

Research Article

Ototoxicity from Combined Cisplatin and Radiation Treatment: An In Vitro Study

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Objective. Combined cisplatin (CDDP) and radiotherapy is increasingly being used to treat advanced head and neck cancers. As both CDDP and radiation can cause hearing loss, it is important to have a better understanding of the cellular and molecular ototoxic mechanisms involved in combined therapy. **Procedure.** The effects of CDDP, radiation, and combined CDDP-radiation on the OC-k3 cochlear cell line were studied using MTS assay, flow cytometry, Western blotting, and microarray analysis. **Results.** Compared to using CDDP or radiation alone, its combined use resulted in enhanced apoptotic cell death and apoptotic-related gene expression, including that of FAS. Phosphorylation of p53 at Ser15 (a marker for p53 pathway activation in response to DNA damage) was observed after treatment with either CDDP or radiation. However, posttreatment activation of p53 occurred earlier in radiation than in CDDP which corresponded to the timings of MDM2 and TP53INP1 expression. **Conclusion.** Enhanced apoptotic-related gene expressions leading to increased apoptotic cell deaths could explain the synergistic ototoxicity seen clinically in combined CDDP-radiation therapy. CDDP and radiation led to differential temporal activation of p53 which suggests that their activation is the result of different upstream processes. These have implications in future antiapoptotic treatments for ototoxicity.

1. Introduction

Combined chemoradiotherapy is increasingly being used to treat advanced head and neck cancers. During radiotherapy, the ear structures are often included in the radiation fields and it is generally accepted that radiation-induced sensorineural hearing loss can result. Cisplatin (CDDP), widely used as an effective antineoplastic drug for these cancers, is also known to cause ototoxicity. In a randomized blinded study, it was demonstrated that patients who had received radiotherapy and concurrent/adjuvant chemotherapy using CDDP experienced greater sensorineural hearing loss compared with patients treated with radiotherapy alone [1]. This was especially so in the high-frequency sounds of the speech range, resulting in significant hearing disability.

In recent years, immortalized cell lines derived from the mouse organ of Corti had been developed and characterized [2]. For example, the OC-k3 cell line was derived from the organ of Corti of the transgenic mouse. It encoded the large

T antigen of the SV40 (simian virus 40), a thermolabile viral protein which drove the cells to proliferate indefinitely at 33°C and in the presence of gamma interferon [3]. This cell line expressed the neuro-epithelial precursor cell marker nestin and the inner ear cell marker OCP2, but did not exhibit markers for glial or neuronal cells. In addition, OC-k3 cells expressed specific auditory sensory cell markers (myosin VIIa and the acetylcholine receptor alpha-9) and the supporting cell marker connexin 26. This and other similar cell lines had been regarded as good models to study the mechanisms of cell fate in the organ of Corti of the cochlea [4].

P53 had been found to play an important role in apoptotic cell death associated with ototoxicity. In a CDDP-induced apoptosis experiment using cochlear organotypic cultures prepared from rats at postnatal days 3-4, significant upregulation of phospho-p53 serine 15 expression was found and apoptosis was suppressed by pifithrin- α , a p53 inhibitor [5]. Other studies have shown that the deletion of the p53

gene protects sensory hair cells from CDDP-induced cell death, caspase-2 activation, and cytochrome c translocation [6]. In radiation-induced ototoxicity, it was found that p53 together with reactive oxidative species (ROS) played an important role in cochlear cell apoptosis [7].

In the combined use of CDDP and radiation, the cellular and molecular mechanisms leading to ototoxicity had not been studied. It is important to have a better understanding of these mechanisms as effective preventive strategies directed at the relevant pathways can potentially be developed. The present study found that although p53 played a role in both CDDP and radiation-induced cochlear cell apoptosis, p53 was activated at different time points after each treatment which corresponded to the time MDM2 and TP53INP1 were expressed. Additional apoptotic-related genes that were not expressed when CDDP or radiation was used alone were expressed when used in combination. This included FAS, an important element involved in the extrinsic apoptotic pathway.

2. Materials and Methods

2.1. Cell Culture. The immortalized OC-k3 cell line derived from the organ of Corti of the transgenic mice (Immortal-mouse H-2Kb-tsA58, Charles Rivers Laboratories, Wilmington, MA) was used. The cell line was cultured in high-glucose Dulbecco's Eagle's medium (DMEM, Gibco, Grand Island, NY) supplemented with 10% fetal bovine serum (FBS, Gibco, Grand Island, NY), 1% penicillin-streptomycin (P/S, Gibco, Grand Island, NY), and 50 U/ml gamma-interferon (mouse recombinant, Sigma-Aldrich, St. Louis, MO) and maintained at 33°C with 10% CO₂. To study the impact of chemoradiation treatment, OC-k3 cells were exposed to 5 Gy of gamma irradiation alone, 0.5 µg/ml of cisplatin alone, or 5 Gy of gamma irradiation in the presence of 0.5 µg/ml cisplatin (Pfizer, Bentley, WA).

2.2. Cell Viability Assay. The OC-k3 cells were seeded in 96-well plates at densities of 5×10^3 cells/well in 200 µl complete medium after being exposed to chemo-irradiation treatment. Cell viability was determined using CellTiter 96 Aqueous One Solution Cell Proliferation Assay (Promega Corp., Madison, WI) containing tetrazolium compound 3-[4,5-dimethylthiazol-2-yl]-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium (MTS) at 3 h, 24 h, 48 h, and 72 h after chemo-irradiation. This test was based on the bioreduction of MTS compound into a soluble and colored formazan product by NADPH or NADH, which is produced by dehydrogenase enzymes in metabolically active cells. Twenty microliters of MTS were added to each well, incubated at 33°C for 3 h, and then the absorbance was recorded at 490 nm with a microplate spectrophotometer (Benchmark Plus, Bio-Rad Laboratories, Hercules, CA).

2.3. Cell Death Analysis. The cells were collected at each time point post CDDP-radiation treatment, fixed in 75% ethanol and stored at 4°C. Upon analysis, the cells were washed with PBS and incubated with 100 µg/ml propidium iodide

(PI) containing 0.1% Triton X-100 and 500 µg/ml RNase A in 50 µl PBS for 30 mins in darkness at 4°C. The DNA contents of cells were analyzed using the flow cytometer CyAn™ ADP Analyser (Beckman Coulter, Fullerton, CA). The magnitudes of the sub-G1 fractions were determined using the Summit 4.3 software (Beckman Coulter, Fullerton, CA). DNA fragmentation resulting from apoptotic cell death would manifest in the sub-G1 fraction.

2.4. Western Blot Analysis. Protein extraction was done by incubating the cells at 4°C for 30 minutes in lysis buffer containing 150 mM NaCl, 10 mM Tris-HCl pH 7.4, 2 mM EDTA, 0.5 mM EGTA, 1 mM sodium orthovanadate, 0.1% sodium deoxycholate, 0.5% NP-40, and 1% Triton X-100 supplemented with 1x complete protease inhibitor mixture (Roche, Basel, Switzerland). Equal amounts of protein samples were denatured separated by 10% SDS-PAGE and transferred onto nitrocellulose membrane by iBlot dry blotting system (Invitrogen, Carlsbad, CA). The membrane was blocked with 5% nonfat milk in PBS with 0.1% Tween-20 (PBST) for 1 h, followed by an overnight incubation of primary antibodies in 5% BSA/PBST at 4°C. Primary antibodies included anti-p53 pAb (NCL-p53-CM5p, Novocastra), anti-phospho-p53 (ser-15) pAb, anti-phospho-c-jun (ser-73) pAb, anti-c-jun (60A8) mAb (Cell Signaling Technology, Inc.), and anti-beta-actin mAb (Sigma-Aldrich, St. Louise, MO). After washing the membrane extensively, incubation with horseradish peroxidase-conjugated antirabbit or antimouse secondary antibody (Cell Signaling Technology, Inc.) was done for 1 h at room temperature. After washing, the membrane was incubated in Immobilon Western chemiluminescent HRP substrate (Millipore, Billerica, MA), and the chemiluminescence signals were detected using UViChem (UViTec, Cambridge, UK), a dedicated chemiluminescence documentation system. For reprobing with a new primary antibody, the membrane was stripped in Re-Blot plus strong solution Western blot stripping buffer (Chemicon, Temecula, CA) at room temperature for 30 minutes and rinsed 3 times with PBST for 10 minutes each time.

2.5. Microarray Analysis. The global changes of gene expression were analyzed at 3 h, 24 h, and 72 h after chemoirradiation, on the GeneChip Mouse Genome 430A 2.0 Array (Affymetrix, Santa Clara, CA). Biological duplicates of experiments were performed. Briefly, RNA was extracted from cells using TRIzol reagent (Invitrogen, Carlsbad, CA) followed by generation of double-stranded cDNA. These were used as templates for synthesis of biotin-labeled cRNA, using the GeneChip IVT labeling kit in accordance with the manufacturer's instructions. The biotinylated cRNA was purified using RNeasy Mini kit (Qiagen, Hilden, Germany) and fragmented before reconstitution in a hybridization cocktail mixture containing eukaryotic hybridization control. The hybridization was performed at 45°C for 16 h in a rotisserie oven set at 60 rpm. Upon completion, the arrays were then loaded onto an Affymetrix Fluidic station, washed according to the standard Affymetrix EukGE-WS2v5 protocol and stained with streptavidin-phycoerythrin

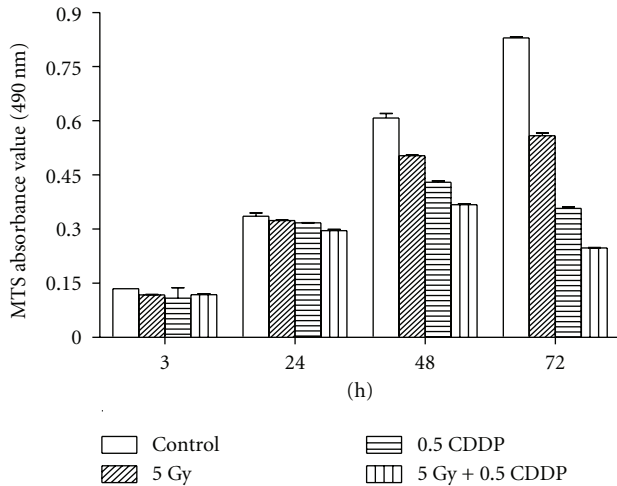


FIGURE 1: Cell viability analysis by MTS assay at different time points (3 h, 24 h, 48 h, and 72 h) after treatment with 5 Gy gamma radiation and 0.5 μ g/ml cisplatin (CDDP). After co-treatment with radiation and CDDP, cell viability was significantly reduced at 72 h. The data shown are the most representative of 3 separate experiments.

(SAPE) solution. After washing and staining, the arrays were scanned with the Gene Array scanner (Affymetrix, Santa Clara, CA). Hybridization intensity data detected by the scanner were automatically acquired and processed by the Affymetrix GeneChip Operating Software (GCOS, Affymetrix, Santa Clara, CA). The average intensity for all the genes was normalized to 100. The statistical algorithms implemented in GCOS software were used for analysis. In a comparison expression analysis, each probe pair on the experimental array was compared to the corresponding probe pair on the baseline array (control). This generated an associated “change” (increased, no change, or decreased) to determine the relative expression of transcripts. To have an overview of gene expression profiles, probe sets showing chemoradiation-induced increased or decreased expressions in both duplicated experiments were retrieved. The differentially expressed genes of chemoradiation treatment were submitted for biological functional analysis using Ingenuity Pathway Analysis (IPA) tools (Ingenuity Systems, <http://www.ingenuity.com>).

3. Results

3.1. Combined CDDP-Radiation Treatment Reduced Cell Viability More than CDDP or Radiation Treatment Alone. Cell viability analysis by MTS assay at different time points revealed that although CDDP and radiation each exerted a negative effect on cell viability, treatment when combined appeared to have a greater effect. These effects were observed at 48 hrs after treatment and became even more marked at 72 hrs after treatment (Figure 1).

3.2. Apoptosis Occurred Predominantly at 72 h after Combined CDDP-Radiation Treatment. At 72 hrs after treatment,

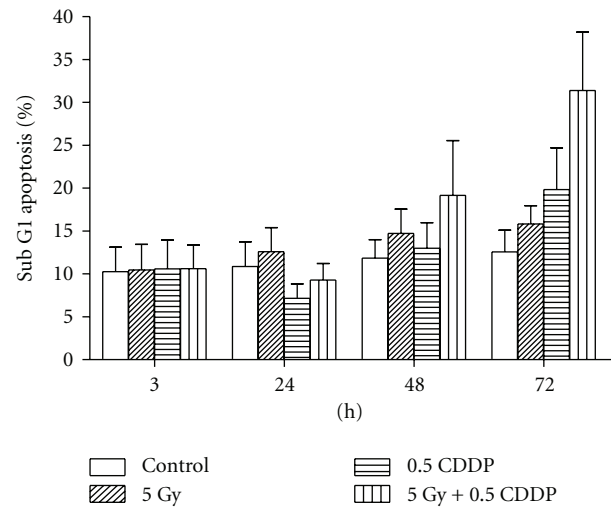


FIGURE 2: Flow cytometric subG1 phase as determined by PI staining at different time points (3 h, 24 h, 48 h and 72 h) after exposure to 5 Gy of gamma radiation and 0.5 μ g/ml of cisplatin (CDDP). Co-treatment with radiation and CDDP resulted in a significant increase in subG1 phase at 72 h. The data shown are the mean + SD of 4 independent experiments.

combined CDDP-radiation led to a greater increase in the sub-G1 phase as compared to using CDDP and radiation alone (Figure 2). As pointed out previously, DNA fragmentation resulting from apoptotic cell death manifests in the sub-G1 fraction.

3.3. Apoptosis-Related Gene Expressions were Enhanced by Combined CDDP-Radiation Treatment. On analyzing the results of molecular and cellular functions under the biological functions of IPA, it was found that among the 3925 probe set IDs which were differentially expressed in at least one treatment, 942 represented 623 unique genes associated with apoptosis (see Table 1). Their distribution at each time point for the different treatment regimes is summarized in Venn diagrams (Figure 3). A subset focusing on the genes, which had a direct upstream or downstream relationship with p53, is shown in Table 2. Combined CDDP-radiation treatment resulted in an increase in the number of gene expressions which was more than merely a summation of the number of expressions resulting from individual treatments (Figure 3, Table 2). At 72 hrs after treatment, 40 out of the 163 genes listed (24.5%) were expressed in combined CDDP-radiation treatment, but not when CDDP or radiation was used alone (Table 2). Among these 40 genes was FAS, an important element of the extrinsic apoptotic pathway.

3.4. Differential Temporal Activation of p53 Occurred with CDDP and Radiation Treatment. It was observed that Post-treatment activation of p53 occurred earlier in radiation than in CDDP (Figure 4). In response to DNA damage, activation of the p53 pathway normally occurs with the phosphorylation of ser-15 in p53. The present study showed radiation-induced phosphorylation of p53 occurred at 3 hrs

TABLE 1: Continued.

Symbol	Probe set ID	Gy	3 h			24 h			72 h		
			CDDP	Gy + CDDP	Gy	CDDP	Gy + CDDP	Gy	CDDP	Gy + CDDP	
C3	1423954.at							I		I	
CACNA1A	1450510.a.at								I		
CACNA1C	1421297.a.at								D	D	
CASP12	1449297.at							D			
CASP2	1448165.at							D			
CASP3	1426165.a.at, 1449839.at			I							
CASP6	1415995.at			I						I	
CASP7	1426062.a.at, 1448659.at							D		D	
CASP9	1426125.a.at	D									
CAST	1426098.a.at, 1435972.at, 1451413.at					I	I	I	I	I	
CAT	1416429.a.at									I	
CAV1	1449145.a.at					D		I	I	I	
CBX5	1421933.at, 1450416.at							D		D	
CCAR1	1436156.at, 1436157.at					I	I				
CCL13	1420380.at							I	I	I	
CCL5	1418126.at				I			I		I	
CCL9	1417936.at, 1448898.at								I	I	
CCNA2	1417910.at, 1417911.at								I	I	
CCNB1	1416076.at, 1419943.s.at, 1448205.at, 1449675.at			D				I	I	I	
CCND1	1417419.at, 1417420.at, 1448698.at	D						I	I	I	
CCND3	1415907.at								I	I	
CCNG1	1420827.a.at, 1450016.at, 1450017.at	I		I	I	I	I	I	I	I	
CD14	1417268.at									I	
CD24	1416034.at, 1437502.x.at, 1448182.a.at			D						I	
CD274	1419714.at							I	D		
CD2AP	1420907.at					I					
CD44	1423760.at, 1434376.at, 1452483.a.at								I	I	
CD47	1419554.at, 1428187.at, 1449507.a.at							D		D	

TABLE 1: Continued.

Symbol	Probe set ID	3 h			24 h			72 h		
		Gy	CDDP	Gy + CDDP	Gy	CDDP	Gy + CDDP	Gy	CDDP	Gy + CDDP
CD80	1432826_a.at									I
CD9	1416066_at			D				I	I	I
CDC20	1416664_at, 1439377_x.at							I	I	I
CDC25B	1421963_a.at								I	I
CDC25C	1422252_a.at, 1456077_x.at								I	I
CDC2L2	1418841_s.at									D
CDC37	1416819_at								I	I
CDC42EP3	1422642_at, 1450700_at							I	I	I
CDC45L	1416575_at							D		D
CDC6	1417019_a.at									D
CDCA2	1437251_at, 1455983_at								I	I
CDH2	1418815_at									I
CDK4	1422439_a.at, 1422440_at, 1422441_x.at							D		D
CDK8	1460389_at									I
CDKN1A	1421679_a.at, 1424638_at	I		I	I	I	I	I	I	I
CDKN1B	1434045_at								D	D
CDKN2A	1450140_a.at							D		
CDKN2C	1416868_at							D		
CEBPB	1427844_a.at			D		D				
CEBPD	1423233_at					D		I	I	I
CENPF	1427161_at									I
CFLAR	1424996_at									I
CHEK1	1439208_at									D
CKAP2	1434748_at					I	I		I	I
CLCF1	1437270_a.at, 1450262_at			D						I
CLU	1418626_a.at, 1437458_x.at, 1437689_x.at, 1454849_x.at							I	I	I
CNN2	1450981_at								I	I
CNP	1418980_a.at, 1437341_x.at								I	I
CNTF	1426327_s.at									D
COPS5	1460171_at									D
CR1	1422563_at									D
CREB3L1	1419295_at								I	I
CRK	1416201_at, 1448248_at			D				I	I	I

TABLE 1: Continued.

Symbol	Probe set ID	Gy	3 h		24 h			72 h		
			CDDP	Gy + CDDP	Gy	CDDP	Gy + CDDP	Gy	CDDP	Gy + CDDP
CROP	1424802.a.at, 1451485.at			I		I				D
CRYAB	1416455.a.at, 1434369.a.at			I					I	I
CSF1	1425154.a.at, 1425155.x.at, 1448914.a.at, 1460220.a.at			D					I	I
CSF2	1427429.at									I
CSNK2A1	1419034.at, 1419035.s.at, 1419036.at, 1419038.a.at								D	D
CST3	1426195.a.at							I		
CTCF	1418330.at, 1449042.at	D								D
CTGF	1416953.at							I	I	I
CTNNA1	1437807.x.at, 1448149.at									I
CTSB	1417490.at, 1417491.at, 1417492.at								I	I
CTSD	1448118.a.at								I	I
CTTN	1421313.s.at, 1421315.s.at, 1423917.a.at, 1433908.a.at								I	I
CUL3	1434717.at							D		
CUL5	1428287.at					I				
CX3CL1	1415803.at									I
CXCL12	1417574.at, 1448823.at					D	D			
CXCL2	1419209.at, 1441855.x.at, 1457644.s.at					D		I	I	I
CXCR7	1417625.s.at					D	D		I	I
CYB5A	1416727.a.at									I
CYB5R3	1422185.a.at, 1422186.s.at, 1425329.a.at								I	I
CYBA	1454268.a.at								I	
CYLD	1429617.at									I
CYR61	1416039.x.at, 1438133.a.at, 1442340.x.at, 1457823.at							I	I	I
DAB2	1420498.a.at, 1423805.at, 1429693.at								I	I
DAP	1423790.at, 1451112.s.at					D	D	I	I	I
DAXX	1419026.at							D	I	

TABLE 1: Continued.

Symbol	Probe set ID	Gy	3 h		24 h			72 h		
			CDDP	Gy + CDDP	Gy	CDDP	Gy + CDDP	Gy	CDDP	Gy + CDDP
DCN	1449368_at									I
DDIT3	1417516_at									D
DDIT4	1428306_at							I	I	I
DDR1	1415797_at, 1415798_at, 1456226_x_at			D					I	I
DDX5	1419653_a_at	I								
DDX58	1436562_at, 1456890_at							D	I	D
DHCR24	1451895_a_at								I	I
DKK3	1417312_at, 1448669_at				D	D		I	I	I
DLC1	1436173_at, 1460602_at									I
DLX2	1448877_at				D					
DNAJC15	1416910_at									D
DNM1L	1428086_at, 1452638_s_at					I				
DTYMK	1438096_a_at			I						
DUSP14	1431422_a_at								I	I
DUSP22	1448985_at								I	
DUSP4	1428834_at			D						
DUSP6	1415834_at									I
DUT	1419270_a_at									D
E2F1	1417878_at							D		
ECOP	1451127_at							D		
EDA2R	1440085_at	I		I				I	I	I
EEF1D	1439439_x_at, 1449506_a_at								D	D
EGR1	1417065_at							I	I	I
EHD4	1449852_a_at								I	
EIF2AK2	1422006_at, 1440866_at				I	I	I			
EIF4E	1450908_at									D
EIF5A	1437859_x_at									D
ELAVL1	1452858_at									D
EMILIN2	1435264_at								I	I
EMP1	1416529_at							I	I	I
EMP3	1417104_at							I	I	I
ENO1	1419022_a_at, 1419023_x_at							I		
EPHA2	1421151_a_at									I
EPHX1	1422438_at							I	I	I
ERCC3	1448497_at									I
ERCC5	1450935_at									I
ESPL1	1433862_at									I
ETS1	1422027_a_at, 1426725_s_at, 1452163_at			D						I

TABLE 1: Continued.

Symbol	Probe set ID	Gy	3 h		24 h			72 h		
			CDDP	Gy + CDDP	Gy	CDDP	Gy + CDDP	Gy	CDDP	Gy + CDDP
GSPT1	1426736_at, 1452168_x_at									D
GSTM1	1416411_at								I	
GSTM5	1448330_at						D			
HBEGF	1418349_at			D						
HELLS	1417541_at					I				
HIP1	1434557_at			D						
HIPK1	1424540_at			D						
HIST1H1C	1416101_a_at, 1436994_a_at					D	D	I	I	
HK1	1420901_a_at								I	I
HK2	1422612_at									I
HMGA1	1416184_s_at							I	I	I
HMGA2	1422851_at, 1450780_s_at, 1450781_at							I	I	I
HMGB1L1	1425048_a_at, 1435324_x_at, 1439463_x_at, 1448235_s_at								D	D
HMGN1	1455897_x_at								D	
HMMR	1425815_a_at, 1427541_x_at, 1450156_a_at, 1450157_a_at					I			I	I
HMOX1	1448239_at									D
HNRNPA1	1423531_a_at, 1430019_a_at, 1430020_x_at							D	D	D
HOXA7	1449499_at									D
HSH2D	1442130_at								I	I
HSP90AA1	1426645_at, 1437497_a_at, 1438902_a_at			I		I	I			
HSP90AB1	1416364_at, 1416365_at									I
HSPA1B	1427127_x_at						D			
HSPA5	1416064_a_at, 1427464_s_at, 1447824_x_at								D	D
HSPB1	1422943_a_at, 1425964_x_at			D				I	I	I
HSPB8	1417014_at									D
HTATIP2	1451814_a_at									I
HUWE1	1415703_at									D
ID1	1425895_a_at									I
ID2	1422537_a_at						D			
IER3	1419647_a_at							I	I	I
IFI16	1419603_at, 1452349_x_at					I		D		

TABLE 1: Continued.

Symbol	Probe set ID	Gy	3 h			24 h			72 h		
			CDDP	Gy + CDDP	Gy	CDDP	Gy + CDDP	Gy	CDDP	Gy + CDDP	
KLF6	1418280_at, 1427742_a_at, 1447448_s_at	D	D	D	I						
LAMP2	1416344_at								I	I	
LCN2	1427747_a_at							I		I	
LDLR	1421821_at						D		I	I	
LGALS3	1426808_at									I	
LGALS3BP	1448380_at								I	I	
LGALS8	1422662_at								I		
LIF	1421207_at									I	
LIMS1	1418232_s_at			D							
LMNA	1421654_a_at, 1425472_a_at, 1457670_s_at			D		D	D	I	I	I	
LPAR1	1426110_a_at, 1448606_at					D	D			I	
LRIG1	1434210_s_at, 1449893_a_at								I	I	
LTBR	1416435_at			D							
MAOA	1428667_at							I	I	I	
MAP2K3	1451714_a_at									I	
MAP3K12	1438908_at	I									
MAP3K4	1459800_s_at								I	I	
MAP3K7	1419988_at									I	
MAPK3	1427060_at			D							
MAPK8	1420932_at									D	
MAPKAP1	1417284_at								I	I	
MAX	1423501_at							D	D	D	
MCF2L	1434140_at			D							
MCL1	1416880_at			D							
MCM2	1448777_at, 1423605_a_at, 1427718_a_at								D	D	
MDM2	1427718_a_at	I		I		I	I		I	I	
MED1	1450402_at			I							
MEF2A	1427186_a_at, 1452347_at				I						
MET	1422990_at, 1434447_at								I	I	
MFGE8	1420911_a_at									I	
MGP	1448416_at					D	D	I		I	
MGST1	1415897_a_at							I	I	I	
MMP2	1416136_at									I	
MMP3	1418945_at							I		I	
MPG	1417571_at, 1417572_at								I	I	
MT1E	1428942_at					D		I	I	I	

TABLE 1: Continued.

Symbol	Probe set ID	Gy	3 h		24 h			72 h		
			CDDP	Gy + CDDP	Gy	CDDP	Gy + CDDP	Gy	CDDP	Gy + CDDP
MT1F	1422557_s.at						D	I		
MTMR6	1425485.at									I
MTPN	1437457_a.at				I					
MX1	1451905_a.at							D	I	
MYC	1424942_a.at			D						
MYO6	1433942.at									I
NAMPT	1417190.at							D		D
NCAM1	1426864_a.at									I
NCAPG2	1417926.at					I				
NDRG1	1420760_s.at,									
	1423413.at,								D	D
	1450976.at,									D
	1456174_x.at									
NDST1	1422044.at,			D					D	
	1460436.at									
NDUFAF4	1427997.at			I						
NDUFV2	1428179.at,									I
	1452692_a.at									
NEDD9	1422818.at									I
NEK2	1417299.at,								I	I
	1437580_s.at									
NEK6	1423596.at,								I	I
	1425850_a.at									
NFAT5	1438999_a.at,			D			I		I	I
	1439805.at									
NFIL3	1418932.at			D						
NFKB1	1427705_a.at									I
NFKB2	1425902_a.at						I			
	1420088.at,									
	1438157_s.at,								I	I
	1448306.at,									
NFKBIA	1449731_s.at									
	1417483.at,								I	I
NFKBIZ	1448728_a.at,								I	I
	1457404.at									
NGF	1419675.at			D						
NME1	1424110_a.at									D
NOD1	1454733.at								I	I
NOTCH2	1455556.at						D			
NP	1416530_a.at,									I
	1453299_a.at									
NQO1	1423627.at								I	I
NQO2	1449983_a.at,									I
	1455590.at									
NR2F1	1418157.at						D			
NR3C1	1421867.at,									I
	1457635_s.at,									
NR4A1	1416505.at			D						I

TABLE 1: Continued.

Symbol	Probe set ID	3 h			24 h			72 h		
		Gy	CDDP	Gy + CDDP	Gy	CDDP	Gy + CDDP	Gy	CDDP	Gy + CDDP
PLD1	1437113_s.at								I	I
PLD2	1417237.at								I	
PLEKHF1	1424671.at								I	I
PLK1	1448191.at			D				I	I	I
PLK3	1434496.at								I	I
PLSCR1	1429527_a.at, 1453181_x.at								I	I
PLSCR3	1449020.at								I	
PMEPA1	1422706.at, 1452295.at			D	D	D				
PML	1448757.at, 1456103.at							D	I	
PNKP	1416378.at								I	I
PNPT1	1452676_a.at							D		
POLK	1449483.at								I	I
PPID	1417057_a.at								D	D
PPM1A	1429501_s.at, 1451943_a.at									D
PPM1F	1454934.at									I
PPP1R13L	1459592_a.at			D						
PPP1R15A	1448325.at								D	D
PPP2R2A	1437730.at, 1453260_a.at							D		D
PRDX5	1416381_a.at									I
PRKAR2B	1438664.at, 1456475_s.at									I
PRKCA	1450945.at									I
PRKD1	1447623_s.at									I
PRMT2	1416844.at								I	
PRPF19	1460633.at						D			
PRR13	1423686_a.at						I		I	I
PSENEN	1415679.at									D
PSIP1	1417166.at, 1460403.at				I	I	I			
PSMG2	1425373_a.at, 1448212.at									D
PTGR1	1417777.at							I	I	I
PTGS1	1436448_a.at									I
PTGS2	1417262.at, 1417263.at							I	I	I
PTMA	1423455.at								D	D
PTPN1	1438670.at			D						
PTPRA	1425340_a.at								I	I
PTPRE	1418540_a.at								I	
PTPRG	1434360_s.at					D	D			
PTRH2	1451845_a.at							D	D	D

TABLE 1: Continued.

Symbol	Probe set ID	Gy	3 h		24 h			72 h		
			CDDP	Gy + CDDP	Gy	CDDP	Gy + CDDP	Gy	CDDP	Gy + CDDP
PTTG1	1419620_at, 1424105_a_at, 1438390_s_at							I	I	I
PXN	1424027_at, 1456135_s_at								I	I
QARS	1423712_a_at, 1456726_x_at								I	I
QKI	1417073_a_at, 1425597_a_at, 1429318_a_at, 1451179_a_at						D	D	D	D
RABGGTB	1419553_a_at									I
RAD18	1451928_a_at					I				
RAD21	1416162_at			D						
RAD54L	1450862_at					I			I	I
RALB	1417744_a_at									I
RARG	1419415_a_at, 1419416_a_at			D						
RASA1	1426476_at, 1426477_at									I
RASSF1	1441737_s_at, 1448855_at						I			
RASSF5	1422637_at									I
RB1	1417850_at					I				
RBBP4	1434892_x_at, 1454791_a_at, 1454875_a_at								D	D
RBBP6	1425114_at									D
RBL1	1424156_at, 1425166_at								D	D
RBP1	1448754_at							I	I	I
RCAN2	1421425_a_at									I
RECK	1450784_at								I	
RFC1	1418342_at, 1449050_at, 1451920_a_at				I	I	I			
RFK	1415737_at, 1416230_at								D	D
RFWD2	1426913_at								I	
RGS3	1425296_a_at, 1425701_a_at								I	I
RIPK1	1419508_at, 1449485_at								I	I
RIPK2	1450173_at									I
RNF34	1415791_at			I						
ROCK1	1423444_at, 1423445_at							I		I
RPS3	1435151_a_at			I						
RPS3A	1422475_a_at			I				I		
RPS6KB1	1454956_at					I				

TABLE 1: Continued.

Symbol	Probe set ID	Gy	3 h			24 h			72 h		
			CDDP	Gy + CDDP	Gy	CDDP	Gy + CDDP	Gy	CDDP	Gy + CDDP	
SLC25A24	1427483.at, 1452717.at								I	I	
SLC2A1	1426599.a.at, 1434773.a.at				D					I	
SLC7A11	1420413.at								I		
SLK	1425977.a.at, 1449336.a.at					I					
SMN1	1426596.a.at								D	D	
SMNDC1	1429043.at								D		
SNRPE	1451294.s.at								D	D	
SOCS3	1416576.at, 1455899.x.at, 1456212.x.at			D					I	I	
SOD2	1417193.at, 1448610.a.at									I	
SOD3	1417633.at								I	I	
SORBS2	1437197.at								I	I	
SOX4	1419155.a.at, 1419156.at, 1419157.at, 1433575.at, 1449370.at					D	D	D	D	D	
SP1	1418180.at, 1454852.at								D	D	
SPP1	1449254.at					D	D	I	I	I	
SRGN	1417426.at								I	I	
STAT1	1420915.at, 1450033.a.at, 1450034.at								D	D	
STAT5A	1421469.a.at, 1450259.a.at								I	I	
STAT6	1426353.at								I	I	
STK24	1426248.at							D			
STMN1	1415849.s.at, 1448113.at								D	D	
STX8	1418089.at								I	I	
SULF1	1436319.at, 1438200.at									I	
TACC3	1417450.a.at, 1436872.at, 1455834.x.at								I	I	
TADA3L	1417467.a.at									I	
TAX1BP1	1420174.s.at, 1448399.at							I		I	
TCF12	1427670.a.at									D	
TCF4	1416724.x.at				I						
TCF7	1433471.at									I	
TENC1	1452264.at			D						I	

TABLE 1: Continued.

Symbol	Probe set ID	Gy	3 h		24 h			72 h		
			CDDP	Gy + CDDP	Gy	CDDP	Gy + CDDP	Gy	CDDP	Gy + CDDP
WISP1	1448593_at, 1448594_at								I	I
WRN	1425982_a.at								I	
WTAP	1454805_at							D		D
WWOX	1416334_at				D	D				
XAF1	1443698_at						I			
XBP1	1420886_a.at, 1437223_s.at				D					
XDH	1451006_at								I	
XPA	1460725_at									I
XRCC2	1455335_at									D
XRCC4	1424601_at								D	D
XRCC6	1417437_at								D	D
YARS	1460638_at									D
YWHAE	1435702_s.at, 1438839_a.at									D
YY1	1435824_at, 1457834_at			D					I	I
ZFP36	1452519_a.at			D					I	
ZFP36L2	1437626_at				D	D				I
ZMAT3	1449353_at			I					I	I
ZNF148	1418381_at, 1449068_at, 1449069_at				I	I				
ZNF622	1438000_x.at								D	D
ZYX	1417240_at							I	I	I

after treatment, compared to CDDP-induced activation which was observed only at 24 hrs or later (Figure 4). These timings corresponded with those observed for the expression of apoptotic-related genes after radiation and CDDP treatment (Figure 3). For example, MDM2 and TP53INP1 were expressed at 3 hrs after radiation. They were however, expressed only at 24 hrs after CDDP (Table 2).

4. Discussion

Combined chemoradiation is increasingly being used to treat advanced head and neck cancers. As radiation and CDDP are both ototoxic, it is of concern that significant sensorineural hearing loss will result. Indeed, patients with nasopharyngeal carcinoma who had received radiotherapy and concurrent/adjuvant chemotherapy using CDDP were found to experience greater sensorineural hearing loss compared with patients treated with radiotherapy alone, especially to high-frequency sounds in the speech range [1]. It is of interest to note that different etiologies of sensorineural hearing loss, such as noise, ototoxic drugs, and aging, result in similar patterns of audiometric changes and cochlear cellular degeneration [8]. The cellular and molecular mechanisms involved in sensorineural hearing loss from diverse causes

appear to lead to a final common pathway which results in apoptosis of cochlear hair cells [6, 9].

In radiation-induced ototoxicity, cochlear cell apoptosis and ROS generation were observed after irradiation, and p53 was thought to play a key role [7]. This phenomenon was dose dependant and occurred predominantly at 72 h after irradiation. Microarray analysis supported these findings, as associated dose-dependant apoptotic gene regulation changes were observed.

The ototoxic manifestations of CDDP are primarily due to its effects on the cochlear hair cells although the spiral ganglion cells and the stria vascularis are also affected to some extent. According to Rybak et al. [10], CDDP ototoxicity appears to be triggered by ROSs that initiate a cascade of molecular events that lead to apoptosis of outer hair cells, resulting in hearing loss. Ototoxic effects on the stria vascularis are transient, resulting in temporary reduction of endocochlear potential associated with stria edema. The endocochlear potential recovers but residual shrinkage of the stria persists. The spiral ganglia are thought to be least affected.

Although the cellular and molecular processes of ototoxicity have been described for radiation and CDDP when used alone, those involved in combined therapy have not been

TABLE 2: Differential expression of apoptosis-related genes which have direct upstream or downstream relationship with p53 in each treatment group [irradiation (Gy), cisplatin (CDDP), or combination of both (Gy + CDDP)] when compared to nontreated control cells at 3 h, 24 h, and 72 h after treatment.

3 hours							
Symbol	Gy	CDDP	Gy+CDDP	Symbol	Gy	CDDP	Gy+CDDP
CCNG1	I		I	KLF6	D	D	D
CDKN1A	I		I	CCND1	D		
MDM2	I		I	CTCF	D		
TP53INP1	I		I	IRS1	D		
BTG2	I			ATF3			D
DDX5	I			AURKA			D
GDF15	I			BID			D
IL6	I			CCNB1			D
C11ORF82			I	CEBPB			D
CASP3			I	DDR1			D
CASP6			I	ETS1			D
CRYAB			I	FHL2			D
HSP90AA1			I	HBEGF			D
MED1			I	HIPK1			D
ZMAT3			I	HSPB1			D
				MAPK3			D
				MCL1			D
				MYC			D
				NR4A1			D
				OSGIN1			D
				PLK1			D
				PMEPA1			D
				PPP1R13L			D
				THBS1			D
				THBS2			D
				YY1			D
24 hours							
Symbol	Gy	CDDP	Gy+CDDP	Symbol	Gy	CDDP	Gy+CDDP
CCNG1	I	I	I	APEX1		D	D
CDKN1A	I	I	I	BHLHE40		D	D
EIF2AK2	I	I	I	BRE		D	D
RFC1	I	I	I	PMEPA1		D	D
BUB1		I	I	S100A4		D	D
CCAR1		I	I	SERPINE1		D	D
CKAP2		I	I	SPP1		D	D
HSP90AA1		I	I	THBS2		D	D
MDM2		I	I	WWOX		D	D
TOP2A		I	I	SLC2A1	D		
TP53INP1		I	I	BTG1		D	
ZNF148		I	I	CAV1		D	
KLF6	I			CEBPB		D	
BLM		I		TIMP3		D	

TABLE 2: Continued.

24 hours							
Symbol	Gy	CDDP	Gy+CDDP	Symbol	Gy	CDDP	Gy+CDDP
BRCA1		I		ATP1A1			D
HMMR		I		CDK4			D
IFI16		I		CDKN2A			D
RAD54L		I		CDKN2C			D
RB1		I		GSTM5			D
TOP1		I		ID2			D
TOPBP1		I		TFAP2A			D
TTK		I					
BTG2			I				
C11ORF82			I				
FUBP1			I				
NFKB2			I				
NUPR1			I				
TOPORS			I				
72 hours							
Symbol	Gy	CDDP	Gy+CDDP	Symbol	Gy	CDDP	Gy+CDDP
ANXA1	I	I	I	LGALS3			I
AURKA	I	I	I	LIF			I
BUB1B	I	I	I	MAP2K3			I
CAV1	I	I	I	MMP2			I
CCNB1	I	I	I	MYO6			I
CCND1	I	I	I	NFKB1			I
CCNG1	I	I	I	PLAUR			I
CDC20	I	I	I	PRKCA			I
CDKN1A	I	I	I	PTGS1			I
CLU	I	I	I	SAT1			I
DDIT4	I	I	I	SERPINE2			I
EGR1	I	I	I	SLC2A1			I
FOSL1	I	I	I	SOD2			I
GLIPR1	I	I	I	TADA3L			I
HSPB1	I	I	I	THBS2			I
IER3	I	I	I	TOPBP1			I
INHBA	I	I	I	TSC2			I
IRS1	I	I	I	VDR			I
NFKBIA	I	I	I	FEN1	D	D	D
PHLDA1	I	I	I	GADD45A	D	D	D
PLK1	I	I	I	JUN	D	D	D
PTGS2	I	I	I	MCM2	D	D	D
PTTG1	I	I	I	NDRG1	D	D	D
S100A4	I	I	I	STAT1	D	D	D
SERPINE1	I	I	I	ATF3	D		D
SGK1	I	I	I	PPP2R2A	D		D
SPP1	I	I	I	TP53BP2	D		D
THBS1	I	I	I	CDK4		D	D
TMSB4X	I	I	I	FUBP1		D	D
UBE2C	I	I	I	GSK3B		D	D
ZYX	I	I	I	HMGB1L1		D	D
BCL3	I		I	HSPA5		D	D
MMP3	I		I	PARK7		D	D
S100A6	I		I	PCNA		D	D

TABLE 2: Continued.

72 hours							
Symbol	Gy	CDDP	Gy+CDDP	Symbol	Gy	CDDP	Gy+CDDP
ABCB1B		I	I	PPP1R15A		D	D
AKAP12		I	I	SMN1		D	D
ATM		I	I	SP1		D	D
BID		I	I	STMN1		D	D
BIRC5		I	I	XRCC6		D	D
BUB1		I	I	DAXX	D	I	
CCNA2		I	I	MX1	D	I	
CCND3		I	I	PML	D	I	
CDC25C		I	I	E2F1	D		
CKAP2		I	I	IFI16	D		
CRYAB		I	I	BRE			D
CTSD		I	I	CDC6			D
DDR1		I	I	CHEK1			D
DHCR24		I	I	COP55			D
EZR		I	I	CTCF			D
FHL2		I	I	DDIT3			D
FOS		I	I	DUT			D
FOXM1		I	I	ELAVL1			D
HMMR		I	I	HOXA7			D
IL6		I	I	HUWE1			D
KAT2B		I	I	KAT5			D
KLF4		I	I	MAPK8			D
MDM2		I	I	NME1			D
MET		I	I	RBBP6			D
NEK2		I	I	TFAP2A			D
NQO1		I	I	TP53			D
NQO2		I	I	VCAN			D
NR3C1		I	I				
PLK3		I	I				
PTPRA		I	I				
RAD54L		I	I				
S100A1		I	I				
SHISA5		I	I				
TACC3		I	I				
TGFB2		I	I				
TIMP3		I	I				
TP53INP1		I	I				
TTK		I	I				
YY1		I	I				
ZMAT3		I	I				
TXN	I						
GDF15		I					
GSTM1		I					
RFWD2		I					
RRM2B		I					
WRN		I					
AFP			I				
AHR			I				
AP2A2			I				
BHLHE40			I				
C11ORF82			I				

TABLE 2: Continued.

72 hours							
Symbol	Gy	CDDP	Gy+CDDP	Symbol	Gy	CDDP	Gy+CDDP
CASP6			I				
CAT			I				
CDK8			I				
CENPF			I				
CFLAR			I				
CSF2			I				
CX3CL1			I				
EPHA2			I				
ERCC3			I				
ERCC5			I				
ETS1			I				
FAS			I				
FASN			I				
GPI			I				
HK2			I				
HSP90AB1			I				
ID1			I				

Differentially expressed apoptotic related genes

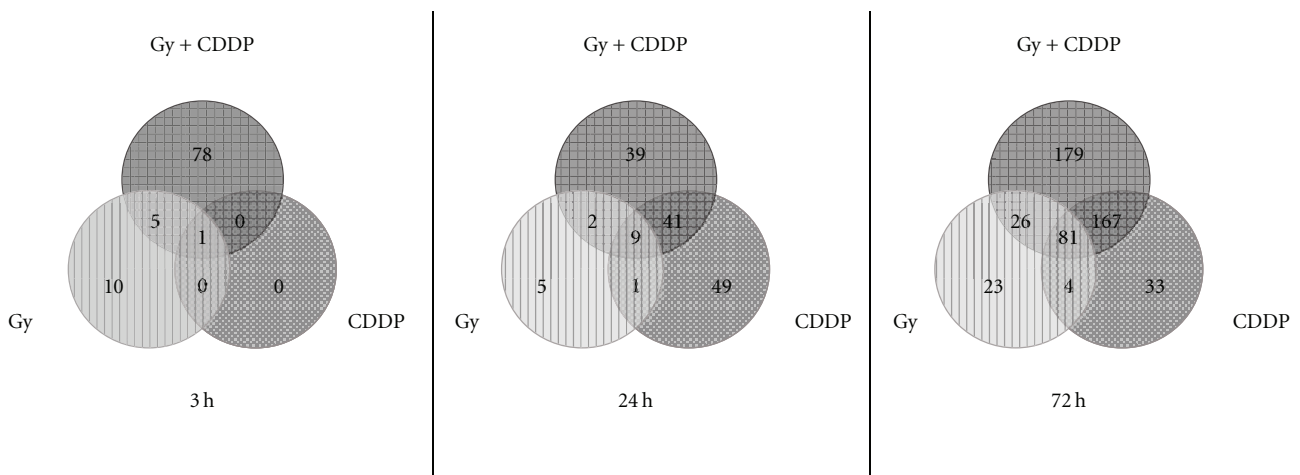


FIGURE 3: Microarray findings are summarized by Venn diagrams which show the distribution of differentially expressed probeset IDs in each treatment group [irradiation (Gy), cisplatin (CDDP) or combination of both (Gy and CDDP)] when compared to nontreated control cells at 3 h, 24 h, and 72 h after treatment.

studied previously. The present study demonstrated that combined therapy led to decreased viability of cochlear cells, with an increase in the subG1 population. These findings support the belief that as in other etiologies of sensorineural loss, apoptosis of cochlear hair cells is important in CDDP-radiation.

It is well established that p53 plays a key role in the cellular response to nuclear DNA damage [11]. It regulates cell cycle arrest and dictates cell fate like senescence, apoptosis, and DNA repair. It is believed that the nature of DNA damage

enables p53 to selectively discriminate between promoters in the induction of target genes, thereby regulating their expression and subsequent cellular outcome [12].

In a study on HEI-OC1 cells derived from the cochlea, CDDP caused an increase in p53 at 3 hrs prior to the activation of Bax, cytochrome-c, and caspase 8 and 9 [13]. In the case of radiation-induced ototoxicity, the role of p53 in triggering apoptotic cell death in cochlear hair cells has also been studied [7]. Based on microarray analysis, the p53 gene was found to be up-regulated after irradiation and p53

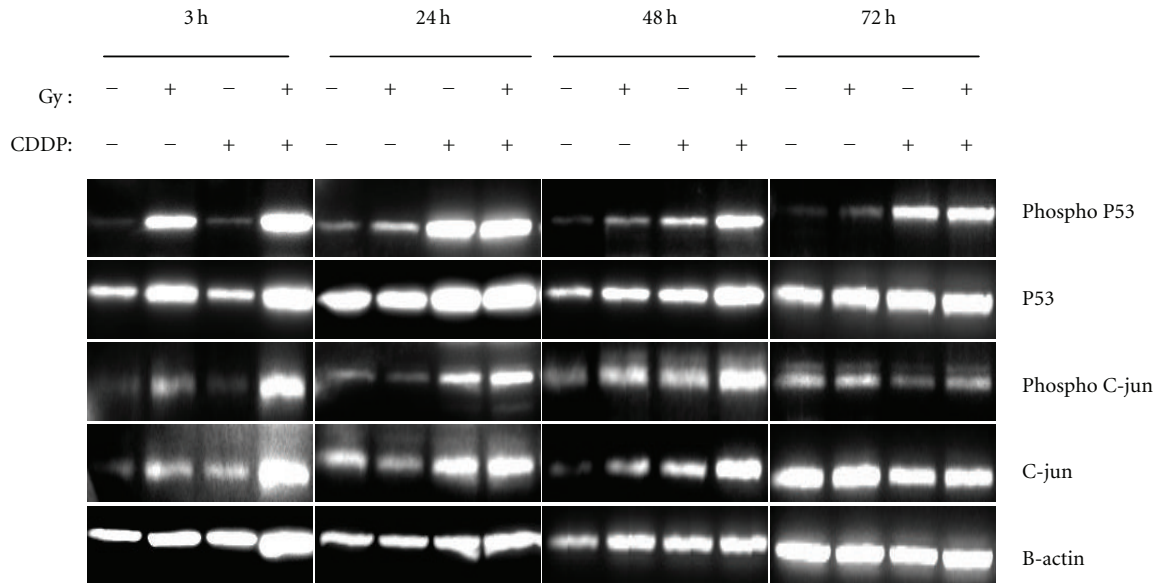


FIGURE 4: Western blot analysis showing p53 and c-jun protein expression and phosphorylation at various time points (3 h, 24 h, 48 h, and 72 h) after 5 Gy of gamma radiation and 0.5 $\mu\text{g/ml}$ pf cisplatin (CDDP). The data are representative of 3 separate experiments.

expression was confirmed by Western blotting. Although p53 plays a role in both CDDP and radiation-induced ototoxicity, the present study showed that p53 was activated at different time points after treatment. Posttreatment phosphorylation of p53 occurred after 24 hrs for CDDP, whereas it occurred as early as 3 hrs for radiation. These timings corresponded to the times MDM2 and TP53INP1 were expressed after treatment with CDDP and radiation respectively. Therefore, although both CDDP and radiation-induced cochlear cell apoptosis appear to involve activation of p53, the upstream processes involved may well be different.

In the present study, combined CDDP-radiation treatment triggered more apoptotic-related gene expressions than those that could be accounted for by a summation of gene expressions resulting from individual treatments. This could explain the synergistic ototoxic effects of combined CDDP-radiation treatment, an observation seen clinically [1]. Interestingly, among the genes which were expressed in combined treatment but not when these entities were used alone was FAS, a key element involved in the extrinsic apoptotic pathway. Although the extrinsic apoptotic pathway has generally been regarded to play only minor role in ototoxicity resulting from the use of CDDP or radiation alone, it may well be important in situations when they are used in combination [14, 15].

The OC-k3 cell line expressed the neuroepithelial precursor cell marker nestin and the inner ear cell marker OCP2, specific auditory sensory cell markers myosin VIIa and the acetylcholine receptor alpha-9 and the supporting cell marker connexin 26. It had been regarded as a good model to study the mechanisms of cell fate in the Organ of Corti of the cochlea [4]. Therefore, the finding that combined treatment actually led to enhanced apoptotic gene expressions including FAS should be further investigated in

in vivo animal studies which may have implications in future antiapoptotic treatments against ototoxicity.

5. Conclusion

Like in other etiologies of sensorineural loss, apoptosis of cochlear hair cells appears to play a role in ototoxicity resulting from combined CDDP-radiation therapy. Differential temporal activation of p53 suggests the possibility of different upstream processes leading to its activation after CDDP and radiation treatment. Enhanced apoptotic gene expressions including that of FAS were observed in combined treatment which could possibly explain the synergistic ototoxic effects seen clinically.

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