# Identification of three new Alu Yb subfamilies by source tracking of recently integrated Alu Yb elements 

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#### Abstract

Background: Alu elements are the most abundant mobile elements in the human genome, with over 1 million copies and constituting more than $10 \%$ of the genome. The majority of these Alu elements were inserted into the primate genome 35 to 60 million years ago, but certain subfamilies of Alu elements are relatively very new and suspected to be still evolving. We attempted to trace the source/master copies of all human-specific members of the Alu Yb lineage using a computational approach by clustering similar Yb elements and constructing an evolutionary relation among the members of a cluster.

Results: We discovered that one copy of Yb8 at 10p14 is the source of several active Yb8 copies, which retrotransposed to generate 712 copies or $54 \%$ of all human-specific Yb8 elements. We detected eight other Yb8 elements that had generated ten or more copies, potentially acting as 'stealth drivers'. One Yb8 element at 14 q 32.31 seemed to act as the source copy for all Yb9 elements tested, having producing 13 active Yb9 elements, and subsequently generated a total of 131 full-length copies. We identified and characterized three new subclasses of Yb elements: Yb8a1, Yb10 and Yb11. Their copy numbers in the reference genome are 75, 8 and 16. We analysed personal genome data from the 1000 Genome Project and detected an additional 6 Yb8a1, 3 Yb10 and 15 Yb 11 copies outside the reference genome. Our analysis indicates that the Yb8a1 subfamily has a similar age to Yb9 (1.93 million years and 2.15 million years, respectively, while Yb10 and Yb11 evolved only 1.4 and 0.71 million years ago, suggesting a linear evolutionary path from Yb8a1 to Yb10 and then to Yb11. Our preliminary data indicate that members in Yb10 and Yb11 are mostly polymorphic, indicating their young age. Conclusions: Our findings suggest that the Yb lineage is still evolving with new subfamilies being formed. Due to their very young age and the high rate of being polymorphic, insertions from these young subfamilies are very useful genetic markers for studying human population genetics and migration patterns, and the trend for mobile element insertions in the human genome.


Keywords: Mobile element, Alu, evolution, Alu Yb8a1, Alu Yb10, Alu Yb11

## Background

Alu elements are the most successful short interspersed elements (SINEs) in primate genomes. Alu elements have proliferated significantly throughout primate evolution and have expanded to more than 1 million copies in the human genome, constituting over $10 \%$ of the genome by mass $[1,2]$. The majority of these elements are

[^0]suspected to have been inserted in the primate genome 35 to 60 million years ago, and since then the proliferation rate has reduced significantly by over 100 fold [3]. Thus, despite the large number of copies present in the human genome, only a small fraction of Alu elements are still active and capable of generating new copies [4-6]. The activity of Alu elements has generated different subfamilies of varying ages, each subfamily being defined and characterized by a set of diagnostic mutations [7]. Each subfamily is thought to have expanded when its master or source copy accumulated a mutation and
then actively transposed to new locations at different rates and time periods of evolution $[8,9]$.

The vast majority of the Alu elements currently found in the human genome were inserted before the divergence of humans and chimps, and thus are shared by all individuals of both species. The small fraction of Alu elements that have been recently inserted into the human genome are mostly restricted to several closely related young subfamilies, with the majority of these young elements being from the Ya 5 and Yb 8 Alu subfamilies [10,11]. Since almost all of these young Alu elements were inserted into the human genome after the humanchimp divergence, they are only found in humans. Some of these young active Alu elements have accumulated new mutations and have acted as source or master copies by generating subsets of elements that are identifiable as new subfamilies. Some of these subfamilies are so recent that they have members that are polymorphic for their presence or absence between individuals and/or populations [12-14]. The availability of a complete human reference genome and large quantities of individual genomic data from the 1000 Genome Project have facilitated the identification of these subfamilies and their level of polymorphism [15,16]. The homoplasy-free nature of Alu elements makes their polymorphic insertions very useful in phylogenetic studies, human population studies, forensics and DNA fingerprinting [9,17-20].

Our study specifically focuses on human-specific Alu elements from the Yb lineage, mainly because they are the second largest young family by the number of copies in the human genome, comprising $40 \%$ of all humanspecific Alu elements with more than $30 \%$ of these copies being polymorphic between individuals and/or populations [15,16,21]. Alu Yb 8 is the major subset of this family. Its high rate of being human-specific and polymorphic among humans and its involvement in human diseases via de novo insertion suggest that this subfamily is still actively retrotransposing [22,23]. The Yb8 subfamily is characterized by a tandem duplication of seven nucleotides from the 246 th to the 252 nd position of the AluY consensus sequence. The concurrent mutation and transposition of certain Yb 8 elements generated the Yb 9 subfamily, which was the latest Yb subfamily identified before this study and characterized by a C to G transversion at the 274th position [9]. In this study, using a computational approach we performed a genome-wide analysis of all human-specific Yb elements to identify their source copies and to track their recent evolutionary pathway. We successfully detected at least one driver copy for Yb 8 and one Yb 8 element that is potentially the source copy for the Yb 9 subfamily. We also identified and characterized three new subfamilies in the Yb lineage: $\mathrm{Yb} 8 \mathrm{a} 1, \mathrm{Yb} 10$, and $\mathrm{Yb} 11 . \mathrm{Yb} 11$ is the youngest Yb subfamily reported to date.

## Results and discussion

## Evolution of recent Alu Yb elements

Of all Yb copies found in the human genome, $80 \%$ (2,545 of 3,179 ) are identified as human-specific ( hsYb ), that is, they became integrated into the human genome after the human-chimp divergence, and they only include members of the Yb 8 and Yb 9 subfamilies (Tang et al., manuscript in preparation). In this study, we included all full-length hsYb elements in an attempt to assess their evolutionary pattern and backtrack their putative source genes. All such hsYb elements were aligned against all $\mathrm{Yb} 7,8$ and 9 sequences in the reference genome to group similar sequences into clusters. For each cluster, a phylogenetic tree was constructed with an outgroup subfamily consensus sequence as its root to assess the evolutionary relation among clusters and members of each cluster. The phylogenetic topology for each cluster can provide information on the potential parent copy for other members in the cluster. In an analysis involving only hsYb8 elements and their best matches, one particular cluster consists of 714 Yb 8 elements. The phylogenetic tree involving all of these elements indicates that one copy of Yb8 (at hg19/chr10:10493416-10493732) seemed to have generated multiple active Yb8 copies that further retrotransposed to produce eventually 713 copies or $54 \%$ of all $1,322 \mathrm{hsYb} 8$ elements studied (Figure 1). This master Yb8 element was most likely the major driver of the Yb8 expansion after the human-chimp divergence. Eight other Yb8 elements were detected that generated at least ten copies of offspring Yb 8 elements. These Y b 8 elements with lower activity level comply with the 'stealth driver' model of Alu evolution, which states that the stealth drivers do not generate as many copies of Alu as the master gene does, but rather function primarily to maintain the genomic retrotransposition capacity over a period of time [24].
A similar approach was taken to track the evolutionary pathway of hsYb9 elements, involving identification and clustering of best-matched sequences from the whole genome. While almost all of the Yb 9 elements tested aligned best with one another, 16 elements aligned best with 16 different Yb 8 elements. When a phylogenetic tree was constructed with all hsYb9 elements and these 16 Yb 8 elements, one particular Yb8 element at chr14:101990881101991202 was found to be the source of all the hsYb9 elements, having generated multiple active Yb 9 elements that subsequently generated 131 additional full-length hsYb9 copies (Figure 2). Along the evolutionary path of hsYb9, shown in Figure 2, some clusters have Yb8 elements, which may have resulted from either reverse mutation to produce Yb 8 elements, or gene conversion or misannotation of Yb 9 copies as Yb8 [25].

## Identification of novel Alu Yb subclasses

Different subfamilies of the Yb lineage are characterized by specific mutations, and the subfamilies are defined


Figure 1 Cladogram with 714 hsYb8 elements constructed by the neighbour joining method. The element marked with a bold line (at hg19/chr10:10493416-10493732) is likely to be the source copy of all others in the tree. The tree was rooted using the Yb8 consensus, which is indicated by the black line.
according to the number of mutation sites with respect to the Alu Y consensus sequence [11]. Identification of new subfamilies is basically the identification of a set of Alu elements that share a particular mutation at a specific site that has not been previously reported. Using a computational approach, we performed a genome-wide analysis of Alu elements that are currently annotated as Yb 8 and Yb 9 , the two most recent subfamilies of the Yb lineage known to date, to investigate whether any specific mutation beyond the Yb 8 and Yb 9 signature mutations is shared by more than one element. To do so, a set of full-length members of the $A l u \mathrm{Yb} 8$ and Yb 9 subfamilies were retrieved from the latest assembly of the human reference genome sequence GRCh37, and multiple sequence alignment was performed after the poly-A segments were removed. Upon careful examination of the alignment data, two specific mutations were observed in multiple Yb 9 and Yb 8 elements at the 201st (insertion of T ) and 259th ( $\mathrm{G} \rightarrow \mathrm{A}$ ) positions, respectively. We also observed that Alu sequences with the single base insertion
after the 200th position always carry the mutation at the 259th position and the Yb 9 diagnostic mutation at the 174th position, but not all sequences with a mutation at the 259th position contain the other two mutations. This is only possible if the sequences with the $259^{G \rightarrow \mathrm{~A}}$ mutation originated from the Yb8 subfamily as the first event and then a subset of these sequences accumulated the Yb9-diagnostic $174^{C \rightarrow G}$ mutation, or vice versa, giving rise to another new subfamily, which subsequently accumulated the $200^{+\mathrm{T}}$ insertion to generate yet another subclass of Yb elements. Following the standard nomenclature of Alus [11], we named the sequences with the $259^{\mathrm{G} \rightarrow \mathrm{A}}$ mutation Alu Yb8al, the sequences with the $259^{\mathrm{G} \rightarrow \mathrm{A}}$ and $174{ }^{C \rightarrow G}$ mutations $A l u \mathrm{Yb} 10$, and the sequences with the $259^{\mathrm{G} \rightarrow \mathrm{A}}$ and $174^{\mathrm{C} \rightarrow \mathrm{G}}$ mutations and the $200^{+\mathrm{T}}$ insertion Alu Yb11 (Figure 3). When a Yb8a1 signatory sequence of 30 bases was constructed and aligned against the human reference genome, 99 Yb 10 copies were identified, among which 75 copies did not have the $174^{\mathrm{C} \rightarrow \mathrm{G}}$ mutation (Yb8a1), 8 had the $174^{\mathrm{C} \rightarrow \mathrm{G}}$ mutation (Yb10), and 16 copies


Figure 2 Cladogram with 131 hs Yb9 and 16 Yb8 elements constructed by the neighbour joining method. Alu Yb9 and Yb8 elements are shown in blue and green, respectively. There is one Yb8a1 element in the cluster that matches best with one of the Yb9 elements, shown in red. The Yb8 copy shown in bold green is likely to be the source of all Yb9 copies in the cladogram. The Yb8 consensus (root) is shown in black.
had both the $174^{\mathrm{C} \rightarrow \mathrm{G}}$ mutation and the $200^{+\mathrm{T}}$ insertion (Yb11). A 24-nucleotide-long signatory sequence was also constructed for Yb 11 , and when this sequence was aligned against the reference genome, 16 matches were detected, all of which overlap with the results from the Yb10 signatory sequence-whole genome alignment, which provides evidence for the accuracy of the method. In the end, we were able to detect $75 \mathrm{Yb} 8 \mathrm{a} 1,8 \mathrm{Yb} 10$ and 16 Yb 11 insertions in the reference genome (Additional file 1: Table S1).
Besides the reference genome, we also analysed 1000 Genome Project (1KGP) data and sequencing trace data from HuRef [26], to identify insertions of the newly identified subfamily members that are absent in the reference genome. We collected all of the Yb 8 and Yb 9 insertions that are absent from the reference genome but present in one or more individual genome sequences in the 1KGP data, for which sufficient insertion sequences could be constructed. Signature sequences for $\mathrm{Yb} 8 \mathrm{a} 1, \mathrm{Yb} 10$ and Y b 11 were then aligned against these sequences and the HuRef sequencing, resulting in the detection of an additional $6 \mathrm{Yb} 8 \mathrm{a} 1,3 \mathrm{Yb} 10$ and 15 Yb 11 insertions outside the reference genome. The insertion of T in the Yb 11 elements outside the reference genome was confirmed by PCR amplification and sequencing for five of these 15 loci and by manually checking the sequencing data from the National Center for Biotechnology Information (NCBI) trace database for three of them (Additional file 2: Figures S1 and S2; Additional file 3: Table S2). Therefore, we were able to identify a total of $81 \mathrm{Yb} 8 \mathrm{a} 1,11 \mathrm{Yb} 10$ and 31 Yb 11
insertions, and we can expect that more of these will be identified after processing more personal genomes.

## Age estimation

Mutation densities were calculated for each subfamily to estimate the approximate age of the new subfamilies. Only full-length or near full-length Alu elements in the reference genome were considered ( 65 Yb 8 a 1 out of 75 , 8 Yb 10 , and 15 Yb 11 out of 16) and the poly-A regions in the middle and at the end were removed. For the 65 elements from the Yb 8 a 1 subfamily, the non-CpG mutation density was $0.29 \%$ ( 43 out of 14,625 total non-CpG bases). Using a neutral rate of evolution of $0.15 \%$ per million years for primate intervening DNA sequences [27] along with the non-CpG mutation density, the average age of the Yb 8 a 1 subfamily was estimated to be 1.93 million years old. For the 8 Yb 10 elements, 5 non-CpG mutations were detected out of a total of 1,904 nonCpG nucleotides constituting only $0.26 \%$ of them, indicating an estimated age of 1.73 million years for Yb 10 . For the Yb 11 subfamily, 15 elements were analysed with a total of 3,720 non-CpG nucleotides; only 4 of these had mutated, yielding a neutral mutation density of $0.107 \%$ and an estimated age of 0.71 million years. To assess how recent these subfamilies are in relation to the already known Yb subfamilies, the age of Yb 9 was also estimated. A total of 166 non-CpG mutations were identified from 254 Alu Yb9 family members containing 51,562 non-CpG nucleotides; 73 members were not included in the calculations

| Y | GGCCGGGCGCGGTGGCTCACGCCTGTAATCCCAGCACTTTGGGAGGCCGAGGCGGGCGGA 60 |
| :---: | :---: |
| Yb8 | .......T... 60 |
| Yb9 | ..T... 60 |
| Yb8a1 | ...T... 60 |
| Yb10 | ..T... 60 |
| Yb11 | ......T... 60 |
|  | 2 . . . 3 |
| Y | TCACGAGGTCAGGAGATCGAGACCATCCTGGCTAACACGGTGAAACCCCGTCTCTACTAA 120 |
| Yb8 | . . T. . . . . . . . . . . . . . . . . . . . . . . . . A. . . . . . . . . . . . . . . . . . . 120 |
| Yb9 | T...........................A. . . . . . . . . . . . . . . . . . 120 |
| Yb8a1 | . T. . . . . . . . . . . . . . . . . . . . . . . A. . . . . . . . . . . . . . . . . . 120 |
| Yb10 | T...............................A. . . . . . . . . . . . . . . . . . 120 |
| Yb11 | .T. . . . . . . . . . . . . . . . . . . . . . . . A. . . . . . . . . . . . . . . . . . . 120 |
|  | . 4 . 4 . 9 |
| Y | AAATACAAAAAATTAGCCGGGCGTGGTGGCGGGCGCCTGTAGTCCCAGCTACTCGGGAGG 180 |
| Yb8 | . . . . C. . . . . . . . . . . . . . . . . . . . . . . . . . . . . 180 |
| Yb9 | .C. . . . . . . . . . . . . . . . . . . . . . . G. . . . . 180 |
| Yb8a1 | . C. . . . . . . . . . . . . . . . . . . . . . . . . . . . . . . . 180 |
| Yb10 | . C. . . . . . . . . . . . . . . . . . . . . . . G. . . . . 180 |
| Yb11 | . .C. . . . . . . . . . . . . . . . . . . . . . G G . . . . 180 |
|  | .11 . 5 . 6 |
| Y | CTGAGGCAGGAGAATGGCGT-GAACCCGGGAGGCGGAGCTTGCAGTGAGCCGAGATCGCG 239 |
| Yb8 | . . . . . . .-. . . . . . . . A. . . . . . . . . . . . . . . . . . . . T. . . 239 |
| Yb9 | -.........A.........................T... 239 |
| Yb8a1 | .A............ . . . . . . . . . . T. . . 239 |
| Yb10 | . A........................T... 239 |
| Yb11 | .T.........A.......................T... 240 |
|  | 7. 8 .8a1 |
| Y | CCACTGCACTCCA-------GCCTGGGCGACAGAGCGAGACTCCGTCTC 281 |
| Yb8 | . . . . . . .G...GCAGTCCG. . . . . . . . . . . . . . . . . . . . . . . . . . 2888 |
| Yb9 | . . . . . . G...GCAGTCCG............................. 288 |
| Yb8a1 | .G....GCAGTCCA. . . . . . . . . . . . . . . . . . . . . . . . . . 288 |
| Yb10 | . G. . . GCAGTCCA. . . . . . . . . . . . . . . . . . . . . . . . . . 288 |
| Yb11 | ........G...GCAGTCCA............................. 289 |

Figure 3 Consensus sequences of Alu Y, Yb8, Yb9, Yb8a1, Yb10, and Yb11. The signatory mutations are numbered in chronological order using Alu $Y$ as the baseline.
due to a $5^{\prime}$ truncation or a large deletion inside the Yb 9 element. Using the same neutral rate of evolution and the non-CpG mutation density of $0.32 \%(166 / 51,562)$, the average age of the Yb 9 subfamily members was estimated to be 2.15 million years. The age of the Yb 9 subfamily estimated in this study is much older than that estimated initially by Roy-Engel et al. [9], mainly because the total number of Yb 9 elements in their study was much smaller than in this study. However, our estimation of the age of Yb 9 is very close to that identified in a similar study, which estimated the age of Yb 9 as 2.32 million years [14]. The estimated age for Yb 8 a 1 indicates that this subfamily originated almost at the same time as Yb 9 , providing evidence that Yb 8 a 1 originated from Yb 8 . The Yb 10 subfamily, which evolved 1.73 million years ago, should be mostly fixed across all human populations, while the Yb 11 subfamily, at only 0.71 million years old, is most likely to be highly polymorphic among human populations because it is the youngest. The level of polymorphism for these newly identified subfamilies with respect to their ages are examined further in the following section.

## Level of polymorphism

The Alu Y family is evolutionarily the 'youngest' Alu family and the Yb lineage was found to be one of the largest and most active lineages of all young Alu elements [12,14,28]. Out of the 2,433 full-length Yb elements found in the human genome, 499 were found to be polymorphic for their presence or absence between individuals and/or populations, and a further 304 Yb copies were identified in individual genome sequences that are not present in the reference genome $[16,29]$. Since the majority of Yb elements became inserted into the human genome 3 to 4 million years ago, we suspect that the very recently evolved subfamilies contribute most to the polymorphism due to the Yb lineage since the divergence of the various human populations from their common ancestor occurred only 100,000 years ago [14]. We assessed the level of polymorphism for all identified $\mathrm{Yb} 8 \mathrm{a} 1, \mathrm{Yb} 10$ and Yb 11 insertions by surveying Alu insertions and deletions in personal genomics data. We compared the insertions that are present in the reference genome with the structural variation data from the 1000 Genome Project [30]. Of these,

13 out of 16 (approximately $81 \%$ ) Yb11 elements and 2 out of $8(25 \%) \mathrm{Yb} 10$ were found to be dimorphic, while 22 out of 75 (approximately 29\%) Yb8a1 present in the reference genome are polymorphic. We then compared these polymorphic insertions with dbRIP to identify how many of them have previously been reported as polymorphic and found that 7 and 2 polymorphic Yb 8 a 1 and Yb 11 elements, respectively, overlap with dbRIP data [6]. Combining insertions both inside and outside the reference genome, a total of 28 out of 31 (approximately 90\%) Yb11 and 5 out of 11 (approximately 45\%) Yb10 were found to be polymorphic, while only 28 out of 81 (approximately $34 \%$ ) of Yb 8 a 1 insertions were identified as polymorphic. The difference in the level of polymorphism is inversely related to the age of the lineage, that is, the higher the polymorphism level among individuals and/or populations, the more evolutionarily recent the lineage. The difference in the fraction of polymorphic members among the three novel subfamilies confirms that Yb11 has evolved more recently than Yb 10 and Yb 8 a . The relative newness of the Yb 11 lineage is further substantiated when we looked at the sequence divergence within the members of each subfamily (Table 1). The mean evolutionary divergence between each pair of sequences in the $\mathrm{Yb} 8 \mathrm{a} 1, \mathrm{Yb} 9$, Yb 10 and Yb 11 subfamilies was estimated to be 0.016 , $0.026,0.015$ and 0.006 , respectively. The divergence value is directly related to the age of the population, that is, the older the set of sequences, the more evolutionarily divergent the sequences are. The mean divergence values provide another line of data suggesting that Yb8a1, Yb10 and Yb 11 evolved chronologically during the evolution of humans.
We also examined the distribution of all polymorphic members of Yb8a1, Yb10 and Yb11 in Yoruban, European, Chinese and Japanese populations. It was observed that $50 \%, 64 \%$ and $59 \%$ of polymorphic elements are present in the Yoruban population for the $\mathrm{Yb} 8 \mathrm{a} 1, \mathrm{Yb} 10$ and Yb 11 subfamilies, respectively (Figure 4). These numbers are higher than the equivalent numbers for the other nonAfrican populations examined. The highest number of polymorphic elements were expected to be present in the Yoruban population as this was the oldest population

Table 1 Estimates of evolutionary divergence between and within full-length Alu Yb9, Yb10 and Yb11 elements

|  | Alu Yb8a1 | Alu Yb9 | Alu Yb10 | Alu Yb11 |
| :--- | :--- | :--- | :--- | :--- |
| Alu Yb8a1 | $0.016^{\text {a }}$ |  |  |  |
| Alu Yb9 | $0.026^{\text {b }}$ | 0.026 |  |  |
| Alu Yb10 | 0.019 | 0.022 | 0.015 |  |
| Alu Yb11 | 0.015 | 0.017 | 0.011 | 0.006 |

[^1]tested in this study [31]. While the presence or absence of some of the polymorphic elements could not be ascertained for the Chinese and Japanese populations (they are flagged as 'unascertained'), the majority of the rest (approximately $66 \%$ ) were present in one or both of the Asian populations. Among these, only one Yb 8 a 1 insertion was found to be specific to the Chinese population and the rest are all shared by one or more other populations. In contrast, $15 \mathrm{Yb} 8 \mathrm{a} 1,5 \mathrm{Yb} 10$ and 10 Yb 11 insertions are specific to the Yoruban population, and 2, 3 and 4 of each of Yb 8 a 1 , Yb 10 and Yb 11 insertions are specific to the European population. This suggests that the number of populationspecific insertions decreases with the age of the population. In other words, the older the population, the more time there has been for active young Alu elements to retrotranspose, creating a direct relation between the number of population-specific Alus and the age of population.

Evolutionary pathways for the three new Alu Yb subfamilies New Alu families are created when a mutation occurs in the master or source active Alu element, which subsequently retrotransposes to give rise to a new lineage of Alu elements that share the same diagnostic mutation. The master gene model is the most widely accepted model for the generation of new Alu subfamilies [8] even though there many doubts about the details of this model [10,32-34]. While this model only gives a hierarchical evolution for the different subfamilies, the specific evolutionary pathways for the generation of different Yb lineages have yet to be characterized. The evolution of Yb9, Yb8 and Yb 7 , the three most recent and abundant subfamilies of the Yb lineage, occurred sequentially [9].
In our study, we predict that the evolution of Yb 11 took a strict sequential linear pathway from Yb 10 since it contains one more mutation than Yb 10 diagnostic mutations, while the Yb10 subfamily evolved from either Yb 8 a 1 or Yb 9 following one or more pathways (Figure 5). A tree using the neighbour joining method was constructed among 25, 181, 65, 8 and 15 full-length $\mathrm{Yb} 8, \mathrm{Yb} 9$, $\mathrm{Yb} 8 \mathrm{a} 1, \mathrm{Yb} 10$ and Yb 11 elements, respectively, rooted with the Yb 8 consensus sequence (Figure 6). The 25 Yb 8 elements were included because these are the only Yb 8 copies that one or more of $\mathrm{Yb} 9, \mathrm{Yb} 8 \mathrm{a} 1, \mathrm{Yb} 10$ and Yb 11 had the best similarity score with. It was observed from the topology that $77 \%$ of all Yb 8 a 1 elements have evolved from one individual Yb 8 copy, and 63 out of 65 Yb 8 a 1 copies tested are evolutionarily closest to members of the Yb8 subfamily. This confirms that Yb8a1 evolved from Yb8 as a separate lineage from Yb 9 . Among the 15 Yb 11 copies included in the phylogenetic analysis, all of them have common nodes with copies from Yb10 elements, confirming their linear evolutionary pathway from the Yb 10 subfamily.
The diagnostic mutations of the Yb 10 subfamily are predicted to have evolved by following one of two pathways:


Figure 4 The level of polymorphisms for the Yb8a1, Yb10 and Yb11 subfamilies. The blue columns at the front indicate the number of polymorphic insertions observed in the population and the orange columns in the middle represent the number of insertions observed in other populations but not in the population. The presence or absence of polymorphic insertions in Chinese and Japanese populations could not be determined and these are labeled as 'unascertained' and represented by grey bars. CEU, Utah residents with European ancestry; CHB, Han Chinese from Beijing areas; JPT, Japanese from the Tokyo area; YRI, Yoruban.
(1) a Yb 9 element obtained the Yb 8 a 1 -specific mutation and retrotransposed to generate the Yb10 subfamily or (2) a Yb8al element obtained the Yb 9 -specific mutation subsequently generating the Yb 10 subfamily. The phylogenetic analysis on its own does seem to favour the latter option since the major branch leading to the $\mathrm{Yb} 10 / \mathrm{Yb} 11$ lineage is closer to the Yb 8 a 1 cluster. For additional evidence, an evolution network was constructed for all fulllength members of the four subfamilies of interest using the median joining method [35]. The network shows that the majority of the Yb 10 elements are linked closer to multiple Yb 8 a 1 elements than to Yb 9 (Additional file 2:


Figure 5 Evolution of the recent $A / u \mathrm{Yb}$ lineage. The subfamilies in black are the current known subfamilies and the subfamilies in red are novel and proposed in this study. The numbers accompanying each subfamily are the total number of copies found in the human reference genome. The dotted line is the less convincing alternative pathway for the evolution of the Yb10 subfamily.

Figure S3), further supporting the prediction that the evolution of Yb 10 was from Yb 8 a 1 by gaining the Yb 9 mutation. The accumulation of the Yb 9 -specific mutation in the Yb 8 a 1 copy parent to create the Yb 10 subfamily may have occurred by gene conversion and requires further analysis for confirmation. A second line of evidence for the evolutionary pathway proposed here is provided by the linear pairwise evolutionary distances calculated for the $\mathrm{Yb} 9, \mathrm{Yb} 8 \mathrm{a} 1, \mathrm{Yb} 10$ and Yb 11 elements (Table 1). The mean evolutionary distance for all sequences between Yb 10 and Yb 11 was calculated as 0.011 , which is lower than the distance between Yb 9 and Yb 11 ( 0.017 ) or Yb 8 a 1 and Yb 11 (0.015) indicating the sequential evolution of Yb 11 from Yb 10 and with Yb 8 a 1 being closer than Yb 9 to Yb 11 .
Each of the Yb8a1, Yb10 and Yb11 subfamilies was also tested using the molecular clock (ML) to assess if all full-length members in each subfamily evolved at a homogeneous rate. A maximum likelihood test of the ML hypothesis was performed separately for each of the $\mathrm{Yb} 8 \mathrm{a} 1, \mathrm{Yb} 10$ and Yb 11 phylogenetic tree topologies and sequence alignments [36]. The ML hypothesis states that all tips of the tree should be equidistant from the root of the tree, or in other words the rate of evolution of all branches in the tree is uniform. The maximum likelihood, $-\ln L$, was calculated to be 990.971 and 907.158 for with-clock and without-clock phylogeny, respectively, for Yb8a1, 466.906 and 455.855 for with-clock and withoutclock phylogeny, respectively, for Yb 10 , and 481.574 and 474.459 for with-clock and without-clock phylogeny, respectively, for Yb 11 . The chi-square test based on the difference in the likelihood ratio between with-clock and without-clock phylogeny rejected the null hypothesis of uniform evolution for both Alu Yb8a1 and Yb10 insertions at a $5 \%$ significance level with $P<0.0001$ and $P<0.001$ for Yb 8 al and Yb 10 , respectively. However, we failed to reject the null hypothesis of an equal evolutionary rate for all Yb11 insertions at a $5 \%$ significance level ( $P<0.43$ ). This


Figure 6 Cladogram of all full-length Yb9, Yb8a1, Yb10, and Yb11 elements using the neighbour joining method. The tree is rooted with the Alu Yb8 consensus sequence, which is shown in black at the top left.
indicates that neither the Yb 8 a 1 nor the Yb 10 subfamily evolved at a uniform evolutionary rate, and that the evolution of the subfamily Yb 11 has been uniform. This provides further evidence that the Yb 8 a 1 and Yb 10 subfamilies are older than the Yb 11 subfamily since evolutionary uniformity is more likely in a recently evolved lineage. Furthermore, when the evolutionary relations for all full-length Yb8a1, $\mathrm{Yb} 9, \mathrm{Yb} 10$ and Yb 11 elements were analysed, more divergence among members of Yb 8 a 1 and Yb 9 was observed than among the members of Yb 10 or Yb 11 (Additional file 2: Figure S4), another indication that the former subfamilies are older than the latter.

## Conclusions

The $A l u \mathrm{Yb}$ lineage has an extended evolutionary history in the human genome. Even though the lineage evolved before the human-chimp divergence, most of the insertions occurred in the last 3 to 4 million years and some copies of this lineage still retain the ability to retrotranspose. One such active Yb 8 copy has generated almost $60 \%$ of all human-specific Yb 8 copies and several others have generated more than ten copies, indicating the presence of both a master copy and stealth drivers for this subset of Yb8 elements.

The tracking of the source copy in this study enabled us to identify the potential master gene of all Yb 9 elements. The relatively higher activity of the Yb lineage than almost all other Alu lineages has generated several subfamilies that were previously undetected and which share a specific pattern of mutations. Three such novel subfamilies proposed in this study are $\mathrm{Yb} 8 \mathrm{a} 1, \mathrm{Yb} 10$ and Yb 11 . Even though Yb8a1 and Yb10 are believed to have evolved within a short time of each other, only eight copies of Yb10 have been detected in the human reference genome compared to 75 copies of Yb 8 a 1 . Furthermore, Yb 9 has been estimated to be only 0.22 million years older than Yb 8 a , yet the number of Yb 9 copies in the human genome is almost five times larger than the number of Yb8a1 copies. This indicates that not all of the Alu subfamilies grew at an equal rate and that some mutation patterns may accelerate the rate of transposition. This is further supported by the fact that the Yb 11 -specific insertional mutation in the Yb 10 sequence has accelerated the rate of retrotransposition resulting in 16 copies of Yb 11 since it first evolved 0.71 million years ago. The possibility that certain mutations accelerate the rate of transposition and their mechanism should be the subject of further study.
Yb11 is the latest subfamily to have evolved in this lineage and it is highly polymorphic among different
individuals and/or populations. The generation of these young subfamilies indicates that Alus are still evolving, and this provides some clues regarding the future trend of Alu activity in the human genome. The homoplasyfree nature of $A l u$ insertions makes these very recent genetic variants a valuable resource in forensics and for studying modern human population genetics and migration patterns.

## Methods

## Source copy tracking

All human-specific Yb elements were retrieved from a separate study (Tang et al., unpublished data). The humanspecific Yb lineage has members from only $\mathrm{Yb} 8, \mathrm{Yb} 9$ and the newly identified subfamilies. Each full-length humanspecific Yb element was aligned against the reference genome using BLAST [37] with the e-value set to $10^{-5}$. Based on the BLAST results, any insertions that match more than one genomic region with equal matching quality were omitted from further analysis as the source copy of these insertions could not be determined. The remaining sequences were divided into clusters based on their similarity with one another. The evolutionary relation between members of each cluster was obtained by constructing a phylogenetic tree using the neighbour joining method rooted with the Yb 8 consensus sequence, and some cases were supplemented with network analysis using the median joining method [35].

## Identification of new Alu Yb subfamilies

Position information for all $A l u \mathrm{Yb} 8$ and Yb 9 elements from the latest major version of the human genome assembly GRCh37 were retrieved from the RepeatMasker track of the UCSC genome browser [38] and the sequence for each insertion was retrieved from the reference genome. The poly-A segments from both the 3' end and the middle were removed manually. The pairwise alignment for all Yb9 sequences was visualized in MEGA5 [39]. A signatory sequence was constructed encompassing each of the signature insertions at the 201st position and the mutation at the 259th position. The sequences were conserved across all $A l u \mathrm{Yb}$ insertions except for the mutation/insertion base. These sequences were aligned against the reference genome using BLAST with an e-value of $10^{-5}$. The resulting matches were filtered using an in-house Perl script to retain only the sequences that have the signature mutation/insertion. To identify additional insertions of the new subfamilies that are absent in the reference genome, genome sequencing and alignment data from the 1000 Genome Project were downloaded to our local server. New insertions for $A l u \mathrm{Yb} 8$ and Yb 9 in the six high coverage genome datasets from phase 1 of the 1000 Genome Project were identified in a separate study [40]; the read cluster for each predicted novel insertion contains all reads from the
inserted region. From the mobile element insertion list generated from the pilot phase 1 data of the 1000 Genome Project [16], we collected 304 Alu Yb 8 and Yb 9 insertions that are absent in the reference genome but were detected in one or more of the test genomes for which a complete insertion sequence could be constructed. A custom BLAST database was created to contain all these new insertion sequences, and the signature sequences were aligned against this custom database using the abovementioned criteria.

## Validation of Yb11 insertions outside the reference genome

 The insertion of T after the 200th nucleotide in Yb 11 can potentially be the result of a sequencing error since the preceding base is also a T. To eliminate the possibility of erroneous results, all reads sequenced by Sanger's method were downloaded from the NCBI trace database to our local server. The Yb11 signatory sequence was aligned against these reads to identify the reads that contain Yb 11 . A total of 130 reads were found to contain the Yb11-specific T insertion. The Phred quality score of the site of the T insertion in each read was analysed using a custom Perl script (Additional file 2: Figure S1). Three out of fifteen loci could be confirmed using these trace data. Of the remaining twelve Yb 11 insertions that are outside the reference genome sequence, primers could be designed for six Alu insertions. Five insertions could be amplified by PCR in DNA samples NA19239 and NA19240 from the Coriell Cell Repositories [41] and an in-house mixed DNA, all of which received approval from the Brock University Research Ethic Board. The amplified products were sequenced using the Sanger method at The Centre for Applied Genomics. The sequencing primers include locus-specific flanking primers and two Alu-internal primers designed from the $5^{\prime}$ and $3^{\prime}$ ends of the Yb 11 consensus sequence, which are TGGCTCACGCCTGTAATC and GACGGAGTCTCGCTCTGTC, respectively. The internal primers help with difficulties in sequencing through the poly-A regions within Alu sequences. The sequences were aligned using clustalW to analyse the Yb 11 -specific site (Additional file 2: Figure S1). All new Alu insertion sequences not covered by dbRIP were processed for deposition into dbRIP [42] under the study ID 2013-02.
## Analyses of the Yb8a1, Yb10 and Yb11 insertion polymorphisms and evolution relations

To assess the level of polymorphism among the insertions of the three new subfamilies, the start and end position of each insertion was compared with structural variation [30] and mobile element insertion [16] data from the 1000 Genome Project and with entries from dbRIP [6]. The phylogenetic tree for all full-length Alu Yb9, Yb8a1, Yb10 and Yb 11 insertions along with the putative source Yb 8 copies obtained from previously mentioned clusters was constructed using the neighbour joining method [43]. All
alignments and phylogenetic trees were visualized using the MEGA software [39]. The evolutionary distance and sequence divergence within and between subfamilies were calculated using the maximum composite likelihood model [44] involving 181 full-length $\mathrm{Yb} 9,65 \mathrm{Yb} 8 \mathrm{a} 1,8 \mathrm{Yb} 10$ and 15 Yb 11 nucleotide sequences without poly-A sequences at the 3 ' end and in the middle.

## Additional files

Additional file 1: Table S1. List of Yb8a1, Yb10 and Yb11 insertions identified in the reference genome.
Additional file 2: Figure $\mathbf{S 1}$ to $\mathbf{S 4}$. Contains Figure S 1 to S 4 to supplement the PCR and evolutionary analysis data presented in the article.
Additional file 3: Table S2. List of Yb8a1, Yb10 and Yb11 insertions identified outside the reference genome.

## Abbreviations

1KGP: 1000 genome project; hsYb: Human-specific A/u Yb; ML: Molecular clock; NCBI: National center for biotechnology information; PCR: Polymerase chain reaction; SINE: Short interspersed element.

## Competing interests

The authors declare that they have no competing interests.

## Authors' contributions

MA collected all data, carried out the analysis and drafted the manuscript. WL performed the PCR analysis and sequencing. PL conceived the study, supervised the design and all analyses performed, and also edited the manuscript to its final version. All authors read and approved the final manuscript for publishing.

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## References

1. Batzer MA, Deininger PL: Alu repeats and human genomic diversity. Nat Rev Genet 2002, 3(5):370-379.
2. Lander ES, Linton LM, Birren B, Nusbaum C, Zody MC, Baldwin J, Devon K, Dewar K, Doyle M, FitzHugh W, Funke R, Gage D, Harris K, Heaford A, Howland J, Kann L, Lehoczky J, LeVine R, McEwan P, McKernan K, Meldrim J, Mesirov JP, Miranda C, Morris W, Naylor J, Raymond C, Rosetti M, Santos R, Sheridan A, Sougnez C, et al: Initial sequencing and analysis of the human genome. Nature 2001, 409(6822):860-921.
3. Shen MR, Batzer MA, Deininger PL: Evolution of the master Alu gene(s). J Mol Evol 1991, 33(4):311-320.
4. Mills RE, Bennett EA, Iskow RC, Luttig CT, Tsui C, Pittard WS, Devine SE: Recently mobilized transposons in the human and chimpanzee genomes. Am J Hum Genet 2006, 78(4):671-679.
5. Mills RE, Bennett EA, Iskow RC, Devine SE: Which transposable elements are active in the human genome? Trends Genet 2007, 23(4):183-191.
6. Wang J, Song L, Grover D, Azrak S, Batzer MA, Liang P: dbRIP: a highly integrated database of retrotransposon insertion polymorphisms in humans. Hum Mutat 2006, 27(4):323-329
7. Jurka J, Milosavljevic A: Reconstruction and analysis of human Alu genes. J Mol Evol 1991, 32(2):105-121.
8. Deininger PL, Batzer MA, Hutchison CA, Edgell MH: Master genes in mammalian repetitive DNA amplification. Trends Genet 1992, 8(9):307-311.
9. Roy-Engel AM, Carroll ML, Vogel E, Garber RK, Nguyen SV, Salem AH, Batzer MA, Deininger PL: Alu insertion polymorphisms for the study of human genomic diversity. Genetics 2001, 159(1):279-290.
10. Batzer MA, Rubin CM, Hellmann-Blumberg U, Alegria-Hartman M, Leeflang EP, Stern JD, Bazan HA, Shaikh TH, Deininger PL, Schmid CW: Dispersion and insertion polymorphism in two small subfamilies of recently amplified human Alu repeats. J Mol Biol 1995, 247(3):418-427.
11. Batzer MA, Deininger PL, Hellmann-Blumberg U, Jurka J, Labuda D, Rubin CM, Schmid CW, Zietkiewicz E, Zuckerkandl E: Standardized nomenclature for Alu repeats. J Mol Evol 1996, 42(1):3-6.
12. Wang J, Song L, Gonder MK, Azrak S, Ray DA, Batzer MA, Tishkoff SA, Liang P: Whole genome computational comparative genomics: a fruitful approach for ascertaining Alu insertion polymorphisms. Gene 2006, 365:11-20.
13. Arcot SS , Fontius JJ, Deininger PL, Batzer MA: Identification and analysis of a 'young' polymorphic Alu element. Biochim Biophys Acta 1995, 1263(1):99-102.
14. Carter AB, Salem AH, Hedges DJ, Keegan CN, Kimball B, Walker JA, Watkins WS, Jorde LB, Batzer MA: Genome-wide analysis of the human Alu Yb-lineage. Hum Genomics 2004, 1(3):167-178.
15. Hormozdiari F, Alkan C, Ventura M, Hajirasouliha I, Malig M, Hach F, Yorukoglu D, Dao P, Bakhshi M, Sahinalp SC, Eichler EE: Alu repeat discovery and characterization within human genomes. Genome Res 2011, 21(6):840-849
16. Stewart C, Kural D, Stromberg MP, Walker JA, Konkel MK, Stutz AM, Urban AE, Grubert F, Lam HY, Lee WP, Busby M, Indap AR, Garrison E, Huff C, Xing J, Snyder MP, Jorde LB, Batzer MA, Korbel JO, Marth GT: 1000 Genomes Project: a comprehensive map of mobile element insertion polymorphisms in humans. PLoS Genet 2011, 7(8):e1002236.
17. Novick GE, Gonzalez T, Garrison J, Novick CC, Batzer MA, Deininger PL, Herrera RJ: The use of polymorphic Alu insertions in human DNA fingerprinting. EXS 1993, 67:283-291.
18. Novick GE, Novick CC, Yunis J, Yunis E, Martinez K, Duncan GG, Troup GM, Deininger PL, Stoneking M, Batzer MA: Polymorphic human specific Alu insertions as markers for human identification. Electrophoresis 1995, 16(9):1596-1601.
19. Batzer MA, Deininger PL: A human-specific subfamily of Alu sequences. Genomics 1991, 9(3):481-487.
20. Batzer MA, Stoneking M, Alegria-Hartman M, Bazan H, Kass DH, Shaikh TH, Novick GE, loannou PA, Scheer WD, Herrera RJ: African origin of humanspecific polymorphic Alu insertions. Proc Natl Acad Sci USA 1994, 91(25):12288-12292.
21. Hedges DJ, Callinan PA, Cordaux R, Xing J, Barnes E, Batzer MA: Differential Alu mobilization and polymorphism among the human and chimpanzee lineages. Genome Res 2004, 14(6):1068-1075.
22. Muratani K, Hada T, Yamamoto Y, Kaneko T, Shigeto Y, Ohue T, Furuyama J, Higashino K: Inactivation of the cholinesterase gene by Alu insertion: possible mechanism for human gene transposition. Proc Natl Acad Sci USA 1991, 88(24):11315-11319.
23. Oldridge M, Zackai EH, McDonald-McGinn DM, Iseki S, Morriss-Kay GM, Twigg SR, Johnson D, Wall SA, Jiang W, Theda C, Jabs EW, Wilkie AO: De novo Alu-element insertions in FGFR2 identify a distinct pathological basis for Apert syndrome. Am J Hum Genet 1999, 64(2):446-461.
24. Han K, Xing J, Wang H, Hedges DJ, Garber RK, Cordaux R, Batzer MA: Under the genomic radar: the stealth model of Alu amplification. Genome Res 2005, 15(5):655-664.
25. Roy AM, Carroll ML, Nguyen SV, Salem AH, Oldridge M, Wilkie AO, Batzer MA, Deininger PL: Potential gene conversion and source genes for recently integrated Alu elements. Genome Res 2000, 10(10):1485-1495.
26. Levy S, Sutton G, Ng PC, Feuk L, Halpern AL, Walenz BP, Axelrod N, Huang J, Kirkness EF, Denisov G, Lin Y, MacDonald JR, Pang AW, Shago M, Stockwell TB, Tsiamouri A, Bafna V, Bansal V, Kravitz SA, Busam DA, Beeson KY, McIntosh TC, Remington KA, Abril JF, Gill J, Borman J, Rogers YH, Frazier ME, Scherer SW, Strausberg RL, et al: The diploid genome sequence of an individual human. PLoS Biol 2007, 5(10):e254
27. Miyamoto MM, Slightom JL, Goodman M: Phylogenetic relations of humans and African apes from DNA sequences in the psi eta-globin region. Science 1987, 238(4825):369-373.
28. Jurka J: A new subfamily of recently retroposed human Alu repeats. Nucleic Acids Res 1993, 21(9):2252.
29. Jurka J, Kapitonov W, Pavlicek A, Klonowski P, Kohany O, Walichiewicz J: Repbase update, a database of eukaryotic repetitive elements. Cytogenet Genome Res 2005, 110(1-4):462-467.
30. Mills RE, Walter K, Stewart C, Handsaker RE, Chen K, Alkan C, Abyzov A, Yoon SC, Ye K, Cheetham RK, Chinwalla A, Conrad DF, Fu Y, Grubert F, Hajirasouliha I, Hormozdiari F, lakoucheva LM, Iqbal Z, Kang S, Kidd JM, Konkel MK, Korn J, Khurana E, Kural D, Lam HY, Leng J, Li R, Li Y, Lin CY, Luo R, et al: 1000 Genomes Project: mapping copy number variation by population-scale genome sequencing. Nature 2011, 470(7332):59-65.
31. Stringer $C B$, Andrews P: Genetic and fossil evidence for the origin of modern humans. Science 1988, 239(4845):1263-1268.
32. Cordaux R, Hedges DJ, Batzer MA: Retrotransposition of Alu elements:
how many sources? Trends Genet 2004, 20(10):464-467.
33. Price AL, Eskin E, Pevzner PA: Whole-genome analysis of Alu repeat elements reveals complex evolutionary history. Genome Res 2004, 14(11):2245-2252.
34. Schmid CW: How many source Alus? Trends Genet 1993, 9(2):39.
35. Bandelt HJ, Forster P, Rohl A: Median-joining networks for inferring intraspecific phylogenies. Mol Biol Evol 1999, 16(1):37-48.
36. Felsenstein J: Evolutionary trees from DNA sequences: a maximum likelihood approach. J Mol Evol 1981, 17(6):368-376.
37. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ: Basic local alignment search tool. J Mol Biol 1990, 215(3):403-410.
38. UCSC Genome Browser. [http://http://genome.ucsc.edu]
39. Tamura K, Peterson D, Peterson N, Stecher G, Nei M, Kumar S: MEGA5: molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. Mol Biol Evol 2011, 28(10):2731-2739.
40. Luo X, Dehne F, Liang P: Identification of transposon insertion polymorphisms (TIPs) by computational comparative analysis of next generation personal genome data. AIP Conference Proceedings 2011, 1368:163-166.
41. Coriell cell repositories. [http://ccr.coriell.org]
42. dbRIP - A database of retrotransposon insertion polymorphisms in humans. [http://dbrip.org]
43. Saitou N, Nei M: The neighbor-joining method: a new method for reconstructing phylogenetic trees. Mol Biol Evol 1987, 4(4):406-425.
44. Tamura K, Nei M, Kumar S: Prospects for inferring very large phylogenies by using the neighbor-joining method. Proc Natl Acad Sci USA 2004, 101(30):11030-11035.

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[^1]:    ${ }^{\text {a }}$ The average of base substitutions per site of all pairwise comparisons within the group.
    ${ }^{\mathrm{b}}$ The average of base substitutions per site of all pairwise comparisons among the members of the two groups compared.

