

Noninvasive Brain Stimulation to Enhance Functional Recovery After Stroke: Studies in Animal Models

Neurorehabilitation and Neural Repair 2018, Vol. 32(11) 927–940 © The Author(s) 2018

Article reuse guidelines: sagepub.com/journals-permissions DOI: 10.1177/1545968318804425 journals.sagepub.com/home/nnr

(\$)SAGE

Julia Boonzaier, MSc¹, Geralda A. F. van Tilborg, PhD¹, Sebastiaan F. W. Neggers, PhD², and Rick M. Dijkhuizen, PhD¹

Abstract

Background. Stroke is the leading cause of adult disability, but treatment options remain limited, leaving most patients with incomplete recovery. Patient and animal studies have shown potential of noninvasive brain stimulation (NIBS) strategies to improve function after stroke. However, mechanisms underlying therapeutic effects of NIBS are unclear and there is no consensus on which NIBS protocols are most effective. Objective. Provide a review of articles that assessed effects and mechanisms of repetitive transcranial magnetic stimulation (rTMS) and transcranial direct current stimulation (tDCS) in animal stroke models. Methods. Articles were searched in PubMed, including cross-references. Results. Nineteen eligible studies reporting effects of rTMS or tDCS after stroke in small rodents were identified. Seventeen of those described improved functional recovery or neuroprotection compared with untreated control or sham-stimulated groups. The effects of rTMS could be related to molecular mechanisms associated with ischemic tolerance, neuroprotection, antiapoptosis, neurogenesis, angiogenesis, or neuroplasticity. Favorable outcome appeared most effectively when using high-frequency (>5 Hz) rTMS or intermittent theta burst stimulation of the ipsilesional hemisphere. tDCS effects were strongly dependent on stimulation polarity and onset time. Although these findings are promising, most studies did not meet Good Laboratory Practice assessment criteria. Conclusions. Despite limited data availability, animal stroke model studies demonstrate potential of NIBS to promote stroke recovery through different working mechanisms. Future studies in animal stroke models should adhere to Good Laboratory Practice guidelines and aim to further develop clinically applicable treatment protocols by identifying most favorable stimulation parameters, treatment onset, adjuvant therapies, and underlying modes of action.

Keywords

cerebrovascular stroke, models, animal, transcranial magnetic stimulation, transcranial direct current stimulation

Introduction

Globally, stroke is a devastating neurological disorder and a leading cause of death and acquired disability.¹ The majority of stroke patients experience motor impairment, which affects movement of the face, leg, and/or arm on one side of the body.² Upper limb motor deficiencies are often persistent and disabling, affecting independent functional activities of daily living.³ Unfortunately, most stroke patients recover incompletely after stroke, despite intensive rehabilitation strategies.^{3,4} Although there is a diverse range of interventions (for overview, see review by Pollock and colleagues⁴) aimed at improving motor outcome after stoke, there is still a pressing need for novel treatment therapies and continued research to reduce disability and improve functional recovery after stroke.

Noninvasive brain stimulation (NIBS) techniques, such as repetitive transcranial magnetic stimulation (rTMS) and transcranial direct current stimulation (tDCS), have shown promising therapeutic potential in stroke patient studies.^{5,6} The rationale behind rTMS or tDCS therapy is to modulate cortical excitability, increase neural plasticity, and improve functional motor outcome. For many studies, this approach has been based on the interhemispheric competition model.⁷ The interhemispheric competition model suggests that functional recovery in stroke patients is hindered due to reduced output from the affected hemisphere and excessive transcallosal inhibition

¹Center for Image Sciences, University Medical Center Utrecht and Utrecht University, Utrecht, Netherlands

²Brain Center Rudolf Magnus, University Medical Center Utrecht and Utrecht University, Utrecht, Netherlands

Corresponding Author:

Rick M. Dijkhuizen, Biomedical Magnetic Resonance Imaging and Spectroscopy Group, Center for Image Sciences, University Medical Center Utrecht, Yalelaan 2, Utrecht 3584 CM, Netherlands. Email: r.m.dijkhuizen@umcutrecht.nl

from the unaffected hemisphere.8 Therefore, improvement in motor deficits may be obtained with NIBS strategies that facilitate excitability in the affected hemisphere or suppress inhibitory activity from the unaffected hemisphere.^{9,10} Depending on the type and duration of the stimulation protocol, both rTMS and tDCS can be used to increase (>5 Hz rTMS; intermittent theta burst stimulation; anodal tDCS) or decrease (≤1 Hz rTMS; continuous theta burst stimulation; cathodal tDCS) cortical excitability, with potentially lasting effects beyond the stimulation period, promoting mechanisms of synaptic plasticity.¹¹ Evidence suggests that rTMS and tDCS techniques are able to induce changes in cortical excitability associated with facilitation or long-term potentiation like plasticity via glutamatergic neurotransmission, or inhibition and long-term GABAergic neurotransmission.12,13 depression via Furthermore, effects of rTMS and tDCS are not restricted to the target region of stimulation, but also affect distantly connected cortical areas, allowing for the modulation of largescale neural networks.14

However, despite accumulating evidence of the potential of NIBS, the precise therapeutic mechanisms of action of rTMS and tDCS are largely unidentified and there is no consensus about standardized treatment protocols. Moreover, when deciding on treatment after stroke with either rTMS or tDCS, the poststroke time and lesion status should be considered, and stimulation intensity and duration must be fine-tuned to prevent further tissue damage or the interruption of beneficial plastic changes.^{15,16} These uncertainties emphasize the critical need for basic understanding of the (patho)physiological processes that are influenced by rTMS and tDCS paradigms after stroke, which may ideally be explored in well-controllable and reproducible experimental animal models.

In animal models of stroke, similar to the human condition, there is a variable degree of spontaneous functional improvement after stroke, associated with a complex cascade of cellular and molecular processes that are activated within minutes after the insult, both in perilesional tissue and remote brain regions.^{17,18} These events include changes in genetic transcriptional and translational processes, alterations in neurotransmitter interactions, altered secretion of growth factors, gliosis, vascular remodeling, and structural changes in axons, dendrites, and synapses.^{19,20} Therefore, assessment of the effects of NIBS on endogenous recovery processes in animal stroke models offer excellent opportunities for the exploration of neuroplastic and neuromodulatory mechanisms, which could aid in the optimization of treatment protocols for clinical applications.

Our goal was to provide an overview of studies that assessed functional outcomes and potential mechanisms of action of rTMS and tDCS in animal models of stroke, which may guide future studies that aim to improve mechanistic insights and therapeutic utilization of NIBS effects after stroke.

Literature Search Strategy and Study Quality Assessment

A bibliographic search was carried out to identify publications on rTMS or tDCS applications in preclinical stroke studies, using specific keywords that are specified in the rTMS and tDCS sections below. The quality of the methods of each study was assessed based on the Good Laboratory Practice (GLP) guidelines provided by Macleod et al,²¹ which have been proposed to prevent the introduction of bias at the bench and the consequent overstatement of neuroprotective efficacy. The GLP guidelines suggest that details of the following 8 points should at least be included in publications: (1) animals (species, strain, source), (2) sample size calculation, (3) inclusion/exclusion criteria, (4) randomization (method), (5) allocation concealment, (6) reporting of animals excluded from analysis, (7) blinded assessment of outcome, and (8) reporting potential conflicts of interest and study funding. The methods of each reviewed article were assessed and scored based on each of the 8 GLP criteria. One point was given for each criterion if all information was present, half a point for partial information, and no point if the information was absent or unclear. The GLP scores, which could range from 0 to 8, for all publications are summarized in Tables 1 and 2.

Repetitive Transcranial Magnetic Stimulation in Stroke Models

In contrast to the majority of scientifically and clinically approved treatments, there is a relative shortage of preclinical nonhuman TMS data.²² This may be explained by the fundamentally noninvasive character of TMS, resulting in approved use of magnetic stimulators for peripheral nerve stimulation in several countries, including the United States, and Food and Drug Administration approval of rTMS to treat depression without animal safety data. Moreover, there is a lack of appropriately sized coils for studies in small animals. Consequently, there are still many uncertainties about the full therapeutic potential of rTMS protocols, and their precise therapeutic mechanism of action in several neurological and psychiatric disorders.

Since the first publication of a TMS study in rats in 1990, there has been an exponential increase in published animal TMS studies, including preclinical studies in animal models of disease.^{23,24} Experiments involving repetitive TMS in animal models of Alzheimer's disease,²⁵ depression,²⁶ epilepsy,²⁷ Huntington's disease,²⁸ Parkinson's disease,²⁹ and stroke^{30,41} have already provided substantial insights into the therapeutic potential of TMS.

An in-depth literature search on PubMed, using combinations of keywords (eg, noninvasive brain stimulation, transcranial magnetic stimulation, rTMS, cerebral/stroke/ ischemia/infarct, disease/animal model, animal, rodent, rat,

GLP Score		I.5	2.5	4.5	4	4	3.5
Results	rTMS protected hippocampal neurons against delayed neuronal death. The extent of neuronal preservation depended on the interval between rTMS and ischemia, the stimulus frequency, and duration. High- frequency (25/50 Hz) rTMS protocols yielded the most beneficial effects.	Significantly improved neurological severity scores, accompanied with increased expression of c-Fos and RDNF	rTMS significantly increased the ATP contents and MAP-2 expression in injured brain area, highest efficacy with high frequency rTMS.	Significantly improved neurological scores and reduced infarct volume. Elevated glucose consumption in the stroke hemisphere, lowered caspase-3 cell numbers, and incressed RcL-2/Bax ratio after rTMS	Reduced cognitive deficits in rTMS group. BDNF and its receptor, TrkB, as well as BcI-2 were upregulated after rTMS treatment, whereas a decrease in the expression of Bax	Improved neurological outcome, and enhanced neurological outcome, the subventricular zone through regulation of the miR-25/p57- signaling pathway.	Improved motor performance on Garcia's motor behavior index in the stimulation group, compared with the control group.
Stimulation State	Conscious	Conscious	Unclear	Conscious	Conscious	Conscious	Ketamine (30 mg/ kg) sedation
Number of Pulses and Sessions	Experiment 1: One session of either 8, 16, 32, 64, 128, or 256 s for each frequency, respectively. One train = 8 s, intertrain intervals of 10 s Experiment 2: One session of 25 Hz for 128 s at different intervals before ischemia	60 (30 pulses twice daily); Animals received rTMS for either 1, 7, 14, 21, or 28 days nossestroke	10 trains of 5 s stimulation, 2 min intertrain interval Animals received rTMS at either 0 h, 12 h, or 36 h after reperfusion; one rTMS cassion auov 12 h	1000 (once daily, for 7 days, 5 s stimulation off, repeated 10 times)	300 (once daily for 7 days; 3 s stimulation on, 50 s stimulation off, repeated 10 times)	300 (once daily for 7 days; 3 s stimulation on, 50 s stimulation off, repeated 10 times)	90 pairs of stimulation at 0.05 Hz (30 min per day, for 5 consecutive days)
T reatment Onset	2-5 days prior to stroke	Directly poststroke	Either 0 h, 12 h, or 36 h after reperfusion	l h after MCAO	24 h poststroke	24 h poststroke	24 h after occlusion
Stimulation Frequency and Intensity	Experiment 1: 5, 10, 25, or 50 Hz at 120% RMT Experiment 2: 25 Hz at 120% RMT	0.5 Hz, unclear RMT	5 Hz at 120% or 200% RMT 20 Hz at 120% or 200% RMT	20 Hz at 100% RMT	10 Hz at 120% RMT	10 Hz at 120% RMT	0.05 Hz at 120% RMT + 6 mA electrical stimulation
Control Condition	Not clearly described, but the authors mention the following control groups: healthy, sham operated, rTMS alone, and BCCAO alone	MCAO without rTMS	Sham rTMS (coil placed perpendicular to head); MCAO without rTMS	MCAO without rTMS; MCAO and sham rTMS	Sham-operated MCAO without rTMS	Sham-operated, MCAO without rTMS	Sham stimulation, electrical stimulation was 0 mA and the TMS coil was elevated I cm above the rat's scalp
Stimulation Target, Coil Type	Target unknown, Circular coil (50 mm diameter)	Target unknown, Circular coil (12 mm outer diamerer)	lpsilesional frontoparietal cortex, F8c (70 mm outer diameter)	Ipsilesional frontoparietal cortex, F8c (70 mm outer diameter)	Ipsilesional cortex, Circular coil (60 mm diameter)	Ipsilesional primary motor cortex, Circular coil (60 mm diameter)	4 hours) after stroke Contralesional hemisphere, FBC (5 cm outer ring diameter) and ipsilesional soleus muscle (electrical stimulation)
Number of Animals	Unclear	stroke 80 (active: 40; control: 40)	90 (active: 60; control: 30)	30 (active: 10; control: 20)	Absent	125 (active: 90; control: 35)	ments acutely (≤2: 54 (active: 19; control: 15; excluded: 20)
Species/ Stroke Model	stroke Mongolian gerbil, BCCAO	(≪24 hours) after SD rats, Permanent MCAO	Wistar rats, Transient MCAO	SD rats, Transient MCAO	SD rats, Transient MCAO	SD rats, Transient MCAO	ed with other treat SD rats, Permanent MCAO
Reference	rTMS before : Fujiki et al (2003) ³⁰	rTMS acutely Zhang et al (2007) ³¹	Feng et al (2008) ³²	Gao et al (2010) ³³	Guo et al (2017) ³⁴	Guo et al (2014) ³⁵	rTMS combin Shin et al (2008) ³⁶

Table 1. TMS Studies in Animal Models of Stroke.

(continued)

GLP Score	2.5	l.5	4	m	'n
Results	Compared to the MCAO model group, improved performance of the EA + rTMS group in the Morris water maze, increased expression of BcI-2 mRNA and decreased expression of caspase-3 24 hours poststroke.	Diminished neurological function and amplified signs of inflammation in particularly the 20 Hz rTMS group combined with G-CSF. No significant differences between treatment and control groups between days 7 and 25 poststroke.	Compared to sham, rTMS group showed improved functional performance in the beam balance test, increased expression of anti-apoptotic Bcl-2 and weakened expression of pro-apoptotic Bax; however, NMDA and MAP-2 did not differ hervean or functions	Both stimulation paradigms improved neurological outcome, reduced the infarct volume, and significantly promoted neurogenesis. Elevated protein levels of BDNF and photshorv/ared-TrKB were defected	All rTMS protocols reduced behavioral deficits and increased expression of genes involved in neurotransmission and plasticity. iTBS was most effective, while cTBS effects were negligible.
Stimulation State	Unclear	Propofol sedation	Conscious	Conscious	Conscious
Number of Pulses and Sessions	60 rTMS pulses (30 pulses, twice daily for 14 days, each session consisted of 1 min of stimulation) combined with 30 min of EA, twice daily, for 14 days	Unclear (10 sessions/2 weeks; 20 min per session)	3500 (10 sessions/2 weeks: 1 session is 7 repetitions of five 1 s trains at a rate of 10 Hz with a 1s intertrain interval)	800/600 (10 sessions/2 weeks; 20 Hz: 40 trains for 1 s, 15 s intertrain interval; ITBS: 20 trains, 10 bursts every 10 s)	2400 (once daily, for 10 days over 2 weeks)
Treatment Onset	6, 12, 24, 48, and 72 h after occlusion	Directly poststroke	4 days poststroke	3 days poststroke	3 days poststroke
Stimulation Frequency and Intensity	0.5 Hz rTMS at unknown RMT + EA (sparse wave 2 Hz, dense wave 30 Hz, current intensity 2 mA)	l Hz or 20 Hz at 100% RMT	10 Hz at 80% RMT	20 Hz or iTBS at 120% or 80% RMT	I Hz, 5 Hz, cTBS or iTBS at 110% RMT
Control Condition	Healthy, sham operated, occlusion alone, all received sham EA + sham rTMS	Saline administration + sham rTMS (coil placed perpendicular to head); G-CSF administration + sham rTMS	Occlusion with sham rTMS	Healthy, sham- operated, sham- operated + TMS	Sham rTMS (coil elevated 15 cm above rat's head); MCAO without rTMS; healthy and sham- operated controls
Stimulation Target, Coil Type	Target unknown, Circular coil (120 mm diameter) EA points: Baihui and Dazhui	Ipsilesional cortex, F8c (25 mm)	Ipsilesional motor cortex, F8c (12 mm inner diameter, 20 mm outer diameter)	Ipsilesional cortex, F8c (22 mm inner diameter, 90 mm outer diameter)	Ipsilesional cortex, F8c
Number of Animals	200 (active: 50; control: 150)	59 (active: 29; control: 30)	r stroke 20 (active: 10; control: 10)	72 (active: 44; control: 28)	149 (active: 36; control: 45)
Species/ Stroke Model	SD rats, Transient MCAO	SD rats, Permanent MCAO	ly (1-7 days) afte SD rats, Transient MCAO	Wistar rats, Transient MCAO	Wistar rats, Transient MCAO
Reference	Li et al (2012) ³⁷	Beom et al (2015) ³⁸	TMS subacutt Yoon et al (2011) ³⁹	Luo et al (2017) ⁴⁰	Ljubisavljevic et al (2015) ⁴¹

colony stimulating factor; GLP, Good Laboratory Practice; iTBS, intermittent theta burst stimulation; MAP-2, microtubule associated protein-2; MCAO, middle cerebral artery occlusion; MO, machine output; RMT, resting motor threshold; rTMS, repetitive transcranial magnetic stimulation; SD, Sprague Dawley.

Table I. (continued)

GLP Score	5	ы. С	3.5	7
Results	In both experiments, C-tDCS reduced infarct volume and the number of spreading depolarizations.	C-tDCS improved functional recovery, reduced lesion volume, decreased number of apoptotic cells, and lowered cortical glutamate and taurine levels. A-tDCS increased infarct volume, disturbed blood-brain barrier integrity, promoted hemorrhage, and elevated numbers of inflammatory and apoptotic cells.	Cathodal tDCS and cathodal tDCS + PSS restored hemodynamics and neural activity in the ischemic region. Additionally, cathodal tDCS + PSS reduced infarct volume, inhibited microglial activation and more effectively preserved grip strength.	Bilateral anodal and cathodal tDCS improved motor function, increased dendritic spine density, and reduced expression of neuronal gap junction, hemichannel pannexin-1 mRNA
Stimulation State	Isoflurane (2%) anesthesia	Isoflurane (1.5/2%) anesthesia	Pentobarbital anesthesia (50 mg/kg bolus for induction and I 5 mg/kg/h for maintenance, ip injections)	Unclear
Treatment Onset	Experiment I: 45 min poststroke Experiment 2: Directy poststroke	30 min or 4.5 h poststroke	Directly poststroke	I day poststroke
Stimulation Intensity, Density, Duration, and Number of Sessions	Experiment 1: 0.2 mA, 28.6 A/m ² , (15 min on and 15 min off, discontinuous for 4) Experiment 2: 0.2 mA, 28.6 A/m ² , (15 min on and 15 min off, discontinuous for 6 h)	0.25 mA, 55 A/m ² , 20 min on and 20 min off Experiment 1: 30 min after onset of anesthesia (no MCAO, 2 sessions Experiment 2: 30 min after MCAO, 2 sessions Experiment 3: 4.5 h after MCAO, 2 sessions	0.4 mÅ, 20.4 Å/m ² , 20 min, I session FSS: Stimulation of 3 FIZ with a 0.2 ms pulse width and 2 mÅ delivered for 90 s, every 15 min for 2 h after the 20 min C-tDCS	0.1 mA, density unclear, daily 30 min sessions for either 3, 7, or 14 days
Control Condition	Experiment 1: MCAO with no tDCS Experiment 2: MCAO with no tDCS Sham MCAO + C-tDCS	Experiment I: Sham MCAO with no tDCS Experiments 2 and 3: MCAO with no tDCS	Photothrombotic group without stimulation	Sham operation group; MCAO without tDCS
Stimulation Polarity and Electrode Location	Cathodal stimulation: ipsilesional hemisphere Active electrode: ipsilesional, 2 mm left and 1 mm post to bregma bregma Counter electrode: ventral side of the thorax	Cathodal or anodal stimulation: ipsilesional hemisphere Active electrode: ipsilesional, left partetal area (2.5 mm left and 0.5 mm post to bregma) counter electrode: ventral thorax	Cathodal stimulation: ipsilesional hemisphere Active electrode: ipsilesional, 5 mm ant and 3 mm lat to bregma Counter electrode: thorax PSS: subdermal needle electrodes (2 in each limb) placed in the fore- and hindpaws contralateral to the ischemic hemisphere	Unclear Active electrode: positioned 5 mm left and 2 mm anterior to the interaural line Counter electrode: Unclear
Number of Animals	53 Experiment I: (active: 12; control: 12) Experiment 2: (active: 17; control: 12)	 137 Experiment 1: (active: 24: control: 12) Experiment 2: (active: 50: control: 25) Experiment 3: (active: 16: control: 10) 	58 (active: 42; control: 16)	90 Number of animals per group was unclear
Species/Stroke Model	24 hours) after stroke SD rats, MCAO	C57BL/6 mice, Transient MCAO	SD rats, Cortical photothrombosis	SD rats, MCAO
Reference	tDCS acutely (≤ Notturno et al (2014) ⁶⁵	Peruzzotti- Jametti et al (2013) ⁶⁶	Liu et al (2017) ⁶⁷	Jiang et al (2012) ⁶⁸

 Table 2. tDCS Studies in Animal Models of Stroke.

(continued)

GLP Score	m	2.5	Ŋ
Results	Early A-tDCS improved motor and cognitive performance, whereas late A-tDCS treatment only improved sensorimotor performance. Early A-tDCS significantly increased expression of MAP- 2. Late A-tDCS significantly enhanced expression of GAP-43.	Repetitive A-tDCS improved motor function and reduced neuronal axon deterioration in the ipsilesional internal capsule. C-tDCS deteriorated motor function. tDCS and exercise did not reduce the infarr tipe	DCS improved motor recovery. C-tDCS led to full recovery in strength and gait, whereas A-tDCS-treated animals did not fully regain limb strength. Beuh A-tDCS and C-tDCS induced neurogenesis in the ipsilesional subventricular zone. Generation and migration of oligodendrocyte precursors from the SVZ to the ischemic lesion was only increased after C-tDCS, accompanied by MI- polarization of microglia.
Stimulation State	2% Isofiurane	Anesthesia with ketamine (1%, 15 mL/kg)	Isoflurane anesthesia (dose unclear)
Treatment Onset	l or 7 days poststroke	2 days poststroke	3 days poststroke
Stimulation Intensity, Density, Duration, and Number of Sessions	0.2 mA, 28.8 A/m ² , 20 min/ day for 5 days, 5 sessions	0.1 mA, 1.3 A/m ² , daily 30 min sessions for 2 weeks	0.5 mA, 142.9 A/m ² , 15 min/day, for a total of 10 days (with a 2 day tDCS-free interval), 10 sessions
Control Condition	MCAO with sham- tDCS	MCAO exercise group; MCAO group with no tDCS	MCAO with sham- tDCS
Stimulation Polarity and Electrode Location	Anodal stimulation: ipsilesional hemisphere Active electrode: ipsilesional border zone of M I Counter electrode: anterior aspect of the chest	Cathodal or anodal stimulation: ipsilesional hemisphere Active electrode: ipsilesional, 3 mm left and 2 mm ant to the interaural line Counter electrode: attached to the trunk of the animal	Cathodal or anodal stimulation: ipsilesional hemisphere Active electrode: ipsilesional, bregma AP + 2.0 mm, ML + 2.0 mm Counter electrode: positioning unclear
Number of Animals	30 (active: 20; control: 10)	61 (20 animals died during the experiment) (active: 20; control: 21)	41 (only 28 fulfilled the inclusion criteria) (active: 19; control: 9)
Species/Stroke Model	y (1-7 doys) after stroke SD rats, Transient MCAO	SD rats, Permanent MCAO	Wistar rats, Transient MCAO
Reference	tDCS subacutel Yoon et al (2012) ⁶⁹	(2010) ⁷⁰ (2010) ⁷⁰	(2016) ⁷¹

Laboratory Practice: Iat, Iateral; MAP-2, microtubule associated protein-2; MCAO, middle cerebral artery occlusion; ML, mediolateral; post, posterior; PSS, peripheral sensory scimulation; SD, Sprague Dawley; SVZ, subventricular zone; tDCS, transcranial direct current stimulation.

Table 2. (continued)

mice/mouse, gerbil, large animals/nonhuman primate), for animal models of stroke involving treatment with rTMS, revealed 12 scientific articles published between 2003 and October 2017.³⁰⁻⁴¹ These articles (summarized in Table 1) applied rTMS after experimental stroke to assess (1) effects on ischemic tolerance,³⁰ (2) underlying therapeutic mechanisms,^{30-35,39,40} (3) the additive effect of TMS when combined with other therapies,³⁶⁻³⁸ and (4) the effect of rTMS on gene expression.⁴¹ These studies applied TMS using coils of different shapes and sizes. Either circular or figure-of-eight coils were used, with outer diameter sizes ranging from 12 to 60 mm or 20 to 70 mm, respectively. Figure-of-eight coils generally provide more focal stimulation⁴²; however, the use of smaller circular TMS coils may improve focality in the small rodent brain.⁴³

rTMS Before Stroke

One study has been published in which rTMS was applied prior to experimental stroke. In this study, Fujiki and colleagues found that rTMS before a transient ischemic insult in adult gerbil brain induced ischemic tolerance preventing delayed neuronal death in the hippocampus.³⁰ The extent of neuronal preservation following rTMS was dependent on the stimulation paradigm, as well as on the interval between stimulation and ischemic stroke. Maximal neuronal preservation and protection was accomplished after the application of 25 Hz rTMS (for at least 128 seconds) at 48 hours before ischemia. This study suggests a potential role for NIBS as a pretreatment in patients undergoing procedures that may induce transient brain ischemia.

rTMS Acutely (≤24 Hours) After Stroke

The application of rTMS directly after permanent middle cerebral artery occlusion (MCAO) in rats has reportedly led to neuroprotection and improved behavioral outcomes.³¹ Following longitudinal treatment with low-frequency (0.5 Hz) rTMS to a nonspecified region of interest for different periods of time (7, 14, 21, and 28 days), Zhang et al found a significant improvement in functional recovery, based on a neurological severity score, as compared with untreated rats. However, these results should be treated with care as statistical testing involved multiple *t*-testing of differences between 10 experimental groups, without a prior ANOVA (analysis of variance) or correction for multiple comparisons, which could have led to type I (false positive) errors. It was also reported that rTMS-treated animals showed a substantial increase in the expression of c-Fos (at days 7, 14, 21, and 28) and brain-derived neurotrophic factor (at days 7, 14, and 21) in cortical tissue surrounding the infarct area. The authors speculated that rTMS-induced c-Fos expression, indicative of neuronal activation, subsequently led to upregulation of brain-derived neurotrophic factor

expression, which could contribute to improved outcome after stroke.^{23,31}

Underlying biological mechanisms of rTMS treatment in the acute phase after ischemic stroke were also explored by Feng and colleagues.³² Their study focused on the effect of rTMS on ATP content and microtubule associated protein-2 expression. Microtubule associated protein-2, a highmolecular-weight protein, mainly present in mature neurons, plays an important role in mitochondrial axonal transport, maintaining the structural integrity of neurons, and acts in synapse formation and dendrite plasticity.^{32,44,45} A reduction of microtubule associated protein-2 expression due to an ischemic lesion might decrease mitochondrial axonal transport, leading to reduced ATP availability and eventually neuronal death.⁴⁶⁻⁴⁹ Rats treated with ipsilesional high-frequency (20 Hz) rTMS paradigms, starting 1 hour after transient MCAO, revealed significantly increased ATP content and microtubule associated protein-2 expression in the affected hemisphere, compared with lower frequency rTMS (5 Hz) and untreated sham/control groups. The increased microtubule associated protein-2 expression following high-frequency rTMS might suggest ongoing processes of neuronal repair.⁵⁰

Repetitive TMS treatment (for 7 days) initiated 1 hour after transient ischemia has also been reported to reduce apoptosis. In a study by Gao and colleagues, ipsilesional high-frequency rTMS (20 Hz) resulted in significantly reduced numbers of caspase-3 positive cells and an increased ratio of anti-apoptotic Bcl-2 over pro-apoptotic Bax in the affected hemisphere, compared with control groups without rTMS treatment.³³ Ipsilesional high-frequency rTMS treatment also reduced the infarct volume and improved the neurological outcome, which could be explained by the blocking of apoptosis and the maintenance of glucose utilization in the ischemic hemisphere as observed with fluoro-deoxyglucose microPET.

Anti-apoptotic effects of ipsilesional high frequency (10 Hz) rTMS (for 7 days) were also observed when introduced 24 hours after stroke.³⁴ Compared with an untreated stroke group, rTMS significantly reduced apoptosis in the CA1 region of the hippocampus after poststroke stimulation. Along with reduced neuronal apoptosis, neurogenesis was enhanced in the hippocampi of the rTMS group, which was accompanied by improved cognitive function in the Morris water maze task. The validity of these positive treatment effects is difficult to judge as the total number of included animals and group sample sizes were vaguely reported for this study.

Repetitive TMS-induced enhancement of poststroke neurogenesis has also been demonstrated by an earlier study from the same research group. In this study, Guo and colleagues applied 10 Hz rTMS treatment (for 7 days) to the lesioned hemisphere starting 24 hours after transient MCAO in rats.³⁵ This significantly increased the proliferation of adult neural stem cells in the ipsilateral subventricular zone and upregulated micro RNA-25 in the ischemic cortex, as compared with sham-operated and untreated model groups.³⁵ Unfortunately, this study lacked a sham rTMS group. Other researchers have demonstrated the potential of rTMS to promote the proliferation of adult neural stem cells in healthy rat brain,⁵¹ which corroborates the hypothesis that rTMS may enhance poststroke neurogenesis.

rTMS Combined With Other Treatments Acutely (≪24 Hours) After Stroke

Some studies have combined TMS with other therapies, such as peripheral nerve stimulation,³⁶ electro-acupuncture,³⁷ and granulocyte-colony stimulating factor (G-CSF) administration,³⁸ for the treatment of stroke. Conjoint electrical stimulation of a peripheral nerve (ipsilesional to the lesion) and contralateral motor cortex stimulation using TMS is also referred to as paired associative stimulation, a method that can induce long-lasting changes in cortical excitability.⁵² Following daily paired associative stimulation for 5 consecutive days, starting 24 hours after permanent MCAO in rats, Shin et al found that the motor behavioral index (7 days poststroke) was significantly higher in the stimulation group than in the sham-stimulated group.³⁶

The combination of rTMS and electro-acupuncture may improve learning and memory abilities in rats, as shown by Li et al.³⁷ In this study, electro-acupuncture electrodes were inserted at Baihui (right midpoint of the parietal bone) and Dazhui (posterior midline) points; however, the rTMS target region/hemisphere was not defined. The combination of 0.5 Hz rTMS with electro-acupuncture treatment (for 14 days) appeared to have an anti-apoptosis effect, by altering the expression of caspase-3 (reduced expression) and Bcl-2 (increased expression) in peri-infarct tissue. Additionally, improved learning and memory abilities in the treatment groups were demonstrated by shorter escape latency times in the Morris water maze task, compared with control groups. Treatment started either at 6, 12, 24, 48, or 72 hours after transient MCAO and was found to be most effective when started 24 hours poststroke.

An attempt to use rTMS to enhance the neuroprotective effects of cytokine G-CSF, by administering treatment directly following permanent MCAO in rats, has been unsuccessful.³⁸ G-CSF and its receptors are widely expressed in the central nervous system and involved in various processes that can contribute to neuroprotection and neurorepair, such as anti-apoptosis, neurogenesis, anti-inflammation, cellular growth, arteriogenesis, anti-oxidation, and stem cell recruitment.⁵³ In animal models, administration of G-CSF has reportedly reduced ischemic infarct volume and facilitated functional recovery, particularly after transient cerebral ischemia.⁵⁴ Beom et al hypothesized that the combination of G-CSF treatment with ipsilesional rTMS would enhance the effects of G-CSF and

reduce its adverse effects (splenomegaly, headache, bone pain, and emergence of bone marrow disease).³⁸ However, the combination therapy of G-CSF (for 5 days) and ipsilesional high-frequency rTMS (for 2 weeks) exerted a deleterious effect on functional recovery. Beom et al speculated this may be related to the reduced expression of angiogenic mechanisms, enhanced inflammatory responses, or inappropriate timing.³⁸ These findings are contradictory to results from other experimental studies that reported improvements in motor function, increased neurogenesis, and reduced apoptosis after single treatment with G-CSF⁵⁴ or rTMS.³³⁻³⁵

rTMS Subacutely (1-7 Days) After Stroke

Neuroprotective and neurotrophic effects of rTMS have also been reported when treatment was initiated in the subacute phase, that is, between 1 and 7 days, after experimental stroke. Yoon and colleagues performed daily ipsilesional high-frequency (10 Hz) rTMS between 4 and 18 days after transient MCAO.³⁹ This therapeutic paradigm resulted in enhanced functional improvement in the beam balance test, and reduced neuronal apoptosis, as compared with a sham stimulation group.³⁹

Three days after transient MCAO in rats, Luo and colleagues started ipsilesional 20 Hz rTMS or intermittent theta burst stimulation for 10 days.⁴⁰ Both stimulation protocols promoted neurogenesis in the ipsilateral subventricular zone, and increased neural progenitor cell migration in the peri-infarct striatum, as compared with a control group, a sham-operated group, and a TMS-stimulated sham-operated group. Assessment of neurological function revealed significant main effects of group and time at 14 days after stroke, but there was no significant interaction between the 2 factors.

Ljubisavljevic and colleagues investigated the effects of 4 different ipsilesional rTMS protocols (1 Hz, 5 Hz, continuous and intermittent theta burst stimulation—starting 3 days poststroke) on gene expression after transient MCAO in rats.⁴¹ Compared with the untreated stroke group, the theta burst stimulation protocols induced significant improvement in behavioral deficit scores, without affecting infarct size, after 2 weeks of treatment. Changes in gene expression were largely dependent on stimulation frequency and pattern. Nonetheless, rTMS generally upregulated a large range of genes involved in neuroprotection, neurotransmission, angiogenesis, neural repair, and neuronal plasticity.

Transcranial Direct Current Stimulation in Stroke Models

Over the past couple of years, the use of tDCS as a therapy for psychiatric and neurological disorders has been increasingly investigated in clinical as well as preclinical studies.55,56 For application of tDCS in animal models, the majority of studies have employed an electrode montage similar to a setup that has been originally described by Liebetanz et al.⁵⁷ In this approach, a small, plastic jacket (3.5 mm² contact area) is fixed onto the cranium using nontoxic cement. Saline and a wire electrode are inserted into the plastic jacket before stimulation. In addition to the unilateral epicranial electrode, a large rubber-plate electrode (counter electrode) is placed onto the thorax of the animal. A weak, constant, electrical current (0.1 µA to 10 mA) can then be applied transcranially.⁵⁸ Safety guidelines for the application of cathodal⁵⁸ and anodal⁵⁹ tDCS protocols in animals have been defined. Unlike TMS, tDCS currents do not evoke action potentials, but rather modify the transmembrane neuronal potential and modulate the firing rate of individual neurons in response to supplementary inputs.60

Transcranial DCS treatment has shown therapeutic potential in animal models of Alzheimer's disease,⁶¹ epilepsy,⁶² neuropathic pain,⁶³ Parkinson's disease,⁶⁴ and stroke.⁶⁵⁻⁷¹ An in-depth literature search, with various combinations of keywords (eg, noninvasive brain stimulation, transcranial direct current stimulation, tDCS, cerebral/stroke/ischemia/infarct, disease/animal model, animal, rodent, rat, mice/mouse, gerbil, large animals/nonhuman primate), for animal models of stroke involving treatment with tDCS, revealed 7 articles published between 2010 and October 2017 (summarized in Table 2). These articles aimed to assess the safety and efficacy of tDCS in acute to subacute stroke,⁶⁵⁻⁶⁸ and to identify which functional⁶⁹ and cellular^{70,71} changes are associated with tDCS-induced recovery after stroke.

tDCS Acutely (≤24 Hours) After Stroke

In the first hours following experimental stroke, cathodal tDCS has been shown to significantly reduce the number of peri-infarct depolarizations,65 which are believed to contribute to infarct growth.⁷² Following discontinuous ipsilesional cathodal stimulation for either 4 hours (starting 45 minutes after MCAO) or 6 hours (starting directly after MCAO) the infarct volume was significantly reduced by 20% or 30%, respectively, compared with nonstimulated MCAO control groups.⁶⁵ Accordingly, the degree of infarct reduction correlated with the extent of cathodal tDCSinduced decrease in peri-infarct depolarizations. In correspondence with these results, Peruzzotti-Jametti and colleagues found that ipsilesional cathodal tDCS for 30 minutes, starting 4.5 hours after transient MCAO in mice, had a significantly favorable treatment effect compared with sham-stimulation and anodal tDCS.⁶⁶ Cathodal tDCS led to reduced edema and inflammation, decreased the number of apoptotic cells, and lowered cortical glutamate, creatine, and taurine levels. Consequently, the cytoarchitecture

of the cerebral cortex was relatively preserved after cathodal tDCS, resulting in smaller infarct volume and better functional recovery.

The combination of ipsilesional cathodal tDCS and peripheral sensory stimulation of the contralateral forelimb has also been shown to be therapeutically beneficial. Mice treated with this therapeutic strategy immediately after photothrombotic stroke in the sensorimotor cortex showed improved perilesional hemodynamics and enhanced recovery of neural activity in the first hours after stroke as compared with untreated animals and tDCS-treated animals without combined peripheral sensory stimulation.⁶⁷ Subsequently, this combination therapy resulted in reduced microglial activation, smaller infarct volumes, and better preserved grip strength 2 days after stroke.

Improved motor function has also been observed by Jiang and colleagues, who allegedly applied repetitive tDCS in rats through the implantation of pericranium electrodes.⁶⁸ However, from the article's Methods section it is unclear whether the authors indeed applied tDCS or a 10 Hz transcranial alternating current stimulation protocol. Nonetheless, the authors found that daily stimulation (starting 1 day after permanent MCAO) resulted in a significant increase of dendritic spine density in the cortex, compared with control groups, on several time points (days 3, 7, and 14) poststroke. In addition, the expression of hemichannel pannexin-1 mRNA, which is potentially involved in hypoxic depolarizations, was reduced.⁶⁸

In contrast to the reported positive effects of cathodal tDCS applied to the ipsilesional hemisphere in the acute phase after stroke, ipsilesional anodal tDCS has been shown to result in elevated numbers of inflammatory cells, augmented instability of the blood-brain barrier, and increased hemorrhage and infarct volume in the first hours after stroke.⁶⁶

tDCS Subacutely (1-7 Days) After Stroke

Several studies have explored the effects of tDCS applied in the subacute phase of stroke. Yoon and colleagues applied repetitive ipsilesional anodal tDCS for 5 days, starting either 1 day (early treatment) or 1 week (late treatment) after transient MCAO in rats.⁶⁹ Both treatment groups showed an improvement in cognitive performance and motor function scores; however, motor function improvement was slightly better in the late treatment group. Their immunohistochemical findings revealed significantly increased levels of microtubule associated protein-2 and growth associated protein-43 (a neuronal plasticity marker) in the perilesional and contralesional cortices in both treatment groups, which correlated with the cognitive and motor improvements. Magnetic resonance imaging and spectroscopy data showed that anodal tDCS did not affect infarct volume or metabolite levels.

Improved motor function was also observed in a study by Kim et al, where ipsilesional anodal tDCS treatment was applied over a period of 2 weeks, starting 2 days after permanent MCAO in rats.⁷⁰ Histologically, no clear changes were observed in infarct volume; nonetheless, axonal integrity in the ipsilesional internal capsule was better preserved. Repetitive ipsilesional cathodal tDCS, however, was associated with diminished functional recovery at 16 days postinfarct. The authors speculated that cathodal tDCS may have contributed to the already decreased excitability of the infarcted brain, thereby suppressing recovery mechanisms. The latter findings are in contrast with a recent study by Braun et al, in which accelerated functional recovery and neurogenesis after transient MCAO in rats were observed in response to ipsilesional delivery of either anodal or cathodal tDCS when applied 3 days after stroke.⁷¹ In this study, different aspects of motor function were influenced depending on the polarity of stimulation. For example, limb strength and gait were fully restored in animals treated with cathodal tDCS, whereas anodal tDCS-treated animals regained their gait, but not their full limb strength. Furthermore, cathodal tDCS triggered the generation and migration of oligodendrocyte precursors from the subventricular zone toward the ischemic lesion alongside an M1-polarization of microglia. The effect of cathodal tDCS on these cellular processes might have resulted in increased functional recovery as compared with the anodal tDCStreated group.

Contradictory findings by Kim et al⁷⁰ and Braun et al⁷¹ on the effects of cathodal tDCS on motor recovery may have been due to differences in the stroke model (permanent vs transient MCAO, respectively), the stimulation protocol, or the anesthesia during tDCS.

Discussion

We reviewed the main findings of 19 studies that applied either rTMS or tDCS transcranially in small rodents after experimental stroke. In general, most articles reported stimulation-induced tissue preservation or functional improvement after stroke, as compared with either untreated stroke control or sham stimulation groups. Several advantageous effects, including ischemic tolerance, neuroprotection, and neurorepair, mediated by molecular mechanisms involved in anti-apoptosis, neurogenesis, and neuroplasticity, were measured after rTMS and tDCS.

Only a few preclinical studies have directly compared the effects of different stimulation paradigms, and so far the majority of published rTMS and tDCS studies only assessed stimulation of the lesioned hemisphere. Moreover, ipsilesional high-frequency rTMS or intermittent theta burst stimulation appear to be more favorable for the induction of ischemic tolerance and expression of factors involved in preservation or recovery of postischemic tissue as compared with ipsilesional low-frequency rTMS or continuous theta burst stimulation.^{30,32,33,35,41} The latter inhibitory paradigms may have more significant therapeutic potential when applied to the contralesional hemisphere, as demonstrated in clinical stroke studies.⁷³⁻⁷⁵ The combination of rTMS with adjunct therapy in experimental stroke models has yielded both positive (paired associative stimulation,³⁶ rTMS plus electro-acupuncture³⁷) and negative results (rTMS plus G-CSF³⁸), and clearly requires further investigation.

The reviewed tDCS studies point toward neuroprotective and neurorestorative effects in animal stroke models, which depend on the polarity and onset of stimulation treatment. Cathodal tDCS of the ipsilesional hemisphere within minutes to hours after stroke reduced progression of ischemic damage.^{65,66} Additionally, the therapeutic benefits of cathodal tDCS may be enhanced when combined with peripheral sensory stimulation, resulting in preservation of neurovascular function and improved functional recovery.⁶⁷ On the other hand, hyperacute ipsilesional anodal tDCS led to progression of degenerative processes.⁶⁶ Repetitive cathodal and/or anodal tDCS of the ipsilesional hemisphere during later stages after stroke may promote various recovery-enhancing factors,⁶⁸⁻⁷¹ although this depends on poststroke timing.^{69,70}

The optimal therapeutic time window, in combination with the preferred stimulation protocol, for poststroke NIBS treatment has yet to be determined. From the current review, it seems that the application of rTMS can have advantageous effects irrespective of treatment onset. The onset of poststroke rTMS treatment will, however, affect the extent to which different neuroprotective and neurorestorative molecular mechanisms are influenced, which depend on various stroke characteristics (eg, type, location, and severity of stroke; age; comorbidities; etc). Similarly, the optimal stimulation parameters and duration may vary.

Similar to NIBS studies in stroke patients, the reviewed studies in animal stroke models employed variable protocols, and treatment efficacy was assessed with different outcome parameters, making it difficult to directly compare interventions and to determine the exact translational value of the applied stimulation protocols.76 Repetitive TMS was applied with either circular or figure-of-eight coils, with outer diameter sizes ranging from 12 to 60 mm or 20 to 70 mm, respectively. Focal stimulation with relatively large figure-of-eight coils can be achieved in rodents by secure fixation and lateralized coil positioning.⁷⁷ However, the majority of the reviewed rTMS studies reported stimulation of the animals while being restraint by hand and conscious, which has most likely negatively affected the focality of the stimulation. Additionally, stimulation frequencies (0.5-50 Hz), intensities (80% to 200% of the resting motor threshold), and number of pulses per session varied extensively between studies (see Table 1). In stroke patients, rTMS above 25 Hz and 130% of the resting motor threshold has been applied infrequently,^{75,76} as stimulation at high frequencies and intensities is considered unsafe and increases the risk of seizures.⁷⁸

Like the rTMS studies, the tDCS studies showed variability in terms of equipment, regions of interest, and stimulation intensity, density, and duration (see Table 2). Notably, unlike the rTMS studies, in all reviewed publications, animals were anesthetized during tDCS. The effect of (different types of) anesthesia on tDCS outcome remains largely unknown and requires further investigation. In the majority of the reported tDCS studies, stimulation parameters were within the safety limits specified by Liebetanz and colleagues.⁵⁸ However, recent anodal tDCS studies have reported the detection of lesions at electrode current densities of 47.8 A/m²⁷⁹ and 20.0 A/m^{2,59} which is significantly below the previously reported safety threshold of 149.9 A/m² using cathodal tDCS.58 This suggests that the safety threshold for lesion induction using tDCS could have been underestimated. The 20.0 A/m² lesion threshold for anodal stimulation is at least more than 10-fold higher than the typical electrode current density of 0.28 to 2.0 A/m² utilized in human studies.^{80,81} The application of lower stimulation parameters is therefore recommended in future tDCS studies of animal models, to improve the validity of the data and to facilitate translation to the clinic.⁸²

We observed that many of the included studies were of relatively low quality based on the GLP assessment criteria²¹ (Tables 1 and 2). Therefore, we cannot rule out that some of the reported findings might have been confounded by bias and could overstate the neuroprotective efficacy of rTMS and tDCS, similar to what has been reported for preclinical stroke drug trials.⁸³ We found that none of the included studies in this review described how the sample size was calculated or whether allocation concealment was implemented. Only 37% of the studies specified inclusion/exclusion criteria based on lesion size, cerebral perfusion status, or behavior poststroke. Exclusion of animals was poorly reported. Only 47% of the studies reported both the strain and source of the animals, and the age of animals was often unclear. The majority of studies reported potential conflicts of interest (79%), blinding of outcome assessment (63%), and random allocation to experimental groups (74%), although the method of randomization was generally not mentioned.

Conclusion

Even though the number of studies that assessed NIBS in animal models of stroke is still limited, the recoveryenhancing effects are encouraging and reflect the translational value of these investigations. Treatments with rTMS and tDCS in animal stroke models have shown that different protocols can have positive influences on functional recovery through ways of neuroprotection or neurorepair. However, the exact therapeutic mechanisms of rTMS and tDCS remain incompletely characterized. Furthermore, it should be mentioned that the limited number of reports of negative or null effects, and the relatively low study quality of several articles, might reflect a publication bias. Preclinical research on modes of action of different NIBS protocols in animal stroke models can provide critical information for the development of NIBS strategies for effective treatment after stroke. Consequently, prospective studies should investigate the effects of multiple stimulation protocols at different time points after stroke, on both the ipsilesional and contralesional hemispheres, to be able to identify optimal treatment protocols that would maximally enhance functional recovery. Prior computer simulations of the induced electrical field can provide essential insights in the location and focality of the stimulation approaches in rodent brain.43,84 Additionally, the rationale and criteria for the selection of the study parameters should be made explicit. Ideally, these studies would follow recent guidelines for preclinical stroke treatment studies, involving randomization and blinded assessments,^{21,85} and include measures of behavioral outcome and (image-based) markers of neuroprotection and neurorepair that are straightforwardly translatable to the clinic.

Declaration of Conflicting Interests

The authors declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Funding

The authors disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: This work was supported by the Netherlands Organization for Scientific Research (VICI 016.130.662).

References

- Bennette DA, Krishnamurthi RV, Barker-Collo S, et al; Global Burden of Diseases, Injuries, and Risk Factors 2010 Study Stroke Expert Group. The global burden of ischemic stroke: findings of the GBD 2010 study. *Glob Heart*. 2014;9:107-112. doi:10.1016/j.gheart.2014.01.001
- Langhorne P, Coupar F, Pollock A. Motor recovery after stroke: a systematic review. *Lancet Neurol*. 2009;8:741-754. doi:10.1016/S1474-4422(09)70150-4
- Claflin ES, Krishnan C, Khot SP. Emerging treatments for motor rehabilitation after stroke. *Neurohospitalist*. 2015;5:77-88. doi:10.1177/1941874414561023
- Pollock A, Farmer SE, Brady MC, et al. Interventions for improving upper limb function after stroke. *Cochrane Database Syst Rev.* 2014;(11):CD010820. doi:10.1002/14651858. CD010820.pub2
- 5. Palm U, Ayache SS, Padberg F, Lefaucheur JP. Non-invasive brain stimulation therapy in multiple sclerosis: a review of

tDCS, rTMS and ECT results. *Brain Stimul*. 2014;7:849-854. doi:10.1016/j.brs.2014.09.014

- Hatem SM, Saussez G, Faille MD, et al. Rehabilitation of motor function after stroke: a multiple systematic review focused on techniques to stimulate upper extremity recovery. *Front Hum Neurosci.* 2016;10:442. doi:10.3389/fnhum.2016.00442
- Takeuchi N, Izumi SI. Noninvasive brain stimulation for motor recovery after stroke: mechanisms and future views. *Stroke Res Treat*. 2012;2012:584727. doi:10.1155/2012/584727
- Murase N, Duque J, Mazzocchio R, Cohen L. Influence of interhemispheric interactions on motor function in chronic stroke. *Ann Neurol.* 2004;55:400-409.
- Nowak DA, Grefkes C, Ameli M, Fink GR. Interhemispheric competition after stroke: brain stimulation to enhance recovery of function of the affected hand. *Neurorehabil Neural Repair*. 2009;23:641-656. doi:10.1177/1545968309336661
- Takeuchi N, Tada T, Toshima M, Matsuo Y, Ikoma K. Repetitive transcranial magnetic stimulation over bilateral hemispheres enhances motor function and training effect of paretic hand in patients after stroke. *J Rehabil Med.* 2009;41:1049-1054. doi:10.2340/16501977-0454
- Dayan E, Censor N, Buch ER, Sandrini M, Cohen LG. Noninvasive brain stimulation: from physiology to network dynamics and back. *Nat Neurosci.* 2013;16:838-844. doi:10.1038/nn.3422
- Pelletier SJ, Cicchetti F. Cellular and molecular mechanisms of action of transcranial direct current stimulation: evidence from in vitro and in vivo models. *Int J Neuropsychopharmacol*. 2015;18(2):pyu047. doi:10.1093/ijnp/pyu047
- Fitzgerald PB, Daskalakis ZJ, eds. The mechanism of action of rTMS. In: *Repetitive Transcranial Magnetic Stimulation Treatment for Depressive Disorders*. Berlin, Germany: Springer; 2013:13-27. doi:10.1007/978-3-642-36467-9
- Shafi MM, Westover MB, Fox MD, Pascual-Leone A. Exploration and modulation of brain network interactions with noninvasive brain stimulation in combination with neuroimaging. *Eur J Neurosci*. 2012;35:805-825. doi:10.1111/ j.1460-9568.2012.08035.x
- Teskey G, Kolb B. Post-stroke recovery therapies in animals. In: Cramer SC, Nudo RJ, eds. *Brain Repair After Stroke*. Cambridge, England: Cambridge University Press; 2010:35-45.
- Schlaug G, Renga V, Nair D. Transcranial direct current stimulation in stroke recovery. *Arch Neurol.* 2008;65:1571-1576. doi:10.1001/archneur.65.12.1571
- Krakauer JW, Carmichael ST, Corbett D, Wittenberg GF. Getting neurorehabilitation right: what can be learned from animal models? *Neurorehabil Neural Repair*. 2012;26:923-931. doi:10.1177/1545968312440745
- Caleo M. Rehabilitation and plasticity following stroke: insights from rodent models. *Neuroscience*. 2015;311:180-194. doi:10.1016/j.neuroscience.2015.10.029
- Carmichael ST. Cellular and molecular mechanisms of neural repair after stroke: making waves. *Ann Neurol*. 2006;59:735-742. doi:10.1002/ana.20845
- Cramer SC. Repairing the human brain after stroke: I. Mechanisms of spontaneous recovery. *Ann Neurol.* 2008;63: 272-287. doi:10.1002/ana.21393

- Macleod MR, Fisher M, O'Collins V, et al. Good laboratory practice: preventing introduction of bias at the bench. *Stroke*. 2009;40:e50-e52. doi:10.1161/STROKEAHA.108.525386
- Wassermann EM, Zimmermann T. Transcranial magnetic brain stimulation: therapeutic promises and scientific gaps. *Pharmacol Ther.* 2012;133:98-107. doi:10.1016/j.pharmthera.2011.09.003
- 23. Tang A, Thickbroom G, Rodger J. Repetitive transcranial magnetic stimulation of the brain: mechanisms from animal and experimental models. *Neuroscientist*. 2015;23:82-94.
- Vahabzadeh-Hagh AM, Muller PA, Gersner R, Zangen A, Rotenberg A. Translational neuromodulation: approximating human transcranial magnetic stimulation protocols in rats. *Neuromodulation*. 2012;15:296-305. doi:10.1111/j.1525-1403.2012.00482.x
- Wang F, Zhang Y, Wang L, et al. Improvement of spatial learning by facilitating large-conductance calcium-activated potassium channel with transcranial magnetic stimulation in Alzheimer's disease model mice. *Neuropharmacology*. 2015;97:210-219. doi:10.1016/j.neuropharm.2015.05.027
- Feng SF, Shi TY, Fan-Yang, Wang WN, Chen YC, Tan QR. Long-lasting effects of chronic rTMS to treat chronic rodent model of depression. *Behav Brain Res.* 2012;232:245-251. doi:10.1016/j.bbr.2012.04.019
- Rotenberg A, Muller P, Birnbaum D, et al. Clinical neurophysiology seizure suppression by EEG-guided repetitive transcranial magnetic stimulation in the rat. *Clin Neurophysiol*. 2008;119:2697-2702. doi:10.1016/j.clinph.2008.09.003
- Tasset I, Pérez-Herrera A, Medina FJ, Arias-Carrión Ó, Drucker-Colín R, Túnez I. Extremely low-frequency electromagnetic fields activate the antioxidant pathway Nrf2 in a Huntington's disease-like rat model. *Brain Stimul*. 2013;6:84-86. doi:10.1016/j.brs.2012.03.015
- Lee JY, Kim SH, Ko AR, et al. Therapeutic effects of repetitive transcranial magnetic stimulation in an animal model of Parkinson's disease. *Brain Res.* 2013;1537:290-302. doi:10.1016/j.brainres.2013.08.051
- Fujiki M, Kobayashi H, Abe T, Kamida T. Repetitive transcranial magnetic stimulation for protection against delayed neuronal death induced by transient ischemia. *J Neurosurg*. 2003;99:1063-1069. doi:10.3171/jns.2003.99.6.1063
- Zhang X, Mei Y, Liu C, Yu S. Effect of transcranial magnetic stimulation on the expression of c-Fos and brainderived neurotrophic factor of the cerebral cortex in rats with cerebral infarct. *J Huazhong Univ Sci Technolog Med Sci*. 2007;27:415-418. doi:10.1007/s11596-007-0416-3
- Feng HL, Yan L, Cui LY. Effects of repetitive transcranial magnetic stimulation on adenosine triphosphate content and microtubule associated protein-2 expression after cerebral ischemia-reperfusion injury in rat brain. *Chin Med J (Engl)*. 2008;121:1307-1312.
- Gao F, Wang S, Guo Y, et al. Protective effects of repetitive transcranial magnetic stimulation in a rat model of transient cerebral ischaemia: a microPET study. *Eur J Nucl Med Mol Imaging*. 2010;37:954-961. doi:10.1007/s00259-009-1342-3
- 34. Guo F, Lou J, Han X, Deng Y, Huang X. Repetitive transcranial magnetic stimulation ameliorates cognitive impairment by enhancing neurogenesis and suppressing apoptosis in

the hippocampus in rats with ischemic stroke. *Front Physiol*. 2017;8:559. doi:10.3389/fphys.2017.00559

- 35. Guo F, Han X, Zhang J, et al. Repetitive transcranial magnetic stimulation promotes neural stem cell proliferation via the regulation of MiR-25 in a rat model of focal cerebral ischemia. *PLoS One*. 2014;9:e109267. doi:10.1371/journal. pone.0109267
- 36. Shin HI, Han TR, Paik NJ. Effect of consecutive application of paired associative stimulation on motor recovery in a rat stroke model: a preliminary study. *Int J Neurosci.* 2008;118:807-820. doi:10.1080/00207450601123480
- Li M, Peng J, Song Y, Liang H, Mei Y. Electro-acupuncture combined with transcranial magnetic stimulation improves learning and memory function of rats with cerebral infarction by inhibiting neuron cell apoptosis. *J Huazhong Univ Sci Technolog Med Sci.* 2012;32:746-749. doi:10.1007/s11596-012-1028-0
- Beom J, Kim W, Han TR, Seo KS, Oh BM. Concurrent use of granulocyte-colony stimulating factor with repetitive transcranial magnetic stimulation did not enhance recovery of function in the early subacute stroke in rats. *Neurol Sci.* 2015;36:771-777. doi:10.1007/s10072-014-2046-4
- 39. Yoon KJ, Lee YT, Han TR. Mechanism of functional recovery after repetitive transcranial magnetic stimulation (rTMS) in the subacute cerebral ischemic rat model: neural plasticity or anti-apoptosis? *Exp brain Res.* 2011;214:549-556. doi:10.1007/s00221-011-2853-2
- 40. Luo J, Zheng H, Zhang L, et al. High-frequency repetitive transcranial magnetic stimulation (rTMS) improves functional recovery by enhancing neurogenesis and activating BDNF/ TrKB signaling in ischemic rats. *Int J Mol Sci.* 2017;18:E455. doi:10.3390/ijms18020455
- Ljubisavljevic MR, Javid A, Oommen J, et al. The effects of different repetitive transcranial magnetic stimulation (rTMS) protocols on cortical gene expression in a rat model of cerebral ischemic-reperfusion injury. *PLoS One.* 2015;10:e0139892. doi:10.1371/journal.pone.0139892
- Cohen LG, Roth BJ, Nilsson J, et al. Effects of coil design on delivery of focal magnetic stimulation. Technical considerations. *Electroencephalogr Clin Neurophysiol*. 1990;75: 350-357.
- Tang AD, Lowe AS, Garrett AR, et al. Construction and evaluation of rodent-specific rTMS coils. *Front Neural Circuits*. 2016;10:47. doi:10.3389/fncir.2016.00047
- Sánchez C, Díaz-Nido J, Avila J. Phosphorylation of microtubule-associated protein 2 (MAP2) and its relevance for the regulation of the neuronal cytoskeleton function. *Prog Neurobiol*. 2000;61:133-168. doi:10.1016/S0301-0082(99)00046-5
- Conde C, Cáceres A. Microtubule assembly, organization and dynamics in axons and dendrites. *Nat Rev Neurosci*. 2009;10:319-332. doi:doi:10.1038/nrn2631
- Kalogeris T, Baines CP, Krenz M, Korthuis RJ. Cell biology of ischemia/reperfusion injury. *Int Rev Cell Mol Biol.* 2012;298:229-317. doi:10.1016/B978-0-12-394309-5.00006-7
- Hollenbeck PJ, Saxton WM. The axonal transport of mitochondria. *J Cell Sci*. 2005;118(pt 23):5411-5419. doi:10.1242/ jcs.02745

- Sims NR, Muyderman H. Mitochondria, oxidative metabolism and cell death in stroke. *Biochim Biophys Acta*. 2010;1802:80-91. doi:10.1016/j.bbadis.2009.09.003
- Errea O, Moreno B, Gonzalez-Franquesa A, Garcia-Roves PM, Villoslada P. The disruption of mitochondrial axonal transport is an early event in neuroinflammation. *J Neuroinflammation*. 2015;12:152. doi:10.1186/s12974-015-0375-8
- Li Y, Jiang N, Powers C, Chopp M. Neuronal damage and plasticity identified by microtubule-associated protein 2, growth-associated protein 43, and cyclin D1 immunoreactivity after focal cerebral ischemia in rats. *Stroke*. 1998;29:1972-1981.
- Ueyama E, Ukai S, Ogawa A, et al. Chronic repetitive transcranial magnetic stimulation increases hippocampal neurogenesis in rats. *Psychiatry Clin Neurosci*. 2011;65:77-81. doi :10.1111/j.1440-1819.2010.02170
- Stefan K, Kunesch E, Benecke R, Cohen LG, Classen J. Mechanisms of enhancement of human motor cortex excitability induced by interventional paired associative stimulation. J Physiol. 2002;543(pt 2):699-708. doi:10.1113/ jphysiol.2002.023317
- SolarogluI, CahillJ, JadhavV, ZhangJH. A novel neuroprotectant granulocyte-colony stimulating factor. *Stroke*. 2006;37:1123-1128. doi:10.1161/01.STR.0000208205.26253.96
- England TJ, Gibson CL, Bath PMW. Granulocyte-colony stimulating factor in experimental stroke and its effects on infarct size and functional outcome: a systematic review. *Brain Res Rev.* 2009;62:71-82. doi:10.1016/j.brainresrev.2009.09.002
- Brunoni AR, Fregni F, Pagano RL. Translational research in transcranial direct current stimulation (tDCS): a systematic review of studies in animals. *Rev Neurosci*. 2011;22:471-481. doi:10.1515/RNS.2011.042
- Brunoni AR, Nitsche MA, Bolognini N, et al. Clinical research with transcranial direct current stimulation (tDCS): challenges and future directions. *Brain Stimul.* 2012;5:175-195. doi:10.1016/j.brs.2011.03.002
- 57. Liebetanz D, Klinker F, Hering D, et al. Anticonvulsant effects of transcranial direct-current stimulation (tDCS) in the rat cortical ramp model of focal epilepsy. *Epilepsia*. 2006;47:1216-1224. doi:10.1111/j.1528-1167.2006.00539
- Liebetanz D, Koch R, Mayenfels S, König F, Paulus W, Nitsche MA. Safety limits of cathodal transcranial direct current stimulation in rats. *Clin Neurophysiol.* 2009;120:1161-1167. doi:10.1016/j.clinph.2009.01.022
- Jackson MP, Truong D, Brownlow ML, et al. Safety parameter considerations of anodal transcranial direct current stimulation in rats. *Brain Behav Immun*. 2017;64:152-161. doi:10.1016/j.bbi.2017.04.008
- Wagner T, Valero-Cabre A, Pascual-Leone A. Noninvasive human brain stimulation. *Annu Rev Biomed Eng.* 2007;9:527-565. doi:10.1146/annurev.bioeng.9.061206.133100
- Yu X, Li Y, Wen H, Zhang Y, Tian X. Intensity-dependent effects of repetitive anodal transcranial direct current stimulation on learning and memory in a rat model of Alzheimer's disease. *Neurobiol Learn Mem.* 2015;123:168-178. doi:10.1016/j.nlm.2015.06.003

- 62. Kamida T, Kong S, Eshima N, Abe T, Fujiki M, Kobayashi H. Transcranial direct current stimulation decreases convulsions and spatial memory deficits following pilocarpine-induced status epilepticus in immature rats. *Behav Brain Res.* 2011;217:99-103. doi:10.1016/j.bbr.2010.08.050
- 63. Filho PRM, Vercelino R, Cioato SG, et al. Transcranial direct current stimulation (tDCS) reverts behavioral alterations and brainstem BDNF level increase induced by neuropathic pain model: long-lasting effect. *Prog NeuroPsychopharmacol Biol Psychiatry*. 2016;64:44-51. doi:10.1016/j.pnpbp.2015.06.016
- 64. Li Y, Tian X, Qian L, Yu X, Jiang W. Anodal transcranial direct current stimulation relieves the unilateral bias of a rat model of Parkinson's disease. *Conf Proc IEEE Eng Med Biol Soc.* 2011;2011:765-768. doi:10.1109/IEMBS.2011.6090175
- Notturno F, Pace M, Zappasodi F, Cam E, Bassetti CL, Uncini A. Neuroprotective effect of cathodal transcranial direct current stimulation in a rat stroke model. *J Neurol Sci.* 2014;342:146-151. doi:10.1016/j.jns.2014.05.017
- Peruzzotti-Jametti L, Cambiaghi M, Bacigaluppi M, et al. Safety and efficacy of transcranial direct current stimulation in acute experimental ischemic stroke. *Stroke*. 2013;44:3166-3174. doi:10.1161/STROKEAHA.113.001687
- Liu YH, Chan SJ, Pan HC, et al. Integrated treatment modality of cathodal-transcranial direct current stimulation with peripheral sensory stimulation affords neuroprotection in a rat stroke model. *Neurophotonics*. 2017;4:045002. doi:10.1117/1.NPh.4.4.045002
- Jiang T, Xu RX, Zhang AW, et al. Effects of transcranial direct current stimulation on hemichannel pannexin-1 and neural plasticity in rat model of cerebral infarction. *Neuroscience*. 2012;226:421-426. doi:10.1016/j.neuroscience.2012.09.035
- Yoon KJ, Oh BM, Kim DY. Functional improvement and neuroplastic effects of anodal transcranial direct current stimulation (tDCS) delivered 1 day vs 1 week after cerebral ischemia in rats. *Brain Res.* 2012;1452:61-72. doi:10.1016/j. brainres.2012.02.062
- Kim SJ, Kim BK, Ko YJ, Bang MS, Kim MH, Han TR. Functional and histologic changes after repeated transcranial direct current stimulation in rat stroke model. *J Korean Med Sci.* 2010;25:1499-1505. doi:10.3346/jkms.2010.25.10.1499
- Braun R, Klein R, Walter HL, et al. Transcranial direct current stimulation accelerates recovery of function, induces neurogenesis and recruits oligodendrocyte precursors in a rat model of stroke. *Exp Neurol*. 2016;279:127-136. doi:10.1016/j.expneurol.2016.02.018
- Dreier JP. The role of spreading depression, spreading depolarization and spreading ischemia in neurological disease. *Nat Med.* 2011;17:439-447. doi:10.1038/nm.2333

- 73. Sebastianelli L, Versace V, Martignago S, et al. Low-frequency rTMS of the unaffected hemisphere in stroke patients: a systematic review. *Acta Neurol Scand.* 2017;136:585-605. doi:10.1111/ane.12773
- Ackerley SJ, Stinear CM, Barber PA, Byblow WD. Combining theta burst stimulation with training. *Stroke*. 2010;41:1568-1572. doi:10.1161/STROKEAHA.110.583278
- Lefaucheur JP, André-Obadia N, Antal A, et al. Evidencebased guidelines on the therapeutic use of repetitive transcranial magnetic stimulation (rTMS). *Clin Neurophysiol.* 2014;125:2150-2206. doi:10.1016/j.clinph.2014.05.021
- Dionísio A, Duarte IC, Patrício M, Castelo-Branco M. The use of repetitive transcranial magnetic stimulation for stroke rehabilitation: a systematic review. J Stroke Cerebrovasc Dis. 2018;27:1-31. doi:10.1016/j.jstrokecerebrovasdis.2017.09.008
- Rotenberg A, Muller PA, Vahabzadeh-Hagh AM, et al. Lateralization of forelimb motor evoked potentials by transcranial magnetic stimulation in rats. *Clin Neurophysiol.* 2010;121:104-108. doi:10.1016/j.clinph.2009.09.008
- Lomarev MP, Kim DY, Richardson SP, Voller B, Hallett M. Safety study of high-frequency transcranial magnetic stimulation in patients with chronic stroke. *Clin Neurophysiol.* 2007;118:2072-2075. doi:10.1016/j.clinph.2007.06.016
- Gellner AK, Reis J, Fritsch B. Glia: a neglected player in noninvasive direct current brain stimulation. *Front Cell Neurosci*. 2016;10:188. doi:10.3389/fncel.2016.00188
- Poreisz C, Boros K, Antal A, Paulus W. Safety aspects of transcranial direct current stimulation concerning healthy subjects and patients. *Brain Res Bull*. 2007;72:208-214. doi:10.1016/j. brainresbull.2007.01.004
- McKinley RA, McIntire L, Bridges N, Goodyear C, Bangera NB, Weisend MP. Acceleration of image analyst training with transcranial direct current stimulation. *Behav Neurosci*. 2013;127:936-946. doi:10.1037/a0034975
- Fritsch B, Gellner AK, Reis J. Transcranial electrical brain stimulation in alert rodents. J Vis Exp. 2017;(129):e56242. doi:10.3791/56242
- Sena ES, van der Worp HB, Bath PMW, Howells DW, Macleod MR. Publication bias in reports of animal stroke studies leads to major overstatement of efficacy. *PLoS Biol.* 2010;8:e1000344. doi:10.1371/journal.pbio.1000344
- Bernabei JM, Lee WH, Peterchev AV. Modeling transcranial electric stimulation in mouse: A high resolution finite element study. *Conf Proc IEEE Eng Med Biol Soc.* 2014;2014:406-409. doi:10.1109/EMBC.2014.6943614
- Fisher M, Feuerstein G, Howells DW, et al. Update of the stroke therapy academic industry roundtable preclinical recommendations. *Stroke*. 2009;40:2244-2250. doi:10.1161/ STROKEAHA.108.541128 (*Cont.*)