## **COMMENTARY**



## Non-linear GABA<sub>A</sub> receptors promote synaptic inhibition in developing neurons

Commentary on Lodge et al. (2021) Sparsification of AP firing in adult-born hippocampal granule cells via voltage-dependent  $\alpha$ 5-GABA $_A$  receptors. Cell Rep 37:109768

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Synaptic inhibition is chiefly mediated by GABA<sub>A</sub> receptors (GABAARs). GABAARs form pentameric anion channels that typically assemble from two  $\alpha$ , two  $\beta$ , and one tertiary  $(\gamma, \delta, \varepsilon, \theta, \pi, \rho)$  subunit, with different subunit combinations conferring different biophysical properties. GABA<sub>A</sub>R-mediated inhibition of action potential (AP) firing results from hyperpolarization and/or an increase in membrane conductance shunting excitatory inputs. Therefore, to ensure efficient inhibition, most mature neurons maintain a low chloride concentration ([Cl<sup>-</sup>]<sub>i</sub>). In contrast, immature neurons accumulate chloride [12], thus favoring depolarizing GABA responses in vivo [7, 14]. Whether excitatory or inhibitory actions of depolarizing GABA predominate is difficult to predict. This is due to complex interactions between the membrane  $(V_{\rm m})$  and chloride reversal potentials, the GABAAR conductance, and the timing and location of synaptic inputs [4, 6, 8, 10]. However, in vivo studies confirmed that depolarizing GABA can effectively inhibit AP firing already in developing neurons [7, 13]. Although these observations suggest that immature neurons can prevent excess excitation through depolarizing GABA, the underlying mechanisms are not fully understood.

In their elegant recent study, Lodge and colleagues [9] discovered a crucial role for  $\alpha 5$  subunit-containing GABA<sub>A</sub>Rs ( $\alpha 5$ -GABA<sub>A</sub>Rs) in this process. Focusing on adult-born hippocampal granule cells (GCs), the authors analyzed GABAergic inputs mediated by soma-targeting parvalbumin (PV) and dendrite-targeting somatostatin (SOM) interneurons. In acute hippocampal slices, in which

young GCs are depolarized by GABA [4], both somatic and dendritic synapses were found to use similar GABA<sub>A</sub>Rs with a striking non-linear voltage-dependence, pharmacologically identified as  $\alpha$ 5-GABA<sub>A</sub>Rs. Their conductance at depolarized potentials close to AP threshold (–35 mV) was fourfold larger than at resting  $V_{\rm m}$  (–80 mV). Previous work demonstrated that  $\alpha$ 5-GABA<sub>A</sub>Rs are high-affinity, slowly desensitizing receptors that can dynamically redistribute between extrasynaptic and synaptic sites in an activity-dependent manner [2, 3], thus mediating both tonic [5] and phasic [11] inhibition.

What are the functional consequences of outwardly rectifying  $\alpha 5$ -GABA<sub>A</sub>Rs? It has long been known that, under certain conditions, depolarizing GABA can enhance the activation of NMDA receptors (NMDARs) and might also facilitate AP firing. This could be of developmental importance, since NMDARs are required for several forms of synaptic plasticity, such as synapse unsilencing [1]. However, as the number of active GABAergic synapses increases, facilitation shifts to inhibition due to increased shunting of glutamatergic currents. Using a rigorous electrophysiological approach in combination with detailed biophysical modeling, the authors demonstrated that outward rectification of α5-GABA<sub>A</sub>Rs results in a dominance of the shunting effect at considerably lower levels of GABAergic activity. Consequently, voltage-dependent  $\alpha 5$ -GABA<sub>A</sub>Rs inhibit NMDARdependent currents and AP firing in a broader range of GABAergic activity. Collectively, the voltage-dependent increase in conductance of α5-GABA<sub>A</sub>Rs around AP threshold is crucial for preventing excess synaptic excitation in young GCs, when GABA is depolarizing.

Comparing the properties of GABAergic synapses in young with those in mature GCs, the authors discovered an intriguing example of developmental regulation. Whereas SOM-GC synapses utilize non-linear  $\alpha 5$ -GABA<sub>A</sub>Rs



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throughout development,  $\alpha 5$ -GABA<sub>A</sub>Rs become developmentally excluded from PV-GC contacts. Thus, as steady-state [Cl<sup>-</sup>]<sub>i</sub> declines during development [4], PV interneurons transition to targeting linear GABA<sub>A</sub>Rs lacking  $\alpha 5$  subunits. These results extend previous work indicating that, during the circuit integration of GCs,  $\alpha 5$ -GABA<sub>A</sub>Rs redistribute from synaptic to extrasynaptic sites, which strengthens their contribution to tonic inhibition [2].

The study by Lodge and colleagues [9] raises exciting questions. (1) What is the impact of non-linear α5-GABA<sub>A</sub>Rs in dendritic synapses of mature neurons? The authors' modeling results indicate that both linear and nonlinear GABA<sub>A</sub>Rs can mediate effective inhibition if steadystate [Cl<sup>-</sup>]; is low. However, this analysis did not consider activity-dependent chloride loads, which could cause substantial [Cl<sup>-</sup>]; shifts, especially in dendritic compartments (due to their high surface-to-volume ratio). (2) How is the abundance of outwardly rectifying vs. linear GABAARs regulated? Such a regulation may represent a powerful mechanism by which (developing) neurons can tune inhibition, independent of changes in [Cl<sup>-</sup>]<sub>i</sub>. (3) Does the use of outwardly rectifying α5-GABA<sub>A</sub>Rs reflect a general principle by which developing neurons prevent excessive excitation? Since α5-GABA<sub>A</sub>Rs inhibit spontaneous cortical network activity in neonatal mice in vivo [7], this seems plausible. In sum, by showing that non-linear α5-GABA<sub>A</sub>Rs promote synaptic inhibition with depolarizing GABA, Lodge and colleagues [9] bring us one step closer to understanding how developing neurons prevent runaway excitation during synaptogenesis and circuit integration.

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## **Declarations**

**Conflict of interest** The author declares no competing interests.

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## References

- Chancey JH, Adlaf EW, Sapp MC, Pugh PC, Wadiche JI, Overstreet-Wadiche LS (2013) GABA depolarization is required for experience-dependent synapse unsilencing in adult-born neurons. J Neurosci 33:6614–6622. https://doi.org/10.1523/JNEUROSCI. 0781-13.2013
- Davenport CM, Rajappa R, Katchan L, Taylor CR, Tsai MC, Smith CM, de Jong JW, Arnold DB, Lammel S, Kramer RH (2021) Relocation of an extrasynaptic GABAA receptor to inhibitory synapses freezes excitatory synaptic strength and preserves memory. Neuron 109(123–134):e124. https://doi.org/10.1016/j. neuron.2020.09.037
- Hausrat TJ, Muhia M, Gerrow K, Thomas P, Hirdes W, Tsukita S, Heisler FF, Herich L, Dubroqua S, Breiden P, Feldon J, Schwarz JR, Yee BK, Smart TG, Triller A, Kneussel M (2015) Radixin regulates synaptic GABAA receptor density and is essential for reversal learning and short-term memory. Nat Commun 6:6872. https://doi.org/10.1038/ncomms7872
- Heigele S, Sultan S, Toni N, Bischofberger J (2016) Bidirectional GABAergic control of action potential firing in newborn hippocampal granule cells. Nat Neurosci 19:263–270. https://doi.org/10.1038/nn.4218
- Holter NI, Zylla MM, Zuber N, Bruehl C, Draguhn A (2010) Tonic GABAergic control of mouse dentate granule cells during postnatal development. Eur J Neurosci 32:1300–1309. https://doi. org/10.1111/j.1460-9568.2010.07331.x
- Jean-Xavier C, Mentis GZ, O'Donovan MJ, Cattaert D, Vinay L (2007) Dual personality of GABA/glycine-mediated depolarizations in immature spinal cord. Proc Natl Acad Sci U S A 104:11477–11482. https://doi.org/10.1073/pnas.0704832104
- Kirmse K, Kummer M, Kovalchuk Y, Witte OW, Garaschuk O, Holthoff K (2015) GABA depolarizes immature neurons and inhibits network activity in the neonatal neocortex in vivo. Nat Commun 6:7750. https://doi.org/10.1038/ncomms8750
- Kolbaev SN, Achilles K, Luhmann HJ, Kilb W (2011) Effect of depolarizing GABA(A)-mediated membrane responses on excitability of Cajal-Retzius cells in the immature rat neocortex. J Neurophysiol 106:2034–2044. https://doi.org/10.1152/jn.00699.2010
- Lodge M, Hernandez MC, Schulz JM, Bischofberger J (2021) Sparsification of AP firing in adult-born hippocampal granule cells via voltage-dependent alpha5-GABAA receptors. Cell Rep 37:109768. https://doi.org/10.1016/j.celrep.2021.109768
- Murata Y, Colonnese MT (2020) GABAergic interneurons excite neonatal hippocampus in vivo. Sci Adv 6:eaba1430. https://doi. org/10.1126/sciadv.aba1430
- Schulz JM, Knoflach F, Hernandez MC, Bischofberger J (2018) Dendrite-targeting interneurons control synaptic NMDA-receptor activation via nonlinear alpha5-GABAA receptors. Nat Commun 9:3576. https://doi.org/10.1038/s41467-018-06004-8



- Sulis Sato S, Artoni P, Landi S, Cozzolino O, Parra R, Pracucci E, Trovato F, Szczurkowska J, Luin S, Arosio D, Beltram F, Cancedda L, Kaila K, Ratto GM (2017) Simultaneous two-photon imaging of intracellular chloride concentration and pH in mouse pyramidal neurons in vivo. Proc Natl Acad Sci U S A 114:E8770–E8779, https://doi.org/10.1073/pnas.1702861114
- Valeeva G, Tressard T, Mukhtarov M, Baude A, Khazipov R (2016) An optogenetic approach for investigation of excitatory and inhibitory network GABA actions in mice expressing channelrhodopsin-2 in GABAergic neurons. J Neurosci 36:5961–5973. https://doi.org/10.1523/JNEUROSCI.3482-15.2016
- van Rheede JJ, Richards BA, Akerman CJ (2015) Sensory-evoked spiking behavior emerges via an experience-dependent plasticity mechanism. Neuron 87:1050–1062. https://doi.org/10.1016/j. neuron.2015.08.021

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