



ELSEVIER

Contents lists available at ScienceDirect

International Journal for Parasitology:  
Parasites and Wildlifejournal homepage: [www.elsevier.com/locate/ijppaw](http://www.elsevier.com/locate/ijppaw)

## Invited Review

A review of neosporosis and pathologic findings of *Neospora caninum* infection in wildlife

Shannon L. Donahoe, Scott A. Lindsay, Mark Krockenberger, David Phalen, Jan Šlapeta \*

Faculty of Veterinary Science, University of Sydney, Sydney, New South Wales, Australia

## ARTICLE INFO

## Article history:

Received 10 March 2015

Revised 16 April 2015

Accepted 17 April 2015

## Keywords:

*Neospora caninum*

Neosporosis

Wildlife

Nondomestic species

Clinical signs

Pathology

## ABSTRACT

*Neospora caninum* is an apicomplexan parasite that is the etiologic agent of neosporosis, a devastating infectious disease regarded as a major cause of reproductive loss in cattle and neuromuscular disease in dogs worldwide. This protozoan pathogen is maintained in the environment by a heteroxenous life cycle that involves a definitive canid host and a wide range of intermediate hosts. In recent years, a number of wildlife species have been investigated for their possible involvement in the *N. caninum* life cycle and many have been implicated as intermediate hosts. However, in many instances these studies have utilized serological and molecular techniques to detect infection in clinically normal animals, and investigation of possible associated morbidity, mortality, and pathology has been neglected. As such, the occurrence and importance of *Neospora*-associated disease in wildlife species are unknown. In order to improve our understanding of the significance of *N. caninum* infection in nondomestic species, the present review provides an up-to-date summary of clinical neosporosis and *N. caninum*-associated pathologic lesions in naturally and experimentally infected wildlife species. We provide a list of all free-ranging and captive wildlife species identified with *N. caninum* infection to date using currently available diagnostic tools. The advantages and disadvantages of diagnostic methods in wildlife are addressed in order to recommend optimal diagnosis of confirming *N. caninum* infection and neosporosis in nondomestic species. Although current data would suggest that *N. caninum* infection does not adversely impact wildlife populations, there is a need for greater international uniformity in the diagnosis of *N. caninum* infection and neosporosis in nondomestic species in order to assess the true consequences of parasite infection.

© 2015 The Authors. Published by Elsevier Ltd on behalf of Australian Society for Parasitology. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

## Contents

1. Introduction .....	217
2. Life cycle and transmission: domestic and sylvatic cycles .....	217
3. What is the outcome of infection in experimental models and is it relevant to neosporosis in wildlife? .....	219
3.1. Host immune response .....	219
3.2. Strain virulence and genetic variability .....	219
4. Neosporosis in domestic animals .....	220
4.1. Overview of disease in cattle .....	220
4.2. Overview of disease in dogs .....	220
5. Naturally acquired neosporosis and pathology in wildlife species .....	221
5.1. Eutherian mammals .....	221
5.1.1. Carnivora .....	221
5.1.2. Artiodactyla .....	223
5.1.3. Perissodactyla .....	225
5.1.4. Rodentia .....	225
5.1.5. Lagomorpha .....	226
5.1.6. Insectivora .....	226
5.1.7. Proboscidea .....	226

\* Corresponding author. Faculty of Veterinary Science, University of Sydney, Sydney, New South Wales, Australia. Tel.: +61 2 9351 2025; fax: +61 2 9351 7348.  
E-mail address: [jan.slapeta@sydney.edu.au](mailto:jan.slapeta@sydney.edu.au) (J. Šlapeta).

5.1.8.	Cetacea .....	226
5.1.9.	Pinnipedia .....	226
5.2.	Metatherian mammals .....	227
5.2.1.	Marsupialia .....	227
5.3.	Birds .....	227
6.	Experimentally acquired neosporosis and pathology in wildlife species .....	227
7.	Animals with no evidence of naturally or experimentally acquired <i>N. caninum</i> exposure .....	230
8.	Diagnostic tools used for detection of <i>N. caninum</i> in wildlife .....	230
8.1.	Isolation .....	230
8.2.	Histopathology and immunohistochemistry .....	230
8.3.	Ultrastructural analysis (transmission electron microscopy) .....	230
8.4.	Seroprevalence studies .....	230
8.5.	Conventional and nested PCR .....	231
8.6.	Quantitative PCR .....	231
8.7.	Mini- and microsatellite analysis – tools for genotyping .....	231
8.8.	Next generation sequencing (NGS) .....	232
9.	Neosporosis in wildlife: the right time for reflection .....	232
9.1.	What is the impact of <i>N. caninum</i> infection on wildlife populations? .....	232
9.2.	How often is <i>N. caninum</i> infection clinically relevant and what factors predispose to pathologic sequelae in nondomestic species? ...	232
9.3.	What are the best practice guidelines for reporting wildlife cases of neosporosis? .....	232
10.	Conclusions .....	233
	Acknowledgements .....	233
	Conflict of interest .....	234
	References .....	234

## 1. Introduction

*Neospora caninum* (Apicomplexa: Coccidia), the etiologic agent of the polysystemic disease neosporosis, is an obligate intracellular tissue cyst-forming coccidian parasite of the phylum Apicomplexa (Dubey et al., 2007; Dubey and Schares, 2011). *Neospora caninum* shares many morphologic and biologic features with its close relative *Toxoplasma gondii* (Dubey et al., 2002, 2007; Dubey and Schares, 2011). Prior to its initial recognition in Norwegian dogs in 1984 (Bjerkas et al., 1984) and consequential classification as a distinct species in 1988 (Dubey et al., 1988), many *N. caninum* infections were misdiagnosed as toxoplasmosis (Dubey et al., 2002; Dubey and Schares, 2011). Key differences were subsequently identified that distinguish the two parasites with regard to their natural host range, antigenicity, virulence factors, and pathogenesis (for reviews, see Dubey and Lindsay, 1996; Dubey et al., 2002; Dubey et al., 2007). Differences between *N. caninum* and *T. gondii* have also been described using comparative genomics and transcriptomics analyses (Reid et al., 2012). In the past two decades *N. caninum* has been extensively investigated due to its importance as a veterinary pathogen. As a result of these studies, it is now known that *N. caninum* has a global distribution and causes severe neuromuscular disease in dogs, and abortion and neonatal mortality in cattle, resulting in devastating economic losses to the beef and dairy industries (Dubey et al., 2007; Dubey and Schares, 2011; Reichel et al., 2013).

Less is known about the epizootiology and impact of this parasite in wildlife (reviewed by Gondim, 2006; Dubey et al., 2007; Dubey and Schares, 2011; Almeria, 2013). Most studies of *N. caninum* infection in wildlife species report on the prevalence of infection using serologic and/or molecular diagnostic assays in asymptomatic animals. While helpful in documenting evidence of exposure to the pathogen amongst wildlife species, these studies do not provide insight into the nature of the host–pathogen interactions in these potential intermediate hosts. In some instances, these analyses are also limited by the uncertainty regarding the sensitivity and specificity of the assays used.

This review provides a critical analysis of clinical neosporosis and related pathologic findings in free-ranging and captive wildlife species for which postmortem analyses of gross and microscopic lesions have been described. Building upon the current literature, this paper aims to improve our knowledge of the host–pathogen interactions in wildlife by (1) reviewing the prevalence of clinical

neosporosis as an outcome of infection with *N. caninum* in nondomestic species and the factors that predispose to pathologic sequelae, (2) examining our current understanding of the impact of *N. caninum* infection on wildlife populations, and (3) formulating best practice guidelines for documenting *N. caninum* infection and neosporosis in wildlife. *Neospora caninum* seroprevalence and molecular diagnostic studies in nondomestic species have been well reviewed (Gondim, 2006; Dubey et al., 2007; Dubey and Schares, 2011; Almeria, 2013) and, unless specifically associated with pathology or clinical disease, the details of these reviews will not be reiterated here.

## 2. Life cycle and transmission: domestic and sylvatic cycles

*Neospora caninum* is characterized by a complex facultative heteroxenous life cycle that involves a definitive canid host in which sexual replication occurs, and a range of intermediate hosts in which asexual replication takes place (Dubey and Lindsay, 1996; Dubey et al., 2006, 2007; Dubey and Schares, 2011). To date, the only confirmed definitive hosts of *N. caninum* are members of the *Canis* genus, including domestic and wild dogs (*Canis familiaris*) (McAllister et al., 1998), coyotes (*Canis latrans*) (Gondim et al., 2004b), gray wolves (*Canis lupus lupus*) (Dubey et al., 2011), and dingoes (*Canis lupus dingo*) (King et al., 2010). Cattle are the most common intermediate host of *N. caninum*; however, in recent years infection has been reported in many warm-blooded vertebrate species – some with the potential to serve as intermediate hosts in domestic and sylvatic transmission cycles (Gondim, 2006; Dubey et al., 2007; Dubey and Schares, 2011; Almeria, 2013). *Neospora caninum* is not considered to be zoonotic despite some serologic evidence of human exposure, particularly in immunocompromised populations (Tranas et al., 1999; Lobato et al., 2006; Barratt et al., 2010).

The *N. caninum* life cycle is characterized by three known infectious life stages: sporozoites within sporulated oocysts, rapidly dividing tachyzoites, and slowly proliferating bradyzoites within tissue cysts (Dubey et al., 2006). Light microscopic and ultrastructural morphology of these stages have been well described, with the notable exception of sporulated oocysts for which ultrastructural description is still lacking (Dubey and Lindsay, 1996; Speer et al., 1999; Dubey et al., 2002, 2006; Dubey, 2003). Oocysts are the environmentally resistant form of the parasite. They are presumably generated by sexual replication in the intestinal epithelial cells of

the definitive host and expelled in the feces in an unsporulated (non-infectious) form, although entero-epithelial sexual stages in the canine alimentary tract have yet to be conclusively demonstrated (Dubey et al., 2004). Outside the host, oocysts undergo sporulation in 24–72 hours and develop two sporocysts, each of which contains four sporozoites, which render them orally infectious (Dubey et al., 2006, 2007; Reichel et al., 2007). Host infection may ensue when sporulated oocysts are ingested. In the gastrointestinal tract, sporozoites are released by excystation and parasitize the intestine, where they transform into tachyzoites (Hemphill et al., 2006). Tachyzoites are capable of infecting a wide range of nucleated host cells, including mononuclear cells that likely aid in parasite dissemination via leukocyte trafficking (Dubey et al., 2006; Hemphill et al., 2006). Within host cells, tachyzoites reside and replicate within a parasitophorous vacuole, an intracellular compartment formed from the host cell membrane with parasite modifications that preclude its fusion with endocytic vesicles (Buxton et al., 2002; Hemphill et al., 2006). During the acute phase of infection, tachyzoites may be found in virtually all host tissues and it is during this stage that progressive cycles of intracellular replication, infected host cell lysis, tachyzoite release and infection of surrounding cells, and related immunopathologic sequelae initiate lesion formation and, in some animals, clinical disease (Dubey et al., 2007). In an immunocompetent host, tachyzoites replicate for an estimated 20 divisions before they differentiate into bradyzoites, the quiescent life stage of the parasite that forms under host immune pressure and produces a tissue cyst (Goodswen et al., 2013).

Tissue cysts shelter bradyzoites from host immunological factors and facilitate long term parasite persistence and chronic asymptomatic infections (Hemphill et al., 2006; Dubey et al., 2007; Dubey and Schares, 2011). Recrudescence of infection may transpire with changes in host immune status (immunomodulation or immunosuppression) that may result in reactivation of bradyzoites and conversion to tachyzoites (Hemphill et al., 2006). This is well documented to occur in pregnant animals and permits tachyzoite spread to other tissues, including dissemination across the placenta and infection of the unborn fetus (Williams et al., 2009).

Natural *N. caninum* infections occur via horizontal or vertical transmission (Dubey et al., 2007). Vertical transmission (also referred to as transplacental or congenital transmission) is the predominant mode of transmission in cattle and other domesticated bovine species. Two forms of vertical transmission are recognized: exogenous and endogenous transplacental transmission (Trees and Williams, 2005; Williams et al., 2009). Exogenous transplacental transmission ensues following ingestion of sporulated oocysts by naïve cattle and is associated with epidemic abortion storms in a herd (Williams et al., 2009). Endogenous transplacental transmission follows recrudescence of infection in a persistently infected cow during pregnancy and is the principle mechanism responsible for maintaining the parasite within cattle populations, resulting in fetal transmission rates as high as 95% (Dubey et al., 2006; Williams et al., 2009; Reichel et al., 2013). Once infected, a cow remains infected for life and may pass infection to consecutive generations of offspring (Dubey and Schares, 2006; Dubey et al., 2006). Seropositive cattle have an increased risk of abortion estimated to be between 1.7 and 7.4 fold (Thurmond and Hietala, 1997). However, abortion risk diminishes with increasing parity, suggesting that some degree of host immunity develops over time, impeding the success of endogenous transplacental transmission (Thurmond and Hietala, 1997; Dubey et al., 2006, 2007). While vertical transmission of the pathogen dominates in cattle and horizontal (postnatal) transmission rates are largely considered infrequent, theoretical mathematical models indicate that some degree of horizontal transmission (ingestion of oocysts from the definitive host) is required to perpetuate *N. caninum* infection in a herd (French et al., 1999; Reichel et al., 2013).

In the definitive canid host and other carnivores, horizontal transmission may result from ingestion of tissue from infected intermediate hosts containing tissue cysts, or from sporulated oocyst-contaminated food or water (Dubey et al., 2006, 2007). Dogs have been shown to shed oocysts following consumption of infected offal or expelled placental membranes from infected cows (Williams et al., 2009). Vertical transmission also occurs in dogs. Subclinically infected bitches can transmit *N. caninum* parasites to their offspring in successive litters; however, transplacental infection on its own is considered less important in sustaining infection in canine populations (Barber and Trees, 1996; Dubey, 2003; Dubey et al., 2005, 2006, 2007; Reichel et al., 2007; Dubey and Schares, 2011; Goodswen et al., 2013).

Experimentally there is evidence that lactogenic transmission occurs in calves, but this mode of transmission has not been demonstrated under natural circumstances in calves or dogs (Davison et al., 2001; Dijkstra et al., 2001; Dubey et al., 2007). *Neospora caninum* DNA, but not viable organisms, has been found in semen collected from naturally exposed bulls and although theoretically possible, venereal transmission via contaminated sperm is considered unlikely (Dubey et al., 2007).

Worldwide, studies indicate that the *N. caninum* life cycle is maintained in a domestic cycle between dogs and cattle (Dubey et al., 2007). They show a strong correlation between the presence of domestic dogs and the incidence of neosporosis on cattle farms (McAllister et al., 1998; Sawada et al., 1998; Wouda et al., 1999; Basso et al., 2001; Moore et al., 2002; Otranto et al., 2003; Hobson et al., 2005; Wanha et al., 2005; Bartels et al., 2007; Dubey et al., 2007; Malmasi et al., 2007; Collantes-Fernandez et al., 2008). Compelling evidence suggests a sylvatic cycle involving domestic and wild canids, and ruminant and herbivore species occur in North America (McAllister et al., 1998; Barling et al., 2000; Gondim et al., 2004b; Gondim, 2006). This is supported by several studies that report high *N. caninum* seroprevalence in North American white-tailed deer, the isolation of viable *N. caninum* from this host, and that dogs fed brain from experimentally infected white-tailed deer shed oocysts infective to cattle (reviewed by Dubey et al., 2013). White-tailed deer are extensively hunted in North America and because most are eviscerated where they are killed, infected deer tissues (e.g., offal) may provide an important source of infection for domestic and wild canid definitive hosts (Dubey et al., 2007). Additionally, North American beef herds grazed in close proximity to coyotes have a higher risk of *N. caninum* infection, signifying that transmission may be occurring between these species (Barling et al., 2000; Gondim et al., 2004b; Dubey et al., 2007). Identification of seropositive cattle from areas where domestic dogs are not known to be present provides additional evidence to support the possibility of a sylvatic transmission cycle between wild canids and cattle (McAllister et al., 1998; Ferroglio et al., 2003; Gondim et al., 2004b).

Less is known about the existence of sylvatic cycles in other countries. In Australia, a sylvatic cycle between dingoes and macropod species has been postulated by King et al. (2011b). Indirect support for this hypothesis is provided by a recent serological survey of western grey kangaroos (*Macropus fuliginosus ocydromus*) from Western Australia that demonstrated a *N. caninum* seroprevalence of 18% (Mayberry et al., 2014), and by experimental *N. caninum* infection in the fat-tailed dunnart (*Sminthopsis crassicaudata*) showing it to be a highly susceptible intermediate host (King et al., 2011a). In Europe, similar sylvatic cycles involving wild ruminants and canids seem plausible but little evidence currently exists to support this hypothesis. One study conducted in Hungary showed farm dogs consuming potentially infected offal from aborted or dead calves, or raw game animals demonstrated a high rate of *N. caninum* seropositivity (Hornok et al., 2006). Furthermore, *N. caninum* antibodies and/or DNA have been detected in a number of European free-ranging carnivore and non-carnivore wildlife species (reviewed by Almeria, 2013).

### 3. What is the outcome of infection in experimental models and is it relevant to neosporosis in wildlife?

Reports of *N. caninum* infection in domestic and nondomestic animals appear to be common, yet there are relatively few reports of clinical neosporosis in any wildlife species (Dubey and Schares, 2011; Goodswen et al., 2013). The reason for this discrepancy is not well understood. Extrapolating from what is known about the more intensively studied *T. gondii*, multiple different host (species, genotype, age, and immune status) and parasite (dose, strain, life stage initiating infection, and route of infection) factors may account for the wide variation in clinical manifestations observed in *N. caninum* infection (Dubey, 2010). In recent years, there has been increasing interest in clarifying the role of the host immune response and parasite strain on infection outcome in order to better understand components influencing control of infection and disease pathogenesis.

#### 3.1. Host immune response

In concordance with other intracellular pathogens, data from numerous *in vitro* and *in vivo* immunological studies in cattle and mice collectively indicate that protective host immunity induced by *N. caninum* infection is typified by a T helper 1 (Th1)-type response. In the acute stage of infection, this is mediated largely by interferon (IFN)- $\gamma$  production from natural killer (NK) cells, and interleukin (IL)-12 primed CD4+ lymphocytes and CD8+ lymphocytes (for reviews, see Innes et al., 2002; Innes et al., 2005; Hemphill et al., 2006; Klevar et al., 2007; Monney and Hemphill, 2014). This pro-inflammatory Th1-type response appears to be essential for restricting parasite replication and induction of latent infection through stage conversion and formation of tissue cysts (Buxton et al., 2002; Innes et al., 2002; Dubey et al., 2006; Hemphill et al., 2006; Williams et al., 2009; Goodswen et al., 2013). In contrast, host immunomodulation, as may occur during pregnancy, favors a bias towards a Th2-type immune response characterized by increased expression of IL-4, and is associated with a bradyzoite to tachyzoite transformation (recrudescence), uncontrolled parasite replication, and corresponding pathologic effects (Buxton et al., 2002; Innes et al., 2002, 2005; Dubey et al., 2006; Hemphill et al., 2006; Williams et al., 2009; Goodswen et al., 2013). Humoral (antibody) immunity probably participates in controlling *N. caninum* infection via mechanisms involving antibody-neutralization of extracellular tachyzoites (Innes et al., 2002; Hemphill et al., 2006; Bartley et al., 2013a; Goodswen et al., 2013).

Immunological studies may be crucial to understanding the pathogenesis of neosporosis and outcome of parasite infection, but investigations of host immunity are challenging. The host immune response to *N. caninum* has been exclusively studied using bovine and murine experimental models, and while valuable for investigating the role of the host immune response, the true significance of findings obtained in these models can only be conclusively assessed in other host species. In many instances, by necessity, experiments have used tachyzoites to infect animals parenterally rather than the natural enteral route of infection by sporocysts or sporozoites because of the extreme difficulty in obtaining *N. caninum* oocysts from experimentally infected dogs (Dubey et al., 2007; Dubey and Schares, 2011). While parasite life stage and mode of infection have been shown to impact the host immune response to *T. gondii* infection, whether or not these factors also modulate the host immune response to *N. caninum* infection is uncertain and necessitates additional investigation (Dubey, 2010). Some of the outstanding questions in the field of the *N. caninum* immunobiology involve understanding how various components of the host immune system interact with each other and with *N. caninum* infected cells to control parasite replication.

The understanding of why different hosts and individuals are more susceptible to neosporosis is growing. It has long been recognized that there is significant variation in species susceptibility to *N. caninum* infection. For example, gerbils are experimentally highly susceptible to developing clinical neosporosis while outbred mice are largely resistant to infection (Pipano et al., 2002). The reason for this difference is poorly understood, although presumably interspecies variation in immunity plays an important role because chemical immunosuppression (i.e., prednisolone treatment) of resistant mice produces disease (Lindsay and Dubey, 1989). Recent studies have provided evidence that intraspecific variations in host immunity can influence outcome of *N. caninum* infection. Differences in the humoral immune response have been described for different breeds of cattle (Santolaria et al., 2011) and strains of mice (Mols-Vorstermans et al., 2013) and, in the latter case, were linked to specific major histocompatibility complex (MHC) class I haplotypes. A retrospective study of Holstein cattle found MHC class II alleles DRB3\*1001 and DRB3\*2703 to be associated with pregnancy loss and resistance, respectively, regardless of serological status (Schwab et al., 2009). Advances in molecular techniques in recent years have enabled genetic characterization of genes associated with the host immune response, and consequently intra- and interspecific differences in innate and adaptive immune responses have been elucidated for a number of species (Litman et al., 2010; Belov et al., 2013; Buchmann, 2014). However, more research is needed to assess how these factors influence the host–pathogen interactions between *N. caninum* infection and wildlife.

#### 3.2. Strain virulence and genetic variability

Significant variations in pathogenicity (virulence) and growth rates for different *N. caninum* isolates have been demonstrated in experimentally infected mice, gerbils, sheep and cattle (for review, see Al-Qassab et al., 2010). For example, Nc-Liverpool-infected mice exhibit more severe central nervous system inflammation and necrosis than mice infected with Nc-Nowra and Nc-SweB1 (Atkinson et al., 1999; Miller et al., 2002), and infection of pregnant cows with NC1 produces fetal death while no fetal death is observed in Nc-Spain 1H-infected cows (Rojo-Montejo et al., 2009). There is also evidence that different strains of *N. caninum* can influence the host immune response and subsequently infection outcome (Atkinson et al., 1999; Costa et al., 2008; Al-Qassab et al., 2010; Dellarupe et al., 2014; Regidor-Cerrillo et al., 2014). Some studies have shown differential immunoglobulin expression in mice and heifers infected with different parasite isolates and, although this appears to be linked to infection dynamics and clinical outcome, a direct causal relationship has yet to be identified (Atkinson et al., 1999; Costa et al., 2008; Rojo-Montejo et al., 2009; Dellarupe et al., 2014).

Interpretation of comparative *N. caninum* strain virulence studies is confounded by several factors which may affect pathogenicity including the parasite life stage used to initiate infection, route of inoculation, inoculum dose, and possible mitigation of virulence and other biological characteristics in cell culture derived parasites (Dubey et al., 2007; Al-Qassab et al., 2010; Dubey and Schares, 2011). As such, direct comparison between the information gleaned from these studies is often not possible, and whether or not virulence studies in mice and cattle will mimic the effect of *N. caninum* infection in other host species requires further investigation.

Most virulence studies have been conducted using isolates from clinically affected animals or neonatal infected animals, and little is known about the differences between virulent and avirulent strains, although as a result of genotyping technology this is beginning to change (Rojo-Montejo et al., 2009; Al-Qassab et al., 2010; Dubey and Schares, 2011; Regidor-Cerrillo et al., 2013). Increasingly, investigations are focusing on identifying genetic components that may impact strain pathogenicity and disease manifestations

(Weiss et al., 1999; Gondim et al., 2004c; McInnes et al., 2006; Al-Qassab et al., 2010; Goodswen et al., 2013). In recent years, multilocus mini- and microsatellite analyses have proven to be useful tools for the genetic characterization of many protozoan organisms, including *T. gondii*, and are currently considered the gold standard for *N. caninum* genotyping (Al-Qassab et al., 2010; King et al., 2012; Goodswen et al., 2013). To date, over 100 different *N. caninum* strains have been analyzed using mini- and microsatellite technology, and data show that this parasite exists as a diverse heterogeneous population and that different strains exhibit extensive genetic diversity worldwide despite sharing similar light microscopic and ultrastructure features (Regidor-Cerrillo et al., 2006, 2013; Basso et al., 2009, 2010; Al-Qassab et al., 2010; Goodswen et al., 2013). An association between this genetic diversity of *N. caninum* isolates and their pathogenicity has not been conclusively demonstrated and it remains unclear if the genetic structure of the parasite is a determinant of host clinical manifestations (Al-Qassab et al., 2010; Goodswen et al., 2013; Regidor-Cerrillo et al., 2013).

Mini- and microsatellite genotyping techniques have been successively applied to epidemiological and population genetic investigations of strains of *N. caninum* in domestic animals (Regidor-Cerrillo et al., 2006, 2013; Basso et al., 2009, 2010; Al-Qassab et al., 2010). Genotyping technology has rarely been applied to wildlife *N. caninum* strains but it has potential to provide valuable insight for future epidemiological investigations in captive and free ranging nondomestic species, establishing the importance of sylvatic and domestic cycles in neosporosis, and evaluating the emergence of new *N. caninum* isolates in wildlife populations.

#### 4. Neosporosis in domestic animals

##### 4.1. Overview of disease in cattle

*Neospora caninum* is a primary pathogen in cattles and abortion is the principal clinical manifestation (Dubey, 2003; Dubey et al., 2007; Dubey and Schares, 2011). Apart from fetal loss, morbidity and mortality are generally not observed in infected cows (Dubey et al., 2006, 2007). The majority of fetuses in seropositive dams will be infected via transplacental transmission and infection outcome is largely dependent on stage of gestation and therefore fetal age (Buxton et al., 2002; Innes et al., 2002; Dubey et al., 2006, 2007; Dubey and Schares, 2011; Goodswen et al., 2013). If infection occurs during the first 100 days of gestation when fetal lymphoid tissues are developing, survival is unlikely and fetal resorption or mummification results (Dubey, 2003; Dubey et al., 2006, 2007; Dubey and Schares, 2011). During the second and third trimesters, the fetus has an increasing capacity to mount an immune response but in many instances it appears insufficient as abortion is the most common sequela of infection in mid-gestation, particularly between 5 and 7 months (Buxton et al., 2002; Reichel et al., 2013). If infection occurs in the third trimester after fetal immunity is more developed, the birth of an infected but clinically normal calf is the most likely outcome (Buxton et al., 2002; Innes et al., 2005; Dubey et al., 2006), although birth of uninfected calves may infrequently occur (Gibney et al., 2008).

Aborted fetuses are predominately autolyzed but may also be expelled fresh or mummified and generally do not exhibit specific gross lesions (Dubey, 2003; Dubey et al., 2006). In some cases, fetal necropsy may reveal hydrocephalus, cerebellar and medulla hypoplasia, and scattered pale to dark foci in the brain, spinal cord, heart, and skeletal muscle that correlate histologically with areas of necrosis and inflammation (Dubey, 2003; Dubey et al., 2006; Schlafer and Miller, 2007). The most significant and distinctive histologic lesions of fetal neosporosis are found in the central nervous system (CNS) and consist of multifocal nonsuppurative encephalomyelitis with perivascular cuffs, necrosis, and microgliosis (Helman et al.,

1998; Slapeta et al., 2003; Dubey et al., 2006). In addition, multifocal nonsuppurative myocarditis and myositis, and nonsuppurative periportal hepatitis with variable hepatic necrosis can be observed (Helman et al., 1998; Dubey, 2003; Dubey and Schares, 2006; Dubey et al., 2006). Nonsuppurative interstitial nephritis, interstitial pneumonia, and adrenal necrosis are reported less frequently (Barr et al., 1991; Dubey et al., 2006; Schlafer and Miller, 2007; Nishimura et al., 2013). The presence of intralésional tissue cysts and/or tachyzoites is inconsistent and may be challenging to detect with standard hematoxylin and eosin (H&E) stains; the use of *N. caninum* specific immunohistochemistry (IHC) greatly facilitates parasite identification (Barr et al., 1991; Dubey and Schares, 2006; Dubey et al., 2006). Protozoan cysts and tachyzoites are most likely to be found in the brain and spinal cord, if present (Dubey et al., 2006; Nishimura et al., 2013). Intracellular tachyzoites can also be recognized in myocytes and myocardial Purkinje fibers (Dubey et al., 1990, 2006; Barr et al., 1991). The placenta may be histologically normal or contain areas of cotyledonary necrosis associated with mononuclear inflammation and occasional clusters of tachyzoites within trophoblasts (Dubey et al., 1990, 2006; Schlafer and Miller, 2007).

Clinically affected congenitally infected calves may exhibit one or more of the following features: small size for gestational age, ataxia, proprioceptive deficits, inability to stand, hyperextension or flexion of forelimbs and/or hindlimbs, exophthalmia, scoliosis, hydrocephalus, and spinal cord narrowing (Dubey, 2003; Dubey et al., 2006; Schlafer and Miller, 2007). Mononuclear encephalomyelitis is the predominant histologic finding (Dubey and Schares, 2006; Dubey et al., 2006). CNS lesions are more frequently detected in the spinal cord than the brain, and tissue cysts are more common than tachyzoites (Dubey et al., 2006). Extraneural lesions are rare but may include myositis, nephritis, and pneumonia (Dubey et al., 2006).

##### 4.2. Overview of disease in dogs

Neosporosis most frequently manifests as neuromuscular disease in young dogs, but dogs of any age may be affected (Ruehlmann et al., 1995; Barber and Trees, 1996; Buxton et al., 2002; Dubey, 2003; Reichel et al., 2007). Two main neurological presentations are recognized: polymyositis-polyradiculoneuritis and encephalomyelitis (Barber and Trees, 1996; Reichel et al., 2007).

Protozoal polymyositis-polyradiculoneuritis is most commonly seen in transplacentally infected young dogs 5 weeks to 6 months of age and may involve multiple littermates (Barber and Trees, 1996; Buxton et al., 2002; Reichel et al., 2007). *Neospora caninum* has a predilection for the lumbosacral spinal nerve roots in young dogs, and consequently pelvic limb muscle atrophy and arthrogryposis develop (Reichel et al., 2007; Garosi et al., 2010). Clinical signs include ascending hindlimb paralysis, muscle atrophy, rigid hyperextension of the pelvic limb, cervical weakness, and dysphagia (Buxton et al., 2002; Dubey, 2003; Reichel et al., 2007; Garosi et al., 2010). Histopathology may reveal nonsuppurative polyradiculoneuritis and polymyositis with intralésional tachyzoites and cysts (Barber and Trees, 1996; Dubey, 2003; Reichel et al., 2007).

In contrast, protozoal encephalomyelitis is more likely to affect adult dogs and is characterized by a more variable clinical presentation that ranges from localized CNS signs to widespread CNS involvement and disseminated disease (Barber and Trees, 1996; Reichel et al., 2007). Reported neurological clinical signs include hind limb paresis and paralysis, head tilt, seizures, ataxia, dysphagia, incontinence, seizures, and ocular abnormalities including miosis, diminished pupillary light reflexes, anisocoria, and enophthalmos (Barber and Trees, 1996; Buxton et al., 2002; Garosi et al., 2010). Gross necropsy lesions are infrequently described, although cerebellar atrophy has been noted in a few cases (Jackson et al., 1995; Cantile and Arispici, 2002; Lorenzo et al., 2002; Garosi et al., 2010).

The most characteristic histopathologic lesion of protozoal encephalomyelitis is a multifocal nonsuppurative variably necrotizing meningoencephalomyelitis with tachyzoites and tissue cysts occasionally identified in neurons and neuropil (Barber and Trees, 1996; Dubey, 2003; Dubey et al., 2006; Brown et al., 2007; Reichel et al., 2007).

Although principally recognized as a neuromuscular disease in dogs, neosporosis can induce a variety of less common lesions including myocarditis, polymyositis, pancreatitis, and interstitial pneumonia with pulmonary edema and alveolitis, depending on the cells parasitized (Barber and Trees, 1996; Buxton et al., 2002; Garosi et al., 2010). Additionally, dermal neosporosis is becoming increasingly recognized as a unique presentation of *N. caninum* infection (Dubey and Lindsay, 1996; La Perle et al., 2001; McInnes et al., 2006; Dubey and Schares, 2011; Dubey et al., 2014b). It has been postulated that the cutaneous manifestation of disease may be precipitated by underlying immunosuppression, possibly related to drug therapy or concurrent disease (La Perle et al., 2001; Buxton et al., 2002; Ordeix et al., 2002; Boyd et al., 2005; McInnes et al., 2006; Dubey et al., 2014b). Macroscopic lesions are those of a multifocal to generalized ulcerative and nodular dermatitis characterized histologically by pyogranulomatous inflammatory infiltrates with variable admixed eosinophilic infiltrates, necrosis and hemorrhage (La Perle et al., 2001; Ordeix et al., 2002; Boyd et al., 2005). Myriad tachyzoites are observed in macrophages, keratinocytes, and neutrophils, and to a lesser extent, endothelial cells and fibroblasts (La Perle et al., 2001; Boyd et al., 2005; Ginn et al., 2007). Tissue cysts are not recognized in lesions of cutaneous neosporosis (La Perle et al., 2001; Ordeix et al., 2002; Boyd et al., 2005; Ginn et al., 2007).

## 5. Naturally acquired neosporosis and pathology in wildlife species

### 5.1. Eutherian mammals

#### 5.1.1. Carnivora

Carnivores have been extensively investigated for shedding *N. caninum* oocysts, implying a potential role as definitive hosts of *N. caninum*. Naturally occurring oocysts of *N. caninum* have been identified only in North American coyotes (*Canis latrans*) (Gondim et al., 2004b; Wapenaar et al., 2006), domestic dog–dingo hybrids living in remote Aboriginal communities in Australia (King et al., 2010, 2012), and gray wolves (*C. lupus lupus*) (Dubey et al., 2011).

In addition to the above confirmed definitive hosts (Barber and Trees, 1996; Lindsay et al., 1996; Gondim et al., 2004a, 2004b; Dubey and Thulliez, 2005; Steinman et al., 2006; Wapenaar et al., 2007; Sobrino et al., 2008; Almborg et al., 2009; Bjorkman et al., 2010; Stevie et al., 2010; Bevins et al., 2013; Dubey et al., 2014a), *N. caninum* antibodies (Ab) and/or DNA have been detected in the following free-ranging carnivores: Iberian wolf (*Canis lupus signatus*) (Ab) (Sobrino et al., 2008), golden jackal (*Canis aureus*) (Ab) (Steinman et al., 2006), African wild dogs (*Lycaon pictus*) (Ab) (Woodroffe et al., 2012), red fox (*Vulpes vulpes*) (Ab, DNA) (Barber et al., 1997; Buxton et al., 1997; Simpson et al., 1997; Schares et al., 2001; Wolfe et al., 2001; Almeria et al., 2002; Hamilton et al., 2005; Hurkova and Modry, 2006; Steinman et al., 2006; Jakubek et al., 2007; Murphy et al., 2007; Wapenaar et al., 2007; Marco et al., 2008; Sobrino et al., 2008; De Craeye et al., 2011; Bartley et al., 2013b; Stuart et al., 2013; Dubey et al., 2014b), Culpeo fox (*Dusicyon culpaeus*) (Ab) (Martino et al., 2004), South American gray fox (*Dusicyon griseus*) (Ab) (Martino et al., 2004), North American gray fox (*Urocyon cinereoventus*) (Ab) (Lindsay et al., 1996), Azara's fox (*Lycalopex gymnocercus*) (Ab) (Canon-Franco et al., 2004), crab-eating fox (*Cerdocyon thous*) (Ab) (Canon-Franco et al., 2004), European brown bear (*Ursus arctos*) (DNA) (Cobadiova et al., 2013), spotted hyena (*Crocuta crocuta*) (Ab) (Ferroglio et al., 2003), raccoon (*Procyon lotor*) (Ab, DNA) (Lindsay

et al., 2001; Lemberger et al., 2005), raccoon dog (*Nyctereute procyonoides*) (Ab) (Kim et al., 2003), stone martin (*Martes foina*) (Ab) (Sobrino et al., 2008), pine martin (*Martes martes*) (Ab) (Sobrino et al., 2008), Eurasian badger (*Meles meles*) (Ab, DNA) (Sobrino et al., 2008; Bartley et al., 2013b), polecat (*Mustella putorius*) (Ab, DNA) (Sobrino et al., 2008; Bartley et al., 2013b), ferret (*Mustela furo*) (DNA) (Bartley et al., 2013b), American mink (*Neovison vison*) (Ab, DNA) (Bartley et al., 2013b; Stuart et al., 2013), European otter (*Lutra lutra*) (DNA) (Stuart et al., 2013), sea otter (*Enhydra lutris nereis*) (Dubey et al., 2003; Miller et al., 2010) common genet (*Genetta genetta*) (Ab) (Sobrino et al., 2008), Egyptian mongoose (*Herpestes ichneumon*) (Ab) (Sobrino et al., 2008; Millan et al., 2009), Eurasian wild cat (*Felis silvestris silvestris*) (Ab) (Sobrino et al., 2008), Iberian lynx (*Lynx pardinus*) (Ab) (Sobrino et al., 2008), cheetah (*Acinonyx jubatus*) (Ab) (Cheadle et al., 1999; Ferroglio et al., 2003), and lion (*Panthera leo*) (Ab) (Cheadle et al., 1999; Ferroglio et al., 2003). Viable *N. caninum* tachyzoites (isolates NcWolfMn1 and NcWolfMc2) have been recently isolated from the brains of two free-ranging gray wolves (Dubey et al., 2014a).

*Neospora caninum* antibodies have been detected in the following captive wild carnivores: European wolf (*Canis lupus lupus*) (Sedlak and Bartova, 2006; Andre et al., 2010), maned wolf (*Chrysocyon brachyurus*) (Vitaliano et al., 2004; Silva et al., 2005; Sedlak and Bartova, 2006; Andre et al., 2010), bush dog (*Speothos venaticus*) (Mattos et al., 2008; Andre et al., 2010), crab-eating fox (*Cerdocyon thous*) (Andre et al., 2010), hoary fox (*Pseudalopex vetulus*) (Andre et al., 2010), blue fox (*Alopex lagopus*) (Yu et al., 2009), Darwin's fox (*Pseudalopex fulvipes*) (Sedlak and Bartova, 2006), fennec fox (*Vulpes zerda*) (Sedlak and Bartova, 2006), fisher (*Martes pennanti*) (Sedlak and Bartova, 2006), red panda (*Ailurus fulgens*) (Qin et al., 2007), cheetah (*Acinonyx jubatus*) (Sedlak and Bartova, 2006), jaguarondi (*Puma yagouaroundi*) (Sedlak and Bartova, 2006; Andre et al., 2010), puma (*Puma concolor*) (Andre et al., 2010), jaguar (*Panthera onca*) (Andre et al., 2010), Indian lion (*Panthera leo goojratensis*) (Sedlak and Bartova, 2006), lion (*Panthera leo*) (Kamga-Waladjo et al., 2009; Andre et al., 2010), tiger (*Panthera tigris*) (Andre et al., 2010), Eurasian lynx (*Lynx lynx*) (Sedlak and Bartova, 2006), ocelot (*Leopardus pardalis*) (Andre et al., 2010), little spotted cat (*Leopardus tigrinus*) (Andre et al., 2010), Pampas cat (*Oncifelis colocolo*) (Andre et al., 2010), caracal (*Caracal caracal*) (Andre et al., 2010), serval (*Letailurus serval*) (Andre et al., 2010), and fishing cat (*Prionailurus viverrinus*) (Andre et al., 2010).

There are only four reports of *N. caninum* infection in wild and/or wild captive carnivores where *N. caninum* parasites were detected histologically, three of which were associated with clinical neosporosis (van der Hage et al., 2002; Lemberger et al., 2005; Yu et al., 2009; Dubey et al., 2014b) (Table 1). Disease in nondomestic carnivores has only been described in young animals and although clinical manifestations and pathologic lesions vary among these cases, similar neurologic and dermatologic presentations have been reported in domestic dogs.

Van der Hage et al. (2002) described a case of fatal neosporosis in a free-ranging juvenile European pine marten (*Martes martes*) (family Mustelidae) from the Netherlands (Table 1). The affected 7-week old pup was taken into care by wildlife rehabilitators following dam abandonment and initially presented with small body size, dehydration, diarrhea, and forelimb and hindlimb ataxia. Over the next two weeks, the pup developed a cough that progressed to dyspnea and tachycardia. Imaging demonstrated cardiomegaly, severely diminished myocardial contractility, and intra-abdominal fluid. The animal was euthanized, and a postmortem examination revealed cardiac biventricular dilation and multiple pale foci scattered throughout the liver. The heart, liver, and lung were evaluated histologically. Microscopic lesions included multifocal nonsuppurative myocarditis with cytoplasmic tachyzoites evident in some mononuclear cells, extensive bridging hepatic necrosis with hydropic

**Table 1**  
Reports of clinically significant neosporosis in wildlife species.

Species	Wild or captive	Age, number	Con-current disease	Tests	Tachyzoites	Tissue-cysts	Animal location	Reference
Order Carnivora Family Mustelidae European Pine Marten ( <i>Martes martes</i> )	Wild <sup>a</sup>	Juvenile, n = 1	No	HP, IHC	Yes	No	The Netherlands (undisclosed location)	van der Hage et al., 2002
Family Canidae Red Fox ( <i>Vulpes vulpes</i> )	Wild <sup>b</sup>	Juvenile, n = 1	Yes <sup>c</sup>	HP, IHC, S, TEM, ISO (neg <sup>d</sup> )	Yes	No	USA (New Jersey)	Dubey et al., 2014a Dubey et al., 2014b
Blue Fox ( <i>Alopex lagopus</i> )	Captive	Newborn, n = 1	No	HP, IHC, S, PCR/Seq	Yes	Yes	China (Hebei province)	Yu et al., 2009
Order Artiodactyla Family Cervidae California Black-Tailed Deer ( <i>Odocoileus hemionus columbianus</i> )	Wild	Juvenile, n = 1	No	HP, IHC	Yes	No	USA (California)	Woods et al., 1994
Fallow deer ( <i>Dama dama</i> )	Captive	Juvenile, n = 1	No	HP, IHC, PCR/Seq	Yes	Yes	Switzerland (undisclosed location)	Soldati et al., 2004
Eld's deer ( <i>Cervus eldi siamensis</i> )	Captive	Stillborn (full-term), n = 1	No	HP, IHC	No	Yes	France (Paris)	Dubey et al., 1996
Axis deer ( <i>Axis axis</i> )	Captive	Neonates, n = 5	No	HP, S, PCR/Seq, ISO, MS	No/NR (1/4)	Yes/NR (1/4)	Argentina (La Plata)	Basso et al., 2014
Family Bovidae Lesser kudu ( <i>Tragelaphus imberbis</i> )	Captive	Stillborn (full term), n = 3	No	HP, IHC (neg), S, PCR	No	No	Germany (Hannover)	Peters et al., 2001
Order Perissodactyla Family Rhinocerotidae White rhinoceros ( <i>Ceratotherium simum</i> )	Captive	Neonate, n = 1	No	HP, TEM	Yes	Yes	South Africa (Lichtenburg)	Williams et al., 2002
	Captive	Adult, n = 1	No	HP, IHC, PCR	Yes	No	Thailand (Chonbari)	Sommanustweechai et al., 2010
	Captive	Fetus, n = 1	No	HP, IHC, PCR/Seq, MS	Yes	No	Australia (New South Wales)	Sangster et al., 2010
Order Marsupialia Family Macropodidae Parma wallaby ( <i>Macropus parma</i> )	Captive	Adult, n = 1	No	HP, IHC, PCR/Seq	Yes	Yes	Austria (Vienna)	Cronstedt-Fell et al., 2012

<sup>a</sup> Free-ranging; maintained in a wildlife rehabilitation center for 3 weeks following dam abandonment.

<sup>b</sup> Free-ranging; maintained 2 weeks in a wildlife rehabilitation center while recovering from vehicular trauma before development of disease.

<sup>c</sup> Concurrent subclinical *T. gondii* infection.

<sup>d</sup> Parasite isolation attempted 26d post clindamycin treatment.

NR, not reported; HP, histopathology; IHC, immunohistochemistry; S, serology; EM, electron microscopy; TEM, transmission electron microscopy; ISO, parasite isolation; PCR, polymerase chain reaction; PCR/Seq, PCR with sequencing; SUB, genotyping; MS, microsatellite analysis; NAT, Neospora agglutination test (direct agglutination test); IFAT, indirect fluorescent antibody tests; IB, immunoblotting.

degeneration of remaining hepatocytes (presumed sequelae of congestive heart failure), and increased numbers of pulmonary alveolar macrophages. Tissue cysts of *N. caninum* were not reported. The diagnosis of neosporosis was based on the detection of *N. caninum* antigen (disseminated tachyzoites) by IHC in all tissues evaluated; parasites were negative for *T. gondii* antigen by IHC. Details of antibiotics used for IHC were not provided.

Yu et al. (2009) reported an outbreak of neosporosis in a group of newborn farm-bred blue foxes (*Alopex lagopus*) (family: Canidae) in China displaying a high rate of morbidity and mortality (Table 1). Of more than 200 pups born during the outbreak, over 60% were affected and of those, over 50% died. Signs included coarse fur, inappetence, emaciation, fever, depression, ataxia, and paralysis. Five animals euthanized for necropsy examination had consistent macroscopic lesions of hydrocephalus and coarse pia mater. Scattered splenic infarcts and foci of renal necrosis and capsular rugosity were also observed. The most significant histologic lesions consisted of multifocal nonsuppurative perivascular encephalitis and marked congestion, and small foci of necrosis in the cerebrum, cerebellum, and spinal cord. Inflammatory foci in the brain were often associated with protozoan cysts and less commonly clustered tachyzoites. Protozoan cysts presumed to be *N. caninum* tissue cysts were also found within renal tubules in two of four foxes with Periodic Acid Schiff (PAS) staining, although parasite identity was not confirmed by IHC. Additional changes were not described. Immunohistochemical analysis using canine and murine polyclonal antibodies raised against

*N. caninum* and *T. gondii*, respectively, revealed the parasites in the brain were positive for *N. caninum* antigen and negative for *T. gondii* antigen. PCR of brain for the *N. caninum* specific Nc5 region was positive in four of five foxes and confirmed by sequencing. Some foxes (28/103 tested) were seropositive for *N. caninum* (c-ELISA, VMRD Laboratories, Pullman, Washington, USA).

Dubey et al. (2014b) reported a case of cutaneous neosporosis in a free-ranging juvenile red fox (*Vulpes vulpes*) (family: Canidae) with concurrent subclinical *T. gondii* infection from New Jersey, USA (Table 1). The fox was in care for two weeks while recovering from vehicular trauma before developing generalized crusty dermal lesions on the ventral thorax, face, and limbs. Histologic examination of a skin biopsy demonstrated marked granulomatous dermatitis with severe parakeratotic hyperkeratosis and neutrophilic crusting, necrotic keratinocytes associated with lymphocytic and neutrophilic infiltrates, epidermal hyperplasia, and myriad individual and clustered protozoan tachyzoites in the epidermis, particularly within keratinocytes. The fox had antibodies to both *T. gondii* (modified agglutination test (MAT), titer 1:3200) and *N. caninum* (Neospora agglutination test, NAT; titer 1:25). Transmission electron microscopy (TEM) of skin lesions revealed tachyzoites with electron dense rhoptries, consistent with *N. caninum*, and the tachyzoites were positive for *N. caninum* antigen and negative for *T. gondii* antigen by IHC using polyclonal rabbit anti-*N. caninum* and anti-*T. gondii* antibodies. Dermal lesions resolved with clindamycin treatment and parasites were not identified with histologic evaluation of skin

**Table 2***Neospora caninum* tissue cysts identified histologically in wildlife species without associated pathologic changes.

Species	Wild or captive	Age	Concurrent disease	Tests	Cysts/Tachyzoites	Animal location	Reference
Order Carnivora Family Procyonidae Raccoon ( <i>Procyon lotor</i> )	Wild	Juvenile	Yes <sup>a</sup>	HP, IHC (PC anti-Nc Ab of unreported sp.)	Yes/No	USA (Illinois)	Lemberger et al, 2005
Order Rodentia Family Muridae Brown rat ( <i>Rattus norvegicus</i> )	Wild	NR	No	HP, IHC, PCR	Yes/No	Mexico (Aguascalientes)	Medina-Esparza et al, 2013
House mouse ( <i>Mus musculus</i> )	Wild	NR	No	HP, IHC, PCR	Yes/No	Mexico (Aguascalientes)	Medina-Esparza et al, 2013
Family Sciuridae Rock squirrel ( <i>Spermophilus variegates</i> )	Wild	NR	No	HP, IHC, PCR	Yes/No	Mexico (Aguascalientes)	Medina-Esparza et al, 2013
Class Aves Order Psittaciformes; family Psittidae Red-and-green macaw ( <i>Ara chloropterus</i> )	NR	NR	No	HP, IHC, S <sup>b</sup>	Yes/No	Brazil	Mineo et al, 2011
Blue-fronted Amazon parrot ( <i>Amazona aestiva</i> )	NR	NR	No	HP, IHC, S <sup>b</sup>	Yes/No	Brazil	Mineo et al, 2011

<sup>a</sup> Canine distemper virus infection.<sup>b</sup> Seronegative by IFAT.

NR, not reported; HP, histopathology; IHC, immunohistochemistry; S, serology; PCR, polymerase chain reaction; IFAT, indirect fluorescent antibody test; Cyst, tissue cyst; Tachy, tachyzoites.

biopsies after 4 weeks of treatment, although viable *T. gondii* (but not *N. caninum*) was isolated from skin biopsies at that time.

Lemberger et al. (2005) identified a single protozoan cyst in the brain of a juvenile free-ranging raccoon (*Procyon lotor*) (family: Procyonidae) euthanized due to canine distemper virus infection from Cook County, Illinois, USA (Table 2). Immunohistochemical staining was performed using polyclonal antibodies and the protozoan cyst was positively labeled with rabbit anti-*N. caninum* antibodies but not anti-*T. gondii* antibodies. Nc5 PCR of DNA derived from the brain confirmed the diagnosis of *N. caninum* infection. The protozoan cyst was considered an incidental finding.

### 5.1.2. Artiodactyla

*Neospora caninum* antibodies and/or DNA have been detected in the following free-ranging nondomestic artiodactyls: white-tailed deer (*Odocoileus virginianus*) (Ab) (Dubey et al., 1999, 2009; Lindsay et al., 2002; Gondim et al., 2004a; Anderson et al., 2007; Gutierrez-Exposito et al., 2012; Olamendi-Portugal et al., 2012), black-tailed deer (*Odocoileus hemionus columbianus*) (Ab, DNA) (Woods et al., 1994; Dubey et al., 2008), mule deer (*Odocoileus hemionus hemionus*) (Ab) (Dubey et al., 2008; Myers et al., 2014), pampas deer (*Ozotoceros bezoarticus*) (Ab) (Tiemann et al., 2005a), roe deer (*Capreolus capreolus*) (Ab) (Bartova et al., 2006; Almeria et al., 2007; Panadero et al., 2010; De Craeye et al., 2011; Malmsten et al., 2011; Candela et al., 2014), fallow deer (*Dama dama*) (Ab) (Bartova et al., 2006; Marco et al., 2008), red deer (*Cervus elaphus* and *C. elephas yarkandensis*) (Ab) (Ferroglia and Rossi, 2001; Bregoli et al., 2006; Almeria et al., 2007; Gozdzik et al., 2010; Billinis, 2013), elk (*Cervus canadensis*) (Ab) (Gutierrez-Exposito et al., 2012), moose (*Alces alces*) (Ab) (Dubey and Thulliez, 2005; Stieve et al., 2010; Malmsten et al., 2011; Gutierrez-Exposito et al., 2012; Moskwa et al., 2014), caribou (*Rangifer tarandus*) (Ab) (Dubey and Thulliez, 2005; Stieve et al., 2010; Gutierrez-Exposito et al., 2012), North American bison (*Bison bison*) (Ab) (Dubey and Thulliez, 2005), European bison (*Bison bonasus bonasus*) (Ab) (Cabaj et al., 2005; Bien et al., 2010), African buffalo (*Syncerus caffer*) (Ab) (Ferroglia et al., 2003), Thompson's gazelle (*Eudorcas thomsonii*) (Ferroglia et al., 2003), impala (*Aepyceros melampus*) (Ab) (Ferroglia et al., 2003), eland (*Taurotragus oryx*) (Ab) (Ferroglia et al., 2003), chamois (*Rupicapra rupicapra*) (Ab, DNA) (Ferroglia and Rossi, 2001; Bregoli et al., 2006; Gaffuri et al., 2006; De Craeye et al., 2011), Alpine ibex (*Capra ibex*) (Ab) (Bregoli et al., 2006), Spanish ibex (*Capra pyrenaica hispanica*

(Ab) (Garcia-Bocanegra et al., 2012), European mouflon (*Ovis musimon*) (Bartova et al., 2006), Barbary sheep (*Ammotragus lervia*) (Ab) (Almeria et al., 2007), musk ox (*Ovibos moschatus*) (Ab) (Dubey and Thulliez, 2005), wild boar (*Sus serofo*) (Ab) (Bartova et al., 2006; Almeria et al., 2007), and warthog (*Phacochoerus aethiopicus*) (Ab) (Ferroglia et al., 2003).

*Neospora caninum* antibodies have also been detected in the following captive nondomestic artiodactyl species: Axis deer (*Axis axis*) (Basso et al., 2014), Eld's deer (*Cervus eldi siamesis*) (Dubey et al., 1996), Thorold's deer (*Cervus albirostris*) (Sedlak and Bartova, 2006), eastern elk (*Cervus elaphus canadensis*) (Sedlak and Bartova, 2006), Vietnam sitka deer (*Cervus Nippon pseudaxis*) (Bartova et al., 2006; Sedlak and Bartova, 2006), Brouchet deer (*Mazama spp.*) (Tiemann et al., 2005b), Pere David's deer (*Elaphurus davidianus*) (Sedlak and Bartova, 2006), antelope (*Tragelaphus iberbis*) (Peters et al., 2001), blackbuck (*Antelope cervicapra*) (Sedlak and Bartova, 2006), lechwa (*Kobus leche*) (Sedlak and Bartova, 2006), African buffalo (*Syncerus caffer caffer*) (Sedlak and Bartova, 2006), eland (*Taurotragus oryx*) (Sedlak and Bartova, 2006), European bison (*Bison bonasus bonasus*) (Sedlak and Bartova, 2006), sitatunga (*Tragelaphus spekei gratus*) (Sedlak and Bartova, 2006), and vicuña (*Vicugna vicugna mensalis*) (Risco-Castillo et al., 2014).

Viable tachyzoites have been isolated from free-ranging white-tailed deer (isolates NC-WTDVA 1–3 (Vianna et al., 2005) and NcWTDm1-2 (Dubey et al., 2013)) and European bison (isolates NC-PoIBb1-2) (Bien et al., 2010) and captive axis deer (isolate Nc-Axis) (Yu et al., 2009). Transplacental transmission has been reported for free-ranging white-tailed deer (Dubey et al., 2013) and captive bred Eld's deer (Dubey et al., 1996), axis deer (Basso et al., 2014), and antelope (Peters et al., 2001).

There are five reports of *N. caninum* infection in artiodactyls (four cervids, one bovine), where parasites have been identified histologically and were associated with lesions (Woods et al., 1994; Dubey et al., 1996; Peters et al., 2001; Soldati et al., 2004; Basso et al., 2014). Disease in artiodactyls is associated with stillbirths and systemic disease in very young animals, and is more commonly described in captive animals. Multiple organ involvement is common and the most consistent lesion is nonsuppurative encephalitis.

Woods et al. (1994) described a case of systemic neosporosis in a free-ranging California black-tailed deer fawn (*O. hemionus columbianus*) (family: Cervidae) from Sacramento County, California (Table 1). The fawn was found dead. Macroscopic lesions included



emaciation, pulmonary edema, and soft pulpy kidneys. Lung, liver, and kidney were evaluated histologically. The lungs were characterized by a diffuse moderate mononuclear interstitial pneumonia with intra-alveolar edema and macrophages, scattered fibrin thrombi, and clusters of interstitial and intra-alveolar tachyzoites were found. Hepatic lesions included moderate bridging portal and central fibrosis, random multifocal coagulation necrosis with mild neutrophilic infiltrates, and occasional moderate portal mononuclear infiltrates. The kidney displayed mild scattered tubular necrosis and mild mononuclear interstitial infiltrates. Tachyzoites were detected in renal tubular epithelial cells and tubule lumina. Parasite tachyzoites were positive for *N. caninum* antigen and negative for *T. gondii* antigen by IHC using polyclonal rabbit antibodies. Tachyzoites were most prevalent in renal medullary tubular epithelial cells with lesser numbers in renal tubules and endothelial cells. In the lungs, tachyzoites were distributed throughout alveolar spaces, alveolar epithelium, and alveolar septal capillary endothelium. In the liver, tachyzoites were concentrated adjacent and peripheral to areas of hepatic necrosis and were also detected within hepatocytes, sinusoidal endothelium, Kupffer cells, and free within sinusoidal spaces. Tissue cysts were not identified.

Soldati et al. (2004) reported neosporosis in a captive bred fallow deer fawn (*Dama dama*) (family: Cervidae) from Switzerland, displaying hindlimb paresis, opisthotonus, pyrexia, tachypnea, and tachycardia (Table 1). The fawn was euthanized. The main gross lesions consisted of patchy brown discoloration throughout the entire length of the spinal cord, multiple dark foci in the brain stem, and pulmonary congestion and edema. The most significant histologic finding was multifocal random necrotizing and granulomatous meningoencephalomyelitis. The most severe lesions were in thoracic segments of spinal cord and were comprised of extensive multifocal necrosis predominately affecting white matter associated with large numbers of gitter cells, microgliosis, astrocytosis, capillary endothelial proliferation, spheroids, and multifocal mineralization. Clustered tachyzoites and few protozoan cysts were detected within inflammatory foci and were positively labeled by IHC using polyclonal caprine anti-*N. caninum* antibodies. Nc5 PCR and sequencing were used to confirm the diagnosis of neosporosis. Parasites were negative for both *T. gondii* and *Sarcocystis neurona* by IHC using polyclonal caprine antibodies. A fawn from the same captive group had been euthanized, with similar clinical signs and histologic lesions one month previously but no tissues were available for further investigation.

Dubey et al. (1996) reported a case of congenital neosporosis in a captive bred full-term stillborn Eld's deer (*Cervus eldi siamensis*) (family: Cervidae) from a zoo in Paris, France. Macroscopic lesions were not detected (Table 1). Microscopic lesions were limited to the brain and included multifocal nonsuppurative encephalitis characterized by perivascular mononuclear infiltrates, microglial nodules with multinucleated macrophages, and necrosis. Lesions were most severe in the midbrain. Protozoan tissue cysts were detected in association with some lesions and in areas of inflammation elsewhere in the brain. Tissue cysts exhibited strong immunoreactivity to monoclonal murine anti-*N. caninum* antibodies and no immunoreactivity to polyclonal rabbit anti-*T. gondii* antibodies.

Basso et al. (2014) identified *N. caninum* as a cause of perinatal mortality in a captive herd of axis deer (*Axis axis*) (family: Cervidae) from a zoo in La Plata, Argentina (Table 1). One fawn died at two weeks of age following a presentation of congenital anal dilation, incontinence, weakness and ataxia. Megacolon and megarectum were noted on postmortem examination. Cytologic preparations of brain smears revealed the presence of protozoan cysts. Histologic lesions included nonsuppurative encephalitis and gliosis, suppurative bronchopneumonia, fibrinonecrotic enteritis, and hepatic degeneration. An additional four neonates were found dead in the

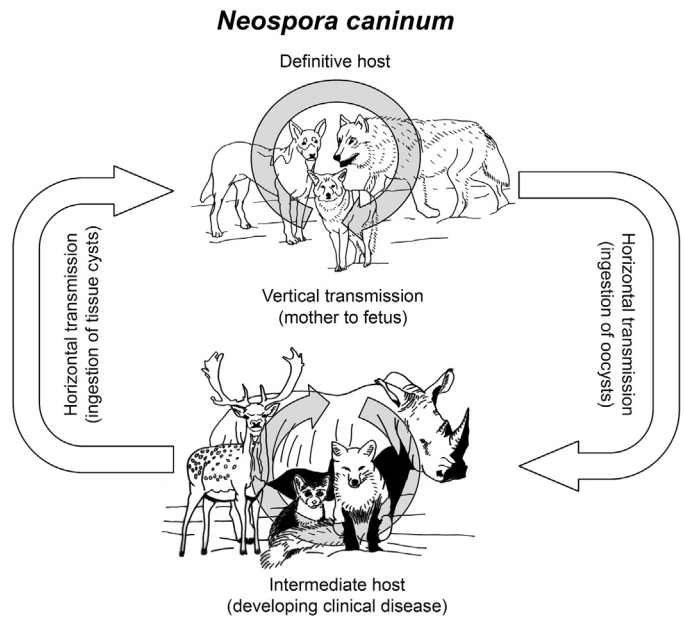


Fig. 1. Life cycle of *Neospora caninum*.

same enclosure within a period of two months before or after the death of the fawn. All were weak at birth and died within one to two days of age. Macroscopic and microscopic lesions were not reported. The fawn and neonates were serologically positive for *N. caninum* and negative for *T. gondii* by IFAT (neonate *N. caninum* IFAT titers were 1:25, 1:400, 1:3200, and 1:6400 and the fawn *N. caninum* IFAT titer was 1:6400). *Neospora*-like cysts were observed in a fresh brain cytologic preparation (see Fig. 1, Soldati et al., 2004). No IHC with anti-*N. caninum* antibodies was reported. Detection of *N. caninum* antibodies in the three neonates that had not ingested colostrum was deemed most consistent with transplacental transmission. *Neospora caninum* DNA was detected by Nc5 PCR in brain samples from the fawn and one of the four neonates and viable *N. caninum* parasites were isolated from fawn brain samples. Multilocus microsatellite-DNA sequence and/or length determination analysis of 9 *N. caninum* microsatellite markers (MS1B, MS2, MS3, MS5, MS6A, MS6B, MS10, MS12 and MS21) revealed an identical microsatellite pattern in *N. caninum* DNA from the fawn, neonate, and isolated parasites, evidencing recent infection from a common point source. Serological analysis of 13 adult deer housed in the same enclosure revealed a 92% seroprevalence rate for *N. caninum* (IFAT titers varied from <1:25 to  $\geq$ 1:6400).

Peters et al. (2001) reported congenital *N. caninum* infection in three full-term stillborn calves in a herd of captive lesser kudu (*Tragelaphus imberbis*) (family: Bovidae) from a zoo in Hanover, Germany (Table 1). Necropsies were performed on the calves, two of which were twins, and no macroscopic lesions were identified. The twin calves both had a moderate multifocal nonsuppurative meningoencephalitis with multiple granulomatous foci, gliosis, and mild mononuclear perivascular cuffs in the neuropil and leptomeninges and mild lymphohistiocytic periportal hepatitis. One calf also had mild lymphocytic myocarditis with focal myocardial mineralization. Parasites were not detected histologically. The twin calves and their dam were seropositive for *N. caninum* (IFAT titers were 1:16, 1:32, and 1:128, respectively; all were immunoblot (IB) positive). The third calf was histologically normal and seronegative by IFAT and IB for *N. caninum* (the dam was not tested). In this calf, *N. caninum* DNA was detected by Nc5 PCR in all tissues tested (brain, lung, heart, liver, and spleen) while in one of the twin calves, *N.*

*caninum* DNA was found only in the brain and lung (the second twin was not tested). All tissue samples were negative for *T. gondii* DNA by B1 PCR. IHC using polyclonal rabbit anti-*N. caninum* antibodies failed to demonstrate *N. caninum* antigen in any of the three calves. The authors postulated that an *N. caninum* seronegative DNA positive calf might be explained by recent infection shortly before stillbirth or transplacental infection early in pregnancy prior to establishment of fetal immunocompetence. The absence of detectable histologic lesions in this calf evidenced that vertical transmission of *N. caninum* infection may result in birth of infected yet clinically normal calves, as is often the case in cattle.

### 5.1.3. *Perissodactyla*

*Neospora caninum* antibodies have been detected in free ranging zebra (*Equus burchielli*) (Ferroglio et al., 2003) and captive white rhinoceros (*Ceratotherium simum*) (Williams et al., 2002; Sangster et al., 2010; Sommanustweechai et al., 2010) with disease and transplacental infection only described in the latter species. White rhinoceros appear to be particularly susceptible to developing fatal neosporosis at any age. Of the three reported cases, all involved captive animals and liver lesions were common.

Williams et al. (2002) described neosporosis in a white rhinoceros calf (family: Rhinocerotidae) from a breeding center in Northern Province, South Africa, that was found dead at 16 days of age (Table 1). Necropsy findings were consistent with congestive heart failure and included marked cardiomegaly, generalized cyanosis, pulmonary congestion and edema, and hepatic congestion. Heart, liver, lung, and kidney were evaluated histologically, with the most significant lesions present in the heart including marked multifocal to diffuse myocarditis with myocardial degeneration and necrosis. Myocardial inflammatory infiltrates were predominately histiocytic with lesser numbers of lymphocytes, plasma cells, and neutrophils. Giemsa staining of heart sections highlighted intralésional protozoan cysts and tachyzoites that were not apparent on H&E. Parasites in the heart were strongly labeled with both monoclonal and polyclonal anti-*N. caninum* antibodies (origin not reported) and were negative for *T. gondii* (antibody details not provided). Additional lesions were marked hepatic centrilobular congestion and mild centrilobular hepatocellular necrosis and fibrosis, diffuse pulmonary congestion and edema with scattered intra-alveolar macrophages, and multifocal atelectasis. Transmission electron microscopy performed on myocardium demonstrated encysted bradyzoites and intracellular tachyzoites compatible with *Neospora* spp.

Sommanustweechai et al. (2010) reported neosporosis in a 16-year old white rhinoceros from a zoological institution in Chonbari, Thailand (Table 1). The animal was found in sternal recumbency shortly before death. Postmortem examination revealed hepatomegaly and extensive multifocal hepatic necrosis involving approximately 60% of the parenchyma. Pulmonary edema and hemorrhage, hemorrhagic splenic infarction, multifocal renal necrosis, mesenteric necrosis, mesenteric lymphadenomegaly, adrenomegaly with diffuse parenchymal ecchymoses, erosive and ulcerative gastritis, and black discoloration and focal hemorrhage of the jejunal mucosa were also observed. Brain, liver, lung, adrenal gland, kidney, spleen, and myocardium were evaluated histologically. Severe, widespread necrotic foci bounded by lymphocytes and plasma cells were present in the liver, adrenal cortex, kidney, and small intestine; intralésional tachyzoites were evident in all tissues except the latter. Additional findings included hepatic fibrosis and granulomas characterized by central necrosis and numerous giant cells and macrophages without surrounding fibrosis, necrotizing lymphadenitis and splenitis with lymphoid depletion and neutrophil infiltration, and pulmonary edema and hemorrhage. Parasites and lesions were not detected in sections of brain or heart. IHC using

polyclonal rabbit anti-*N. caninum* antibodies positively labeled tachyzoites in the liver, adrenal gland, kidney and small intestine; immunopositive cells included Kupffer cells, hepatocytes, sinusoidal endothelium, and biliary and intestinal epithelium. Authors showed *Neospora*-like cysts in the liver and adrenal gland on H&E (see fig. 1BCD in Sommanustweechai et al., 2010). Nc5 PCR of liver was positive. Diagnostic tests to rule out *T. gondii* infection were not reported.

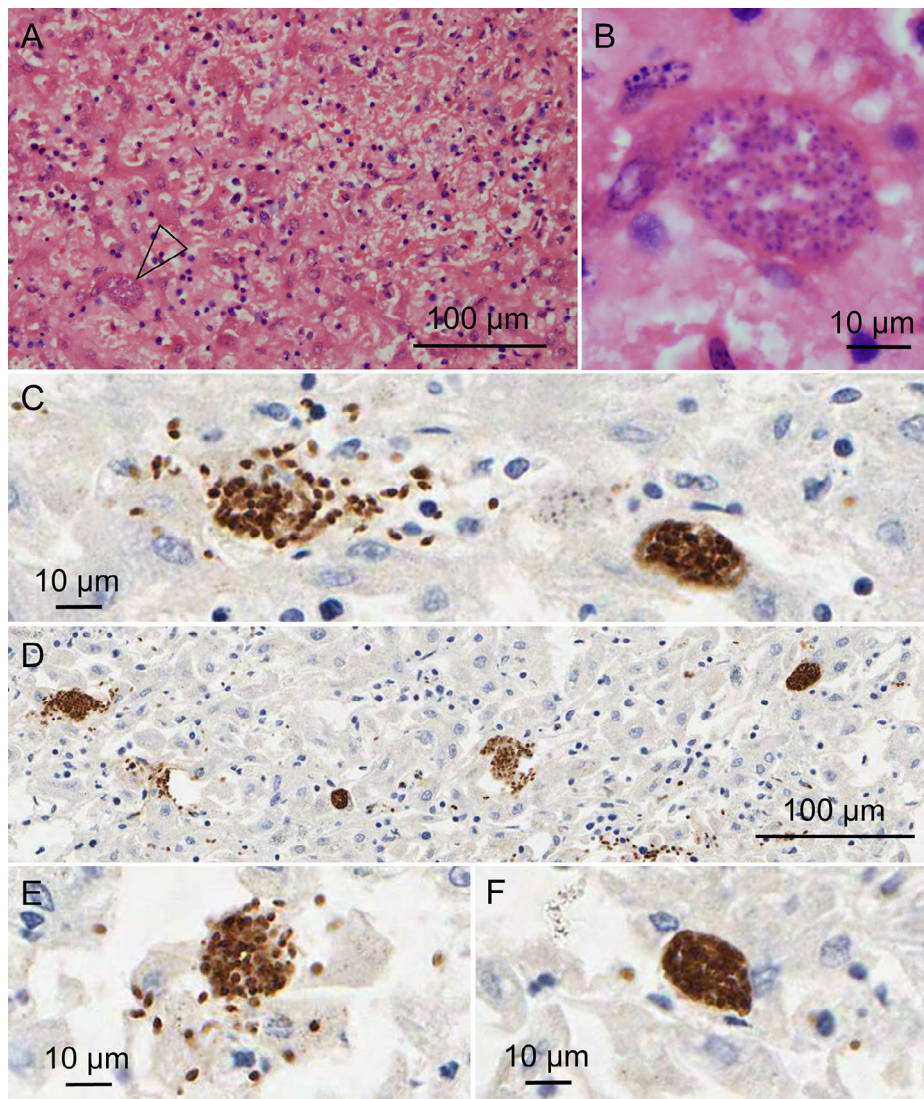
Sangster et al. (2010) identified *N. caninum* as the etiologic agent of abortion in a southern white rhinoceros fetus from a zoo in Dubbo, New South Wales, Australia (Table 1). Necropsy of the aborted fetus demonstrated hepatomegaly and herniation of a small segment of intestine through the umbilical opening. Microscopic lesions included multifocal hepatic necrosis with small numbers of mixed inflammatory cells and clustered extracellular and intrahepatocellular tachyzoites and cyst-like structures on *N. caninum* IHC (Figure 2). Similar tachyzoites were also present in the cerebellum with minimal inflammation. Polyclonal anti-*N. caninum* and anti-*T. gondii* antibodies (origin not reported) identified *N. caninum* but not *T. gondii* antigen in areas of hepatic necrosis. PCR of liver DNA targeting multiple markers (Nc5, SSU rDNA, LSU rDNA, ITS1, and ITS1-5.8S-ITS2 rDNA) and sequencing of the latter four PCR amplification products confirmed the presence of *N. caninum* DNA. Microsatellite analysis using the MS10 marker detected a unique pattern of trinucleotide repeats distinct from any characterized *N. caninum* isolates.

### 5.1.4. *Rodentia*

*Neospora caninum* antibodies and/or DNA have been detected in the following free ranging rodents: brown rat (*Rattus norvegicus*) (Ab, DNA) (Huang et al., 2004; Ferroglio et al., 2007; Jenkins et al., 2007; Medina-Esparza et al., 2013), house mouse (*Mus musculus*) (Ab, DNA) (Ferroglio et al., 2007; Jenkins et al., 2007; Barratt et al., 2008; Thomasson et al., 2011; Medina-Esparza et al., 2013), field mouse (*Apodemus sylvaticus*) (Ab, DNA) (Ferroglio et al., 2007; Thomasson et al., 2011), harvest mouse (*Micromys minutus*) (DNA) (Meerburg et al., 2012), common vole (*Microtus arvalis*) (DNA) (Fuehrer et al., 2010), water vole (*Arvicola terrestris*) (DNA) (Fuehrer et al., 2010), rock squirrel (*Spermophilus variegates*) (DNA) (Medina-Esparza et al., 2013), and capybara (*Hydrochaeris hydrochaeris*) (Ab, DNA) (Yai et al., 2008; Truppel et al., 2010; Valadas et al., 2010).

There are no reports of naturally occurring neosporosis in mice or other rodents; however, parasite tissue cysts have been identified in several species. These cases will be addressed briefly below. Some authors suggest that wild rodents contribute to the *N. caninum* sylvatic life cycle by serving as infected reservoir hosts for canid hosts (Huang et al., 2004; Ferroglio et al., 2007; Jenkins et al., 2007; Meerburg et al., 2012; Medina-Esparza et al., 2013). To our knowledge, viable *N. caninum* parasites have yet to be isolated from any species of wild rodents to date (Gondim, 2006; Dubey et al., 2007; Jenkins et al., 2007; Dubey and Schares, 2011; Medina-Esparza et al., 2013). As such, the putative role of rodents in the epidemiology of *N. caninum* requires further investigation.

Medina-Esparza et al. (2013) screened brown rats (*R. norvegicus*) (family: Muridae), house mice (*M. musculus*) (family: Muridae) and rock squirrels (*S. variegates*) (family: Sciuridae) for *N. caninum* infection by assaying samples of brain, spinal cord, and liver using *N. caninum* specific ITS1 PCR and IHC employing polyclonal caprine anti-*N. caninum* antibodies (Table 2). Liver was the tissue in which *N. caninum* cysts and parasite DNA were detected with greatest frequency and hepatitis (severity not reported) was evident histologically in all *N. caninum* positive animals, although it was also evident in some negative control animals. *Neospora caninum* immunopositive cysts were rarely found in the brain (1/13 mice, 0/6 rats, 0/14 squirrels) and heart (0/13 mice, 0/6 rats, 2/14 squirrels).



**Fig. 2.** Liver from an aborted white rhinoceros fetus with naturally acquired congenital *N. caninum* infection. Multifocal hepatic necrosis with intralosomal intracellular protozoan cyst-like structure (open arrow). H&E (A). Intracellular protozoan cyst-like structure. H&E (B). Immunohistochemistry (IHC) using polyclonal caprine anti-*N. caninum* showing clustered free and intracellular protozoan tachyzoites (C, D, E) and intracellular protozoan cyst-like structures (C, D, F). H&E photomicrographs (A, B) courtesy of Cheryl Sangster, Taronga Conservation Society Australia.

Previous attempts to identify parasites by IHC in seropositive and DNA positive brown rats were unsuccessful (Hughes et al., 2006).

#### 5.1.5. Lagomorpha

*Neospora caninum* antibodies and/or DNA have been detected in the following free-ranging lagomorphs: rabbit (*Oryctolagus cuniculus*) (Ab,DNA) (Ibrahim et al., 2009), Iberian hare (*Lepus granatensis*) (Ab) (Almeria et al., 2007; Hughes et al., 2008), brown hare (*Lepus europaeus*) (Ab) (Ezio and Anna, 2003; Bartova et al., 2010), and eastern cottontail rabbits (*Sylvilagus floridanus*) (DNA) (Zanet et al., 2013). Neosporosis has not been reported in lagomorphs.

#### 5.1.6. Insectivora

*Neospora caninum* DNA has been detected in the common shrew (*Sorex araneus*) (Meerburg et al., 2012) and the white-toothed shrew (*Crocidura russula*) (Meerburg et al., 2012). There are no reports of neosporosis in insectivores.

#### 5.1.7. Proboscidea

*Neospora caninum* antibodies have been detected in captive Indian elephants (*Elephas maximus indicus*) (Wiengcharoen et al., 2012). There are no reports of neosporosis in elephants.

#### 5.1.8. Cetacea

The bottlenose dolphin (*Tursiops truncatus*) is the only cetacean species in which *N. caninum* antibodies have been detected (Dubey et al., 2003). Neosporosis in cetaceans has not been reported.

#### 5.1.9. Pinnipedia

*Neospora caninum* antibodies have been detected in free-ranging and captive California sea lions (*Zalophus californianus*) (Dubey et al., 2003), and free-ranging walrus (*Odobenus rosmarus*) (Dubey et al., 2003), ringed seals (*Phoca hispida*) (Dubey et al., 2003), bearded seals (*Erignathus barbatus*) (Dubey et al., 2003), harbor seals (*Phoca vitulina*) (Dubey et al., 2003), ribbon seals (*Phoca fasciata*) (Dubey et al., 2003), spotted seals (*Phoca largha*) (Dubey et al., 2003; Fujii et al.,

2007), and Kuril harbor seals (*Phoca vitulina stejnergeri*) (Fujii et al., 2007). Neosporosis in pinnipeds has not been reported.

## 5.2. Metatherian mammals

### 5.2.1. Marsupialia

Reports of naturally acquired *N. caninum* infection in marsupials are rare, with antibodies and/or DNA identified in just three marsupial species: free-ranging South American opossums (*Didelphis marsupialis*) (Ab) (Yai et al., 2003) and western grey kangaroos (*Macropus fuliginosus ocydromus*) (Ab) (Mayberry et al., 2014), and a captive Parma wallaby (*Macropus parma*) (DNA) (Cronstedt-Fell et al., 2012). Current dogma suggests that marsupials are prone to developing overwhelming toxoplasmosis and it has been proposed that they may be equally susceptible to pathologic sequelae of *N. caninum* infection (King et al., 2011a); however, this hypothesis does not appear to be supported by current data. Mayberry et al. (2014) recently investigated 102 asymptomatic western grey female kangaroos from Western Australia for *N. caninum* exposure using an indirect fluorescent-antibody test (IFAT) modified for use in macropods and reported an overall seroprevalence of 18% (IFAT titers not reported; cut-off 1:50). Importantly, this study did not indicate that *N. caninum* exposure negatively impacted the reproductive performance of any of these doe(s) is/are female kangaroos.

To date, there is only a single case of naturally occurring neosporosis in a marsupial. Cronstedt-Fell et al. (2012) reported a case of neosporosis in a captive adult Parma wallaby (*M. parma*) from a zoo in Vienna, Austria that was found dead. Postmortem examination revealed marked eccentric hypertrophy of the left side of the heart and moderate dilation of the right heart chambers. Significant histologic lesions were present in the heart and consisted of a multifocal interstitial lymphocytic myocarditis and moderate multifocal myocardial degeneration and necrosis associated with mild histiocytic infiltrates and minimal hemorrhage. Throughout the myocardium, cardiomyocytes occasionally contained clusters of protozoan zoites but these were not initially apparent in areas of inflammation and necrosis. Additional findings included moderate pulmonary edema, emphysema and congestion, moderate multifocal ulcerative gastritis and mild catarrhal colitis. The diagnosis of neosporosis was confirmed by IHC using polyclonal caprine anti-*N. caninum* and anti-*T. gondii* antibodies, which revealed numerous *N. caninum* immunopositive and *T. gondii* immunonegative free and intracellular protozoan tachyzoites and random cysts in areas of myocardial necrosis. Findings were supported by the detection of *N. caninum* DNA from cardiac tissue using ITS1 PCR followed by sequencing.

### 5.3. Birds

The role of birds in the *N. caninum* life cycle is uncertain. A positive correlation has been shown between the presence of birds on cattle farms and increased seroprevalence and risk of *N. caninum*-associated abortion storms, suggesting that birds may contribute to parasite transmission in sylvatic cycles either as mechanical vectors or as intermediate hosts (Bartels et al., 1999; Ould-Amrouche et al., 1999; Otranto et al., 2003). Until the recent identification of domestic chickens (*Gallus domesticus*) as intermediate hosts (Costa et al., 2008) and the discovery that domestic dogs fed *N. caninum* inoculated embryonated chicken eggs shed oocysts (Furuta et al., 2007), there was little evidence that avian species could function as natural hosts for the parasite. The putative role of birds as intermediate hosts is further supported by the identification of *N. caninum* antibodies and/or DNA in the following free-ranging birds: house sparrow (*Passer domesticus*) (DNA) (Gondim et al., 2010), common raven (*Corvus corax*) (Ab) (Molina-Lopez et al., 2012), magpie (*Pica pica*)

(DNA) (Darwich et al., 2012), and common buzzard (*Buteo buteo*) (DNA) (Darwich et al., 2012).

Histologic evidence of *N. caninum* infection in birds is rare and naturally occurring neosporosis is not reported. South American parrots are the only birds in which protozoan cysts presumed to be *N. caninum* have been identified (Mineo et al., 2011). Histopathologic evaluation of a red-and-green macaw (*Ara chloropterus*) and a blue-fronted Amazon parrot (*Amazona aestiva*), both of which died from unrelated causes, revealed protozoan tissue cysts in pericloacal and cervical skeletal muscle, respectively (Table 2). IHC using polyclonal murine anti-*N. caninum* and anti-*T. gondii* antibodies and mAb 74.1.8, a monoclonal antibody directed against the bradyzoites specific antigen BAG1 (commonly expressed in both *N. caninum* and *T. gondii* tissue cysts (Weiss et al., 1999)) demonstrated that parasitic cysts were immunopositive for *N. caninum* and BAG1 and immunonegative for *T. gondii*. When the same antibodies were tested on pigeon tissues bearing *Sarcocystis* spp. cysts, a complete absence of staining was observed. Unfortunately, fresh tissue from the parrots was not available for further investigation and DNA extraction from formalin fixed paraffin embedded tissue was unsuccessful. Protozoan cysts in both cases were not associated with any significant histologic lesions and were considered incidental findings.

## 6. Experimentally acquired neosporosis and pathology in wildlife species

Experimentally induced *N. caninum* infections have been reported for a number of different species under a variety of experimental conditions. The purpose of this manuscript is not to detail the various experimental approaches used in these studies. For additional details, we encourage readers to seek out the references provided. Instead, we focus on (1) the pathologic changes associated with *N. caninum* infection, and (2) parasite detection in different wildlife laboratory models. By doing so, our aim is to better understand neosporosis in these species and ascertain optimal sample collection to facilitate diagnosis when *N. caninum* infection is suspected in wildlife cases.

Multiple species of rodents have been used to study the pathology of *Neospora* infection (Lindsay and Dubey, 1989; Dubey and Lindsay, 2000; Pipano et al., 2002; Uchida et al., 2003; Ramamoorthy et al., 2005; Hurkova-Hofmannova et al., 2007; Kang et al., 2009; Mols-Vorstermans et al., 2013; Bottari et al., 2014). Less traditional laboratory rodent species, considered here as wildlife species despite originating from breeding colonies, in which experimental infections were conducted and disease demonstrated, include the following: multimammate rat (*Mastomys natalensis*) (family muridae, subfamily murinae) (Hurkova-Hofmannova et al., 2007), sand rat (*Psammomys obesus*) (family muridae, subfamily gerbillinae) (Pipano et al., 2002), Tristam's jird (*Meriones tristrami*) (family muridae, subfamily gerbillinae) (Hurkova-Hofmannova et al., 2007), Wagner's gerbil (*Gerbilus dasyuris*) (family muridae, subfamily gerbillinae) (Hurkova-Hofmannova et al., 2007), Mongolian gerbils (*Meriones unguiculatus*) (family muridae, subfamily gerbillinae) (Cuddon et al., 1992; Gondim et al., 1999, 2001; Dubey and Lindsay, 2000; Ramamoorthy et al., 2005; Kang et al., 2009; Bottari et al., 2014), and Djungarian hamsters (*Phodopus sungorus*) (family cricetidae) (Uchida et al., 2003). Of these species, gerbiline rodents have proven to be highly susceptible to *N. caninum* infection and the Mongolian gerbil is currently the common immunocompetent experimental host used for the study of neosporosis (Dubey and Lindsay, 2000; Gondim et al., 2001; Ramamoorthy et al., 2005; Hurkova-Hofmannova et al., 2007; Kang et al., 2009). Clinical signs of disease are similar among different susceptible rodent genera and include roughened fur, apathy/depression, inappetence, weight loss, incoordination, and hindlimb paresis (Dubey and Lindsay, 2000; Gondim et al., 2001; Pipano et al., 2002; Uchida et al., 2003;

Ramamoorthy et al., 2005; Hurkova and Modry, 2006). Although not always evident, the most commonly observed macroscopic lesions are fibrinous peritonitis with ascites containing tachyzoites (likely due to intraperitoneal method of inoculation), splenomegaly, and hepatomegaly (Dubey and Lindsay, 2000; Pipano et al., 2002; Ramamoorthy et al., 2005; Hurkova-Hofmannova et al., 2007; Bottari et al., 2014). Histopathologic changes are somewhat more variable and difficult to compare between studies due to inconsistencies in tissue evaluation and lesion description, differences in inoculum dose and route, and variations in disease chronicity in evaluated animals. Reported microscopic lesions include miliary hepatitis and hepatic necrosis, granulomatous hepatitis, splenic lymphoid and reticular cell hyperplasia, granulomatous splenitis, myocarditis, myositis, gastroenteritis (more severe in animals inoculated orally with sporulated oocysts), bronchopneumonia, and meningoencephalitis, with intralesional tachyzoites often evident in areas of inflammation (Dubey and Lindsay, 2000; Gondim et al., 2001; Hurkova-Hofmannova et al., 2007; Kang et al., 2009; Bottari et al., 2014). Of the species listed above, tissue cysts have only been described in Mongolian gerbils and Djungarian hamsters (Gondim et al., 2001; Uchida et al., 2003). Kang et al. (2009) used Nc5 PCR to investigate *N. caninum* tissue distribution in Mongolian gerbils during acute infection resulting from intraperitoneal inoculation with tachyzoites. Tissue screening showed tissues were targeted in the following order, from early to late: liver, spleen, and kidney; heart, skeletal muscle, spinal cord and blood; lung; and then brain. In this study, *N. caninum* DNA was detected from day six in the liver and spleen, and only from day eight onwards throughout the remainder of the body, suggesting that liver and spleen may be the optimal tissues for PCR detection of suspected infection (Kang et al., 2009).

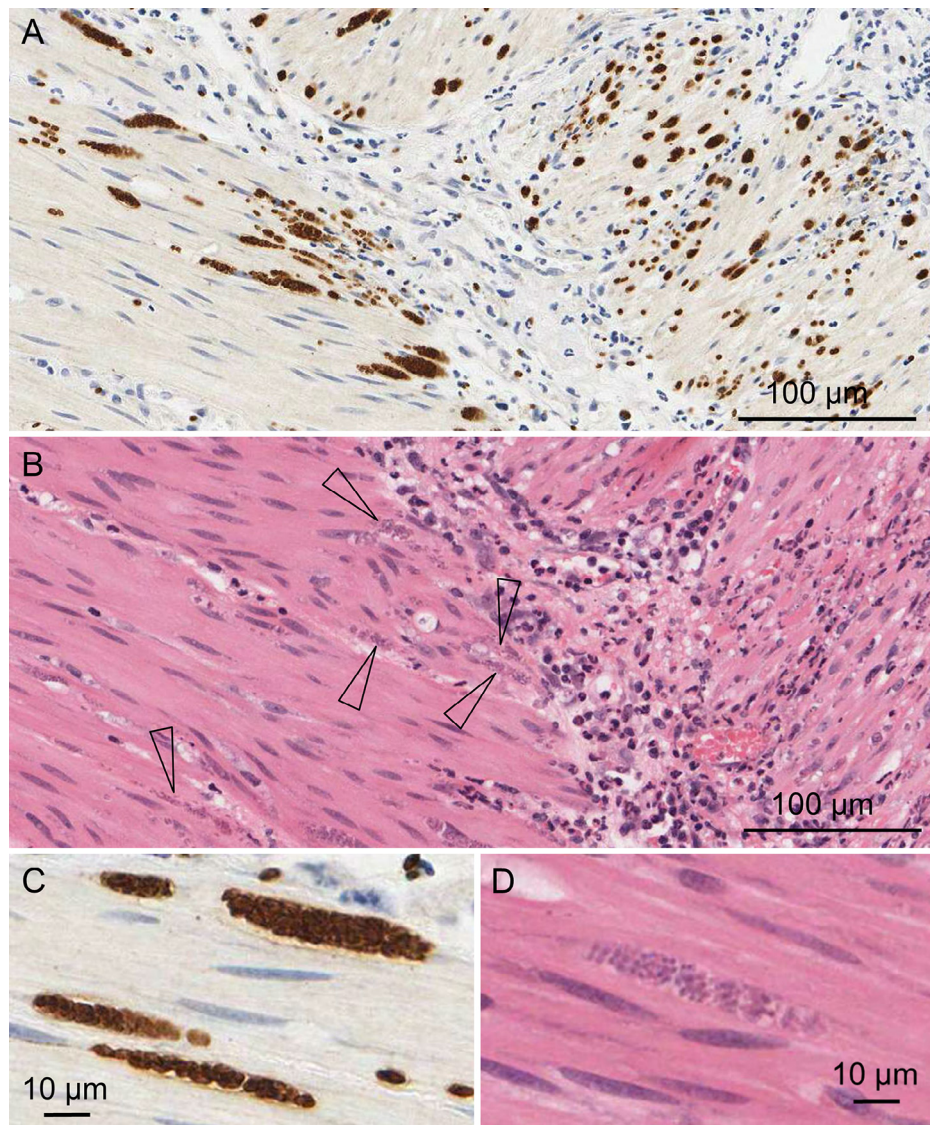
Recent investigations of experimental neosporosis in the fawn-tailed dunnart (*Sminthopsis crassicaudata*), a native Australian carnivorous marsupial, have demonstrated this species to be a susceptible intermediate host to *N. caninum* and that resultant disease is severe and dominated by tissue cyst production (King et al., 2011a). Dunnarts inoculated intraperitoneally with  $10^5$  tachyzoites were euthanized between 13 and 18 days post-inoculation due to severe clinical signs, which included roughened fur, lethargy, reduced activity, inappetence, progressive hindlimb paralysis, incontinence, and weight loss. The only consistent gross finding at necropsy was marked atrophy of adipose tissue. Microscopic lesions were consistent with disseminated protozoal infection and included severe necrosuppurative pancreatitis, widespread myonecrosis and myodegeneration of hindlimb skeletal muscle and urinary bladder detrusor muscle, suppurative cystitis, peritonitis, adrenal necrosis, and granulomatous to pyogranulomatous interstitial pneumonia. Intralesional tachyzoites and tissue cysts were common and displayed strong immunoreactivity to polyclonal caprine anti-*N. caninum* antibodies (Figure 3). Parasites were most numerous in the urinary bladder detrusor muscle, cardiac and skeletal striated muscle, and pancreas, with distinct replicating tachyzoites present in pancreatic acinar cells. IHC-positive parasites were also found in spleen, brain, accessory sex gland, mesenteric lymph node, gall bladder and gastric muscular tunics, and intestinal villi/enterocytes; parasites were not detected in the eye, kidney, or spinal cord. Bradyzoite specific IHC using monoclonal anti-*T. gondii* BAG5 (also referred to as BAG1) antibodies that cross react with their *N. caninum* equivalents confirmed the presence of *N. caninum* tissue cysts in the heart, lung, pancreas, adrenal gland, urinary bladder, and skeletal muscle. All dunnarts seroconverted to *N. caninum* by 14 days post inoculation as determined by cELISA, and *N. caninum* DNA was detected in the brain by Nc5 and ITS1 PCR. In contrast, dunnarts infected with  $10^4$  *N. caninum* tachyzoites did not demonstrate clinical signs or histologic lesions by 28 days post inoculation and *N. caninum* DNA was not detected in the brain, liver, and lung, although two animals had evidence of seroconversion. Dunnarts dosed orally with 20–40 *N.*

*caninum* oocysts did not develop disease, histologic lesions, or seroconvert; *N. caninum* DNA was detected in the brain of one animal and in the other, *N. caninum* antigen was identified in the spleen by IHC.

Barr et al. (1994) inoculated fetal and pregnant female rhesus macaques (*Macaca mulatta*) with *N. caninum* tachyzoites in order to assess primate susceptibility to infection. Two fetuses (experiment A) received direct intramuscular inoculations of  $1 \times 10^6$  tachyzoites in utero on gestational day 65 and two pregnant dams (experiment B) received intravenous and intramuscular inoculations with a total of  $1.6 \times 10^7$  tachyzoites on gestational day 43. This study confirmed that nonhuman primate fetuses could become infected by direct in utero inoculation and by transplacental transmission. Hysterotomy and fetal necropsy was conducted at 13–22 days (experiment A) and 67–70 days post inoculation (experiment B). Gross lesions in experiment A were oligohydramnios, placental edema, and complete absence of chorioamniotic fusion. Histologic lesions were consistent with systemic neosporosis and included pyogranulomatous deciduitis, fibrinous and pyogranulomatous variably necrotizing placentitis, multifocal superficial mixed dermatitis with small foci of parakeratotic hyperkeratosis, random focal pyogranulomatous to mononuclear encephalitis with occasional foci of necrosis, multifocal mononuclear interstitial pneumonia, and diffuse suppurative interstitial nephritis; intralesional tachyzoites were detected in all tissues but the latter. Additionally, rare infiltrates of mononuclear cells and/or protozoan tachyzoites were also noted in the skeletal muscle, liver, heart, spleen, and intestinal wall. Gross lesions in experiment B included multiple small cavitations and foci of discoloration in the cerebrum, and in one animal, the umbilical interstitium contained a small opaque pale tan focus. Microscopic lesions consisted of severe multifocal to coalescing granulomatous variably necrotizing encephalomyelitis, multifocal necrotizing and fibrinous placentitis, focal necrotizing omphalitis, and multifocal perivascular mononuclear deciduitis; intralesional tachyzoites were evident in all tissues except the latter. In all infected fetuses, *Neospora*-specific antibodies were detected by IFAT (IgM titers were <2 to 128, IgG titers were 160 – 5,120, and IgA titers were <40 to 1,280); tissue cysts were not identified; and tachyzoites were reisolated from fetal brain, placenta, and amniotic fluid. Parasites were stained immunohistochemically with polyclonal rabbit anti-*N. caninum* antibodies.

Over the years, birds have been intensively investigated as possible reservoir hosts for *N. caninum*. Evidence of *N. caninum* exposure in domestic fowl (Costa et al., 2008) and wild bird populations (Gondim et al., 2010; Mineo et al., 2011; Darwich et al., 2012; Molina-Lopez et al., 2012) has prompted questions about the role avian species may play in maintaining the parasite in the sylvatic life cycle. To address this gap in our knowledge of *N. caninum* epidemiology, experimental investigations in domestic pigeons (*Columba livia*) (McGuire et al., 1999; Mineo et al., 2009), quail (*Coturnix coturnix japonica*) (de Oliveira et al., 2013), and zebra finches (*Poephila guttata*) (McGuire et al., 1999) have aimed to examine the consequences of *N. caninum* infection in birds. These studies have shown that avian species differ in their susceptibility to *N. caninum* infection, similar to what is seen in mammals. Pigeons appear to be the most susceptible to infection while quail and zebra finches are more resistant.

Mineo et al. (2009) inoculated four pigeons intraperitoneally with  $10^7$  *N. caninum* tachyzoites and found that all birds rapidly seroconverted with peak *N. caninum*-specific IgG levels detected between 10 and 20 days post inoculation (IFAT titers varied from 1: <200 to 1:640; cut-off 1:20). One pigeon died 25 days post inoculation, with evidence of widespread parasite dissemination; *N. caninum* (life stage not specified) was identified by immunohistochemical staining with polyclonal murine anti-*N. caninum* antibodies in the lungs, heart, central nervous system, and kidney. Additional light microscopic findings were not reported and whether or not



**Fig. 3.** Urinary bladder from a fat-tailed dunnart (*Sminthopsis crassicaudata*) experimentally infected with *N. caninum*. IHC using polyclonal caprine anti-*N. caninum* showing marked necrotizing and widespread degeneration and necrosis of the detrusor muscle with many neutrophils and macrophages and intralysosomal protozoan cysts and tachyzoites (A). H&E of the same tissue; open arrows indicate intracellular protozoan organisms (B). *N. caninum* IHC (C) and H&E (D) of protozoan tissue cysts.

parasites were associated with significant lesions is therefore unclear. Surviving pigeons were seronegative and IHC negative for *N. caninum* at 45 days post inoculation, suggesting that active infection may be resolved within several weeks and seroconversion is of limited duration.

These findings are in contrast to a previous study by McGuire et al. (1999) in which three pigeons were inoculated intraperitoneally with  $10^4$ – $10^6$  tachyzoites, with the two birds receiving the higher inoculum dose being seropositive (IFAT titers 1:800 and 1:400) at 6 weeks post inoculation (study conclusion). All infected birds remained clinically normal, and had detectable *N. caninum* DNA in samples of brain with Nc5 PCR and viable tachyzoites reisolated from brain homogenates (McGuire et al., 1999). Histologic lesions were found in the brain (only tissue examined) of the pigeon infected with  $10^6$  tachyzoites and consisted of multifocal mononuclear perivascular cuffs in the cerebral cortex, midbrain, and meninges, and a moderate focal granulomatous and heterophilic neurohypophysitis (McGuire et al., 1999). Zebra finches receiving the same inoculum dose and route remained refractory and developed no evidence of infection (McGuire et al., 1999).

Quail inoculated subcutaneously with  $3.5$ – $5 \times 10^5$  *N. caninum* tachyzoites demonstrated no evidence of morbidity or mortality despite evidence of infection (de Oliveira et al., 2013). Tissues were screened for *N. caninum* at different time points throughout the experiment by histopathology, IHC using polyclonal bovine anti-*N. caninum* antibodies, and Nc5 PCR. In some birds, the liver contained mononuclear infiltrates associated with *N. caninum* antigen but this was detected only during the first week of infection (de Oliveira et al., 2013). Tissues that most frequently tested positive for *N. caninum* by IHC and PCR were the bursa of Fabricius, brain, spleen, liver, and lungs, demonstrating that parasite dissemination occurred post-inoculation. Tissue cysts were not detected. Infected birds seroconverted (IFAT titers not reported; cut-off 1:10) after a week, with peak titers detected between weeks two and three; however, rapid decline of the *N. caninum* antibody titer to undetectable levels by one month suggested that this species was able to adequately control infection.

Based on these studies, antibody production against *N. caninum* parasites in quail and pigeons, at least in some cases, may be sharply transient, and similar findings have also been reported for chickens

(Furuta et al., 2007; Mineo et al., 2009). Speculatively, this may also be the case for other species of wild birds and thus should be considered when interpreting findings of serological surveys in avian species. Tissue cysts were not described in any of these reports and it remains unclear whether or not *N. caninum* establishes chronic infections in birds.

## 7. Animals with no evidence of naturally or experimentally acquired *N. caninum* exposure

To our knowledge, *N. caninum* exposure and/or infection have yet to be reported in the following orders of eutherian mammals: primates (primates), chiroptera (bats), dermoptera (colugos or flying lemurs), edentata (toothless mammals), tubulidentata (aardvarks), hyracoidae (hyraxes, dassies), pholidata (pangolins), and sirenina (dugongs and manatees).

There is no evidence to date of *N. caninum* exposure and/or infection in prototherian species (platypus and echidna) or any ectotherm species (reptiles, amphibians, and fish).

## 8. Diagnostic tools used for detection of *N. caninum* in wildlife

Several diagnostic assays have been employed with varying degrees of success to identify *N. caninum* infection in nondomestic species in order to better understand the potential role of wildlife in the epizootiology of *N. caninum*. Most of these reports have utilized serological techniques, with direct detection of parasite specific DNA using PCR-based molecular methods increasingly reported. Unfortunately, in many instances the interpretation and comparison of study results are confounded by many factors including variation in study design and methodology, sample size, and data interpretation. It is important to note that the identification of *N. caninum* antibodies, DNA, and/or tissue cysts is not necessarily analogous with the presence of viable parasites and data should be interpreted cautiously, particularly in those cases where only a single diagnostic test has been used (Dubey et al., 2007; Dubey and Schares, 2011). Specific procedures for many of the techniques described in this chapter may be found in “Protozoal abortion in farm ruminants: guidelines for diagnosis and control” (Ortega-Mora, 2007).

### 8.1. Isolation

Definitive identification of intermediate host species requires isolation of viable *N. caninum* parasites by bioassays in animals and/or cell culture. Unfortunately, *N. caninum* is notoriously difficult to isolate from naturally infected animals due to lack of parasite viability and/or low numbers, and it may take longer than a month to visualize parasite life stages in primary cultures (Conrad et al., 1993; Kim et al., 2000). Additionally, some isolates (e.g., white-tailed deer isolate NC-WTDVA-2) do not grow in cell culture (Vianna et al., 2005). Bioassays in rodents are costly, involve ethical considerations, and require colonies of immunosuppressed or susceptible species such as cortisone-treated outbred mice, IFN- $\gamma$  gene knockout mice or gerbils (Dubey and Schares, 2006; Dubey et al., 2007). Consequently, parasite isolation is predominately used for research purposes when strain characterization is desired. That said, there is a clear need for the more frequent isolation of *N. caninum* globally, particularly from wildlife, because of existing uncertainties over virulence and sylvatic cycles as discussed above. To achieve this, national centers with infrastructure capable of culturing *N. caninum* are required to respond adequately to the unpredictable emergence of cases in wildlife.

Brain is the most common tissue for isolation studies but isolation has also been achieved using spinal cord, blood, heart or other infected tissues, as well as skin lesion biopsies (Dubey et al., 2007, 2014a; Dubey and Schares, 2011).

### 8.2. Histopathology and immunohistochemistry

Histopathology has a low sensitivity for detection of organisms compared to other available diagnostic tests because parasite life stages are not always present or readily observed in tissue section (Dubey and Schares, 2006). However, histopathology remains an extremely valuable diagnostic tool that is essential for improving our understanding of infection outcome in wildlife species. It enables lesion description, tissue distribution analysis, and identification of concurrent pathology with intralésional parasites, documenting an association between parasite presence and disease. Because it is not possible to definitively distinguish the different tissue-cyst forming coccidia using light microscopic examination of tissue alone, additional confirmatory diagnostic tests should be conducted in conjunction with histopathology.

IHC with *N. caninum* specific antibodies can facilitate diagnosis by identifying parasite-specific antigen (Dubey and Schares, 2006). In general, polyclonal antibodies are preferred over monoclonal antibodies for IHC diagnosis of neosporosis because they exhibit higher sensitivity in detecting *N. caninum* antigen when present (Dubey and Schares, 2006; Dubey et al., 2007). For this purpose, the commercial polyclonal antibodies (in our laboratory we use *N. caninum* antiserum, #PAB-NC, VMRD, Pullman, WA, USA) are globally distributed and suitable for protocol validation by individual histopathology laboratories. Polyclonal antibodies are less specific than monoclonal antibodies and may exhibit cross-reactivity with other closely related apicomplexan parasites (Dubey and Schares, 2006; Dubey et al., 2007). Although not used for routine diagnosis of *N. caninum*, bradyzoite specific antibodies (BAG1/BAG5) may be applied to differentiate tissue cysts from clustered tachyzoites (Dubey and Schares, 2006). The absence of commercially available bradyzoite/tachyzoite-specific antibodies makes these assays less suitable for routine analysis.

It is our opinion that postmortem examination should be conducted on (1) any wild animal euthanized for suspected neosporosis or (2) for investigation of *N. caninum* infection, and a variety of tissues be collected and fixed in 10% formalin for histopathology (processed into paraffin blocks within 7 days if IHC is considered). For that matter, best practice suggests that wherever possible, routine post mortem examination should be performed on any wild animal dying or euthanized for unknown cause, given the variability in signs associated with neosporosis. Based on our review of wildlife cases in which *N. caninum* parasites have been detected histologically, sampled tissues should include, at the minimum, brain and spinal cord (if possible), lung, heart, liver, skeletal muscle, and placenta (if applicable). For cases of suspected disease, spleen, lymph node, kidney, adrenal gland, urinary bladder, pancreas, representative sections of gastrointestinal tract, and reproductive structures should also be assessed to provide a complete picture. Even if all of these samples are not evaluated initially for a given research project, collection and proper sample storage will make future research projects possible and enable ancillary diagnostic tests to be conducted if warranted based on unusual health findings.

### 8.3. Ultrastructural analysis (transmission electron microscopy)

Transmission electron microscopy (TEM) can be used to identify features of different parasite species as well as differentiate between tachyzoites and cysts and identify infected cells. Due to advances in diagnostic techniques in recent years, it is no longer a method of choice for routine diagnosis of *N. caninum* infection.

### 8.4. Seroprevalence studies

Serological assays are useful diagnostic tools to assess wild animals for evidence of *N. caninum* exposure, and techniques may

be applied antemortem or postmortem on sera and other biological fluids (e.g., CSF, thoracic fluid). In recent years, numerous investigations of *N. caninum* seroprevalence have been conducted in wildlife. Unfortunately, the data are not comparable between many studies due to the use of different techniques and lack of test validation in nondomestic species and results should be interpreted cautiously, particularly when only one serological method has been used (Gondim, 2006; Dubey et al., 2007; Dubey and Schares, 2011; Almeria, 2013). Many factors might influence whether or not antibodies are detected in wildlife by a given assay including postmortem degradation of immunoglobulins (if samples are obtained at necropsy), lack of appropriate species-specific secondary antibodies or conjugates, and potential for serological cross-reaction with closely related apicomplexan parasites (Dubey and Schares, 2006; Gondim, 2006; Almeria, 2013). Studies in cattle have shown that *N. caninum* specific antibodies can fluctuate during pregnancy and may fall below detectable levels (Dubey and Schares, 2011). Furthermore, infected animals do not always develop a detectable antibody response (De Marez et al., 1999; Lindsay et al., 1999). It is interesting to speculate that this may also be true for wild animals, especially given the above experimental results in birds, although additional research is required to identify if this is the case. Despite their limitations, serological studies have provided compelling evidence of *N. caninum* exposure in wildlife species.

Serological techniques for specific detection of *N. caninum* in domestic animals include the indirect fluorescent antibody test (IFAT), immunoblotting (IB), *N. caninum* direct agglutination test (NAT), and a variety of enzyme linked immunosorbent assays (ELISAs) (reviewed by Dubey and Schares, 2006). In wildlife, the most commonly used serological tests are the competitive enzyme-linked immunosorbent assay (cELISA) and *N. caninum*-agglutination test (NAT) because these techniques do not require the use of species specific secondary antibodies (Almeria, 2013). The usefulness and application of different serological techniques in wildlife species have been recently extensively reviewed (Almeria, 2013) and we encourage readers to seek out additional details from the provided reference. The limitation of these studies is that homologous true positive and negative sera are extremely difficult and often impossible to obtain, implying that cut-offs remain speculative.

### 8.5. Conventional and nested PCR

PCR is a highly sensitive and specific technique for detecting *N. caninum* DNA that may be applied to tissues and other clinical samples such as blood, CSF, and other body fluids. The *N. caninum* specific repetitive Nc5 gene and the internal transcribed spacer 1 (ITS1) region of the rRNA gene are the most common markers used for routine PCR-based *N. caninum* detection (Dubey and Schares, 2006). The ITS1 gene is a useful target for phylogenetic analysis between closely related taxa (e.g., *Toxoplasma*, *Hammondia*, and *Neospora*) because it is present at high copy number in the genome and exhibits interspecies variability and intraspecies conservation, allowing differentiation between *N. caninum* and other coccidian species (Holmdahl and Mattsson, 1996; Dubey et al., 2002; Gondim et al., 2004c; Al-Qassab et al., 2010). The D2 domain of the large subunit (D2-LSU) rDNA is also a phylogenetically important marker used to distinguish between closely related coccidian genera and species (Ellis et al., 1998, 1999; Slapeta et al., 2002). Amplification of ITS1/D2-LSU rDNA needs to be followed by direct bidirectional DNA sequencing to fully ascertain sequence identity. Since originally described by Muller et al. (1996), Holmdahl and Mattsson (1996), and Ellis et al. (1998), Nc5- and ITS1/D2-LSU rDNA-PCR protocols have been modified in many laboratories using a variety of primers and two-step nested PCR approaches targeting these markers have also been developed to enhance parasite detection in low level infections (Ellis et al., 1999; Slapeta et al., 2002, 2003; Dubey and

Schares, 2006). Although some *N. caninum* isolates have minor sequence variations in the Nc5 and ITS1/D2-LSU rDNA regions, sufficient polymorphisms have not been identified to allow these genes to be useful for strain differentiation (Slapeta et al., 2002; Gondim et al., 2004c; Al-Qassab et al., 2010).

In low-level infections, the ability to detect *N. caninum* by PCR depends largely upon sampling strategy. Because *N. caninum* is assumed to encyst most commonly in the brain (Dubey et al., 2007; Dubey and Schares, 2011), it is the tissue used most frequently in molecular surveys of *N. caninum* infection in wildlife. Multiple studies have identified parasite DNA in other tissues, in some instances with greater frequency than the brain (Hughes et al., 2006; Ferroglio et al., 2007; Kang et al., 2009; Gondim et al., 2010; Sangster et al., 2010; Truppel et al., 2010; Bartley et al., 2013b; Medina-Esparza et al., 2013). Analysis of multiple tissues has been shown to increase the sensitivity of *N. caninum* detection and additional tissues in which *N. caninum* DNA has been commonly recognized include skeletal muscle, heart, liver, kidney, spinal cord, and lung (Hughes et al., 2006; Ferroglio et al., 2007; Kang et al., 2009; Gondim et al., 2010; Sangster et al., 2010; Truppel et al., 2010; Bartley et al., 2013b; Medina-Esparza et al., 2013), with skeletal muscle and liver in rodents (Ferroglio et al., 2007; Kang et al., 2009; Truppel et al., 2010; Medina-Esparza et al., 2013) and heart in sparrows (Gondim et al., 2010) identified by some studies as the tissues most likely to be PCR positive for *N. caninum*. Studies have also shown that testing more than one section of brain significantly increases the likelihood of finding *N. caninum* DNA (Hughes et al., 2008; Stuart et al., 2013). The tissue distribution of parasites in wildlife is not known, so to preclude underestimation of infection, we advise sampling multiple tissues (listed above) and at least two samples of brain to assess for *N. caninum* infection in molecular investigations. Tissues for molecular studies should be aseptically collected and stored frozen at  $-20^{\circ}\text{C}$  until used. It is important to note that in some studies, tissue samples were PCR positive for the ITS1 region but did not amplify the Nc5 region, possibly due to variation in copy number within the genome, which emphasizes the importance of using more than one marker for molecular investigations (Jenkins et al., 2007; Truppel et al., 2010).

### 8.6. Quantitative PCR

At present, quantitative PCR for *N. caninum* is only used in academic research but we mention it here because it has the potential to be a valuable tool for investigations of *N. caninum* infection and pathogenesis of neosporosis in wildlife. Compared with conventional and nested PCR, qPCR has greater sensitivity, does not require as much starting material, and allows both detection and quantitative estimation of *N. caninum* in biological samples. Several different qPCR techniques targeting the Nc5 gene are described (Collantes-Fernandez et al., 2002; Muller et al., 2002; Okeoma et al., 2005; Ghalmi et al., 2008). To our knowledge, qPCR has not been used in any wildlife species.

### 8.7. Mini- and microsatellite analysis – tools for genotyping

Mini- and microsatellites are short repetitive DNA elements ubiquitously distributed throughout the genomes of eukaryotic and prokaryotic organisms that are being used with increasing frequency for studies of genetic diversity in many organisms, including protozoa (Basso et al., 2009, 2010; Al-Qassab et al., 2010; Regidor-Cerrillo et al., 2013). These tandemly repetitive DNA motifs vary from 2–6 nucleotides (microsatellites) to 8–100 nucleotides (minisatellites) in length (Al-Qassab et al., 2010). Unique genetic patterns have been established for different *N. caninum* strains by analysis of length polymorphisms in microsatellite and minisatellite containing areas, and this technology is currently considered the gold



standard for evaluating *N. caninum* genetic diversity (Al-Qassab et al., 2010; Goodswen et al., 2013). The microsatellite-containing region MS10 is a highly polymorphic locus that is particularly useful for characterizing divergence between different *N. caninum* strains and has shown excellent discriminatory power in detecting novel alleles (Basso et al., 2009; Al-Qassab et al., 2010). Recent description of multiplex microsatellite genotyping methods will enable the rapid assessment of *N. caninum* strain genetic diversity (Al-Qassab et al., 2010; Regidor-Cerrillo et al., 2013). Current applications include epidemiological investigation of abortion storms in cattle, differentiating between challenge and vaccine strains in vaccine development research, and population genetic diversity studies (Al-Qassab et al., 2010; Goodswen et al., 2013; Regidor-Cerrillo et al., 2013).

At present, most molecular investigations of *N. caninum* in wildlife have focused on detecting parasite DNA in tissue and little has been done to further characterize the parasite. Molecular genotyping techniques utilizing mini- and microsatellite technologies will be useful for epidemiological studies involving wildlife species and emerging new isolates and provide valuable information about the importance of sylvatic cycles in the biology of this parasite (Al-Qassab et al., 2010; Goodswen et al., 2013; Regidor-Cerrillo et al., 2013). Availability of the *N. caninum* draft genome sequence should be used to revisit the assay of markers and possibly develop a new array that may better define the *N. caninum* population structure.

### 8.8. Next generation sequencing (NGS)

Next generation sequencing (NGS) is an emerging high-throughput diagnostic technique that utilizes generic primers to amplify a conservative region of DNA found in all eukaryotic species followed by sequencing of numerous short strands of both host and microbial DNA (Sun et al., 2011; Fournier et al., 2013). In recent years, NGS technology has developed rapidly and become an important analytical tool used in many research investigations including human infectious disease investigations and assessments of microbial community composition in a variety of environments (Sun et al., 2011; Fournier et al., 2013; Slapeta and Linares, 2013). Advantages of this technology as compared to other diagnostic tests include identification of co-infections with other pathogens, and discriminatory capacity for genotyping and molecular epidemiologic analysis (Fournier et al., 2013; Slapeta and Linares, 2013). Disadvantages include cost of sequencing (although this is decreasing), unknown sensitivity of the generic primers for *N. caninum* and need for bioinformatics for sequence analysis of tens of thousands of sequence reads generated in a single assay (Fournier et al., 2013). NGS is not currently used in the standard diagnosis of *N. caninum* infection but it has been used successfully to identify the presence of apicomplexan protists (Slapeta and Linares, 2013) and represents a valuable diagnostic technique for investigations of wildlife disease and microbial community DNA analysis.

## 9. Neosporosis in wildlife: the right time for reflection

### 9.1. What is the impact of *N. caninum* infection on wildlife populations?

To date, an extensive number of wildlife species have been investigated for their possible role in the *N. caninum* life cycle and many have been implicated as intermediate hosts; however, the occurrence and importance of disease due to infection in nondomestic animals remain poorly understood. Most reports of *N. caninum* infection in wildlife species are based on serologic and/or molecular evidence in asymptomatic animals and, in many instances, investigations of possible associated morbidity, mortality, and pathology have been neglected. Histopathologic assessment to screen infected animals for evidence of pathologic changes and chronic infection

(i.e., presence of tissue cysts) is a critical factor in building a complete picture of infection outcome and identifying wild animal intermediate hosts. Although current data would indicate that *N. caninum* does not appear to adversely impact wildlife populations, as evidenced by the low number of animals reported with clinical versus subclinical infection, additional investigation is warranted to clarify the significance of infection in nondomestic species and the role it may play in possible parasite transmission in natural conditions. Because *N. caninum* tissue cysts can easily be mistaken for other tissue cyst forming apicomplexan parasites, the detection of a protozoan tissue cyst should be followed up with IHC.

### 9.2. How often is *N. caninum* infection clinically relevant and what factors predispose to pathologic sequelae in nondomestic species?

There are only 12 reports of neosporosis in wildlife species (summarized in Table 1). White rhinoceros are over-represented and account for 25% (3/12) of reported cases. Of the additional nine reports, all involved different species. Five cases were described in artiodactyls (four cervids and one bovine), three reports were in carnivores (two canids and one mustelid), and one report was in a marsupial (macropod). With the exception of one case, all reports of neosporosis in nondomestic species were in captive animals (9/12 cases; 75%) or free-ranging wildlife species maintained in rehabilitation facilities at the time of diagnosis (2/12 cases; 12.7%). Captivity induced stress is believed to play a role in the development of toxoplasmosis (Dubey, 2010), and although it is tempting to conclude that captivity is also a risk factor for *N. caninum*-associated disease, these findings may be more reflective of sampling bias because captive animals are far more likely to receive a complete postmortem assessment. Similar to domestic dogs and cattle, most reports of neosporosis are in young or fetal animals, with only 2/12 (16.7%) cases described in adults (Dubey et al., 2006, 2007; Dubey and Schares, 2011). Concurrent disease, identified as a risk factor for neosporosis in cattle (Dubey et al., 2007; Dubey and Schares, 2011), was only reported in a fox (8.3%); however, additional diagnostic tests were inconsistently described and unidentified concurrent subclinical disease processes cannot be ruled out.

Clinical disease and pathologic changes associated with *N. caninum* infection in wildlife species are comparable to what has been reported for domestic dogs and cattle. In summary, affected nondomestic species are commonly found dead (7/12 cases; 58.3%) or exhibit neuromuscular signs (4/12 cases; 33%). Encephalitis (6/12 cases; 50%), myocarditis (4/12 cases; 33%), and hepatic necrosis (3/12; 25%) are frequently reported lesions.

### 9.3. What are the best practice guidelines for reporting wildlife cases of neosporosis?

At present, there are no generally accepted recommendations for the optimal tissues to collect or tests to perform in order to diagnose *N. caninum* infection in nondomestic animals. With the goal of establishing greater international uniformity in making a diagnosis of neosporosis in wildlife, we propose that the following minimum criteria should be met whenever cases are suspected (Table 3).

A best practice diagnosis for *N. caninum* infection in wildlife should include the following:

- (1) A thorough clinical history and baseline health assessment should be obtained (whenever possible) on any animal exhibiting clinical disease or being handled for *N. caninum* screening purposes.
- (2) A necropsy including histopathology is always recommended for animals with suspected neosporosis. At necropsy, a variety of tissues should be collected and (a) stored frozen at  $-20^{\circ}\text{C}$

**Table 3**

Best practice protocol: criteria for reporting cases of neosporosis in wildlife species.

Technique	Minimum	Optimum	Aspirational	Comments
Clinical history	X	X	X	A thorough clinical history and baseline health assessment on any animal exhibiting clinical disease or being handled for <i>N. caninum</i> screening purposes. Block submitted to public repository.
Histopathology	X	X	X	A necropsy and histopathology for animals with suspected neosporosis. Lesions and parasites are most commonly identified in the brain, spinal cord, heart, skeletal muscle, liver, and placenta.
IHC	X	X	X	IHC increases the likelihood of detecting parasites over histopathology alone. To rule out presence of closely related parasites such as <i>Toxoplasma gondii</i> and antibodies cross reactivity, IHC for <i>T. gondii</i> in conjunction with IHC for <i>N. caninum</i> is done.*
Serology		X	X	cELISA and NAT as they do not require species specific secondary antibodies.**
PCR		X	X	<i>N. caninum</i> specific PCR (e.g., Nc5). PCR amplification of phylogenetically informative genes region (e.g., ITS1, D2 LSU rDNA) directly bidirectionally sequenced and submitted to public repository. Tissues in which <i>N. caninum</i> DNA is most commonly detected by PCR are brain, spinal cord, skeletal muscle, heart, liver, kidney, and lung. Due to uncertainty regarding parasite tissue distribution in wildlife, to maximize likelihood of parasite detection, PCR should be done on multiple tissues, including two sections or more of brain.
Multilocus genotyping			X	Available mini- and microsatellite <i>N. caninum</i> genotyping PCR assay applied to asses if multiple genotypes are present in wildlife cases.
Isolation			X	Brain is most commonly used for parasite isolation studies.
TEM			X	Rarely done.
Microbiome / NGS			X	Not reported to date.

For additional details on various diagnostic tests and sample collection, please refer to [Section 8](#).

\* At the Veterinary Pathology Diagnostics Services, the following antibodies have been routinely utilized: for IHC-*N.caninum* – polyclonal caprine anti-*N. caninum* antibodies (Product Code PAB-NC, VMRD, Inc., Veterinary Medical Research and Development, Pullman, WA, USA); for IHC-*T. gondii* – polyclonal rabbit anti-*T. gondii* antibodies (Product Code RB-282-A, Thermo-Fisher Scientific Australia).

\*\* Commercially available *N. caninum* serological tests are reviewed by [Dubey and Schares \(2006\)](#).

H&E, hematoxylin and eosin; IHC, immunohistochemistry; cELISA, competitive enzyme linked immunosorbent assay; NAT, Neospora agglutination test; TEM, transmission electron microscopy.

for future evaluation, and (b) fixed in 10% formalin and processed into paraffin blocks within 7 days. Tissue specimen collection should follow the guidelines outlined in [section 8.2](#) and [8.5](#).

- (3) Diagnosis of *N. caninum* infection necessitates the detection of parasite antigen and DNA using both IHC and PCR, respectively. Molecular diagnostics based on PCR should include at least two independent markers. Primers for cyst forming coccidia should be used to rule out presence of similar parasites (e.g., *Toxoplasma*, *Sarcocystis*, *Besnoitia*, and *Hammondia*). Tissue cyst presence should be reported and cross-reactivity with anti-*T. gondii* antibody confirmed. Correlation of immunological status with additional diagnostic tests should be implemented whenever possible to strengthen the diagnosis of neosporosis.
- (4) Materials such as paraffin blocks, frozen material and full case documentation should be deposited in a public depository for wide dissemination and retrieval.

This best practice protocol for reporting cases of *N. caninum* infection will optimize our knowledge about *N. caninum* infection in nondomestic species and enable us to better assess the consequences of infection, incidence of clinical disease, and impact of this parasite on wildlife populations. While the above recommendations are mentioned as aspirational, we believe that those reports that do not include such data should be returned to researchers to complete appropriate analyses.

The incidence of neosporosis is low and consequently it may not be on top of mind in wildlife deaths. Wild animals, especially prey species, are renowned for dying without displaying clinical signs. Optimal sample collection and storage will facilitate ancillary diagnostic tests and confirmation of *N. caninum* infection even when it may not be initially suspected.

## 10. Conclusions

Investigations to date of *N. caninum* infection in free-ranging and captive wildlife have opened a productive research agenda. The present paper has highlighted multiple gaps in our knowledge base and raised questions about the effects of this parasite in wild animal

health. Studies aimed at elucidating the role of wild animals in maintaining the parasite sylvatic cycle have identified many species as possible intermediate hosts and enhanced our knowledge about the parasite in wildlife. However, available data largely refer to detection of *N. caninum* antibodies and/or DNA, and there remain few reports of disease or associated pathological findings. It remains unclear whether or not *N. caninum* infection poses a substantial health risk to individuals within the nondomestic species, and the impact this has on a population basis. Furthermore, the disjoin between an expanding number of reports of *N. caninum* exposure and infection and the limited number (12) of studies reporting clinical neosporosis continue to increase. Reports must be interpreted conservatively when only one or a limited number of diagnostic tests are used to identify infection, particularly when histopathology is not available to assess host–parasite interaction and associated pathology. In order to ascertain the pathological impact of the parasite in wildlife populations and more accurately determine how often clinical disease attributable to *N. caninum* infection occurs, there is a need for greater uniformity in establishing a diagnosis of *N. caninum* infection. Therefore, our recommended protocol for reporting cases of neosporosis in nondomestic species offers an integrated approach for parasite detection and is intended to standardize future studies and allow comparisons between laboratories.

Unraveling the circumstances under which *N. caninum* infection triggers disease and chronic infections in nondomestic species continues to be a more difficult challenge. Immunological and pathogenicity investigations in wild animals are inherently challenging due to the lack of suitable experimental models. New animal models using susceptible animals are needed. Utilizing available *N. caninum* draft genome data and advanced diagnostic techniques in wildlife studies, such as microsatellite analysis and NGS technologies, will enhance our knowledge of *N. caninum* infection by facilitating epidemiological investigation, thereby allowing clinical manifestations of disease to be associated with parasite genotype and virulence, and the presence of co-infection with other microbial agents.

## Acknowledgements

The authors thank John Ellis, Milton McAllister, Bronwyn McAllen, Graeme Brown, Cheryl Sangster, Michelle Campbell-Ward, and Benn

Bryant for discussion and valuable insight on this topic. The authors also acknowledge the Taronga Conservation Society Australia for the use of photomicrographs and the University of Sydney Veterinary Pathology Diagnostic Services for excellent technical assistance. This research was supported by funding from The Whitehead Bequest and the Dr. Williams Award in Veterinary Pathology (University of Sydney). Shannon L. Donahoe is supported by the International Postgraduate Research Scholarship (IPRS) and an Australian Postgraduate Award (APA) tenable at The University of Sydney. The authors extend their apologies to those authors whose work could not be cited owing to space limitations.

### Conflict of interest

The authors declared that there is no conflict of interest.

### References

- Al-Qassab, S., Reichel, M.P., Ellis, J., 2010. On the biological and genetic diversity in *Neospora caninum*. *Diversity* 2, 411–438.
- Almberg, E.S., Mech, L.D., Smith, D.W., Sheldon, J.W., Crabtree, R.L., 2009. A serological survey of infectious disease in Yellowstone National Park's canid community. *PLoS ONE* 4, e7042.
- Almeria, S., 2013. *Neospora caninum* and wildlife. *ISRN Parasitol.* 2013, 1–23.
- Almeria, S., Ferrer, D., Pabon, M., Castilla, J., Manas, S., 2002. Red foxes (*Vulpes vulpes*) are a natural intermediate host of *Neospora caninum*. *Vet. Parasitol.* 107, 287–294.
- Almeria, S., Vidal, D., Ferrer, D., Pabon, M., Fernandez-de-Mera, M.I., Ruiz-Fons, F., et al., 2007. Seroprevalence of *Neospora caninum* in non-carnivorous wildlife from Spain. *Vet. Parasitol.* 143, 21–28.
- Anderson, T., DeJardin, A., Howe, D.K., Dubey, J.P., Michalski, M.L., 2007. *Neospora caninum* antibodies detected in Midwestern white-tailed deer (*Odocoileus virginianus*) by Western blot and ELISA. *Vet. Parasitol.* 145, 152–155.
- Andre, M.R., Adania, C.H., Teixeira, R.H., Silva, K.F., Jusi, M.M., Machado, S.T., et al., 2010. Antibodies to *Toxoplasma gondii* and *Neospora caninum* in captive neotropical and exotic wild canids and felids. *J. Parasitol.* 96, 1007–1009.
- Atkinson, R., Harper, P.A., Ryce, C., Morrison, D.A., Ellis, J.T., 1999. Comparison of the biological characteristics of two isolates of *Neospora caninum*. *Parasitology* 118 (Pt 4), 363–370.
- Barber, J.S., Trees, A.J., 1996. Clinical aspects of 27 cases of neosporosis in dogs. *Vet. Rec.* 139, 439–443.
- Barber, J.S., Gasser, R.B., Ellis, J., Reichel, M.P., McMillan, D., Trees, A.J., 1997. Prevalence of antibodies to *Neospora caninum* in different canid populations. *J. Parasitol.* 83, 1056–1058.
- Barling, K.S., Sherman, M., Peterson, M.J., Thompson, J.A., McNeill, J.W., Craig, T.M., et al., 2000. Spatial associations among density of cattle, abundance of wild canids, and seroprevalence to *Neospora caninum* in a population of beef calves. *J. Am. Vet. Med. Assoc.* 217, 1361–1365.
- Barr, B.C., Anderson, M.L., Dubey, J.P., Conrad, P.A., 1991. *Neospora*-like protozoal infections associated with bovine abortions. *Vet. Pathol.* 28, 110–116.
- Barr, B.C., Conrad, P.A., Sverlow, K.W., Tarantal, A.F., Hendrickx, A.G., 1994. Experimental fetal and transplacental *Neospora* infection in the nonhuman primate. *Lab. Invest.* 71, 236–242.
- Barratt, J., Al Qassab, S., Reichel, M.P., Ellis, J.T., 2008. The development and evaluation of a nested PCR assay for detection of *Neospora caninum* and *Hammondia heydorni* in feral mouse tissues. *Mol. Cell. Probes* 22, 228–233.
- Barratt, J.L., Harkness, J., Marriott, D., Ellis, J.T., Stark, D., 2010. Importance of nonenteric protozoan infections in immunocompromised people. *Clin. Microbiol. Rev.* 23, 795–836.
- Bartels, C.J., Wouda, W., Schukken, Y.H., 1999. Risk factors for *Neospora caninum*-associated abortion storms in dairy herds in the Netherlands (1995 to 1997). *Theriogenology* 52, 247–257.
- Bartels, C.J., Huinink, I., Beiboer, M.L., van Schaik, G., Wouda, W., Dijkstra, T., et al., 2007. Quantification of vertical and horizontal transmission of *Neospora caninum* infection in Dutch dairy herds. *Vet. Parasitol.* 148, 83–92.
- Bartley, P.M., Katzer, F., Rocchi, M.S., Maley, S.W., Benavides, J., Nath, M., et al., 2013a. Development of maternal and foetal immune responses in cattle following experimental challenge with *Neospora caninum* at day 210 of gestation. *Vet. Res.* 44, 91.
- Bartley, P.M., Wright, S.E., Zimmer, I.A., Roy, S., Kitchener, A.C., Meredith, A., et al., 2013b. Detection of *Neospora caninum* in wild carnivores in Great Britain. *Vet. Parasitol.* 192, 279–283.
- Bartova, E., Sedlak, K., Literak, I., 2006. Prevalence of *Toxoplasma gondii* and *Neospora caninum* antibodies in wild boars in the Czech Republic. *Vet. Parasitol.* 142, 150–153.
- Bartova, E., Sedlak, K., Tremil, F., Holko, I., Literak, I., 2010. *Neospora caninum* and *Toxoplasma gondii* antibodies in European brown hares in the Czech Republic, Slovakia and Austria. *Vet. Parasitol.* 171, 155–158.
- Basso, W., Venturini, L., Venturini, M.C., Moore, P., Rambeau, M., Unzaga, J.M., et al., 2001. Prevalence of *Neospora caninum* infection in dogs from beef-cattle farms, dairy farms, and from urban areas of Argentina. *J. Parasitol.* 87, 906–907.
- Basso, W., Schares, S., Barwald, A., Herrmann, D.C., Conrath, F.J., Pantchev, N., et al., 2009. Molecular comparison of *Neospora caninum* oocyst isolates from naturally infected dogs with cell culture-derived tachyzoites of the same isolates using nested polymerase chain reaction to amplify microsatellite markers. *Vet. Parasitol.* 160, 43–50.
- Basso, W., Schares, S., Minke, L., Barwald, A., Maksimov, A., Peters, M., et al., 2010. Microsatellite typing and avidity analysis suggest a common source of infection in herds with epidemic *Neospora caninum*-associated bovine abortion. *Vet. Parasitol.* 173, 24–31.
- Basso, W., More, G., Quiroga, M.A., Balducci, D., Schares, G., Venturini, M.C., 2014. *Neospora caninum* is a cause of perinatal mortality in axis deer (*Axis axis*). *Vet. Parasitol.* 199, 255–258.
- Belov, K., Miller, R.D., Old, J.M., Young, L.J., 2013. Marsupial immunology bounding ahead. *Aust. J. Zool.* 61, 24–40.
- Bevins, S., Blizard, E., Bazan, L., Whitley, P., 2013. *Neospora caninum* exposure in overlapping populations of coyotes (*Canis latrans*) and feral swine (*Sus scrofa*). *J. Wildl. Dis.* 49, 1028–1032.
- Bien, J., Moskwa, B., Cabaj, W., 2010. In vitro isolation and identification of the first *Neospora caninum* isolate from European bison (*Bison bonasus L.*). *Vet. Parasitol.* 173, 200–205.
- Billinis, C., 2013. Wildlife diseases that pose a risk to small ruminants and their farmers. *Small Ruminant Res.* 110, 67–70.
- Bjerkas, I., Mohn, S.F., Presthus, J., 1984. Unidentified cyst-forming sporozoon causing encephalomyelitis and myositis in dogs. *Z. Parasitenkd* 70, 271–274.
- Bjorkman, C., Jakubek, E.B., Arnemo, J.M., Malmsten, J., 2010. Seroprevalence of *Neospora caninum* in gray wolves in Scandinavia. *Vet. Parasitol.* 173, 139–142.
- Bottari, N.B., Tonin, A.A., Figuera, R., Flores, M.M., Franca, R.T., Camillo, G., et al., 2014. *Neospora caninum* and *Toxoplasma gondii*: relationship between hepatic lesions, cytological and biochemical analysis of the cavity liquid during the acute phase of the diseases in experimental models. *Exp. Parasitol.* 136, 68–73.
- Boyd, S.P., Barr, P.A., Brooks, H.W., Orr, J.P., 2005. Neosporosis in a young dog presenting with dermatitis and neuromuscular signs. *J. Small Anim. Pract.* 46, 85–88.
- Bregoli, M., Gioia, C., Stefano, N., Mariapia, C., Claudio, P., 2006. Serological survey of *Neospora caninum* in free-ranging wild ruminants. *Vet. Arhiv.* 76, S111–S115.
- Brown, C.B., Baker, D.C., Barker, I.K., 2007. The alimentary system. In: Maxie, M.G. (Ed.), *Jubb, Kennedy, and Palmer's Pathology of Domestic Animals*. Elsevier Saunders, Philadelphia, PA, pp. 272–273.
- Buchmann, K., 2014. Evolution of innate immunity: clues from invertebrates via fish to mammals. *Front Immunol.* 5, 459.
- Buxton, D., Maley, S.W., Pastoret, P.P., Brochier, B., Innes, E.A., 1997. Examination of red foxes (*Vulpes vulpes*) from Belgium for antibody to *Neospora caninum* and *Toxoplasma gondii*. *Vet. Rec.* 141, 308–309.
- Buxton, D., McAllister, M.M., Dubey, J.P., 2002. The comparative pathogenesis of neosporosis. *Trends Parasitol.* 18, 546–552.
- Cabaj, W., Moskwa, B., Pastusiak, K., Gill, J., 2005. Antibodies to *Neospora caninum* in the blood of European bison (*Bison bonasus bonasus L.*) living in Poland. *Vet. Parasitol.* 128, 163–168.
- Candela, M.G., Serrano, E., Sevilla, J., Leon, L., Caro, M.R., Verheyden, H., 2014. Pathogens of zoonotic and biological importance in roe deer (*Capreolus capreolus*): seroprevalence in an agro-system population in France. *Vet. Sci.* 96, 254–259.
- Canon-Franco, W.A., Yai, L.E., Souza, S.L., Santos, L.C., Farias, N.A., Ruas, J., et al., 2004. Detection of antibodies to *Neospora caninum* in two species of wild canids, *Lycalopex gymnocercus* and *Cerdocoyon thous* from Brazil. *Vet. Parasitol.* 123, 275–277.
- Cantile, C., Arispici, M., 2002. Necrotizing cerebellitis due to *Neospora caninum* infection in an old dog. *J. Vet. Med. A Physiol. Pathol. Clin. Med.* 49, 47–50.
- Cheadle, M.A., Spencer, J.A., Blagburn, B.L., 1999. Seroprevalences of *Neospora caninum* and *Toxoplasma gondii* in nondomestic felids from southern Africa. *J. Zoo Wildl. Med.* 30, 248–251.
- Cobadiova, A., Vichova, B., Majlathova, V., Reiterova, K., 2013. First molecular detection of *Neospora caninum* in European brown bear (*Ursus arctos*). *Vet. Parasitol.* 197, 346–349.
- Collantes-Fernandez, E., Zaballos, A., Alvarez-Garcia, G., Ortega-Mora, L.M., 2002. Quantitative detection of *Neospora caninum* in bovine aborted fetuses and experimentally infected mice by real-time PCR. *J. Clin. Microbiol.* 40, 1194–1198.
- Collantes-Fernandez, E., Gomez-Bautista, M., Miro, G., Alvarez-Garcia, G., Pereira-Bueno, J., Frisuelos, C., et al., 2008. Seroprevalence and risk factors associated with *Neospora caninum* infection in different dog populations in Spain. *Vet. Parasitol.* 152, 148–151.
- Conrad, P.A., Barr, B.C., Sverlow, K.W., Anderson, M., Daft, B., Kinde, H., et al., 1993. In vitro isolation and characterization of a *Neospora* sp. from aborted bovine foetuses. *Parasitology* 106 (Pt 3), 239–249.
- Costa, K.S., Santos, S.L., Uzeda, R.S., Pinheiro, A.M., Almeida, M.A., Araujo, F.R., et al., 2008. Chickens (*Gallus domesticus*) are natural intermediate hosts of *Neospora caninum*. *Int. J. Parasitol.* 38, 157–159.
- Cronstedt-Fell, A., Richter, B., Voracek, T., Kubber-Heiss, A., 2012. Neosporosis in a captive Parma wallaby (*Macropus parma*). *J. Comp. Pathol.* 146, 274–277.
- Cuddon, P., Lin, D.S., Bowman, D.D., Lindsay, D.S., Miller, T.K., Duncan, I.D., et al., 1992. *Neospora caninum* infection in English Springer Spaniel littermates. Diagnostic evaluation and organism isolation. *J. Vet. Intern. Med.* 6, 325–332.
- de Oliveira, U.V., de Magalhaes, V.C., Almeida, C.P., Santos Idos, A., Mota, D.A., Macedo, L.S., et al., 2013. Quails are resistant to infection with *Neospora caninum* tachyzoites. *Vet. Parasitol.* 198, 209–213.

- Darwich, L., Cabezon, O., Echeverria, I., Pabon, M., Marco, I., Molina-Lopez, R., et al., 2012. Presence of *Toxoplasma gondii* and *Neospora caninum* DNA in the brain of wild birds. *Vet. Parasitol.* 183, 377–381.
- Davison, H.C., Guy, C.S., McGarry, J.W., Guy, F., Williams, D.J., Kelly, D.F., et al., 2001. Experimental studies on the transmission of *Neospora caninum* between cattle. *Res. Vet. Sci.* 70, 163–168.
- De Craeye, S., Speybroeck, N., Ajzenberg, D., Darde, M.L., Collinet, F., Tavernier, P., et al., 2011. *Toxoplasma gondii* and *Neospora caninum* in wildlife: common parasites in Belgian foxes and Cervidae? *Vet. Parasitol.* 178, 64–69.
- De Marez, T., Liddell, S., Dubey, J.P., Jenkins, M.C., Gasbarre, L., 1999. Oral infection of calves with *Neospora caninum* oocysts from dogs: humoral and cellular immune responses. *Int. J. Parasitol.* 29, 1647–1657.
- Dellarupe, A., Regidor-Cerrillo, J., Jimenez-Ruiz, E., Schares, G., Unzaga, J.M., Venturini, M.C., et al., 2014. Clinical outcome and vertical transmission variability among canine *Neospora caninum* isolates in a pregnant mouse model of infection. *Parasitology* 141, 356–366.
- Dijkstra, T., Eysker, M., Schares, G., Conraths, F.J., Wouda, W., Barkema, H.W., 2001. Dogs shed *Neospora caninum* oocysts after ingestion of naturally infected bovine placenta but not after ingestion of colostrum spiked with *Neospora caninum* tachyzoites. *Int. J. Parasitol.* 31, 747–752.
- Dubey, J.P., 2003. Review of *Neospora caninum* and neosporosis in animals. *Korean J. Parasitol.* 41, 1–16.
- Dubey, J.P., 2010. *Toxoplasmosis of Animals and Humans*, second ed. CRC Press, Boca Raton, FL.
- Dubey, J.P., Lindsay, D.S., 1996. A review of *Neospora caninum* and neosporosis. *Vet. Parasitol.* 67, 1–59.
- Dubey, J.P., Lindsay, D.S., 2000. Gerbils (*Meriones unguiculatus*) are highly susceptible to oral infection with *Neospora caninum* oocysts. *Parasitol. Res.* 86, 165–168.
- Dubey, J.P., Schares, G., 2006. Diagnosis of bovine neosporosis. *Vet. Parasitol.* 140, 1–34.
- Dubey, J.P., Schares, G., 2011. Neosporosis in animals—the last five years. *Vet. Parasitol.* 180, 90–108.
- Dubey, J.P., Thulliez, P., 2005. Prevalence of antibodies to *Neospora caninum* in wild animals. *J. Parasitol.* 91, 1217–1218.
- Dubey, J.P., Carpenter, J.L., Speer, C.A., Topper, M.J., Uggla, A., 1988. Newly recognized fatal protozoan disease of dogs. *J. Am. Vet. Med. Assoc.* 192, 1269–1285.
- Dubey, J.P., Miller, S., Lindsay, D.S., Topper, M.J., 1990. *Neospora caninum*-associated myocarditis and encephalitis in an aborted calf. *J. Vet. Diagn. Invest.* 2, 66–69.
- Dubey, J.P., Rigoulet, J., Lagourette, P., George, C., Longeart, L., LeNet, J.L., 1996. Fatal transplacental neosporosis in a deer (*Cervus eldi siamensis*). *J. Parasitol.* 82, 338–339.
- Dubey, J.P., Hollis, K., Romand, S., Thulliez, P., Kwok, O.C., Hungerford, L., et al., 1999. High prevalence of antibodies to *Neospora caninum* in white-tailed deer (*Odocoileus virginianus*). *Int. J. Parasitol.* 29, 1709–1711.
- Dubey, J.P., Barr, B.C., Barta, J.R., Bjerkas, I., Bjorkman, C., Blagburn, B.L., et al., 2002. Redescription of *Neospora caninum* and its differentiation from related coccidia. *Int. J. Parasitol.* 32, 929–946.
- Dubey, J.P., Zarnke, R., Thomas, N.J., Wong, S.K., Van Bonn, W., Briggs, M., et al., 2003. *Toxoplasma gondii*, *Neospora caninum*, *Sarcocystis neurona*, and *Sarcocystis canis*-like infections in marine mammals. *Vet. Parasitol.* 116, 275–296.
- Dubey, J.P., Sreekumar, C., Knickman, E., Miska, K.B., Vianna, M.C., Kwok, O.C., et al., 2004. Biologic, morphologic, and molecular characterisation of *Neospora caninum* isolates from littermate dogs. *Int. J. Parasitol.* 34, 1157–1167.
- Dubey, J.P., Knickman, E., Greene, C.E., 2005. Neonatal *Neospora caninum* infections in dogs. *Acta Parasitol.* 50, 176–179.
- Dubey, J.P., Buxton, D., Wouda, W., 2006. Pathogenesis of bovine neosporosis. *J. Comp. Pathol.* 134, 267–289.
- Dubey, J.P., Schares, G., Ortega-Mora, L.M., 2007. Epidemiology and control of neosporosis and *Neospora caninum*. *Clin. Microbiol. Rev.* 20, 323–367.
- Dubey, J.P., Mansfield, K., Hall, B., Kwok, O.C., Thulliez, P., 2008. Seroprevalence of *Neospora caninum* and *Toxoplasma gondii* in black-tailed deer (*Odocoileus hemionus columbianus*) and mule deer (*Odocoileus hemionus hemionus*). *Vet. Parasitol.* 156, 310–313.
- Dubey, J.P., Jenkins, M.C., Kwok, O.C., Zink, R.L., Michalski, M.L., Ulrich, V., et al., 2009. Seroprevalence of *Neospora caninum* and *Toxoplasma gondii* antibodies in white-tailed deer (*Odocoileus virginianus*) from Iowa and Minnesota using four serologic tests. *Vet. Parasitol.* 161, 330–334.
- Dubey, J.P., Jenkins, M.C., Rajendran, C., Miska, K., Ferreira, L.R., Martins, J., et al., 2011. Gray wolf (*Canis lupus*) is a natural definitive host for *Neospora caninum*. *Vet. Parasitol.* 181, 382–387.
- Dubey, J.P., Jenkins, M.C., Kwok, O.C., Ferreira, L.R., Choudhary, S., Verma, S.K., et al., 2013. Congenital transmission of *Neospora caninum* in white-tailed deer (*Odocoileus virginianus*). *Vet. Parasitol.* 196, 519–522.
- Dubey, J.P., Jenkins, M.C., Ferreira, L.R., Choudhary, S., Verma, S.K., Kwok, O.C., et al., 2014a. Isolation of viable *Neospora caninum* from brains of wild gray wolves (*Canis lupus*). *Vet. Parasitol.* 201, 150–153.
- Dubey, J.P., Whitesell, L.E., Culp, W.E., Daye, S., 2014b. Diagnosis and treatment of *Neospora caninum*-associated dermatitis in a red fox (*Vulpes vulpes*) with concurrent *Toxoplasma gondii* infection. *J. Zoo Wildl. Med.* 45, 454–457.
- Ellis, J.T., Amoyal, G., Ryce, C., Harper, P.A., Clough, K.A., Homan, W.L., et al., 1998. Comparison of the large subunit ribosomal DNA of *Neospora* and *Toxoplasma* and development of a new genetic marker for their differentiation based on the D2 domain. *Mol. Cell. Probes* 12, 1–13.
- Ellis, J.T., Morrison, D.A., Liddell, S., Jenkins, M.C., Mohammed, O.B., Ryce, C., et al., 1999. The genus *Hammondia* is paraphyletic. *Parasitology* 118 (Pt 4), 357–362.
- Ezio, F., Anna, T., 2003. Antibodies to *Neospora caninum* in European brown hare (*Lepus europaeus*). *Vet. Parasitol.* 115, 75–78.
- Ferroglio, E., Rossi, L., 2001. Prevalence of *Neospora caninum* antibodies in wild ruminants from the Italian Alps. *Vet. Rec.* 148, 754–755.
- Ferroglio, E., Wambwa, E., Castiello, M., Trisciuglio, A., Prouteau, A., Pradere, E., et al., 2003. Antibodies to *Neospora caninum* in wild animals from Kenya, East Africa. *Vet. Parasitol.* 118, 43–49.
- Ferroglio, E., Pasino, M., Romano, A., Grande, D., Pregel, P., Trisciuglio, A., 2007. Evidence of *Neospora caninum* DNA in wild rodents. *Vet. Parasitol.* 148, 346–349.
- Fournier, P.-E., Drancourt, M., Colson, P., Rolain, J.-M., Scola, B.L., Raoult, D., 2013. Modern clinical microbiology: new challenges and solutions. *Nat. Rev. Microbiol.* 11, 574–585.
- French, N.P., Clancy, D., Davison, H.C., Trees, A.J., 1999. Mathematical models of *Neospora caninum* infection in dairy cattle: transmission and options for control. *Int. J. Parasitol.* 29, 1691–1704.
- Fuehrer, H.P., Bloschl, I., Siehs, C., Hassl, A., 2010. Detection of *Toxoplasma gondii*, *Neospora caninum*, and *Encephalitozoon cuniculi* in the brains of common voles (*Microtus arvalis*) and water voles (*Arvicola terrestris*) by gene amplification techniques in western Austria (Vorarlberg). *Parasitol. Res.* 107, 469–473.
- Fujii, K., Kakumoto, C., Kobayashi, M., Saito, S., Kariya, T., Watanabe, Y., et al., 2007. Seroprevalence of *Toxoplasma gondii* and *Neospora caninum* in seals around Hokkaido, Japan. *J. Vet. Med. Sci.* 69, 393–398.
- Furuta, P.I., Mineo, T.W., Carrasco, A.O., Godoy, G.S., Pinto, A.A., Machado, R.Z., 2007. *Neospora caninum* infection in birds: experimental infections in chicken and embryonated eggs. *Parasitology* 134, 1931–1939.
- Gaffuri, A., Giacometti, M., Tranquillo, V.M., Magnino, S., Cordioli, P., Lanfranchi, P., 2006. Serosurvey of roe deer, chamois and domestic sheep in the central Italian Alps. *J. Wildl. Dis.* 42, 685–690.
- Garcia-Bocanegra, I., Cabezon, O., Pabon, M., Gomez-Guillamon, F., Arenas, A., Alcaide, E., et al., 2012. Prevalence of *Toxoplasma gondii* and *Neospora caninum* antibodies in Spanish ibex (*Capra pyrenaica hispanica*). *Vet. J.* 191, 257–260.
- Garosi, L., Dawson, A., Couturier, J., Matiasek, L., de Stefani, A., Davies, E., et al., 2010. Necrotizing cerebellitis and cerebellar atrophy caused by *Neospora caninum* infection: magnetic resonance imaging and clinicopathologic findings in seven dogs. *J. Vet. Intern. Med.* 24, 571–578.
- Ghalmi, F., China, B., Kaidi, R., Daube, G., Losson, B., 2008. Detection of *Neospora caninum* in dog organs using real time PCR systems. *Vet. Parasitol.* 155, 161–167.
- Gibney, E.H., Kipar, A., Rosbottom, A., Guy, C.S., Smith, R.F., Hetzel, U., et al., 2008. The extent of parasite-associated necrosis in the placenta and foetal tissues of cattle following *Neospora caninum* infection in early and late gestation correlates with foetal death. *Int. J. Parasitol.* 38, 579–588.
- Ginn, P.E., Mansell, J.E.K., Rakich, P.M., 2007. The skin and appendages. In: Maxie, M.G. (Ed.), *Jub, Kennedy, and Palmer's Pathology of Domestic Animals*. Elsevier Saunders, Philadelphia, PA, p. 711.
- Gondim, L.F., 2006. *Neospora caninum* in wildlife. *Trends Parasitol.* 22, 247–252.
- Gondim, L.F., Saeki, H., Onaga, H., Haritani, M., Yamane, I., 1999. Maintenance of *Neospora caninum* tachyzoites using Mongolian gerbils (*Meriones unguiculatus*). *N. Z. Vet. J.* 47, 36.
- Gondim, L.F., Pinheiro, A.M., Santos, P.O., Jesus, E.E., Ribeiro, M.B., Fernandes, H.S., et al., 2001. Isolation of *Neospora caninum* from the brain of a naturally infected dog, and production of encysted bradyzoites in gerbils. *Vet. Parasitol.* 101, 1–7.
- Gondim, L.F., McAllister, M.M., Mateus-Pinilla, N.E., Pitt, W.C., Mech, L.D., Nelson, M.E., 2004a. Transmission of *Neospora caninum* between wild and domestic animals. *J. Parasitol.* 90, 1361–1365.
- Gondim, L.F., McAllister, M.M., Pitt, W.C., Zemlicka, D.E., 2004b. Coyotes (*Canis latrans*) are definitive hosts of *Neospora caninum*. *Int. J. Parasitol.* 34, 159–161.
- Gondim, L.F., Laski, P., Gao, L., McAllister, M.M., 2004c. Variation of the internal transcribed spacer 1 sequence within individual strains and among different strains of *Neospora caninum*. *J. Parasitol.* 90, 119–122.
- Gondim, L.S., Abe-Sandes, K., Uzeda, R.S., Silva, M.S., Santos, S.L., Mota, R.A., et al., 2010. *Toxoplasma gondii* and *Neospora caninum* in sparrows (*Passer domesticus*) in the Northeast of Brazil. *Vet. Parasitol.* 168, 121–124.
- Goodswen, S.J., Kennedy, P.J., Ellis, J.T., 2013. A review of the infection, genetics, and evolution of *Neospora caninum*: from the past to the present. *Infect. Genet. Evol.* 13, 133–150.
- Goździk, K., Jakubek, E.B., Bjorkman, C., Bien, J., Moskwa, B., Cabaj, W., 2010. Seroprevalence of *Neospora caninum* in free living and farmed red deer (*Cervus elaphus*) in Poland. *Pol. J. Vet. Sci.* 13, 117–120.
- Gutierrez-Exposito, D., Miguel Ortega-Mora, L., Gajadhar, A.A., Garcia-Lunar, P., Dubey, J.P., Alvarez-Garcia, G., 2012. Serological evidence of *Besnoitia* spp. infection in Canadian wild ruminants and strong cross-reaction between *Besnoitia besnoiti* and *Besnoitia tarandi*. *Vet. Parasitol.* 190, 19–28.
- Hamilton, C.M., Gray, R., Wright, S.E., Gangadharan, B., Laurenson, K., Innes, E.A., 2005. Prevalence of antibodies to *Toxoplasma gondii* and *Neospora caninum* in red foxes (*Vulpes vulpes*) from around the UK. *Vet. Parasitol.* 130, 169–173.
- Helman, R.G., Stair, E.L., Lehenbauer, T.W., Rodgers, S., Saliki, J.T., 1998. Neosporal abortion in Oklahoma cattle with emphasis on the distribution of brain lesions in aborted fetuses. *J. Vet. Diagn. Invest.* 10, 292–295.
- Hemphill, A., Vonlaufen, N., Naguleswaran, A., 2006. Cellular and immunological basis of the host-parasite relationship during infection with *Neospora caninum*. *Parasitology* 133, 261–278.
- Hobson, J.C., Duffield, T.F., Keltou, D., Lissemore, K., Hietala, S.K., Leslie, K.E., et al., 2005. Risk factors associated with *Neospora caninum* abortion in Ontario Holstein dairy herds. *Vet. Parasitol.* 127, 177–188.

- Holmdahl, O.J., Mattsson, J.G., 1996. Rapid and sensitive identification of *Neospora caninum* by in vitro amplification of the internal transcribed spacer 1. *Parasitology* 112 (Pt 2), 177–182.
- Hornok, S., Edelhofer, R., Fok, E., Berta, K., Fejes, P., Repasi, A., et al., 2006. Canine neosporosis in Hungary: screening for seroconversion of household, herding and stray dogs. *Vet. Parasitol.* 137, 197–201.
- Huang, C.C., Yang, C.H., Watanabe, Y., Liao, Y.K., Ooi, H.K., 2004. Finding of *Neospora caninum* in the wild brown rat (*Rattus norvegicus*). *Vet. Res.* 35, 283–290.
- Hughes, J.M., Williams, R.H., Morley, E.K., Cook, D.A., Terry, R.S., Murphy, R.G., et al., 2006. The prevalence of *Neospora caninum* and co-infection with *Toxoplasma gondii* by PCR analysis in naturally occurring mammal populations. *Parasitology* 132, 29–36.
- Hughes, J.M., Thomasson, D., Craig, P.S., Georgin, S., Pickles, A., Hide, G., 2008. *Neospora caninum*: detection in wild rabbits and investigation of co-infection with *Toxoplasma gondii* by PCR analysis. *Exp. Parasitol.* 120, 255–260.
- Hurkova, L., Modry, D., 2006. PCR detection of *Neospora caninum*, *Toxoplasma gondii* and *Encephalitozoon cuniculi* in brains of wild carnivores. *Vet. Parasitol.* 137, 150–154.
- Hurkova-Hofmannova, L., Vaclavek, P., Skoric, M., Fictum, P., Modry, D., 2007. Multimammate rat (*Mastomys natalensis*), Tristram's jird (*Meriones tristrami*) and Wagner's gerbil (*Gerbillus dasyurus*) as laboratory models of acute neosporosis. *Res. Vet. Sci.* 82, 377–381.
- Ibrahim, H.M., Huang, P., Salem, T.A., Talaat, R.M., Nasr, M.I., Xuan, X., et al., 2009. Short report: prevalence of *Neospora caninum* and *Toxoplasma gondii* antibodies in northern Egypt. *Am. J. Trop. Med. Hyg.* 80, 263–267.
- Innes, E.A., Andrianarivo, A.G., Bjorkman, C., Williams, D.J., Conrad, P.A., 2002. Immune responses to *Neospora caninum* and prospects for vaccination. *Trends Parasitol.* 18, 497–504.
- Innes, E.A., Wright, S., Bartley, P., Maley, S., Macalodow, C., Esteban-Redondo, I., et al., 2005. The host-parasite relationship in bovine neosporosis. *Vet. Immunol. Immunopathol.* 108, 29–36.
- Jackson, W., de Lahunta, A., Adaska, J., Cooper, B., Dubey, J.P., 1995. *Neospora caninum* in an adult dog with progressive cerebellar signs. *Prog. Vet. Neurol.* 6, 124–127.
- Jakubek, E.B., Farkas, R., Palfi, V., Mattsson, J.G., 2007. Prevalence of antibodies against *Toxoplasma gondii* and *Neospora caninum* in Hungarian red foxes (*Vulpes vulpes*). *Vet. Parasitol.* 144, 39–44.
- Jenkins, M.C., Parker, C., Hill, D., Pinckney, R.D., Dyer, R., Dubey, J.P., 2007. *Neospora caninum* detected in feral rodents. *Vet. Parasitol.* 143, 161–165.
- Kamga-Waladjo, A.R., Gbati, O.B., Kone, P., Lapo, R.A., Dombou, E., Chatagnon, G., et al., 2009. *Neospora caninum* and *Toxoplasma gondii* in lion (*Panthera leo*) from Senegal, West Africa. *Asian J. Anim. Vet. Adv.* 4, 346–349.
- Kang, S.W., Park, S.S., Choe, S.E., Jean, Y.H., Jung, S.C., Kim, K., et al., 2009. Characterization of tissue distribution and histopathological lesions in *Neospora caninum* experimentally infected gerbils. *Parasitol. Res.* 104, 1261–1268.
- Kim, J.H., Sohn, H.J., Hwang, W.S., Hwang, E.K., Jean, Y.H., Yamane, I., et al., 2000. In vitro isolation and characterization of bovine *Neospora caninum* in Korea. *Vet. Parasitol.* 90, 147–154.
- Kim, J.H., Kang, M.S., Lee, B.C., Hwang, W.S., Lee, C.W., So, B.J., et al., 2003. Seroprevalence of antibodies to *Neospora caninum* in dogs and raccoon dogs in Korea. *Korean J. Parasitol.* 41, 243–245.
- King, J.S., Slapeta, J., Jenkins, D.J., Al-Qassab, S.E., Ellis, J.T., Windsor, P.A., 2010. Australian dingoes are definitive hosts of *Neospora caninum*. *Int. J. Parasitol.* 40, 945–950.
- King, J.S., McAllan, B., Spielman, D.S., Lindsay, S.A., Hurkova-Hofmannova, L., Hartigan, A., et al., 2011a. Extensive production of *Neospora caninum* tissue cysts in a carnivorous marsupial succumbing to experimental neosporosis. *Vet. Res.* 42, 75.
- King, J.S., Jenkins, D.J., Ellis, J.T., Fleming, P., Windsor, P.A., Slapeta, J., 2011b. Implications of wild dog ecology on the sylvatic and domestic life cycle of *Neospora caninum* in Australia. *Vet. J.* 188, 24–33.
- King, J.S., Brown, G.K., Jenkins, D.J., Ellis, J.T., Fleming, P.J., Windsor, P.A., et al., 2012. Oocysts and high seroprevalence of *Neospora caninum* in dogs living in remote Aboriginal communities and wild dogs in Australia. *Vet. Parasitol.* 187, 85–92.
- Klevar, S., Kulberg, S., Boysen, P., Storset, A.K., Moldal, T., Bjorkman, C., et al., 2007. Natural killer cells act as early responders in an experimental infection with *Neospora caninum* in calves. *Int. J. Parasitol.* 37, 329–339.
- La Perle, K.M., Del Piero, F., Carr, R.F., Harris, C., Stromberg, P.C., 2001. Cutaneous neosporosis in two adult dogs on chronic immunosuppressive therapy. *J. Vet. Diagn. Invest.* 13, 252–255.
- Lemberger, K.Y., Gondim, L.F., Pessier, A.P., McAllister, M.M., Kinsel, M.J., 2005. *Neospora caninum* infection in a free-ranging raccoon (*Procyon lotor*) with concurrent canine distemper virus infection. *J. Parasitol.* 91, 960–961.
- Lindsay, D.S., Dubey, J.P., 1989. *Neospora caninum* (Protozoa: apicomplexa) infections in mice. *J. Parasitol.* 75, 772–779.
- Lindsay, D.S., Kelly, E.J., McKown, R.D., Stein, F.J., Plozer, J., Herman, J., et al., 1996. Prevalence of *Neospora caninum* and *Toxoplasma gondii* antibodies in coyotes (*Canis latrans*) and experimental infections of coyotes with *Neospora caninum*. *J. Parasitol.* 82, 657–659.
- Lindsay, D.S., Dubey, J.P., Duncan, R.B., 1999. Confirmation that the dog is a definitive host for *Neospora caninum*. *Vet. Parasitol.* 82, 327–333.
- Lindsay, D.S., Spencer, J., Rupprecht, C., Blagburn, B.L., 2001. Prevalence of agglutinating antibodies to *Neospora caninum* in raccoons, *Procyon lotor*. *J. Parasitol.* 87, 1197–1198.
- Lindsay, D.S., Little, S.E., Davidson, W.R., 2002. Prevalence of antibodies to *Neospora caninum* in white-tailed deer, *Odocoileus virginianus*, from the southeastern United States. *J. Parasitol.* 88, 415–417.
- Litman, G.W., Rast, J.P., Fugmann, S.D., 2010. The origins of vertebrate adaptive immunity. *Nat. Rev. Immunol.* 10, 543–553.
- Lobato, J., Silva, D.A., Mineo, T.W., Amaral, J.D., Segundo, G.R., Costa-Cruz, J.M., et al., 2006. Detection of immunoglobulin G antibodies to *Neospora caninum* in humans: high seropositivity rates in patients who are infected by human immunodeficiency virus or have neurological disorders. *Clin. Vaccine Immunol.* 13, 84–89.
- Lorenzo, V., Pumarola, M., Siso, S., 2002. Neosporosis with cerebellar involvement in an adult dog. *J. Small Anim. Pract.* 43, 76–79.
- Malmasi, A., Hosseini, M., Haddadzadeh, H., Badii, A., Bahonar, A., 2007. Serologic study of anti-*Neospora caninum* antibodies in household dogs and dogs living in dairy and beef cattle farms in Tehran, Iran. *Parasitol. Res.* 100, 1143–1145.
- Malmsten, J., Jakubek, E.B., Bjorkman, C., 2011. Prevalence of antibodies against *Toxoplasma gondii* and *Neospora caninum* in moose (*Alces alces*) and roe deer (*Capreolus capreolus*) in Sweden. *Vet. Parasitol.* 177, 275–280.
- Marco, I., Ferroglia, E., Lopez-Olvera, J.R., Montane, J., Lavin, S., 2008. High seroprevalence of *Neospora caninum* in the red fox (*Vulpes vulpes*) in the Pyrenees (NE Spain). *Vet. Parasitol.* 152, 321–324.
- Martino, P.E., Montenegro, J.L., Preziosi, J.A., Venturini, C., Bacigalupe, D., Stanchi, N.O., et al., 2004. Serological survey of selected pathogens of free-ranging foxes in southern Argentina, 1998–2001. *Rev. Sci. Tech.* 23, 801–806.
- Mattos, B.C., Patricio, L.L., Plugge, N.F., Lange, R.R., Richartz, R.R., Dittrich, R.L., 2008. [Seroprevalence of antibodies anti-*Neospora caninum* and anti-*Toxoplasma gondii* in captive wild canids]. *Rev. Bras. Parasitol. Vet.* 17 (Suppl. 1), 267–272.
- Mayberry, C., Maloney, S.K., Mitchell, J., Mawson, P.R., Bencini, R., 2014. Reproductive implications of exposure to *Toxoplasma gondii* and *Neospora caninum* in western grey kangaroos (*Macropus fuliginosus ocydromus*). *J. Wildl. Dis.* 50, 364–368.
- McAllister, M.M., Dubey, J.P., Lindsay, D.S., Jolley, W.R., Wills, R.A., McGuire, A.M., 1998. Dogs are definitive hosts of *Neospora caninum*. *Int. J. Parasitol.* 28, 1473–1478.
- McGuire, A.M., McAllister, M., Wills, R.A., Tranas, J.D., 1999. Experimental inoculation of domestic pigeons (*Columba livia*) and zebra finches (*Poephila guttata*) with *Neospora caninum* tachyzoites. *Int. J. Parasitol.* 29, 1525–1529.
- McInnes, L.M., Irwin, P., Palmer, D.G., Ryan, U.M., 2006. In vitro isolation and characterisation of the first canine *Neospora caninum* isolate in Australia. *Vet. Parasitol.* 137, 355–363.
- Medina-Esparza, L., Macias, L., Ramos-Parra, M., Morales-Salinas, E., Quezada, T., Cruz-Vazquez, C., 2013. Frequency of infection by *Neospora caninum* in wild rodents associated with dairy farms in Aguascalientes, Mexico. *Vet. Parasitol.* 191, 11–14.
- Meerburg, B.G., De Craeye, S., Dierick, K., Kijlstra, A., 2012. *Neospora caninum* and *Toxoplasma gondii* in brain tissue of feral rodents and insectivores caught on farms in the Netherlands. *Vet. Parasitol.* 184, 317–320.
- Millan, J., Candela, M.G., Palomares, F., Cubero, M.J., Rodriguez, A., Barral, M., et al., 2009. Disease threats to the endangered Iberian lynx (*Lynx pardinus*). *Vet. J.* 182, 114–124.
- Miller, C.M., Quinn, H.E., Windsor, P.A., Ellis, J.T., 2002. Characterisation of the first Australian isolate of *Neospora caninum* from cattle. *Aust. Vet. J.* 80, 620–625.
- Miller, M.A., Conrad, P.A., Harris, M., Hatfield, B., Langlois, G., Jessup, D.A., et al., 2010. A protozoal-associated epizootic impacting marine wildlife: mass-mortality of southern sea otters (*Enhydra lutris nereis*) due to *Sarcocystis neurona* infection. *Vet. Parasitol.* 172, 183–194.
- Mineo, T.W., Carrasco, A.O., Marciano, J.A., Werther, K., Pinto, A.A., Machado, R.Z., 2009. Pigeons (*Columba livia*) are a suitable experimental model for *Neospora caninum* infection in birds. *Vet. Parasitol.* 159, 149–153.
- Mineo, T.W., Carrasco, A.O., Raso, T.F., Werther, K., Pinto, A.A., Machado, R.Z., 2011. Survey for natural *Neospora caninum* infection in wild and captive birds. *Vet. Parasitol.* 182, 352–355.
- Molina-Lopez, R., Cabezon, O., Pabon, M., Darwich, L., Obon, E., Lopez-Gatius, F., et al., 2012. High seroprevalence of *Toxoplasma gondii* and *Neospora caninum* in the common raven (*Corvus corax*) in the Northeast of Spain. *Res. Vet. Sci.* 93, 300–302.
- Mols-Vorstermans, T., Hemphill, A., Monney, T., Schaap, D., Boerhout, E., 2013. Differential effects on survival, humoral immune responses and brain lesions in inbred BALB/C, CBA/CA, and C57BL/6 mice experimentally infected with *Neospora caninum* tachyzoites. *ISRN Parasitol.* 2013, 11.
- Monney, T., Hemphill, A., 2014. Vaccines against neosporosis: what can we learn from the past studies? *Exp. Parasitol.* 140, 52–70.
- Moore, D.P., Campero, C.M., Odeon, A.C., Posso, M.A., Cano, D., Leunda, M.R., et al., 2002. Seroprevalence of beef and dairy herds and fetal study of *Neospora caninum* in Argentina. *Vet. Parasitol.* 107, 303–316.
- Moskwa, B., Gozdziak, K., Bien, J., Kornacka, A., Cybulska, A., Reiterova, K., et al., 2014. Detection of antibodies to *Neospora caninum* in moose (*Alces alces*): the first report in Europe. *Folia Parasitol. (Praha)* 61, 34–36.
- Muller, N., Zimmermann, V., Hentrich, B., Gottstein, B., 1996. Diagnosis of *Neospora caninum* and *Toxoplasma gondii* infection by PCR and DNA hybridization immunoassay. *J. Clin. Microbiol.* 34, 2850–2852.
- Muller, N., Vonlaufen, N., Gianinazzi, C., Leib, S.L., Hemphill, A., 2002. Application of real-time fluorescent PCR for quantitative assessment of *Neospora caninum* infections in organotypic slice cultures of rat central nervous system tissue. *J. Clin. Microbiol.* 40, 252–255.
- Murphy, T.M., Walochnik, J., Hassl, A., Moriarty, J., Mooney, J., Toolan, D., et al., 2007. Study on the prevalence of *Toxoplasma gondii* and *Neospora caninum* and molecular evidence of *Encephalitozoon cuniculi* and *Encephalitozoon (Septata) intestinalis* infections in red foxes (*Vulpes vulpes*) in rural Ireland. *Vet. Parasitol.* 146, 227–234.

- Myers, W.L., Foreyt, W.J., Talcott, P.A., Evermann, J.F., Chang, W.Y., 2014. Serologic, trace element, and fecal parasite survey of free-ranging, female mule deer (*Odocoileus hemionus*) in Eastern Washington, USA. *J. Wildl. Dis.* 51, 125–136.
- Nishimura, M., Kohara, J., Hiasa, J., Muroi, Y., Yokoyama, N., Kida, K., et al., 2013. Tissue distribution of *Neospora caninum* in experimentally infected cattle. *Clin. Vaccine Immunol.* 20, 309–312.
- Okeoma, C.M., Stowell, K.M., Williamson, N.B., Pomroy, W.E., 2005. *Neospora caninum*: quantification of DNA in the blood of naturally infected aborted and pregnant cows using real-time PCR. *Exp. Parasitol.* 110, 48–55.
- Olamendi-Portugal, M., Caballero-Ortega, H., Correa, D., Sanchez-Aleman, M.A., Cruz-Vazquez, C., Medina-Esparza, L., et al., 2012. Serosurvey of antibodies against *Toxoplasma gondii* and *Neospora caninum* in white-tailed deer from Northern Mexico. *Vet. Parasitol.* 189, 369–373.
- Ordeix, L., Lloret, A., Fondevila, D., Dubey, J.P., Ferrer, L., Fondati, A., 2002. Cutaneous neosporosis during treatment of pemphigus foliaceus in a dog. *J. Am. Anim. Hosp. Assoc.* 38, 415–419.
- Ortega-Mora, L., 2007. Protozoal Abortion in Farm Ruminants: Guidelines for Diagnosis and Control. CABI, Wallingford, England.
- Otranto, D., Lazari, A., Testini, G., Traversa, D., Frangipane di Regalbono, A., Badan, M., et al., 2003. Seroprevalence and associated risk factors of neosporosis in beef and dairy cattle in Italy. *Vet. Parasitol.* 118, 7–18.
- Ould-Amrouche, A., Klein, F., Osdoit, C., Mohammed, H.O., Touratier, A., Sanaa, M., et al., 1999. Estimation of *Neospora caninum* seroprevalence in dairy cattle from Normandy, France. *Vet. Res.* 30, 531–538.
- Panadero, R., Panceira, A., Lopez, C., Vazquez, L., Paz, A., Diaz, P., et al., 2010. Seroprevalence of *Toxoplasma gondii* and *Neospora caninum* in wild and domestic ruminants sharing pastures in Galicia (Northwest Spain). *Res. Vet. Sci.* 88, 111–115.
- Peters, M., Wohlsein, P., Knieriem, A., Schares, G., 2001. *Neospora caninum* infection associated with stillbirths in captive antelopes (*Tragelaphus imberbis*). *Vet. Parasitol.* 97, 153–157.
- Pipano, E., Shkap, V., Fish, L., Savitsky, I., Perl, S., Orgad, U., 2002. Susceptibility of *Psammomys obesus* and *Meriones tristrami* to tachyzoites of *Neospora caninum*. *J. Parasitol.* 88, 314–319.
- Qin, Q., Wei, F., Li, M., Dubovi, E.J., Loeffler, I.K., 2007. Serosurvey of infectious disease agents of carnivores in captive red pandas (*Ailurus fulgens*) in China. *J. Zoo Wildl. Med.* 38, 42–50.
- Ramamoorthy, S., Sriranganathan, N., Lindsay, D.S., 2005. Gerbil model of acute neosporosis. *Vet. Parasitol.* 127, 111–114.
- Regidor-Cerrillo, J., Pedraza-Diaz, S., Gomez-Bautista, M., Ortega-Mora, L.M., 2006. Multilocus microsatellite analysis reveals extensive genetic diversity in *Neospora caninum*. *J. Parasitol.* 92, 517–524.
- Regidor-Cerrillo, J., Diez-Fuertes, F., Garcia-Culebras, A., Moore, D.P., Gonzalez-Warleta, M., Cuevas, C., et al., 2013. Genetic diversity and geographic population structure of bovine *Neospora caninum* determined by microsatellite genotyping analysis. *PLoS ONE* 8, e72678.
- Regidor-Cerrillo, J., Arranz-Solis, D., Benavides, J., Gomez-Bautista, M., Castro-Hermida, J.A., Mezo, M., et al., 2014. *Neospora caninum* infection during early pregnancy in cattle: how the isolate influences infection dynamics, clinical outcome and peripheral and local immune responses. *Vet. Res.* 45, 10.
- Reichel, M.P., Ellis, J.T., Dubey, J.P., 2007. Neosporosis and hammondiosis in dogs. *J. Small Anim. Pract.* 48, 308–312.
- Reichel, M.P., Alejandra Ayanegui-Alcerrec, M., Gondim, L.F., Ellis, J.T., 2013. What is the global economic impact of *Neospora caninum* in cattle – the billion dollar question. *Int. J. Parasitol.* 43, 133–142.
- Reid, A.J., Vermont, S.J., Cotton, J.A., Harris, D., Hill-Cawthorne, G.A., Konen-Waisman, S., et al., 2012. Comparative genomics of the apicomplexan parasites *Toxoplasma gondii* and *Neospora caninum*: coccidia differing in host range and transmission strategy. *PLoS Pathog.* 8, e1002567.
- Risco-Castillo, V., Wheeler, J.C., Rosadio, R., Garcia-Pena, F.J., Arnaiz-Seco, I., Hoces, D., et al., 2014. Health impact evaluation of alternative management systems in vicuna (*Vicugna vicugna mensalis*) populations in Peru. *Trop. Anim. Health Prod.* 46, 641–646.
- Rojo-Montejo, S., Collantes-Fernandez, E., Regidor-Cerrillo, J., Alvarez-Garcia, G., Marugan-Hernandez, V., Pedraza-Diaz, S., et al., 2009. Isolation and characterization of a bovine isolate of *Neospora caninum* with low virulence. *Vet. Parasitol.* 159, 7–16.
- Ruehlmann, D., Podell, M., Oglesbee, M., Dubey, J.P., 1995. Canine neosporosis: a case report and literature review. *J. Am. Anim. Hosp. Assoc.* 31, 174–183.
- Sangster, C., Bryant, B., Campbell-Ward, M., King, J.S., Slapeta, J., 2010. Neosporosis in an aborted southern white rhinoceros (*Ceratotherium simum simum*) fetus. *J. Zoo Wildl. Med.* 41, 725–728.
- Santolaria, P., Almeria, S., Martinez-Bello, D., Nogareda, C., Mezo, M., Gonzalez-Warleta, M., et al., 2011. Different humoral mechanisms against *Neospora caninum* infection in purebred and crossbred beef/dairy cattle pregnancies. *Vet. Parasitol.* 178, 70–76.
- Sawada, M., Park, C.H., Kondo, H., Morita, T., Shimada, A., Yamane, I., et al., 1998. Serological survey of antibody to *Neospora caninum* in Japanese dogs. *J. Vet. Med. Sci.* 60, 853–854.
- Schares, G., Wenzel, U., Muller, T., Conraths, F.J., 2001. Serological evidence for naturally occurring transmission of *Neospora caninum* among foxes (*Vulpes vulpes*). *Int. J. Parasitol.* 31, 418–423.
- Schlafer, D.H., Miller, R.B., 2007. The female genital system. In: Jubb, Kennedy, and Palmer's Pathology of Domestic Animals, fifth ed. Elsevier Saunders, Philadelphia, PA, pp. 514–516.
- Schwab, A.E., Geary, T.G., Baillargeon, P., Schwab, A.J., Fecteau, G., 2009. Association of BoLA DRB3 and DQA1 alleles with susceptibility to *Neospora caninum* and reproductive outcome in Quebec Holstein cattle. *Vet. Parasitol.* 165, 136–140.
- Sedlak, K., Bartova, E., 2006. Seroprevalences of antibodies to *Neospora caninum* and *Toxoplasma gondii* in zoo animals. *Vet. Parasitol.* 136, 223–231.
- Silva, D.A., Vitaliano, S.N., Mineo, T.W., Ferreira, R.A., Bevilacqua, E., Mineo, J.R., 2005. Evaluation of homologous, heterologous, and affinity conjugates for the serodiagnosis of *Toxoplasma gondii* and *Neospora caninum* in maned wolves (*Chrysocyon brachyurus*). *J. Parasitol.* 91, 1212–1216.
- Simpson, V.R., Monies, R.J., Riley, P., Cromey, D.S., 1997. Foxes and neosporosis. *Vet. Rec.* 141, 503.
- Slapeta, J., Linares, M.C., 2013. Combined amplicon pyrosequencing assays reveal presence of the apicomplexan “type-N” (cf. *Gemmocystis cylindrus*) and *Chromera velia* on the Great Barrier Reef, Australia. *PLoS ONE* 8, e76095.
- Slapeta, J.R., Modry, D., Kyselova, I., Horejs, R., Lukes, J., Koudela, B., 2002. Dog shedding oocysts of *Neospora caninum*: PCR diagnosis and molecular phylogenetic approach. *Vet. Parasitol.* 109, 157–167.
- Slapeta, J.R., Modry, D., Votypka, J., Jirku, M., Lukes, J., Koudela, B., 2003. Evolutionary relationships among cyst-forming coccidia *Sarcocystis* spp. (Alveolata: Apicomplexa: Coccidia) in endemic African tree vipers and perspective for evolution of heteroxenous life cycle. *Mol. Phylogenet. Evol.* 27, 464–475.
- Sobrinho, R., Dubey, J.P., Pabon, M., Linares, N., Kwok, O.C., Millan, J., et al., 2008. *Neospora caninum* antibodies in wild carnivores from Spain. *Vet. Parasitol.* 155, 190–197.
- Soldati, S., Kiupel, M., Wise, A., Maes, R., Botteron, C., Robert, N., 2004. Meningoencephalomyelitis caused by *Neospora caninum* in a juvenile fallow deer (*Dama dama*). *J. Vet. Med. A Physiol. Pathol. Clin. Med.* 51, 280–283.
- Sommanustweechai, A., Vongpakorn, M., Kasantikul, T., Taewnean, J., Siriaronrat, B., Bush, M., et al., 2010. Systemic neosporosis in a white rhinoceros. *J. Zoo Wildl. Med.* 41, 165–168.
- Speer, C.A., Dubey, J.P., McAllister, M.M., Blixt, J.A., 1999. Comparative ultrastructure of tachyzoites, bradyzoites, and tissue cysts of *Neospora caninum* and *Toxoplasma gondii*. *Int. J. Parasitol.* 29, 1509–1519.
- Steinman, A., Shpigel, N.Y., Mazar, S., King, R., Baneth, G., Savitsky, I., et al., 2006. Low seroprevalence of antibodies to *Neospora caninum* in wild canids in Israel. *Vet. Parasitol.* 137, 155–158.
- Stieve, E., Beckmen, K., Kania, S.A., Widner, A., Patton, S., 2010. *Neospora caninum* and *Toxoplasma gondii* antibody prevalence in Alaska wildlife. *J. Wildl. Dis.* 46, 348–355.
- Stuart, P., Zintl, A., De Waal, T., Mulcahy, G., Hawkins, C., Lawton, C., 2013. Investigating the role of wild carnivores in the epidemiology of bovine neosporosis. *Parasitology* 140, 296–302.
- Sun, Y., Wolcott, R., Dowd, S., 2011. Tag-encoded FLX amplicon pyrosequencing for the elucidation of microbial and functional gene diversity in any environment. In: Kwon, Y.M., Ricke, S.C. (Eds.), High-Throughput Next Generation Sequencing. Humana Press, New York, USA, pp. 129–141.
- Thomasson, D., Wright, E.A., Hughes, J.M., Dodd, N.S., Cox, A.P., Boyce, K., et al., 2011. Prevalence and co-infection of *Toxoplasma gondii* and *Neospora caninum* in *Apodemus sylvaticus* in an area relatively free of cats. *Parasitology* 138, 1117–1123.
- Thurmond, M.C., Hietala, S.K., 1997. Effect of congenitally acquired *Neospora caninum* infection on risk of abortion and subsequent abortions in dairy cattle. *Am. J. Vet. Res.* 58, 1381–1385.
- Tiemann, J.C., Souza, S.L., Rodrigues, A.A., Duarte, J.M., Gennari, S.M., 2005a. Environmental effect on the occurrence of anti-*Neospora caninum* antibodies in pampas-deer (*Ozotoceros bezoarticus*). *Vet. Parasitol.* 134, 73–76.
- Tiemann, J.C., Rodrigues, A.A., de Souza, S.L., Duarte, J.M., Gennari, S.M., 2005b. Occurrence of anti-*Neospora caninum* antibodies in Brazilian cervids kept in captivity. *Vet. Parasitol.* 129, 341–343.
- Tranas, J., Heinzen, R.A., Weiss, L.M., McAllister, M.M., 1999. Serological evidence of human infection with the protozoan *Neospora caninum*. *Clin. Diagn. Lab. Immunol.* 6, 765–767.
- Trees, A.J., Williams, D.J., 2005. Endogenous and exogenous transplacental infection in *Neospora caninum* and *Toxoplasma gondii*. *Trends Parasitol.* 21, 558–561.
- Truppel, J.H., Montiani-Ferreira, F., Lange, R.R., Vilani, R.G., Reifur, L., Boerger, W., et al., 2010. Detection of *Neospora caninum* DNA in capybaras and phylogenetic analysis. *Parasitol. Int.* 59, 376–379.
- Uchida, Y., Ike, K., Kurotaki, T., Takeshi, M., Imai, S., 2003. Susceptibility of Djungarian hamsters (*Phodopus sungorus*) to *Neospora caninum* infection. *J. Vet. Med. Sci.* 65, 401–403.
- van der Hage, M.H., Kik, M.J.L., Dorrestein, G.M., 2002. *Neospora caninum*: myocarditis in a European pine marten. In: 4th Scientific Meeting of the European Association of Zoo and Wildlife Veterinarians and European Wildlife Disease Association, European Wildlife Disease Association, Heidelberg, Germany, pp. 217–220.
- Valadas, S., Gennari, S.M., Yai, L.E., Rosypal, A.C., Lindsay, D.S., 2010. Prevalence of antibodies to *Trypanosoma cruzi*, *Leishmania infantum*, *Encephalitozoon cuniculi*, *Sarcocystis neurona*, and *Neospora caninum* in capybara, *Hydrochoerus hydrochaeris*, from Sao Paulo State, Brazil. *J. Parasitol.* 96, 521–524.
- Vianna, M.C., Sreekumar, C., Miska, K.B., Hill, D.E., Dubey, J.P., 2005. Isolation of *Neospora caninum* from naturally infected white-tailed deer (*Odocoileus virginianus*). *Vet. Parasitol.* 129, 253–257.
- Vitaliano, S.N., Silva, D.A., Mineo, T.W., Ferreira, R.A., Bevilacqua, E., Mineo, J.R., 2004. Seroprevalence of *Toxoplasma gondii* and *Neospora caninum* in captive maned wolves (*Chrysocyon brachyurus*) from southeastern and midwestern regions of Brazil. *Vet. Parasitol.* 122, 253–260.

- Wanha, K., Edelhofer, R., Gabler-Eduardo, C., Prosl, H., 2005. Prevalence of antibodies against *Neospora caninum* and *Toxoplasma gondii* in dogs and foxes in Austria. *Vet. Parasitol.* 128, 189–193.
- Wapenaar, W., Jenkins, M.C., O'Handley, R.M., Barkema, H.W., 2006. *Neospora caninum*-like oocysts observed in feces of free-ranging red foxes (*Vulpes vulpes*) and coyotes (*Canis latrans*). *J. Parasitol.* 92, 1270–1274.
- Wapenaar, W., Barkema, H.W., Schares, G., Rouvinen-Watt, K., Zeijlemaker, L., Poorter, B., et al., 2007. Evaluation of four serological techniques to determine the seroprevalence of *Neospora caninum* in foxes (*Vulpes vulpes*) and coyotes (*Canis latrans*) on Prince Edward Island, Canada. *Vet. Parasitol.* 145, 51–58.
- Weiss, L.M., Ma, Y.F., Halonen, S., McAllister, M.M., Zhang, Y.W., 1999. The in vitro development of *Neospora caninum* bradyzoites. *Int. J. Parasitol.* 29, 1713–1723.
- Wiengcharoen, J., Nokkaew, W., Prasithpon, S., Prasomtong, P., Sukthana, Y., 2012. *Neospora caninum* and *Toxoplasma gondii* antibodies in captive elephants (*Elephas maximus indicus*) in Kanchanaburi Province. *Thai. J. Vet. Med.* 42, 235–240.
- Williams, D.J., Hartley, C.S., Bjorkman, C., Trees, A.J., 2009. Endogenous and exogenous transplacental transmission of *Neospora caninum* – how the route of transmission impacts on epidemiology and control of disease. *Parasitology* 136, 1895–1900.
- Williams, J.H., Espie, I., van Wilpe, E., Matthee, A., 2002. Neosporosis in a white rhinoceros (*Ceratotherium simum*) calf. *J. S. Afr. Vet. Assoc.* 73, 38–43.
- Wolfe, A., Hogan, S., Maguire, D., Fitzpatrick, C., Vaughan, L., Wall, D., et al., 2001. Red foxes (*Vulpes vulpes*) in Ireland as hosts for parasites of potential zoonotic and veterinary significance. *Vet. Rec.* 149, 759–763.
- Woodroffe, R., Prager, K.C., Munson, L., Conrad, P.A., Dubovi, E.J., Mazet, J.A., 2012. Contact with domestic dogs increases pathogen exposure in endangered African wild dogs (*Lycaon pictus*). *PLoS ONE* 7, e30099.
- Woods, L.W., Anderson, M.L., Swift, P.K., Sverlow, K.W., 1994. Systemic neosporosis in a California black-tailed deer (*Odocoileus hemionus columbianus*). *J. Vet. Diagn. Invest.* 6, 508–510.
- Wouda, W., Dijkstra, T., Kramer, A.M., van Maanen, C., Brinkhof, J.M., 1999. Seroepidemiological evidence for a relationship between *Neospora caninum* infections in dogs and cattle. *Int. J. Parasitol.* 29, 1677–1682.
- Yai, L.E., Canon-Franco, W.A., Gerald, V.C., Summa, M.E., Camargo, M.C., Dubey, J.P., et al., 2003. Seroprevalence of *Neospora caninum* and *Toxoplasma gondii* antibodies in the South American opossum (*Didelphis marsupialis*) from the city of Sao Paulo, Brazil. *J. Parasitol.* 89, 870–871.
- Yai, L.E., Ragozo, A.M., Canon-Franco, W.A., Dubey, J.P., Gennari, S.M., 2008. Occurrence of *Neospora caninum* antibodies in capybaras (*Hydrochaeris hydrochaeris*) from Sao Paulo State, Brazil. *J. Parasitol.* 94, 766.
- Yu, X., Chen, N., Hu, D., Zhang, W., Li, X., Wang, B., et al., 2009. Detection of *Neospora caninum* from farm-bred young blue foxes (*Alopex lagopus*) in China. *J. Vet. Med. Sci.* 71, 113–115.
- Zanet, S., Palese, V., Trisciuglio, A., Canton Alonso, C., Ferroglio, E., 2013. *Encephalitozoon cuniculi*, *Toxoplasma gondii* and *Neospora caninum* infection in invasive Eastern cottontail rabbits *Sylvilagus floridanus* in Northwestern Italy. *Vet. Parasitol.* 197, 682–684.