RESEARCH Open Access

High prevalence, intensity, and genetic diversity of *Trichinella* spp. in wolverine (*Gulo gulo*) from Yukon, Canada



Rajnish Sharma¹, N. Jane Harms², Piia M. Kukka², Thomas S. Jung^{2,3}, Sarah E. Parker⁴, Sasha Ross¹, Peter Thompson⁵, Benjamin Rosenthal⁵, Eric P. Hoberg^{6,7} and Emily J. Jenkins^{1*}

Abstract

Background: Species of *Trichinella* are globally important foodborne parasites infecting a number of domestic and wild vertebrates, including humans. Free-ranging carnivores can act as sentinel species for detection of *Trichinella* spp. Knowledge of the epidemiology of these parasites may help prevent *Trichinella* spp. infections in northern Canadian animals and people. Previous research on *Trichinella* spp. in wildlife from Yukon did not identify risk factors associated with infection, or the diversity and identity of species of *Trichinella* in regional circulation, based on geographically extensive sampling with large sample sizes.

Methods: In a cross-sectional study, we determined the prevalence, infection intensity, risk factors, and species or genotypes of *Trichinella* in wolverine (*Gulo gulo*) in two regions of Yukon, Canada, from 2013–2017. A double separatory funnel digestion method followed by mutiplex PCR and PCR-RFLP were used to recover and identify species of *Trichinella*, respectively.

Results: We found larvae of *Trichinella* in the tongues of 78% (95% CI 73–82) of 338 wolverine sampled. The odds of adult (\geq 2 years) and yearling (1 year) wolverine being *Trichinella* spp.-positive were four and two times higher, respectively, compared to juveniles (<1 year). The odds of *Trichinella* spp. presence were three times higher in wolverine from southeast than northwest Yukon. The mean intensity of infection was 22.6 ± 39 (SD, range 0.1–295) larvae per gram. *Trichinella* T6 was the predominant genotype (76%), followed by *T. nativa* (8%); mixed infections with *Trichinella* T6 and *T. nativa* (12%) were observed. In addition, *T. spiralis* was detected in one wolverine. Out of 22 isolates initially identified as *T. nativa* in multiplex PCR, 14 were analyzed by PCR-RFLP to distinguish them from *T. chanchalensis*, a recently discovered cryptic species, which cannot be distinguished from the *T. nativa* on multiplex PCR. Ten isolates were identified either as *T. chanchalensis* alone (n = 7), or mixed infection with *T. chanchalensis* and *T. nativa* (n = 2) or *T. chanchalensis* and *Trichinella* T6 (n = 1)].

Conclusions: Wolverine hosted high prevalence, high larval intensity, and multiple species of *Trichinella*, likely due to their scavenging habits, apex position in the food chain, and wide home range. Wolverine (especially adult males) should be considered as a sentinel species for surveys for *Trichinella* spp. across their distributional range.

Keywords: Trichinella spp., Wolverine, Canada, Prevalence, T. nativa, Trichinella T6, T. chanchalensis

Full list of author information is available at the end of the article

Background

Species of *Trichinella* nematodes are among the most significant foodborne parasites listed by the World Health Organization/United Nations Food and Agriculture



© The Author(s) 2021. This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

^{*}Correspondence: ejj266@mail.usask.ca

¹ Department of Veterinary Microbiology, Western College of Veterinary Medicine, University of Saskatchewan, 52 Campus Drive, Saskatoon, SK S7N 5B4, Canada

Sharma et al. Parasites Vectors (2021) 14:146 Page 2 of 9

Organization [1]. All Trichinella species and genotypes have been reported in mammals, whereas T. pseudospiralis also develops in avian hosts, and T. papuae and T. zimbabwensis have also been reported from reptilian hosts. All species and genotypes of Trichinella are zoonotic and transmitted by consuming raw or insufficiently cooked meat [2]. There are 13 species of Trichinella classified into two clades: encapsulated and nonencapsulated. A survey in Canada conducted over a period of 10 years on 15 wildlife species showed an overall prevalence of 20% for multiple species of *Trichinella* among tested animals [3], and identified T. nativa, Trichinella T6, T. murrelli, and T. pseudospiralis [3]. Trichinella chanchalensis, a cryptic encapsulated species, has recently been discovered and described in wolverine (Gulo gulo) from northwestern Canada [4].

Globally, most human outbreaks of trichinellosis have been attributed to domestic pigs infected with T. spiralis [5]. However, almost all human outbreaks in the last 20 years in Canada involved T. nativa and Trichinella T6 linked to consumption of raw or insufficiently cooked game meat, commonly from walrus (*Odobenus rosmarus*) and bears (Ursus spp.) [6-9]. For example, 15 trichinellosis outbreaks have occurred in Nunavik (northern Québec, Canada) from 1982-2009; of these, nine were linked to walrus meat [9]. One of the largest outbreaks reported in Canada was from Saskatchewan, where 78 people became sick after consuming meat from a black bear (Ursus americanus) infected with T. nativa [7]. A recent outbreak of human trichinellosis due to T. nativa in Ontario, Canada was associated with consumption of dried meat of black bear and resulted in rare thrombotic complications [8]. Trichinella T6 has also been identified in an outbreak in Ohio, USA, linked to consumption of bear meat from Canada [6]. Knowledge of the epidemiology of the disease and pathways for exposure from wild animals can help prevent *Trichinella* spp. infections in humans and animals.

Free-ranging carnivores may be reservoirs of *Trichinella* spp., and may act as sources of infection for humans and as sentinel species for parasite detection. For example, various species of canids have been previously proposed as sentinel species for *Trichinella* spp. [10–12]. An ideal sentinel animal host should possess the following features: adequate availability (relative population stability), measurable response (such as parasites in tissues), earlier response than people or sympatric wild species (high chance of exposure early in life and a relatively short life span), and high levels of exposure [13, 14]. Wolverine (*Gulo gulo*) may act as an ideal sentinel species for detecting *Trichinella* spp. in northwestern North America because (1) they are legally killed for their fur and carcasses are available from fur harvesters [15], (2)

initial studies have reported relatively high prevalence and intensity of *Trichinella* spp. infection in wolverine, suggesting that they may be a key species in the epidemiology of infection [3, 16], and (3) they are a facultative scavenger [17–19] near the top of terrestrial food webs, and as such may be bio-accumulators of foodborne parasites such as *Trichinella* spp.

There are few published studies on species of Trichinella infecting wild carnivores from the Yukon [3, 20, 21]. The two older studies (45 years ago) reported Trichinella spp. in 71% of 21 grizzly bear (Ursus arctos) from the Yukon [21] and 47% of 127 gray wolves (Canis lupus) from Yukon and the Northwest Territories [20]. Larvae of Trichinella spp. were detected in Arctic fox (Vulpes lagopus), wolf, and wolverine from the Yukon in a survey conducted 10 years ago [3]. Previously, we published two studies, one where we compared tongue and diaphragm muscles as predilection sites [22] and another where we discovered a new species of Trichinella [4] in a portion of wolverine carcasses used in the present study. Previous research on Trichinella in wildlife from Yukon did not involve geographically extensive and site-intensive sampling, or large sample sizes. To date, no epidemiological study has associated risk factors or identified recovered larvae to species level. To fill these knowledge gaps, we sampled a large number of wolverine across Yukon, Canada.

To determine the suitability of wolverine as a sentinel species, we therefore sought (1) to identify the species/genotypes of *Trichinella* and their distribution in Yukon; (2) to determine the prevalence and intensity of infections with species of *Trichinella*; and (3) to explore the associations between potential risk factors and *Trichinella* spp. positivity and larval intensity in wolverine.

Methods

Wolverine sampling

We obtained wolverine carcasses from across Yukon, Canada, which is ~483,000 km² and sparsely populated by people (~ 0.08 people\km²), with 76% of the human population living in the city of Whitehorse [23]. The topography of Yukon is rugged and characterized by mountain ranges, plateaus, valleys, and lowlands. Climate is subarctic continental, with annual precipitation ranging from 250-600 mm, most of which falls as snow from October to May. The mean daily temperature ranges from −15 °C to −30 °C in January and from 10 °C to 15 °C in July. Yukon is characterized by boreal forest in valley bottoms, and shrub communities and alpine tundra at and above the treeline, respectively [24]. Wolverine are legally harvested throughout Yukon by fur trappers during the winter [25-27], with an average of 132 ± 31 (standard deviation [SD]) animals harvested

Sharma et al. Parasites Vectors (2021) 14:146 Page 3 of 9

annually [25]. Wolverine carcasses were submitted by fur trappers to Environment Yukon (Government of Yukon) and kept frozen at −20°C for 6−10 months prior to necropsy. The harvest location and sex of each animal were recorded, and age was determined by cementum analysis of a premolar tooth at a commercial laboratory (Matson's Laboratory LLC, Milltown, MT, USA). Wolverine were classified as juveniles (<1 year), yearlings (1 year), or adults (≥ 2 years) based on cementum-derived ages. A body condition index (BCI) for each wolverine was calculated using predetermined regression equations for wolverine from our study area that incorporated the mass of select fat depots with body size and sex, where higher BCIs corresponded to greater mass-specific fat levels, for each sex, than lower BCIs [27]. Tongues were collected in plastic bags and stored at -20 °C until further analyses. The harvest location of each wolverine was assigned to one of two broad geographic areas (northwest Yukon or southeast Yukon; Fig 1), because these geographic areas correspond to broad phylogeographic divisions for wolverine that could influence the genetic diversity of wolverine parasites [28, 29].

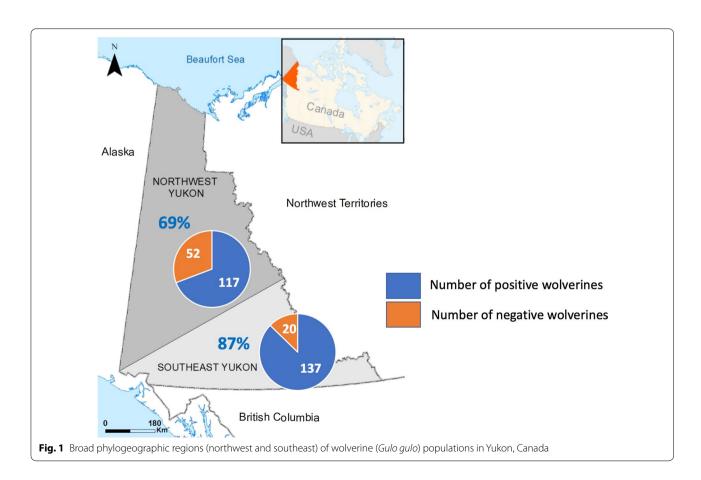
The number of wolverine in our study area was unknown but was estimated to range between 3500 and

4000 [30]. We used OpenEpi version 3.01 [31] to estimate the sample size necessary to be representative of the wolverine population in Yukon. We parameterized OpenEpi by using the midpoint of the population size estimate range (3750) and an expected prevalence of *Trichinella* spp. of 80%, based on earlier reports of 88% and 77% [3, 16] in wolverine from northern Canada. Based on these values, the calculated sample size necessary for our study was 231 with a confidence interval of 95%. We obtained 338 wolverine carcasses from animals harvested during 2013–2017, which exceeded the estimated sufficient sample size for our study.

Because we obtained samples from animals that were harvested for purposes other than our research, this was considered exempt from animal care committee review at the University of Saskatchewan. We obtained a wildlife research permit and appropriate export permits from the Government of Yukon.

Recovery of Trichinella spp. larvae

In a previous study, we compared tongues and diaphragms obtained from wolverine harvested in 2012–2013 and 2013–2014 and found that the tongue was a better predilection site for *Trichinella* spp. [22]. Thus,



Sharma et al. Parasites Vectors (2021) 14:146 Page 4 of 9

only tongues were collected for recovery of larvae in the current study. Tongues were thawed at room temperature and cut into 0.5–1.0 cm cubes, which were mixed and a portion randomly selected to make up to 10 g. Connective tissue and fat were removed and ≤ 10 g of muscle was digested using the double separatory funnel digestion method [32]. Larvae per gram (LPG) of each positive sample were calculated by dividing total number of larvae by weight of digested tissue. To identify *Trichinella* species, larvae were collected in 1X polymerase chain reaction (PCR) buffer and stored at -20 °C until DNA extraction [33].

Molecular identification of *Trichinella* spp. by multiplex PCR

From each wolverine positive for Trichinella spp., parasite genomic DNA was extracted from five individual larvae as well as a pool of ten larvae using a Proteinase K extraction method [33, 34]. In the case of availability of less than 15 larvae from a positive animal, five individual larvae and the rest of the larvae in the pool were processed. To identify species or genotypes of *Trichinella* larvae, primers amplifying internal transcribed spacer regions (ITS 1 and 2) as well as the expansion segment V of the large subunit ribosomal DNA were used in a multiplex PCR assay, as previously described [33, 34]. PCR reactions were performed with master mix, containing 1X AmpliTag Gold 360 Master Mix (Applied Biosystems, CA, USA), 50 µM primers, 2 µl of GC Enhancer, 2.5 µl of DNA, and ultrapure water to a total volume of 25 μl. The thermocycler (Bio-Rad, USA) conditions were 95 °C for 10 min followed by 40 cycles of 95 °C for 15 s, 55 °C for 30 s, 72 °C for 90 s, and final elongation at 72 °C for 7 min. Species were identified based on the band pattern developed on 2.5% agarose gel when analyzed under UV light using a gel doc system (Alpha Innotech AlphaImager digital imaging system). DNA from six species of Trichinella (T. spiralis, T. nativa, T. britovi, T pseudospiralis, T murrelli, and Trichinella T6) were used as positive controls. Positive controls, negative template control, and DNA extraction control (only 10X PCR buffer) were also included in each PCR. Trichinella nativa and T. chanchalensis showed similar banding patterns on the multiplex PCR (a band of 127 bp); therefore, a subsequent PCRrestriction fragment length polymorphism (RFLP) was performed to differentiate them, as previously described **[4]**.

Data analysis

A wolverine was considered positive for *Trichinella* spp. if ≥ 1 larva was recovered from tissue samples. Using Epitools epidemiological calculators [35], prevalence with 95% confidence intervals (CI) was calculated from the

proportion of wolverine that were positive for any species of Trichinella. The intensity of infection was measured as LPG, with "low" and "high" intensity defined as ≤10 LPG and >10 LPG, respectively. We built binary logistic regression models to independently test for (1) presence of Trichinella spp. (present or absent) and (2) intensity of Trichinella spp. infection (low or high). We evaluated the following potential predictors for inclusion in a final multivariable model using univariable logistic regression analysis with a relaxed level of significance ($p \le 0.20$): age (juvenile, yearling, or adult), sex (female or male), harvest location (southeast Yukon or northwest Yukon), harvest season (winters 2013-2014, 2014-2015, 2015-2016, or 2016-2017), and BCI (continuous variable). The weight of muscle tissue processed (< 5 g, 5.0-9.9 g, or \geq 10 g) was also included as a potential predictor in both models. For outcome "Trichinella spp. larval intensity (high/ low)," single vs. co-infection status was also included as a predictor (single infection means infection with any one of the species Trichinella T6, T. nativa, T. chanchalensis, or T spiralis; co-infection means co-infection with either Trichinella T6 and T. nativa, Trichinella T6 and T. chanchalensis, or T. nativa and T. chanchalensis). Stepwise forward multivariable logistic regressions were performed to build the final model. Goodness of fit of the final model was evaluated by the Hosmer-Lemeshow test. Variables with a significance level of p < 0.05 were retained in the model. To estimate the degree of the association between each predictor and Trichinella status, odds ratios and respective 95% confidence intervals were calculated. All statistical analyses were performed using IBM SPSS software (version 24; Armonk, NY, USA).

Results

Descriptive analysis

The mean age and BCI of sampled wolverine was 1.6 years (SD = 2.05, range <1-10) and 8.88 (SD = 5.24, range <1-30), respectively. Age was not estimated for 3 of 338 wolverine (0.88%), and harvest location was not available for 12 of 338 wolverine (3.5%). Most carcasses submitted were male, with a male-to-female ratio of 2:1. The age-class distribution of wolverine that we sampled was similar among juveniles (37%), yearlings (29%), and adults (34%). Wolverine were obtained in relatively equal percentages (23-27%) from each of the 4 years of sampling as well as from the two phylogeographic regions; 50% and 46% from the northwest and southeast Yukon, respectively.

Trichinella spp. prevalence

Trichinella spp. larvae were detected in 78% (262 of 338) of wolverine tested (95% CI 73–82). Larvae of *Trichinella* spp. were detected most frequently in adult wolverine

Sharma et al. Parasites Vectors (2021) 14:146 Page 5 of 9

(87%, 99 of 114, 95% CI 79–92), followed by yearlings (82%, 79 of 96, 95% CI 73–89) and juveniles (65%, 81 of 125, 95% CI 56–73). More males (80%, 95% CI 74–84) than females (73%, 95% CI 64–81) were positive for *Trichinella* spp. (Table 1).

Our univariable logistic regressions revealed that age (p < 0.001), sex (p = 0.184), and phylogeographic region (p

Table 1 Prevalence (% positive) of *Trichinella* spp. in 338 wolverine (*Gulo gulo*) in Yukon, Canada

	Number of positive wolverine	Total animals examined	Prevalence	95% confidence interval		
Age						
Juvenile	81	125	64.8	56.1-72.6		
Yearling	79	96	82.3	73.5-88.6		
Adult	99	114	86.8	79.4-91.9		
Sex						
Female	82	112	73.2	64.3-80.6		
Male	180	226	79.6	73.9-84.4		
Harvest location						
Northwest	117	169	69.2	61.9-75.7		
Southeast	137	157	87.3	81.1-91.6		
Harvest year	Harvest year					
2013-2014	69	86	80.0	70.6-87.3		
2014-2015	66	91	72.5	62.6-80.6		
2015-2016	60	77	77.9	67.5-85.7		
2016-2017	67	84	79.8	70.0-87.0		
Weight of tongue processed						
Less than 5 g	24	36	66.7	50.3-80.0		
5-9.9 g	22	28	78.6	60.5-89.8		
10 g	216	274	78.8	73.6–83.3		

< 0.001) were significantly associated with the presence of *Trichinella* spp. in wolverine. Our stepwise multivariable regression suggested that age and phylogeographic region were significantly associated with *Trichinella* spp. prevalence (Table 2). The odds of *Trichinella* spp. presence were two times (odds ratio = 2.25, 95% CI 1.16–4.36; p=0.016) and four times (odds ratio = 3.76, 95% CI 1.86–7.60; p=0.001) higher in yearlings and adults, respectively, than in juveniles (<1 year). The odds of the presence of *Trichinella* spp. were three times (odds ratio = 2.87, 95% CI 1.60–5.16; p=0.001) higher in wolverine from southeast Yukon than northwest Yukon.

Intensity of Trichinella spp. infection

The median intensity of *Trichinella* spp. larvae was 8.4 (range 0.1–295) LPG. Larval intensity > 1 was observed in 89% (232 of 262 animals sampled) wolverine. The median larval intensity of *Trichinella* spp. was higher in juveniles (14 LPG) than in yearlings (7 LPG) and adult wolverine (7 LPG). Age and co-infection was significantly associated with high larval intensity in both univariable and stepwise multivariable regression (Table 2). The odds of high larval intensity were two times (odds ratio = 2.41, 95% CI 1.28–4.54; p=0.012) higher in yearlings than adult wolverine. The odds of high larval intensity were three times (odds ratio = 3.33, 95% CI 1.53–7.24; p=0.002) higher in wolverine that were co-infected than those infected with a single species of *Trichinella* (Table 2).

Genetic diversity of Trichinella spp.

Using multiplex PCR, *Trichinella* spp. larvae from 254 of 262 (97%) infected wolverine were identified to species/genotype. Overall, 85% (222 of 262) of wolverine

Table 2 Risk factors for *Trichinella* spp. prevalence and intensity in wolverine (*Gulo gulo*) identified in final stepwise logistic regression; odd ratios (OR) and its 95% confidence intervals and probability (*p*)

Model (outcome) (n)	Predictors in the final model		OR	95 CI	р
Trichinella spp. infection (yes/no) [323]	Age	Juvenile			0.001
		Yearling	2.25	1.16-4.36	0.016
		Adult	3.76	1.86-7.60	0.001
	Harvest location	Northwest			0.001
		Southeast	2.87	1.60-5.16	0.001
Trichinella spp. larval burden (high/low) [254] ^a	Age	Adult			0.012
		Yearling	2.41	1.28-4.54	0.007
		Juvenile	1.07	0.57-2.00	0.840
	Single vs. co-infection status ^b	Trichinella-single species infection			0.002
		Trichinella-mixed infection	3.33	1.53-7.24	

n Animals with complete record for respective model

^a Only positive samples with *Trichinella* species were included in the model

b Single infection means infection with any one of the listed species *Trichinella* T6, *T. nativa*, *T. chanchalensis*, or *T spiralis*; co-infection means co-infection with either *Trichinella* T6 and *T. nativa*, *Trichinella* T6 and *T. chanchalensis*, or *T. nativa* and *T. chanchalensis*

Sharma et al. Parasites Vectors (2021) 14:146 Page 6 of 9

were infected with a single species (Trichinella T6, T. nativa, or T. spiralis). Trichinella T6 was the predominant genotype (76%; 199 of 262), followed by T. nativa (8%; 22 of 262). Mixed infections with both *Trichinella* T6 and T. nativa were detected in 12% (32 of 262). Larvae of T. spiralis were present in one wolverine. Out of 22 pure T. nativa positive isolates (without any indication of co-infection on multiplex PCR), 14 were subjected to RFLP (DNA was not amplified from 8 isolates due to either disintegrated larvae or lack of sufficient larvae). Among the samples tested by RFLP, ten were positive for T. chanchalensis, with only three having mixed infection with either T. nativa or genotype T6. (Table 3). Overall (based on multiplex PCR and/PCR-RFLP), 86% (219/254) of wolverine were infected with a single species (either Trichinella T6 or T. nativa or T. chanchalensis or T. spiralis) and 14% were co-infected with two species (either T2 + T6, T2 + T13, T6 + 13).

Discussion

We report high prevalence (78%) of *Trichinella* spp. (predominantly T6) in wolverine in Yukon, Canada, based on the largest sampling effort yet for this species (n=338 wolverine). The prevalence of *Trichinella* spp. appears to be substantially higher in wolverine than all other carnivores tested so far in northern Canada or Alaska, with the possible exception of grizzly bears (*Ursus arctos*; Additional file 1: Table S1). The prevalence of *Trichinella* spp. in wolverine in our study was comparable to other reports from northern Canada, where prevalence of 77% and 88% had been noted [3, 16], and higher than that observed for this species in other countries (0% in Sweden, 30% in Kamchatka, Russia, 50% in Alaska, USA) [36–38] (Additional file 1: Table S1). Variation in

Table 3 Species of *Trichinella* identified in wolverine (*Gulo gulo*) in the Yukon

Species diagnosis	By multiplex PCR	By PCR-RFLP ^a
Trichinella T6 only	199	NA
T. nativa only	22	4 (T. nativa only)
		7 (T. chanchalensis only)
		2 (T. chanchalensis and T. nativa)
		1 (<i>T. chanchalensis</i> and <i>Trichinella</i> T6)
		8 NA
Trichinella T6 and T. nativa	32	NA
T. spiralis	1	NA
Species not identified	8	NA

NA isolates of Trichinella not tested by PCR-RFLP

prevalence documented across studies may be due to differences in sample size, type of muscle sampled (tongue, diaphragm, cheek muscles, or leg muscles), or methodology (trichinoscopy versus muscle digestion) [37, 39–41]. In the current study, we used the pepsin-HCl digestion method, which has higher sensitivity than trichinoscopy [37, 39, 40]. Digestion is now considered the gold standard for detection of larval *Trichinella* spp. in animal tissues [42]. We used tongue tissue, which has been demonstrated to be a more sensitive sampling site than the diaphragm for detection of larval *Trichinella* spp. in wolverine [22]. Finally, we used relatively large samples of tongue tissue for examination (89% of samples were ≥ 5 g), as recommended for studies of wild carnivores [43].

Our data indicate that prevalence increases with age, as demonstrated for other wildlife species in Europe [44, 45] but not for a similar study in wolverine from Nunavut, Canada [16]. The reason for higher prevalence of *Trichinella* spp. infection in older animals likely reflects an accumulation of diet-based exposure risk, and enduring chronic infection thereafter. A higher prevalence of *Trichinella* spp. in male than female wolverine was observed, consistent with findings from a similar survey in the Northwest Territories, Canada [46], whereas almost equal prevalence rates (89% in male vs. 86% in female wolverine) were reported in wolverine from Nunavut, Canada [16]. Males have larger home ranges, and disperse further, than females [47], increasing opportunities for exposure.

Trichinella spp. were widely distributed in wolverine in Yukon; however, prevalence was higher in the southeast region than in the northwest region. There may be regional variation in wolverine diet, which may play a role in varying prevalence among regions. Other factors that can affect regional differences in prevalence of Trichinella spp. may include environmental factors such as snow cover, altitude, air temperature or humidity, or anthropogenic factors. For example, the number of snow cover days was positively associated with the incidence of Trichinella spp. infections among wild boars from Latvia [48]. However, based on this assumption alone, a greater number of snow cover days in northern Yukon suggests that prevalence should have been higher in the northern regions.

In our study we observed a median infection intensity (8 LPG) that was twice as high as that reported from an earlier study in northwestern North America (including British Columbia, Northwest Territories, and Yukon; 3.7 LPG; Additional file 1: Table S2; [3]). In contrast to the differences in age-related prevalence, we observed higher median larval intensity in juveniles than yearlings and adults. This suggests that while risk of exposure increases with age, the severity of infection

^a Only 14 of 22 *T. nativa* isolates (identified on multiplex PCR) were tested by PCR-RFLP [4]

Sharma et al. Parasites Vectors (2021) 14:146 Page 7 of 9

is higher in young animals, which could be due to juveniles having relatively undeveloped immunity.

We determined the genotype and/or species of *Trichinella* circulating among wildlife in Yukon. Our surveys revealed two sylvatic species of *Trichinella* (*Trichinella* T6 and *T. nativa*) and, unexpectedly, a species typically considered to be restricted to domestic circulation (*T. spiralis*). Moreover, *T. chanchalensis* (a previously unknown and cryptic species of *Trichinella*) was also reported, occurring at a greater prevalence in single infections than *T. nativa*, from which it must be distinguished by PCR-RFLP [4].

Due to their freeze tolerance, larvae of *T. nativa*, *Trichinella* T6, and *T. chanchalensis* are expected in arctic and subarctic regions. We report higher prevalence of *Trichinella* T6 than *T. nativa* in wolverine from Yukon, consistent with observations elsewhere in the western Canadian Arctic (Nunavut) [16]. *Trichinella* T6 is the most common genotype of *Trichinella* observed in Canadian wildlife, with wider host and geographic ranges than *T. nativa* [3]. Differences in infectivity and host range among genotype or species of *Trichinella* warrant further study.

Trichinella spiralis is a reportable pathogen in domestic animals in Canada. Currently, Canada is considered free of *T. spiralis* in swine raised for commercial purposes, and a single case does not immediately implicate its circulation among free-ranging wildlife hosts. Although the source of infection remains unknown for this isolated observation, it is unlikely that wolverine play a significant role in transmission of *T. spiralis* (although see discussion about ecological fitting and otherwise incidental hosts in the circulation of parasites [49, 50]). Further, because *T. spiralis* is not freeze-tolerant, the significance of this finding remains uncertain. Additional studies are required to explore the degree to which wolverine and other wildlife may serve as reservoirs for *T. spiralis*.

Our results show that *Trichinella* T6, *T. chanchalensis*, and *T. nativa* occur in sympatry geographically (Yukon) and in the same individual host(s) (wolverine). Co-infection with *T. nativa* and *Trichinella* T6 has been previously reported in wolverine [16], and hybridization between the two species has been observed in Alaska [51]. The multiplex PCR (which targets nuclear DNA only) does not distinguish hybrids of *Trichinella* T6 and *T. nativa*, which requires microsatellite and/or mitochondrial DNA analysis [52]. Based on a study in wolverine from Nunavut, microsatellite and mitochondrial DNA analysis showed that hybrids appear as *T. nativa* on the multiplex PCR. We report mixed infection of *T. chanchalensis* with either *Trichinella* T6 or *T. nativa* as well.

Routes of transmission of *Trichinella* larvae among wildlife include scavenging, predation, and cannibalism

[53-55]. The overall high prevalence, intensity, and diversity of Trichinella spp. in wolverine may indicate that they become infected by multiple pathways. Hunting and fur trapping practices such as leaving carcasses in the field, or using meat of wild carnivores as bait, may facilitate transmission [56]. While clinical disease due to Trichinella is usually absent in animals, surveying wildlife is important in terms of zoonoses and food safety [50]. For example, well-organized testing programs in Nunavik (northern Québec, Canada) for Trichinella in walruses have been developed to limit the dissemination of infected meat before distribution among communities [9, 57]. Monitoring wildlife can also be important to determine epidemiological and genetic baselines for endemic Trichinella species, and to detect spillover of T. spiralis or introduced species into wildlife. For example, in Europe, especially in regions with no recognized Trichinella infection in pigs, monitoring of wildlife using suitable sentinel species is mandatory [58].

Conclusion

Wolverine, especially adult males, are good sentinels for Trichinella spp. in northwestern North America because they have a high prevalence and diversity (at least 3 endemic species) of Trichinella. Moreover, carcasses obtained from fur trappers provide relatively large and consistent sample sizes, and engage local people in surveillance programs. As such, we suggest using wolverine for monitoring changes in Trichinella spp. infection in wildlife of northwestern Canada, and our data can be used as a baseline. Larval intensity ≥ 1 LPG is considered a significant risk for food safety [3], and 89% of wolverine positive for *Trichinella* spp. in our study met or exceeded this threshold. Although wolverine are not consumed as food by people, universal precautions (namely wearing gloves and thoroughly washing hands before eating, and storing wolverine carcasses away from other meat intended as food, and away from pets) should be practiced when handling wolverine carcasses to prevent transmission. Practices such as leaving carcasses in the field or using wolverine meat as bait for trapping or hunting may also facilitate transmission, even in winter, as larvae of sylvatic species of Trichinella present in arctic and subarctic regions survive freezing.

Supplementary Information

The online version contains supplementary material available at https://doi.org/10.1186/s13071-021-04636-2.

Additional file 1:Table S1. Prevalence of *Trichinella* spp. in wild carnivores in Canada and Alaska. **Table S2.** Prevalence and larval burden (larvae per gram) of *Trichinella* spp. in wolverine from different geographical locations.

Sharma et al. Parasites Vectors (2021) 14:146 Page 8 of 9

Acknowledgements

We are grateful to the Yukon fur trappers that submitted wolverine carcasses for this study; without their involvement this project would have not been possible. We thank the many staff and students who assisted with obtaining and sampling the carcasses, particularly Meghan Larivee (Government of Yukon). We thank the Canadian Food Inspection Agency (CFAP) for providing larvae of six species/genotypes of *Trichinella*. We also acknowledge Brent Wagner and Cherise Hedlin (Western College of Veterinary Medicine), and Brad Scandrett and Kelly Konecsni (CFAP) for technical support and reviewing this manuscript (BW, BS, KK). We dedicate this manuscript to Robert L. Rausch and Virginia R. Rausch for their significant contributions towards Arctic parasitology.

Authors' contributions

RS: study design, sample and data collection, laboratory methodology, data analysis and results interpretation, manuscript writing. NJH, PMK, TSJ: study design, sample and data collection, results interpretation, article review. SEP: study design, data analysis and results interpretation, article review. SR: sample and data collection, laboratory methodology, article review. PT, BR, EPH: results interpretation, article review and supervision. EJ: study design, results interpretation, article review and supervision. All co-authors contributed intellectually to the article. All authors read and approved the final manuscript.

Funding

Funding for this study was provided by the Western College of Veterinary Medicine (University of Saskatchewan), Government of Yukon, and a Discovery Grant and Northern Research Supplement from the National Science and Engineering Research Council (NSERC). We have no competing interests to declare.

Availability of data and materials

The data sets generated and/or analyzed during the current study are available from the corresponding author on reasonable request.

Ethics approval and consent to participate

As animals were harvested for purposes other than research, this was considered Category A and exempt from animal research ethics review at the University of Saskatchewan. Relevant wildlife research and export permits were obtained from the government wildlife management agency.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no conflict of interest.

Author detail:

¹ Department of Veterinary Microbiology, Western College of Veterinary Medicine, University of Saskatchewan, 52 Campus Drive, Saskatoon, SK S7N 5B4, Canada. ² Department of Environment, Government of Yukon, P.O. Box 2703, Whitehorse, YT Y1A 2C6, Canada. ³ Department of Renewable Resources, University of Alberta, 351 General Services Building, Edmonton, AB T6H 3T1, Canada. ⁴ Centre for Applied Epidemiology, Large Animal Clinical Sciences, Western College of Veterinary Medicine, University of Saskatchewan, 52 Campus Drive, Saskatoon, SK S7N 5B4, Canada. ⁵ USDA-Agricultural Research Service, Animal Parasitic Diseases Laboratory, Beltsville Agricultural Research Center, 10300 Baltimore Avenue, Beltsville, MD 20705, USA. ⁶ Museum of Southwestern Biology and Department of Biology, University of New Mexico, Albuquerque, NM 87131-0001, USA. ⁷ Department of Pathobiological Sciences, School of Veterinary Medicine, University of Wisconsin-Madison, Madison, WI 53706, USA.

Received: 7 October 2020 Accepted: 11 February 2021 Published online: 08 March 2021

References

 FAO/WHO. Multicriteria-based ranking for risk management of foodborne parasites: report of a joint FAO/WHO expert meeting, 3–7 September 2012, FAO Headquarters, Rome, Italy. 2014, p. 302

- Zarlenga DS, Rosenthal BM, La Rosa G, Pozio E, Hoberg EP. Post-Miocene expansion, colonization, and host switching drove speciation among extant nematodes of the archaic genus *Trichinella*. Proc Natl Acad Sci USA. 2006;103:7354–9.
- Gajadhar AA, Forbes LB. A 10-year wildlife survey of 15 species of Canadian carnivores identifies new hosts or geographic locations for Trichinella genotypes T2, T4, T5, and T6, Vet Parasitol. 2010;168:78–83.
- Sharma R, Thompson PC, Hoberg EP, Brad Scandrett W, Konecsni K, Harms NJ, et al. Hiding in plain sight: discovery and phylogeography of a cryptic species of *Trichinella* (Nematoda: Trichinellidae) in wolverine (*Gulo gulo*). Int J Parasitol. 2020;50:277–87.
- Darwin Murrell K, Pozio E. Worldwide occurrence and impact of human trichinellosis, 1986–2009. Emerg Infect Dis. 2011;17:2194–202.
- Nelson M, Wright TL, Pierce A, Krogwold RA. A common-source outbreak of trichinosis from consumption of bear meat. J Environ Health. 2003;65:16–9
- Schellenberg RS, Tan BJK, Irvine JD, Stockdale DR, Gajadhar AA, Serhir B, et al. An outbreak of trichinellosis due to consumption of bear meat infected with *Trichinella nativa*, in 2 northern Saskatchewan communities. J Infect Dis. 2003;188:835–43.
- Dalcin D, Zarlenga DS, Larter NC, Hoberg E, Boucher DA, Merrifield S, et al. Trichinella nativa outbreak with rare thrombotic complications associated with meat from a black bear hunted in Northern Ontario. Clin Infect Dis. 2017;64:1367–73.
- Larrat S, Simard M, Lair S, Bélanger D, Proulx JF. From science to action and from action to science: the Nunavik trichinellosis prevention program. Int J Circumpolar Health. 2012;71:18595.
- Aoun O, Lacour SA, Levieuge A, Lou MJ, Vallée I, Davoust B. Screening for Trichinella britovi infection in red fox (Vulpes vulpes) and wild boar (Sus scrofa) in Southeastern France. J Wildl Dis. 2012;48:223–5.
- Badagliacca P, Di Sabatino D, Salucci S, Romeo G, Cipriani M, Sulli N, et al. The role of the wolf in endemic sylvatic *Trichinella britovi* infection in the Abruzzi region of Central Italy. Vet Parasitol. 2016;231:124–7.
- Shamsian A, Pozio E, Fata A, Navi Z, Moghaddas E. The Golden jackal (Canis aureus) as an indicator animal for Trichinella britovi in Iran. Parasite. 2018;25:94–7.
- NRCCAMEH. National Research Council (US) Committee on Animals as Monitors of Environmental Hazards. In: Animals as sentinels of environmental health hazards. Washington (DC); 1991.
- Halliday JEB, Meredith AL, Knobel DL, Shaw DJ, Bronsvoort BMDC, Cleaveland S. A framework for evaluating animals as sentinels for infectious disease surveillance. J R Soc Interface. 2007;4:973–84.
- COSEWIC Committee on the Status of Endangered Wildlife in Canada.
 Ottawa. COSEWIC assessment and status report on the Wolverine Gulo gulo in Canada. http://www.registrelepsararegistry.gc.ca/default_e.cfm). xi+76. p. 2014.
- Reichard MV, Torretti L, Snider TA, Garvon JM, Marucci G, Pozio E. *Trichinella* T6 and *Trichinella nativa* in wolverines (*Gulo gulo*) from Nunavut. Canada Parasitol Res. 2008;103:657–61.
- Dalerum F, Shults B, Kunkel K. Estimating sustainable harvest in wolverine populations using logistic regression. J Wildl Manage. 2008;72:1125–32.
- Lofroth EC, Krebs JA, Harrower WL, Lewis D. Food habits of wolverine Gulo gulo in montane ecosystems of British Columbia Canada. Wildlife Biol. 2007;13:31–7.
- Van Dijk J, Andersen T, May R, Andersen R, Andersen R, Landa A. Foraging strategies of wolverines within a predator guild. Can J Zool. 2009;26:066-75.
- Choquette LP, Gibson GG, Kuyt E, Pearson AM. Helminths of wolves, Canis lupus L., in the Yukon and Northwest Territories. Can J Zool. 1973;51:1087–91.
- 21. Choquette LP, Gibson GG, Pearson AM. Helminths of the grizzly bear, *Ursus arctos* L., in northern Canada. Can J Zool. 1969;47:167–70.
- 22. Sharma R, Harms NJ, Kukka PM, Parker SE, Gajadhar AA, Jung TS, et al. Tongue has higher larval burden of *Trichinella* spp. than diaphragm in wolverines (*Gulo qulo*). Vet Parasitol. 2018;253:94–7.
- 23. Yukon Bureau of Statistics. Population report June 2014. Information sheet no. 58.49. Government of Yukon, Whitehorse, Canada; 2014.
- Smith CAS, Meikle JC RC. Ecoregions of the Yukon Territory: biophysical properties of Yukon landscapes.: Agriculture and Agri-Food Canada: Summerland, British Columbia; 2004.

Sharma et al. Parasites Vectors (2021) 14:146 Page 9 of 9

- Kukka PM, Jung TS, Robitaille JF, Schmiegelow FKA. Temporal variation in the population characteristics of harvested wolverine (*Gulo gulo*) in northwestern Canada. Wildl Res. 2017;44:497–503.
- Oakley MP, Jung TS, Kukka PM, Robitaille JF. Prevalence of renal calculi in wolverine (*Gulo gulo*) from northwestern Canada. Mamm Biol. 2016;81:189–93.
- Robitaille JF, Villano L, Jung TS, Slama HP, Oakley MP. Fat dynamics and development of body condition indices for harvested populations of wolverine *Gulo gulo*. Wildlife Biol. 2012;18:35–45.
- Kresja DM, Talbot SL, Sage GK, Sonsthagen SA, Jung TS, Magoun AJ, Cook JA. Dynamic landscapes in northwestern North America structured populations of wolverine (*Gulo gulo*). J Mammal. 2021; in press.
- Cook JA, Galbreath KE, Bell KC, Campbell ML, Carrière S, Colella JP, et al.
 The Beringian Coevolution Project: holistic collections of mammals and associated parasites reveal novel perspectives on evolutionary and environmental change in the North. Arct Sci. 2017;3:585–617.
- Slough BG. Status of the wolverine Gulo gulo in Canada. Wildlife Biol. 2007;13:76–82.
- 31. Dean AG, Sullivan KM SM. OpenEpi: open source epidemiologic statistics for public health, version. http://www.OpenEpi.com; 2013.
- 32. Forbes LB, Gajadhar AA. A validated *Trichinella* digestion assay and an associated sampling and quality assurance system for use in testing pork and horse meat. J Food Prot. 1999;62:1308–13.
- Zarlenga DS, Chute MB, Martin A, Kapel CMO. A multiplex PCR for unequivocal differentiation of all encapsulated and non-encapsulated genotypes of *Trichinella*. Int J Parasitol. 1999;29:159–67.
- Scandrett B, Konecsni K, Lalonde L, Boireau P, Vallée I. Detection of natural *Trichinella murrelli* and *Trichinella spiralis* infections in horses by routine post-slaughter food safety testing. Food Waterborne Parasitol. 2018;11:1–5.
- Sergeant E. Epitools epidemiological calculators. Ausvet Pty Ltd; 2019. http://epitools.ausvet.com.au.
- Britov A. *Trichinellosis* in Kamchatka. Wiadomości Parazytol. 1997:43:287–8.
- Mörner T, Eriksson H, Bröjer C, Nilsson K, Uhlhorn H, Ågren E, et al. Diseases and mortality in free-ranging brown bear (*Ursus arctos*), gray wolf (*Canis lupus*), and Wolverine (*Gulo gulo*) in Sweden. J Wildl Dis. 2005;41:298–303.
- Rausch R, Babero BB, Rausch RV, Schiller EL. Studies on the helminth fauna of Alaska. XXVII. The occurrence of larvae of *Trichinella spiralis* in Alaskan mammals. J Parasitol. 1956;42:259–71.
- 39. Addison EM, Boles B. Helminth parasites of wolverine, *Gulo gulo*, from the district of Mackenzie Northwest Territories. Can J Zool. 1978;56:2241–2.
- Hoberg EP, Aubry KB, Brittell JD. Helminth parasitism in martens (Martes americana) and ermines (Mustela erminea) from Washington, with comments on the distribution of Trichinella spiralis. J Wildl Dis. 1990;26:447–52.
- 41. Schmitt N, Saville JM, Friis L, Stovell PL. Trichinosis in British Columbia wildlife. Can J Public Heal. 1976;67:21–4.
- 42. Gottstein B, Pozio E, Nöckler K. Epidemiology, diagnosis, treatment, and control of trichinellosis. Clin Microbiol Rev. 2009;22:127–45.
- 43. Gamble HR, Bessonov AS, Cuperlovic K, Gajadhar AA, Van Knapen F, Noeckler K, et al. International Commission on *Trichinellosis*:

- recommendations on methods for the control of *Trichinella* in domestic and wild animals intended for human consumption. Vet Parasitol. 2000:93:393–408.
- 44. Oksanen A, Lindgren E, Tunkkari P. Epidemiology of trichinellosis in lynx in Finland. J Helminthol. 1998;72:47–53.
- Frey CF, Schuppers ME, Müller N, Ryser-Degiorgis MP, Gottstein B. Assessment of the prevalence of *Trichinella* spp. in red foxes and Eurasian lynxes from Switzerland. Vet Parasitol. 2009;159:295–9.
- 46. Larter N, Elkin B, Allaire D. Prevalence of *Trichinella* spp. in Wildlife of the Dehcho. Government Northwest Territ: 2016.
- 47. Banci V. Ecology and behaviour of wolverine in Yukon. M.Sc. Thesis. Simon Fraser University, Burnaby, BC; 1987.
- 48. Kirjušina M, Deksne G, Marucci G, Bakasejevs E, Jahundovića I, Daukšte A, et al. A 38-year study on *Trichinella* spp. in wild boar (*Sus scrofa*) of Latvia shows a stable incidence with an increased parasite biomass in the last decade. Parasit Vectors. 2015;8:137.
- 49. Brooks DR, Hoberg EPBW. The Stockholm Paradigm: Climate Change and Emerging Disease. Chicago: University of Chicago Press; 2019.
- Brooks DR, Hoberg EP, Boeger WA, Gardner SL, Galbreath KE, Herczeg D, et al. Finding them before they find Us: Informatics, parasites, and environments in accelerating climate change. Comp Parasitol. 2014:81:155–64.
- La Rosa G, Marucci G, Zarlenga DS, Casulli A, Zarnke RL, Pozio E. Molecular identification of natural hybrids between *Trichinella nativa* and *Trichinella* T6 provides evidence of gene flow and ongoing genetic divergence. Int J Parasitol. 2003;33:209–16.
- Dunams-Morel DB, Reichard MV, Torretti L, Zarlenga DS, Rosenthal BM.
 Discernible but limited introgression has occurred where *Trichinella nativa* and the T6 genotype occur in sympatry. Infect Genet Evol. 2012;12:530–8.
- Campbell WC. Trichinosis revisited-another look at modes of transmission. Parasitol Today. 1988;4:83–6.
- Rah H, Chomel BB, Follmann EH, Kasten RW, Hew CH, Farver TB, et al. Serosurvey of selected zoonotic agents in polar bears (*Ursus maritimus*). Vet Rec. 2005;156:7–13.
- Forbes LB. The occurrence and ecology of *Trichinella* in marine mammals. Vet Parasitol. 2000;93:321–34.
- Pozio E, Casulli A, Bologov VV, Marucci G, La Rosa G. Hunting practices increase the prevalence of *Trichinella* infection in wolves from European Russia. J Parasitol. 2001;87:1498–501.
- 57. Proulx JF, MacLean JD, Gyorkos TW, Leclair D, Richter AK, Serhir B, et al. Novel prevention program for trichinellosis in Inuit communities. Clin Infect Dis. 2002;34:1508–14.
- 58. Zimmer IA, Hunter SJ, Morgan CP, Hunt KR, Smith GC, Howell M, et al. Detection and surveillance for animal trichinellosis in GB. Vet Parasitol. 2008;151:233–41.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- $\bullet\,$ thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

