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OPEN Effects of the mode of resocialization after juvenile social isolation on medial prefrontal cortex myelination and function

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Social isolation is an important factor in the development of psychiatric disorders. It is necessary to develop an effective psychological treatment, such as cognitive rehabilitation, for children who have already suffered from social isolation, such as neglect and social rejection. We used socially isolated mice to validate whether elaborate re-socialization after invenile social isolation can restore hypomyelination in the medial prefrontal cortex (mPFC) and the attendant functions manifested in socially isolated mice. While mice who underwent re-socialization with socially isolated mice after juvenile social isolation (Re-IS mice) demonstrated less mPFC activity during exposure to a strange mouse, as well as thinner myelin in the mPFC than controls, mice who underwent re-socialization with socially housed mice after juvenile social isolation (Re-SH mice) caught up with the controls in terms of most mPFC functions, as well as myelination. Moreover, social interaction of Re-IS mice was reduced as compared to controls, but Re-SH mice showed an amount of social interaction comparable to that of controls. These results suggest that the mode of re-socialization after juvenile social isolation has significant effects on myelination in the mPFC and the attendant functions in mice, indicating the importance of appropriate psychosocial intervention after social isolation.

Multiple lines of evidence have shown that social experience during the juvenile period substantially affects brain function and behavior in rodents, dogs, and humans¹⁻⁴. Interestingly, juvenile social experience changes not only brain function, but also brain structure, by affecting myelination⁵⁻⁷. A number of studies have indicated that aberrant juvenile social experiences have long-lasting effects on brain function and structure, extending into adulthood in rodents^{6, 8, 9} and humans^{5, 10}. In addition, other studies have suggested that interventions, such as re-socialization after social isolation, and an enriched environment after adverse juvenile experience can recover these brain abnormalities in rodents^{7,11} and humans¹².

Social isolation, such as neglect and social rejection, which is a form of social experience shutdown in terms of experiencing being disliked or not disliked¹³, is a key factor in the development or exacerbation of psychiatric disorders, such as mood disorders, anxiety disorders, personality disorders, attention-deficit hyperactivity disorder, and autism spectrum disorder (ASD)¹⁴⁻¹⁶. While the most important intervention is to prevent neglect¹⁷, it is also necessary to develop an effective treatment for children who have already suffered from social isolation. Recently, psychosocial interventions, such as cognitive behavioral therapy, have attracted increasing attention in clinical psychiatry^{18, 19}, even for patients with conditions that are regarded as refractory or involving permanent disability, e.g., schizophrenia and ASD, as well as for abused children²⁰. Whereas the main purpose of psychosocial interventions is to improve brain function, recent studies have suggested that such interventions change both brain structure and function^{21, 22}.

Recent studies have shown that myelination in the central nervous system is essential for acquiring motor skills and that myelination in the medial prefrontal cortex (mPFC) is necessary for sociability in mice^{23, 24}. These

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Figure 1. Experimental designs.

findings indicate that myelination in the mPFC can be one of the critical elements in mPFC functions. A previous study has shown that re-socialization with socially isolated mice does not alter the hypomyelination in the mPFC caused by juvenile social isolation⁶, which indicates that social interaction between socially isolated mice is not sufficient to restore hypomyelination. In this study, we employed socially isolated mice as a model of neglect and social rejection and sought to investigate whether re-socialization with socially housed mice, rather than with socially isolated mice, after juvenile social isolation (Fig. 1), can restore hypomyelination in the mPFC and the attendant brain dysfunction, even after myelin impairment is manifest in mice due to juvenile social isolation.

Materials and Methods

Mice and housing conditions. Male C57BL/6J mice were used for each experiment and there was no duplication of mice between experiments. Mice were maintained in a fixed 12-h light-dark cycle (light period, 8:00–20:00; dark period, 20:00–8:00). After weaning at postnatal day 21 (P21), four littermates were reared together in a cage until P65 (control, Ctrl mice) or a mouse was housed in a cage until P35 or P65 (isolation, IS mice). Then, each isolated mouse for 2 weeks after weaning was housed together with three other male isolated mice from the same litter (re-socialization) in a cage, from P35 until P65 (re-socialization with isolated mice; Re-IS mice). On the other hand, from P21 until P35, one mouse was housed individually and three other mice from the same litter were socially housed, and these four mice were then housed together in a cage from P35 until P65 (re-socialization with socially housed mice; Re-SH mice) (Fig. 1a,c). Experimental protocols were approved by the Animal Care Committee of Nara Medical University and all methods were carried out in accordance with the policies of the National Institutes of Health Guide for the Care and Use of Laboratory Animals.

Response of IEGs in the mPFC to social exposure. Experimental pairs of mice (P65) were set up for monitoring IEG mRNA expression in experimental mice in the presence and absence of a strange mouse (that was socially housed) in order to measure neuronal activity in the mPFC during social interaction. A male age-matched mouse was placed into a cage in which four male age-matched experimental littermates (Ctrl, Re-IS, or Re-SH) were housed, and 30 min later, each randomly numbered experimental mouse (No. 1–4) was decapitated to measure IEG mRNA expression in the mPFC ("IEG mRNA expression with exposure"). Alternatively, four co-housed Ctrl, Re-IS, or Re-SH mice (No. 1–4) were decapitated to measure IEG mRNA expression in the mPFC ("IEG mRNA expression without exposure"). The ratio of mPFC neuronal activity (IEG mRNA expression with exposure/IEG mRNA expression without exposure) of randomly paired mice (No. 1 from mice with exposure) was measured. This experiment was performed in dim light during the dark period.

Quantitative reverse transcription polymerase chain reaction (qRT-PCR). mRNA levels of *cFos*, *Arc*, and *Npas4* were measured in layers V and VI of the mouse mPFC at P65 by means of qRT-PCR. The details have previously been described⁶. Coronal sections (1-mm-thick) of layer V and VI in the prelimbic and infralimbic areas of the mPFC were dissected around Bregma 1.70 mm²⁵. RNA was isolated using RNeasy Mini Kit (Qiagen; Valencia, CA, USA) following the manufacturer's protocol, and any genomic DNA contamination was eliminated by DNase treatment. RNA quantity was determined by absorbance at 260 nm. cDNA was then synthesized from 500 ng of RNA using an iScript kit (Bio-Rad Laboratories; Hercules, CA, USA) as recommended by the supplier. qRT-PCR was performed using SYBR *Premix Ex Taq*TM II (Takara Bio Inc., Shiga, Japan) according to the manufacturer's recommendations. The 18S ribosomal RNA was chosen as an internal control for the delta-delta C_T method. Primer sequences are available upon request. *In vivo* electroencephalogram (EEG). In order to measure δ - (0.5–4Hz), θ - (4–8Hz), α - (8–12Hz), σ - (12–16Hz), β - (16–24Hz), and γ - (25–55Hz) oscillations in the mPFC, we used a telemetry implant (PhysioTel[®] transmitter TA10ETA-F20; DSI, Lexington, KY, USA) and a receiver (RPC-1, DSI). Mice were anesthetized with chloral hydrate (400 mg/kg) (Nacalai Tesque Inc., Kyoto, Japan). An electric probe was stereotaxically inserted into layer V within the prelimbic and infralimbic areas of mPFC (Bregma: 1.9 mm anterior, 0.5 mm lateral [right], 3.5 mm ventral) and the other electrode was placed in the longitudinal cerebral fissure (Bregma: 6 mm posterior, 0-mm lateral). DataquestTM A.R.T.TM (DSI) software was used to acquire and analyze EEG data. EEGs were recorded during the habituation period, in which mice were placed in a novel cage for 30 min, and during subsequent social interaction, in which an age-matched male mouse was inserted into the cage for 5 min. The ratio of oscillation power during social interaction (per 5 min) to the average of oscillation power during habituation (per 5 min) was measured to evaluate mPFC function in response to social interaction. The randomization of mice for examination was performed as in the IEG response experiments (Fig. 1b). This experiment was performed in darkness during the dark period.

Electron microscopy. Four mice per condition (Ctrl, IS, Re-IS, and Re-SH), at P65, were perfused intracardially with fixative (2% paraformaldehyde, 2.5% glutaraldehyde, and 0.1% picric acid in 100 mM cacodylate buffer); the brains were removed from the skulls and fixed in the same solution overnight. Coronal sections (500-µm-thick) were then obtained to isolate the mPFC (between Bregma 1.70 mm and 1.98 mm), embedded in epoxy resin, and examined by transmission electron microscope in 50-nm sections. Using this approach, multiple, non-overlapping regions of the brain from each mouse were imaged. Three mice were excluded from the study due to insufficient myelin fixation. A g-ratio was calculated as the axon perimeter/(axon + myelin sheath) perimeter, and axons with circularity ($4 \times \pi \times$ area/perimeter²) under 6.0 were excluded from analysis, because the perimeter does not precisely reflect myelin thickness in cases where the axons were cut at a slant. The average value of the g-ratio from each mouse was used for statistical analysis.

Social interaction test. Two clear perforated boxes, one of which contained a male mouse at about P120, and which had not been housed with experimental mice, were placed in an open field $(40 \text{ cm} \times 40 \text{ cm})$. The Ctrl, IS, Re-IS, and Re-SH mice were placed in the open field and was allowed to explore the apparatus freely for 5 min. A video tracking system traced the nose point and center point of their bodies to measure social interaction and locomotion, respectively, and then the time spent within 2.5 cm of each box was recorded (TopScan Suite, Clever Sys Inc., Reston, VA, USA). Social interaction was analyzed as the ratio of time spent exploring the box containing the mouse to the total time approaching both boxes. This experiment was performed in dim light during the dark period.

Statistical analyses. Analysis of variance (ANOVA) followed by the Newman-Keuls multiple comparison test was used for statistical analyses with GraphPad Prism. Data are presented as mean \pm standard error of mean. P values less than 0.05 were considered statistically significant.

Results

The mode of re-socialization after juvenile social isolation affects mPFC activity. Since hypoactive mPFC function after child maltreatment has been reported²⁶, we hypothesized that mPFC function would be affected by the mode of re-socialization. Immediate early gene (IEG) responses in the mPFC during exposure to a strange mouse were measured as an index of mPFC function (IEG mRNA expression with exposure/IEG mRNA expression without exposure). In this study, we employed *cFos* as a general marker of neuronal activity²⁷, *Arc* as a marker of excitatory synapse²⁸, and *Npas4* as a marker of inhibitory synapse²⁹. As expected, changes in *cFos*, *Arc*, and *Npas4* expression were much less marked in Re-IS mice than in Ctrl mice, but were not significantly different between Ctrl mice and Re-SH mice, except for that of *cFos*, while Re-SH mice showed higher expression levels of these genes than did Re-IS mice (*cFos*, n = 4/group, F(2, 9) = 28.34; *Arc*, n = 4/group, F(2, 9) = 16.01; *Npas4*, n = 4/group, F(2, 9) = 4.878) (Fig. 2a). On the other hand, there were no differences in terms of changes in IEG expression after exposure to a strange mouse in the motor cortex (*cFos*, n = 4/group, F(2, 9) = 1.650; *Arc*, n = 4/group, F(2, 9) = 0.4589; *Npas4*, n = 4/group, F(2, 9) = 2.344) (Fig. 2b). These results suggest that the mode of re-socialization after juvenile social isolation affects mPFC activity during the exposure to a strange mouse.

The mode of re-socialization after juvenile social isolation affects the power of γ -oscillation and β -oscillation. Based on these findings, we further aimed to investigate mPFC function by evaluating changes in the power of electric oscillations (δ , θ , α , σ , β , γ) in the mPFC in response to a strange mouse, which could be an electrophysiological measure for social interaction³⁰. Interestingly, the responses in γ -oscillation paralleled expression of one of the IEGs (*cFos*). Re-IS mice had lower responses in γ -oscillation than did Ctrl mice and Re-SH mice, while Re-SH mice had lower responses than Ctrl mice (n = 3/group, F(2, 6) = 17.21) (Fig. 3f). In contrast, the power of β -oscillation was markedly higher in Re-IS mice than in Ctrl and Re-SH mice (n = 3/group, F(2, 6) = 96.68) (Fig. 3e). The powers of δ -, θ -, α -, σ -oscillations were not different between the three groups (n = 3/group, F(2, 6) = 1.100; n = 3/group, F(2, 6) = 0.4345; n = 3/group, F(2, 6) = 0.3958; n = 3/group, F(2, 6) = 1.890), respectively) (Fig. 3a-d).

The mode of re-socialization after juvenile social isolation affects myelination in the mPFC. In order to evaluate whether the mode of re-socialization after juvenile social isolation affected myelination in the mPFC, an index of myelin thickness, the g-ratio, in layer V of the mPFC, was compared. We statistically compared the average g-ratio of more than 50 axons from each mouse (n = 3/group). While IS and Re-IS mice had thinner myelin in the mPFC than did Ctrl mice, Re-SH mice had a myelin thickness comparable to that of Ctrl mice (F(3, 8) = 19.29) (Fig. 4a,b), indicating that myelination in the mPFC is sensitive to the mode of re-socialization after juvenile social isolation.





The mode of re-socialization after juvenile social isolation affects social interaction. A recent study has shown that restoration of myelination in the mPFC can improve impaired social interaction of mice exposed to protracted social isolation²⁴, which suggests that myelin can control social interaction. In the current study, IS and Re-IS mice with hypomyelination in the mPFC showed poor social interaction as compared to Ctrl mice with intact myelination, while Re-SH mice with intact myelination demonstrated social interaction comparable to that of Ctrl mice. Re-SH mice showed higher social interaction than IS and Re-IS mice (Ctrl, n = 16; IS, n = 16; Re-IS, n = 16; Re-SH, n = 12, F(3, 56) = 4.558) (Fig. 5a). There were no differences in the locomotive distance of mice during the social interaction test (Ctrl, n = 16; IS, n = 16; Re-SH, n = 12, F(3, 56) = 0.2461) (Fig. 5b). These results indicated a possibility that appropriate re-socialization among others, after juvenile social isolation, can improve social interaction by restoring myelination.



Figure 3. The powers of β - and γ -oscillation in the medial prefrontal cortex (mPFC) are dependent on the mode of re-socialization. The powers of δ -, θ -, α -, σ -, β -, and γ -oscillation in the m PFC during exposure to a strange mouse for 5 min (relative to baseline) were measured. (**a**-**d**) The power of either δ -, θ -, α -, or σ -oscillation was not different between Ctrl, Re-IS, and Re-SH mice. (**e**) The power of β -oscillation was markedly increased in Re-IS mice as compared to Ctrl and Re-SH mice. (**f**) The increase in γ -oscillation power in Re-IS mice was significantly less than that in Ctrl and Re-SH mice, while the amount of change in Re-SH mice was less than that in Ctrl mice. **n** = 3/group, ***p** < 0.05, ****p** < 0.01, *****p** < 0.01. Error bars = SEM.



Figure 4. Myelin thickness in the medial prefrontal cortex (mPFC) depends on the mode of re-socialization. (a) Images of myelinated axons in layer V of the mPFC. (b) The average g-ratio of axons from each mouse (n = 3) was used as a statistical value. The statistical comparison of Ctrl, IS, Re-IS, and Re-SH mice showed that the myelin of IS and Re-IS mice was thinner than that of Ctrl and Re-SH mice, while Ctrl and Re-SH mice showed comparable myelin thickness (more than 50 axons/each mouse, n = 3/group), **p < 0.01, ***p < 0.001, scale bar = 0.5 μ m. Error bars = SEM.



Figure 5. Social interaction depends on the mode of re-socialization. The ratio of time taken to approach a box with a mouse to that taken to approach a box without a mouse was measured. (**a**) Social interaction of IS and Re-IS mice was reduced compared to that of Ctrl, while Ctrl and Re-SH mice showed a comparable score for social interaction. Re-SH mice showed higher social interaction than IS and Re-IS mice. (**b**) There were no differences in the distance of locomotion during social interaction test. Ctrl; n = 16, IS; n = 16, Re-IS; n = 16, Re-SH; n = 12, *p < 0.05. Error bars = SEM.

Discussion

Neglect affects children devastatingly and deeply, and possibly influences their mental state for the rest of their life. Children suffering neglect often present with language deficits, which may be due to insufficient language exposure to others. The language problem may exaggerate the extent of their social isolation. Those children are also likely to have mental problems, such as anxiety, depression, somatization, paranoia, and hostility³¹. Based on these clinical studies, we investigated whether psychosocial interventions after juvenile social isolation, leading to hypomyelination, could affect myelination in the mPFC and the attendant functions, whereas it remains unclear whether social isolated during the juvenile period and then re-socialized with its three littermates that had also been socially isolated. Considering that the behaviors of socially isolated mice appear abnormal, we considered that re-socialization. Therefore, in the current study, we also isolated a mouse during the juvenile period and then placed it back with three littermates that had been socially housed. We then investigated the phenotypes created by these seemingly subtle differences in social interaction.

While social interaction between socially isolated mice did not change myelination, re-socializing these mice with socially housed mice resulted in restoration of hypomyelination in the mPFC, as well as recovery of mPFC dysfunctions caused by juvenile social isolation from P21 to P35. In contrast to the mPFC, social experience had no effects on locomotor function (Fig. 5b) or myelination in the motor cortex⁶, indicative of a lack of a motor function response as well as myelination in the motor cortex to social experience. These findings may indicate that appropriate interventions can be beneficial, even after brain structure has been damaged by juvenile social isolation, implying that psychosocial approaches can be effective for the treatment of abused children, and can even affect aspects of brain structure, such as myelination, in the mPFC.

Myelin speeds up the axonal conduction velocity by inducing saltatory conduction³². If myelination is impaired, and axonal conduction velocity consequently altered, the brain function in the affected region is likely to be disordered due to loss of optimal synchrony of the neuronal networks³³. In this study, we focused on myelination in layer V of the mPFC, because myelination is much more substantial in layers V and VI than in layers II, III, and IV³⁴. The differences in myelination may explain the variations in the IEG responses and in γ -oscillation in layers V and VI of the mPFC, between Re-IS mice and Re-SH mice. Several reports have provided evidence that myelin impairment may be a primary cause of psychiatric symptoms. Transgenic mice with a primary oligodendrocyte deficit show schizophrenia-like symptoms³⁵, while the restoration of social isolation-induced hypomyelination improves social interaction ability²⁴. Taken together, our findings indicate that the reduced sociability of Re-IS mice may be caused by hypomyelination in the mPFC.

On the other hand, it has also been shown that neuronal activity changes myelination³⁶. Oligodendrocytes express membrane receptors for various growth factors and, interestingly, also express ion channels and neurotransmitter receptors. In addition to growth factors, adenosine tyrosine kinase, adenosine, gamma-aminobutyric acid, and glutamate have been shown to address oligodendrocytes and affect myelination^{33, 36, 37}. Neurotransmitters, such as glutamate, have traditionally been regarded as molecules that are released at synapses. However, recent studies have shown that neurotransmitters can also be released from axons through membrane channels³⁷. Therefore, oligodendrocytes can directly receive such molecules from axons, forming activity-dependent myelination along the axons. Based on these findings, the restoration of mPFC myelination in Re-SH mice can possibly be attributed to high neuronal activities in the mPFC as compared to Re-IS mice, as shown in this study. Furthermore, a recent article has also indicated that the extent of γ -oscillation might be markedly subject to activity-dependent myelination³⁸, suggesting that myelination may shape neural networks.

In the current study, two types of measurements were employed to investigate mPFC function, viz., IEG responses and γ -oscillations. We chose to investigate *cFos*, *Arc*, and *Npas4* among the IEGs. *cFos* mRNA expression acts as a general marker of recent neuronal activity, as cFos expression is elevated immediately after neuronal activation³⁹. The levels of Arc and Npas4 mRNA are markers of excitatory and inhibitory transmission in neurons, respectively^{40, 41}, and they are also critical molecules for the formation of excitatory and inhibitory synapses, respectively. Since the expression of all these three IEGs in Ctrl and RE-SH mice was higher than that in Re-IS mice, both excitatory and inhibitory transmission in neurons were apparently increased by re-socialization with socially housed mice as compared to social isolated mice. On the other hand, γ -oscillation is a rhythmic γ -band neural activity, and in the mPFC, γ -oscillation has been shown to play a critical role in maintaining the excitatory-inhibitory balance (E-I balance) that is related to emotion, cognition, and social interaction $30, 42, 43, \gamma$ -Oscillation is controlled by parvalbumin-positive cells, which are inhibitory neurons⁴². Our current results suggest that re-socialization with socially housed mice may increase the function of the parvalbumin-positive cells in the mPFC, which may alter the E-I balance and thereby social interaction. The elevated function of the parvalbumin-positive cells may accelerate axonal activity-dependent myelination on the basis of recent finding that axons of inhibitory neurons are substantially myelinated³⁴. Further electrophysiological and histological experiments are needed to dissect the E-I balance and myelination in the mPFC after re-socialization.

Both β - and γ -oscillation in the mPFC are related to attention; top-down attention tasks are mediated by β -oscillation and the power of γ -oscillation increases during bottom-up attention tasks^{44, 45}. Based on these findings, Re-IS mice and Re-SH mice might have different modalities of attention due to altered frequencies of oscillations; this may underlie our social interaction test findings. Whereas the mechanism underlying the switches between β - and γ -oscillation in the mPFC remains unknown, recent studies have shown that the competition between these oscillations depends on sensory inputs and centrifugal inputs in the olfactory bulb. This indicates that different natural conditions, such as odor features and behaviors, affect the occurrence of both oscillations⁴⁶. Based on these findings, we speculate that different inputs into the mPFC between Re-IS and Re-SH mice might control β - and γ -oscillation in the brain region. Although it is interesting to investigate oscillations of IS mice as a reference to Re-IS and Re-SH mice, it is not possible to examine mPFC responses to social exposure of IS mice after the current experimental protocol, in which four littermates in a home cage were exposed to a strange mouse; this protocol is not applicable to IS mice individually housed in a cage.

Altogether, we found that the two types of re-socialization led to different neuronal activities and myelination in the mPFC; nevertheless, further studies are needed to observe the exact differences in social interaction during re-socialization. We speculated that the amount of social interaction between isolated mice (Re-IS) was less than that between the isolated mice and socially housed mice (Re-SH). Based on the current findings, it is reasonable to consider that the differences in myelin formation are dependent on neuronal activities in this brain region, although it is not possible to deduce whether myelination or neuronal activity occurred first. As exercise accelerates myelination and new myelination is necessary to acquire exercise skill²³, we hypothesize that, analogously, more "intense" re-socialization after juvenile social isolation may accelerate myelination in the mPFC, and that this novel myelination is necessary for acquiring social skills. These findings strongly support the possibility that an appropriate psychosocial approach can be used as intervention for abused children, even when they have already developed psychiatric symptoms, as it alters myelination and the attendant functions in the mPFC. This study warrants further investigations to assess whether changes in myelination or neuronal function is the primary cause of the impaired social interaction that is caused by social isolation of juveniles.

References

- Han, X., Wang, W., Xue, X., Shao, F. & Li, N. Brief social isolation in early adolescence affects reversal learning and forebrain BDNF expression in adult rats. *Brain research bulletin* 86, 173–178, doi:10.1016/j.brainresbull.2011.07.008 (2011).
- 2. Freedman, D. G., King, J. A. & Elliot, O. Critical period in the social development of dogs. Science 133, 1016–1017 (1961).
- Chugani, H. T. et al. Local brain functional activity following early deprivation: a study of postinstitutionalized Romanian orphans. NeuroImage 14, 1290–1301, doi:10.1006/nimg.2001.0917 (2001).
- Bick, J., Fox, N., Zeanah, C. & Nelson, C. A. Early deprivation, atypical brain development, and internalizing symptoms in late childhood. *Neuroscience*, doi:10.1016/j.neuroscience.2015.09.026 (2015).
- Eluvathingal, T. J. et al. Abnormal brain connectivity in children after early severe socioemotional deprivation: a diffusion tensor imaging study. Pediatrics 117, 2093–2100, doi:10.1542/peds.2005-1727 (2006).
- Makinodan, M., Rosen, K. M., Ito, S. & Corfas, G. A critical period for social experience-dependent oligodendrocyte maturation and myelination. *Science* 337, 1357–1360, doi:10.1126/science.1220845 (2012).
- 7. Liu, J. *et al.* Impaired adult myelination in the prefrontal cortex of socially isolated mice. *Nature neuroscience* **15**, 1621–1623, doi:10.1038/nn.3263 (2012).
- Carlyle, B. C. *et al.* Maternal separation with early weaning: a rodent model providing novel insights into neglect associated developmental deficits. *Development and psychopathology* 24, 1401–1416, doi:10.1017/S095457941200079X (2012).
- 9. Weaver, I. C. *et al.* Epigenetic programming by maternal behavior. *Nature neuroscience* 7, 847–854, doi:10.1038/nn1276 (2004).
- Li, M., D'Arcy, C. & Meng, X. Maltreatment in childhood substantially increases the risk of adult depression and anxiety in prospective cohort studies: systematic review, meta-analysis, and proportional attributable fractions. *Psychological medicine* 1–14, doi:10.1017/S0033291715002743 (2015).
- 11. Bredy, T. W., Humpartzoomian, R. A., Cain, D. P. & Meaney, M. J. Partial reversal of the effect of maternal care on cognitive function through environmental enrichment. *Neuroscience* 118, 571–576 (2003).

- 12. Le Mare, L. & Audet, K. A longitudinal study of the physical growth and health of postinstitutionalized Romanian adoptees. *Paediatrics & child health* 11, 85–91 (2006).
- Dodge, K. A., Coie, J. D. & Brakke, N. P. Behavior patterns of socially rejected and neglected preadolescents: the roles of social approach and aggression. *Journal of abnormal child psychology* 10, 389–409 (1982).
- 14. Maguire, S. A. *et al.* A systematic review of the emotional, behavioural and cognitive features exhibited by school-aged children experiencing neglect or emotional abuse. *Child: care, health and development* **41**, 641–653, doi:10.1111/cch.12227 (2015).
- Carr, C. P., Martins, C. M., Stingel, A. M., Lemgruber, V. B. & Juruena, M. F. The role of early life stress in adult psychiatric disorders: a systematic review according to childhood trauma subtypes. *The Journal of nervous and mental disease* 201, 1007–1020, doi:10.1097/ NMD.000000000000049 (2013).
- Platt, B., Cohen Kadosh, K. & Lau, J. Y. The role of peer rejection in adolescent depression. *Depression and anxiety* 30, 809–821, doi:10.1002/da.22120 (2013).
- 17. Casillas, K. L., Fauchier, A., Derkash, B. T. & Garrido, E. F. Implementation of evidence-based home visiting programs aimed at reducing child maltreatment: A meta-analytic review. *Child abuse & neglect*, doi:10.1016/j.chiabu.2015.10.009 (2015).
- Kasari, C., Shire, S., Factor, R. & McCracken, C. Psychosocial treatments for individuals with autism spectrum disorder across the lifespan: new developments and underlying mechanisms. *Current psychiatry reports* 16, 512, doi:10.1007/s11920-014-0512-6 (2014).
- Chien, W. T., Leung, S. F., Yeung, F. K. & Wong, W. K. Current approaches to treatments for schizophrenia spectrum disorders, part II: psychosocial interventions and patient-focused perspectives in psychiatric care. *Neuropsychiatric disease and treatment* 9, 1463–1481, doi:10.2147/NDT.S49263 (2013).
- Leenarts, L. E., Diehle, J., Doreleijers, T. A., Jansma, E. P. & Lindauer, R. J. Evidence-based treatments for children with traumarelated psychopathology as a result of childhood maltreatment: a systematic review. *European child & adolescent psychiatry* 22, 269–283, doi:10.1007/s00787-012-0367-5 (2013).
- Eack, S. M. *et al.* Neuroprotective effects of cognitive enhancement therapy against gray matter loss in early schizophrenia: results from a 2-year randomized controlled trial. *Archives of general psychiatry* 67, 674–682, doi:10.1001/archgenpsychiatry.2010.63 (2010).
- Keller, T. A. & Just, M. A. Altering cortical connectivity: remediation-induced changes in the white matter of poor readers. *Neuron* 64, 624–631, doi:10.1016/j.neuron.2009.10.018 (2009).
- 23. McKenzie, I. A. *et al.* Motor skill learning requires active central myelination. *Science* **346**, 318–322, doi:10.1126/science.1254960 (2014).
- Liu, J. et al. Clemastine Enhances Myelination in the Prefrontal Cortex and Rescues Behavioral Changes in Socially Isolated Mice. The Journal of neuroscience: the official journal of the Society for Neuroscience 36, 957–962, doi:10.1523/JNEUROSCI.3608-15.2016 (2016).
- Van De Werd, H. J. & Uylings, H. B. Comparison of (stereotactic) parcellations in mouse prefrontal cortex. Brain structure & function 219, 433-459, doi:10.1007/s00429-013-0630-7 (2014).
- van Harmelen, A. L. et al. Hypoactive medial prefrontal cortex functioning in adults reporting childhood emotional maltreatment. Social cognitive and affective neuroscience 9, 2026–2033, doi:10.1093/scan/nsu008 (2014).
- Cruz, F. C. et al. New technologies for examining the role of neuronal ensembles in drug addiction and fear. Nature reviews. Neuroscience 14, 743–754, doi:10.1038/nrn3597 (2013).
- Chowdhury, S. et al. Arc/Arg3.1 interacts with the endocytic machinery to regulate AMPA receptor trafficking. Neuron 52, 445–459, doi:10.1016/j.neuron.2006.08.033 (2006).
- Bloodgood, B. L., Sharma, N., Browne, H. A., Trepman, A. Z. & Greenberg, M. E. The activity-dependent transcription factor NPAS4 regulates domain-specific inhibition. *Nature* 503, 121–125, doi:10.1038/nature12743 (2013).
- Yizhar, O. et al. Neocortical excitation/inhibition balance in information processing and social dysfunction. Nature 477, 171–178, doi:10.1038/nature10360 (2011).
- Gauthier, L., Stollak, G., Messe, L. & Aronoff, J. Recall of childhood neglect and physical abuse as differential predictors of current psychological functioning. *Child abuse & neglect* 20, 549–559 (1996).
- Tolhurst, D. J. & Lewis, P. R. Effect of myelination on the conduction velocity of optic nerve fibres. Ophthalmic & physiological optics: the journal of the British College of Ophthalmic Opticians 12, 241–243 (1992).
- Fields, R. D. A new mechanism of nervous system plasticity: activity-dependent myelination. *Nature reviews. Neuroscience* 16, 756–767, doi:10.1038/nrn4023 (2015).
- Micheva, K. D. et al. A large fraction of neocortical myelin ensheathes axons of local inhibitory neurons. eLife 5, doi:10.7554/ eLife.15784 (2016).
- Roy, K. et al. Loss of erbB signaling in oligodendrocytes alters myelin and dopaminergic function, a potential mechanism for neuropsychiatric disorders. Proceedings of the National Academy of Sciences of the United States of America 104, 8131–8136, doi:10.1073/pnas.0702157104 (2007).
- Wake, H., Moorhouse, A. J., Jinno, S., Kohsaka, S. & Nabekura, J. Resting microglia directly monitor the functional state of synapses in vivo and determine the fate of ischemic terminals. *The Journal of neuroscience: the official journal of the Society for Neuroscience* 29, 3974–3980, doi:10.1523/jneurosci.4363-08.2009 (2009).
- Lundgaard, I. *et al.* Neuregulin and BDNF induce a switch to NMDA receptor-dependent myelination by oligodendrocytes. *PLoS biology* 11, e1001743, doi:10.1371/journal.pbio.1001743 (2013).
- Pajevic, S., Basser, P. J. & Fields, R. D. Role of myelin plasticity in oscillations and synchrony of neuronal activity. *Neuroscience* 276, 135–147, doi:10.1016/j.neuroscience.2013.11.007 (2014).
- Northcutt, K. V. & Lonstein, J. S. Social contact elicits immediate-early gene expression in dopaminergic cells of the male prairie vole extended olfactory amygdala. *Neuroscience* 163, 9–22, doi:10.1016/j.neuroscience.2009.06.018 (2009).
- 40. Steward, O. & Worley, P. F. Selective targeting of newly synthesized Arc mRNA to active synapses requires NMDA receptor activation. *Neuron* **30**, 227–240 (2001).
- Lin, Y. et al. Activity-dependent regulation of inhibitory synapse development by Npas4. Nature 455, 1198–1204, doi:10.1038/ nature07319 (2008).
- Canetta, S. et al. Maternal immune activation leads to selective functional deficits in offspring parvalbumin interneurons. Molecular psychiatry. doi:10.1038/mp.2015.222 (2016).
- Nakamura, T. *et al.* Relationships among parvalbumin-immunoreactive neuron density, phase-locked gamma oscillations, and autistic/schizophrenic symptoms in PDGFR-beta knock-out and control mice. *PloS one* 10, e0119258, doi:10.1371/journal. pone.0119258 (2015).
- Benchenane, K., Tiesinga, P. H. & Battaglia, F. P. Oscillations in the prefrontal cortex: a gateway to memory and attention. *Current opinion in neurobiology* 21, 475–485, doi:10.1016/j.conb.2011.01.004 (2011).
- 45. Pinto, Y., van der Leij, A. R., Sligte, I. G., Lamme, V. A. & Scholte, H. S. Bottom-up and top-down attention are independent. *Journal of vision* 13, 16, doi:10.1167/13.3.16 (2013).
- 46. David, F., Courtiol, E., Buonviso, N. & Fourcaud-Trocme, N. Competing Mechanisms of Gamma and Beta Oscillations in the Olfactory Bulb Based on Multimodal Inhibition of Mitral Cells Over a Respiratory Cycle. *eNeuro* 2, doi:10.1523/ ENEURO.0018-15.2015 (2015).

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Author Contributions

M.M., D.I., K.Y., Y.Y., M.T., S.K., T.Y., K.O., T.K., S.F., and H.Y. performed experiments. M.M., S.K., A.W., and T.K. made plans for this study. M.M. and T.K. wrote this manuscript.

Additional Information

Competing Interests: The authors declare that they have no competing interests.

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