induce epithelial-mesenchymal transition of bladder cancer cells through paracrine IL-6 signalling. BMC Cancer. 2019;19(1):137.
- Fiori ME, Di Franco S, Villanova L, Bianca P, Stassi G, De Maria R. Cancer-associated fibroblasts as abettors of tumor progression at the crossroads of EMT and therapy resistance. *Mol Cancer*. 2019;18(1):70.
- Chen QK, Lee K, Radisky DC, Nelson CM. Extracellular matrix proteins regulate epithelial-mesenchymal transition in mammary epithelial cells. *Differentiation*. 2013;86(3):126-132.
- 103. Li H, Xu F, Li S, Zhong A, Meng X, Lai M. The tumor microenvironment: An irreplaceable element of tumor budding and epithelial-mesenchymal transition-mediated cancer metastasis. *Cell Adh Migr.* 2016;10(4):434-446.
- 104. White BH, Ewer J. Neural and hormonal control of postecdysial behaviors in insects. *Annu Rev Entomol.* 2014;59:363-381.
- 105. Chong CR, Janne PA. The quest to overcome resistance to EGFRtargeted therapies in cancer. *Nat Med*. 2013;19(11):1389-1400.
- Roma-Rodrigues C, Mendes R, Baptista PV, Fernandes AR. Targeting tumor microenvironment for cancer therapy. Int J Mol Sci. 2019;20(4):840.
- 107. Belli C, Trapani D, Viale G, et al. Targeting the microenvironment in solid tumors. *Cancer Treat Rev.* 2018;65:22-32.
- Darvin P, Toor SM, Sasidharan Nair V, Elkord E. Immune checkpoint inhibitors: recent progress and potential biomarkers. *Exp Mol Med.* 2018;50(12):1-11.
- 109. Capasso A, Lang J, Pitts TM, et al. Characterization of immune responses to anti-PD-1 mono and combination immunotherapy in hematopoietic humanized mice implanted with tumor xenografts. *J Immunotherapy Cancer*. 2019;7(1):37.
- 110. Morecki S, Yacovlev E, Diab A, Slavin S. Allogeneic cell therapy for a murine mammary carcinoma. *Can Res.* 1998;58(17):3891-3895.
- 111. Dumauthioz N, Labiano S, Romero P. Tumor resident memory T cells: new players in immune surveillance and therapy. *Front Immunol*. 2018;9:2076.
- 112. Jackson HJ, Rafiq S, Brentjens RJ. Driving CAR T-cells forward. Nat Rev Clin Oncol. 2016;13(6):370-383.
- 113. Zhang C, Liu J, Zhong JF, Zhang X. Engineering CAR-T cells. Biomark Res. 2017;5:22.
- 114. Arabi F, Torabi-Rahvar M, Shariati A, Ahmadbeigi N, Naderi M. Antigenic targets of CAR T Cell Therapy. A retrospective view on clinical trials. *Exp Cell Res.* 2018;369(1):1-10.
- Porter DL, Levine BL, Kalos M, Bagg A, June CH. Chimeric antigen receptor-modified T cells in chronic lymphoid leukemia. New Engl J Med. 2011;365(8):725-733.
- 116. Porter DL, Hwang W-T, Frey NV, et al. Chimeric antigen receptor T cells persist and induce sustained remissions in relapsed refractory chronic lymphocytic leukemia. *Sci Transl Med.* 2015;7(303):303ra139.
- 117. Ellis TM, Simms PE, Slivnick DJ, Jack HM, Fisher RI. CD30 is a signal-transducing molecule that defines a subset of human activated CD45RO+ T cells. J Immunol. 1993;151(5):2380-2389.
- 118. Louis CU, Savoldo B, Dotti G, et al. Antitumor activity and longterm fate of chimeric antigen receptor-positive T cells in patients with neuroblastoma. *Blood.* 2011;118(23):6050-6056.
- 119. Gao H, Li K, Tu H, et al. Development of T cells redirected to glypican-3 for the treatment of hepatocellular carcinoma. *Clin Cancer Res.* 2014;20(24):6418-6428.
- 120. Singh AK, McGuirk JP. CAR T cells: continuation in a revolution of immunotherapy. *Lancet Oncol.* 2020;21(3):e168-e178.

EY-Proliferation

- June CH, O'Connor RS, Kawalekar OU, Ghassemi S, Milone MC. CAR T cell immunotherapy for human cancer. *Science*. 2018;359(6382):1361-1365.
- 122. Bedoya F, Frigault MJ, Maus MV. The flipside of the power of engineered T cells: observed and potential toxicities of genetically modified T cells as therapy. *Mol Ther.* 2017;25(2):314-320.
- 123. Björklund AT, Carlsten M, Sohlberg E, et al. Complete remission with reduction of high-risk clones following haploidentical NK-cell therapy against MDS and AML. *Clin Cancer Res.* 2018;24(8):1834-1844.
- 124. Fu W, Lei C, Liu S, et al. CAR exosomes derived from effector CAR-T cells have potent antitumour effects and low toxicity. *Nat Commun.* 2019;10(1):4355.
- 125. Narang P, Mittal V, Menon V, Bhaduri A, Chaudhuri BR, Honavar SG. Primary limbal stem cell transplantation in the surgical management of extensive ocular surface squamous neoplasia involving the limbus. *Indian J Ophthalmol.* 2018;66(11):1569-1573.
- 126. Gaut D, Schiller GJ. Hematopoietic stem cell transplantation in primary central nervous system lymphoma: a review of the literature. *Int J Hematol.* 2019;109(3):260-277.
- 127. Hammert LC, Ball ED. Purging autologous bone marrow with monoclonal antibodies for transplantation in acute myelogenous leukemia. *Blood Rev.* 1997;11(2):80-90.
- 128. Stuart RK. Autologous bone marrow transplantation for leukemia. *Semin Oncol.* 1993;20(5 Suppl 6):40-54.
- Pettengell R. Autologous stem cell transplantation in follicular non-Hodgkin's lymphoma. *Bone Marrow Transplant*. 2002;29(Suppl 1):S1-4.
- 130. Giralt S. Stem cell transplantation for multiple myeloma: current and future status. *Hematology*. 2011;2011:191-196.
- Chopra N, Choudhury S, Bhargava S, Wajid S, Ganguly NK. Potentials of "stem cell-therapy" in pancreatic cancer: an update. *Pancreatology*. 2019;19(8):1034-1042.
- Singh AK, McGuirk JP. Allogeneic stem cell transplantation: a historical and scientific overview. *Can Res.* 2016;76(22):6445-6451.
- Gündüz M, Özen M, Şahin U, et al. Subsequent malignancies after allogeneic hematopoietic stem cell transplantation. *Clin Transplant*. 2017;31(7):e12987.
- Ades L, Guardiola P, Socie G. Second malignancies after allogeneic hematopoietic stem cell transplantation: new insight and current problems. *Blood Rev.* 2002;16(2):135-146.
- Danylesko I, Shimoni A. Second malignancies after hematopoietic stem cell transplantation. *Curr Treat Options Oncol.* 2018;19(2):9.
- Munoz J, Shah N, Rezvani K, et al. Concise review: umbilical cord blood transplantation: past, present, and future. *Stem Cells Transl Med.* 2014;3(12):1435-1443.
- Anagnostopoulos A, Giralt S. Critical review on non-myeloablative stem cell transplantation (NST). *Critical Rev Oncol/Hematol.* 2002;44(2):175-190.
- 138. Bregni M, Bernardi M, Ciceri F, Peccatori J. Allogeneic stem cell transplantation for the treatment of advanced solid tumors. *Springer Semin Immunopathol.* 2004;26(1–2):95-108.
- Morecki S, Moshel Y, Gelfend Y, Pugatsch T, Slavin S. Induction of graft vs. tumor effect in a murine model of mammary adenocarcinoma. *Int J Cancer*. 1997;71(1):59-63.
- 140. Cote GM, Hochberg EP, Muzikansky A, et al. Autologous stem cell transplantation with thiotepa, busulfan, and cyclophosphamide (TBC) conditioning in patients with CNS involvement by non-Hodgkin lymphoma. *Biol Blood Marrow Transplant*. 2012;18(1):76-83.
- 141. Soussain C, Suzan F, Hoang-Xuan K, et al. Results of intensive chemotherapy followed by hematopoietic stem-cell rescue in 22 patients with refractory or recurrent primary CNS lymphoma or intraocular lymphoma. *J Clin Oncol.* 2001;19(3):742-749.

- 142. Arya M, Chao D, Patel HR. Allogeneic hematopoietic stem-cell transplantation: the next generation of therapy for metastatic renal cell cancer. *Nat Clin Pract Onco*. 2004;1(1):32-38.
- Bregni M, Herr W, Blaise D. Solid Tumor Working Party of E. Allogeneic stem cell transplantation for renal cell carcinoma. *Expert Rev Anticancer Ther.* 2011;11(6):901-911.
- 144. Chamberlain G, Fox J, Ashton B, Middleton J. Concise review: mesenchymal stem cells: their phenotype, differentiation capacity, immunological features, and potential for homing. *Stem Cells*. 2007;25(11):2739-2749.
- 145. Kern S, Eichler H, Stoeve J, Kluter H, Bieback K. Comparative analysis of mesenchymal stem cells from bone marrow, umbilical cord blood, or adipose tissue. *Stem Cells*. 2006;24(5):1294-1301.
- Caplan AI. Adult mesenchymal stem cells for tissue engineering versus regenerative medicine. J Cell Physiol. 2007;213(2):341-347.
- 147. Jin H, Bae Y, Kim M, et al. Comparative analysis of human mesenchymal stem cells from bone marrow, adipose tissue, and umbilical cord blood as sources of cell therapy. *Int J Mol Sci.* 2013;14(9):17986-18001.
- 148. Song N, Gao L, Qiu H, et al. Mouse bone marrow-derived mesenchymal stem cells inhibit leukemia/lymphoma cell proliferation in vitro and in a mouse model of allogeneic bone marrow transplant. *Int J Mol Med.* 2015;36(1):139-149.
- 149. Liang R, Huang G-S, Wang Z, et al. Effects of human bone marrow stromal cell line (HFCL) on the proliferation, differentiation and apoptosis of acute myeloid leukemia cell lines U937, HL-60 and HL-60/VCR. Int J Hematol. 2008;87(2):152-166.
- 150. Maj M, Bajek A, Nalejska E, et al. Influence of mesenchymal stem cells conditioned media on proliferation of urinary tract cancer cell lines and their sensitivity to ciprofloxacin. J Cell Biochem. 2017;118(6):1361-1368.
- 151. Farahmand L, Esmaeili R, Eini L, Majidzadeh AK. The effect of mesenchymal stem cell-conditioned medium on proliferation and apoptosis of breast cancer cell line. J Cancer Res Therapeutics. 2018;14(2):341-344.
- 152. Mortezaee K. CXCL12/CXCR4 axis in the microenvironment of solid tumors: a critical mediator of metastasis. *Life Sci.* 2020;249:117534.
- 153. Vallabhaneni KC, Hassler M-Y, Abraham A, et al. Mesenchymal stem/stromal cells under stress increase osteosarcoma migration and apoptosis resistance via extracellular vesicle mediated communication. *PLoS One.* 2016;11(11):e0166027.
- Lis R, Touboul C, Mirshahi P, et al. Tumor associated mesenchymal stem cells protects ovarian cancer cells from hyperthermia through CXCL12. *Int J Cancer*. 2011;128(3):715-725.
- Huang WH, Chang MC, Tsai KS, Hung MC, Chen HL, Hung SC. Mesenchymal stem cells promote growth and angiogenesis of tumors in mice. *Oncogene*. 2013;32(37):4343-4354.
- Karnoub AE, Dash AB, Vo AP, et al. Mesenchymal stem cells within tumour stroma promote breast cancer metastasis. *Nature*. 2007;449(7162):557-563.
- 157. Albarenque SM, Zwacka RM, Mohr A. Both human and mouse mesenchymal stem cells promote breast cancer metastasis. *Stem Cell Res.* 2011;7(2):163-171.
- Zheng H, Zou W, Shen J, et al. Opposite effects of coinjection and distant injection of mesenchymal stem cells on breast tumor cell growth. Stem Cells Transl Med. 2016;5(9):1216-1228.
- 159. O'Malley G, Treacy O, Lynch K, et al. Stromal cell PD-L1 inhibits CD8(+) T-cell antitumor immune responses and promotes colon cancer. *Cancer Immunol Res*. 2018;6(11):1426-1441.
- 160. Hendijani F, Javanmard SH, Sadeghi-aliabadi H. Human Wharton's jelly mesenchymal stem cell secretome display antiproliferative effect on leukemia cell line and produce additive cytotoxic effect in combination with doxorubicin. *Tissue Cell*. 2015;47(3):229-234.

17 of 17

- Wu S, Ju GQ, Du T, Zhu YJ, Liu GH. Microvesicles derived from human umbilical cord Wharton's jelly mesenchymal stem cells attenuate bladder tumor cell growth in vitro and in vivo. *PLoS One*. 2013;8(4):e61366.
- Bruno S, Collino F, Deregibus MC, Grange C, Tetta C, Camussi G. Microvesicles derived from human bone marrow mesenchymal stem cells inhibit tumor growth. *Stem Cells Dev.* 2013;22(5):758-771.
- 163. Du T, Ju G, Wu S, et al. Microvesicles derived from human Wharton's jelly mesenchymal stem cells promote human renal cancer cell growth and aggressiveness through induction of hepatocyte growth factor. *PLoS One*. 2014;9(5):e96836.
- 164. Studeny M, Marini FC, Champlin RE, Zompetta C, Fidler IJ, Andreeff M. Bone marrow-derived mesenchymal stem cells as vehicles for interferon-beta delivery into tumors. *Can Res.* 2002;62(13):3603-3608.
- 165. Jiang X, Fitch S, Wang C, et al. Nanoparticle engineered TRAILoverexpressing adipose-derived stem cells target and eradicate glioblastoma via intracranial delivery. *Proc Natl Acad Sci USA*. 2016;113(48):13857-13862.
- Moniri MR, Sun X-Y, Rayat J, et al. TRAIL-engineered pancreas-derived mesenchymal stem cells: characterization and cytotoxic effects on pancreatic cancer cells. *Cancer Gene Ther.* 2012;19(9):652-658.
- 167. Houthuijzen JM, Daenen LG, Roodhart JM, Voest EE. The role of mesenchymal stem cells in anti-cancer drug resistance and tumour progression. *Br J Cancer*. 2012;106(12):1901-1906.
- 168. Paiboon N, Kamprom W, Manochantr S, et al. Gestational tissue-derived human mesenchymal stem cells use distinct combinations of bioactive molecules to suppress the proliferation of human hepatoblastoma and colorectal cancer cells. *Stem Cells Int.* 2019;2019:9748795.
- Conrad C, Hüsemann Y, Niess H, et al. Linking transgene expression of engineered mesenchymal stem cells and angiopoietin-1-induced differentiation to target cancer angiogenesis. *Ann Surg.* 2011;253(3):566-571.
- 170. Niess H, Bao QI, Conrad C, et al. Selective targeting of genetically engineered mesenchymal stem cells to tumor stroma microenvironments using tissue-specific suicide gene expression suppresses growth of hepatocellular carcinoma. *Ann Surg.* 2011;254(5):767-774; discussion 774-765.
- Kauer TM, Figueiredo JL, Hingtgen S, Shah K. Encapsulated therapeutic stem cells implanted in the tumor resection cavity induce cell death in gliomas. *Nat Neurosci.* 2011;15(2):197-204.
- Jiang X, Wang C, Fitch S, Yang F. Targeting tumor hypoxia using nanoparticle-engineered CXCR4-overexpressing adipose-derived stem cells. *Theranostics*. 2018;8(5):1350-1360.
- 173. Zhou S, Abdouh M, Arena V, Arena M, Arena GO. Reprogramming malignant cancer cells toward a benign phenotype following exposure to human embryonic stem cell microenvironment. *PLoS One*. 2017;12(1):e0169899.

- Ladstatter S, Tachibana-Konwalski K. A surveillance mechanism ensures repair of DNA lesions during zygotic reprogramming. *Cell*. 2016;167(7):1774-1787.e13.
- 175. Lee LM, Seftor EA, Bonde G, Cornell RA, Hendrix MJ. The fate of human malignant melanoma cells transplanted into zebrafish embryos: assessment of migration and cell division in the absence of tumor formation. *Developmental*. 2005;233(4):1560-1570.
- Hendrix MJ, Seftor EA, Seftor RE, Kasemeier-Kulesa J, Kulesa PM, Postovit LM. Reprogramming metastatic tumour cells with embryonic microenvironments. *Nat Rev Cancer*. 2007;7(4):246-255.
- 177. Durr M, Harder F, Merkel A, Bug G, Henschler R, Muller AM. Chimaerism and erythroid marker expression after microinjection of human acute myeloid leukaemia cells into murine blastocysts. *Oncogene*. 2003;22(57):9185-9191.
- 178. Postovit L-M, Margaryan NV, Seftor EA, et al. Human embryonic stem cell microenvironment suppresses the tumorigenic phenotype of aggressive cancer cells. *Proc Natl Acad Sci U S A*. 2008;105(11):4329-4334.
- 179. Zhou C, Huang Z, Li P, et al. Safety and efficacy of embryonic stem cell microenvironment in a leukemia mouse model. *Stem Cells Dev.* 2014;23(15):1741-1754.
- Liu J, Huang Z, Yang L, et al. Embryonic stem cells modulate the cancer-permissive microenvironment of human uveal melanoma. *Theranostics*. 2019;9(16):4764-4778.
- Wang C, Wang X, Liu J, et al. Embryonic stem cell microenvironment suppresses the malignancy of cutaneous melanoma cells by down-regulating PI3K/AKT pathway. *Cancer Med.* 2019;8(9):4265-4277.
- 182. Proietti S, Cucina A, Pensotti A, et al. Active fraction from embryo fish extracts induces reversion of the malignant invasive phenotype in breast cancer through down-regulation of TCTP and modulation of E-cadherin/beta-catenin pathway. *Int J Mol Sci.* 2019;20(9):2151.
- He N, Feng G, Li Y, et al. Embryonic stem cell preconditioned microenvironment suppresses tumorigenic properties in breast cancer. Stem Cell Res Ther. 2016;7(1):95.
- 184. Lan G, Lin Z, Zhang J, et al. Notch pathway is involved in the suppression of colorectal cancer by embryonic stem cell microenvironment. Onco Targets Ther. 2019;12:2869-2878.

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