

Synthesis, Antifungal Ergosterol Inhibition, Antibiofilm Activities, and Molecular Docking on β -Tubulin and Sterol 14-Alpha Demethylase along with DFT-Based Quantum Mechanical Calculation of Pyrazole Containing Fused Pyridine–Pyrimidine Derivatives

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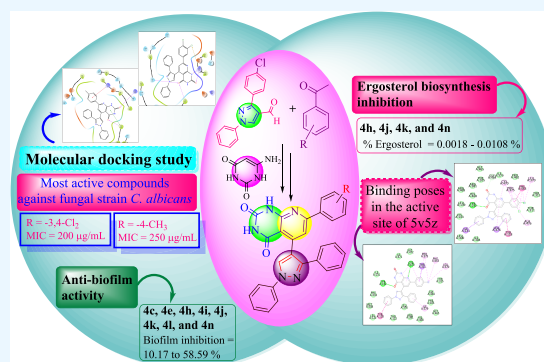
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ABSTRACT: Multidrug-resistant fungal infections have become much more common in recent years, especially in immune-compromised patients. Therefore, researchers and pharmaceutical professionals have focused on the development of novel antifungal agents that can tackle the problem of resistance. In continuation to this, a novel series of pyrazole-bearing pyrido[2,3-*d*]pyrimidine-2,4(1*H*,3*H*)-dione derivatives (**4a–4o**) have been developed. These compounds have been screened against *Candida albicans*, *Aspergillus niger*, and *Aspergillus clavatus*. The synthesized compounds were characterized by well-known spectroscopic techniques, i.e., IR, ^1H NMR, ^{13}C NMR, and mass spectrometry. *In vitro* antifungal results revealed that compound **4n** showed activity against *C. albicans* having MIC value of 200 $\mu\text{g}/\text{mL}$. To know the plausible mode of action, the active derivatives were screened for anti-biofilm and ergosterol biosynthesis inhibition activities. The compounds **4h**, **4j**, **4k**, and **4n** showed greater ergosterol biosynthesis inhibition than the control DMSO. To comprehend how molecules interact with the receptor, studies of molecular docking of **4k** and **4n** have been performed on the homology-modeled protein of β -tubulin. The molecular docking revealed that the active compounds **4h**, **4j**, **4k**, **4l**, and **4n** interacting with the active site amino acid of sterol 14-alpha demethylase (PDB ID: 5v5z) indicate one of the possible modes of action of ergosterol inhibition activity. The synthesized compounds **4c**, **4e**, **4h**, **4i**, **4j**, **4k**, **4l**, and **4n** inhibited biofilm formation and possessed the potential for anti-biofilm activity. DFT-based quantum mechanical calculations were carried out to optimize, predict, and compare the vibration modes of the molecule **4a**.



1. INTRODUCTION

Invasive fungal infections are becoming more common as a result of stem cell transplantation, organ transplantation, chemotherapy, and the human immunodeficiency virus.^{1,2} The most common pathogenic strains that cause systemic fungal infections are *Candida albicans*, *Cryptococcus neoformans*, and *Aspergillus fumigatus*.³ Fluconazole, voriconazole, itraconazole, and miconazole are azole-based broad-spectrum antifungal drugs that are widely used (Figure 1).⁴ They have broad-spectrum antifungal activity against the majority of filamentous fungi, but several of them are ineffective against invasive aspergillosis and have substantial drug resistance.⁵ Furthermore, widespread usage of existing antifungal medications has resulted in significant drug resistance.⁶ As a result, novel antifungal agents with outstanding action against a wide range of clinical fungal species are urgently needed.

Heterocycles are suggestively a wealthy target for the development of bioactive agents with a wide range of biological activities acting as oxygen carriers (hemoglobin), constituents of DNA, energy storage (adenosine triphosphate), and natural antimicrobial agents (diketopiperazines).⁷ Thus, these compounds with various functional moieties have been explored for their antimicrobial characteristics.^{8,9} Compounds with nitrogen-rich five- and six-membered heterocyclic structures, such as

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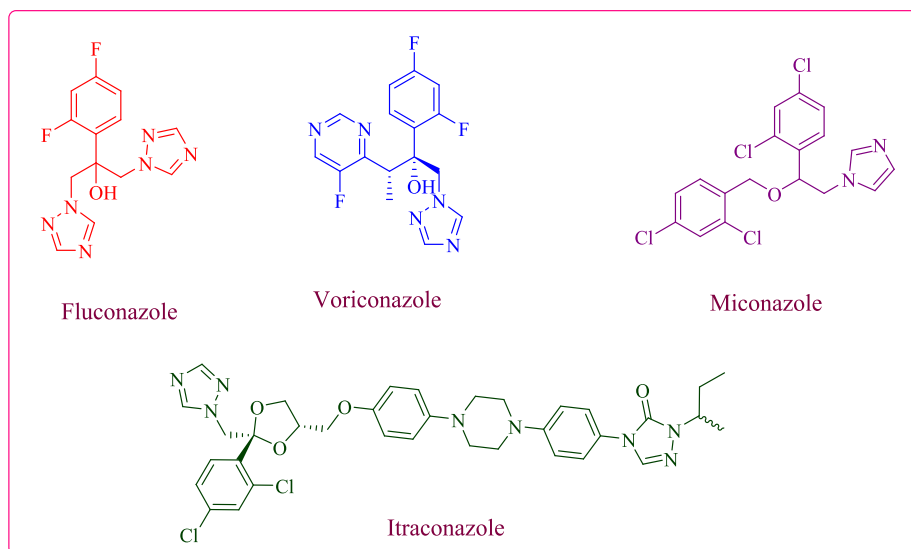


Figure 1. Commercially available azole-based antifungal drugs.

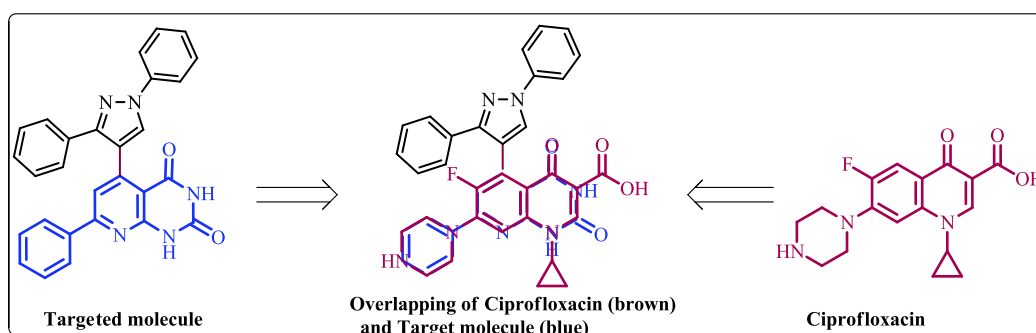


Figure 2. Design of target molecule based on the commercially available drug ciprofloxacin.

pyrazoles, pyridine, and pyrimidine, are a vital source of biologically active components for a variety of applications.¹⁰

Pyrazole contains two nitrogen atoms in its five-membered ring and is chemically 1,2-diazoles. Pyrazole-containing compounds have been effectively synthesized as they possess various biological activities like antimicrobial,¹¹ anti-inflammatory,¹² anticancer,¹³ and antitubercular.¹⁴ Moreover, pyridine is a six-membered heterocyclic entity containing nitrogen atoms while pyrimidine has two nitrogen atoms at its adjacent carbon. Pyridine and pyrimidine derivatives have a wide range of biological activities, i.e., antimicrobial,^{15,16} anti-inflammatory,¹⁷ antitumor,¹⁸ anticancer,¹⁹ antitubercular,^{20,21} etc.

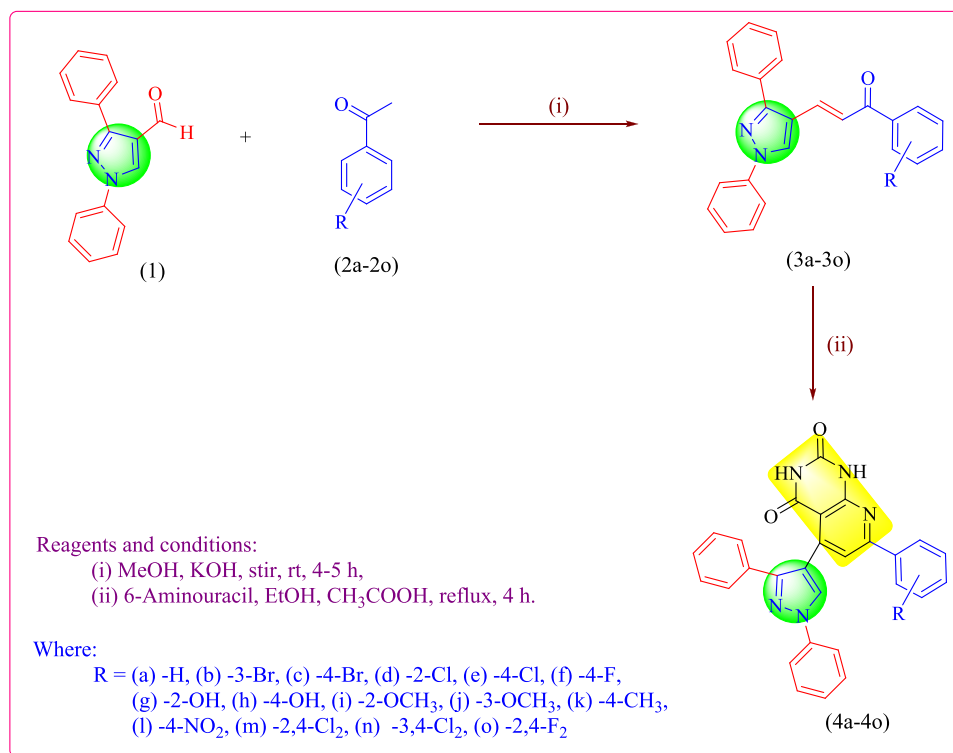
We have used heterocycles to invent our novel hybrid heterocyclic compounds focusing on different fused pyridine–pyrimidines with pyrazole motifs, as noted in the present study. The synthesized compounds were uniformly auspicious to the medicinal chemist for correlation of the similarities with commercially available medicine consisting of pyridine scaffolds. The pyridine and pyrimidine scaffolds clubbed with the pyrazole showed potential antimicrobial activities. The pyrazole ring-containing compounds showed antifungal activity via ergosterol biosynthesis inhibition²² and biofilm inhibition. The structural modification of the ciprofloxacin to 7-phenylpyrido[2,3-*d*]-pyrimidine-2,4(1*H*,3*H*)-dione core highlighted in Figure 2 indicates that the pyrido[2,3-*d*]pyrimidine-2,4(1*H*,3*H*)-dione pharmacophore matches with the quinolin-4(1*H*)-one core of the ciprofloxacin and the phenyl ring at 7-position of the target

scaffold matches with the piperazine ring. Also, the 5-position of pyrido[2,3-*d*]pyrimidine-2,4(1*H*,3*H*)-dione was modified by substituting the pyrazole for enhancing the activity. These things enlightened us the synthesis of 5-(1,3-diphenyl-1*H*-pyrazol-4-yl)-7-phenylpyrido[2,3-*d*]pyrimidine-2,4(1*H*,3*H*)-dione as an antimicrobial agent. The possible mode of action of antifungal activity was also evaluated by ergosterol biosynthesis inhibition and biofilm inhibition activities.

2. RESULTS AND DISCUSSION

2.1. Chemistry. The titled molecules (4a–4o) were synthesized by multi-step reactions. Chalcone derivatives (3a–3o) were prepared by using an aldol condensation of 1,3-diphenyl-1*H*-pyrazole-4-carbaldehyde (1) and substituted acetophenones (2a–2o). Final compounds (4a–4o) were prepared from chalcone derivatives, which was reacted with 6-aminouracil using acetic acid as a catalyst. The synthetic root adopted for the synthesis of compounds (4a–4o) is illustrated in Scheme 1. IR spectra of compound 4a, at frequencies of 3178 and 2811 cm⁻¹, confirmed the presence of –CH– and –CH=CH– groups, respectively. The presence of >C=O was confirmed at 1755 cm⁻¹. Stretching vibrations at 1689, 1659, and 1276 cm⁻¹ confirmed the presence of –C=N, –C=C–, and –C–N groups, respectively. The proton of secondary amine in the pyrimidine ring was confirmed at $\delta = 12.09$ –10.39 ppm while the proton of pyrazole was confirmed at 8.55 ppm, and the signal observed at 8.36 ppm confirmed the proton of the

Scheme 1. Synthetic Pathways of Synthesized Compounds 4a–4o



pyridine ring. Moreover, the signal appeared in the range of $\delta = 8.04\text{--}7.45$ ppm confirming 15 aromatic protons. In ¹³C NMR, chemical shifts at 159, 153.5, and 151.3 ppm agreed with the carbon of the pyrimidine ring. Furthermore, carbons of the pyridine ring were confirmed at 150.5, 149.0, 112.4, and 110.3 ppm. Signals that appeared at 145.8, 131.6, and 101.3 ppm confirmed the presence of a heterocyclic ring of pyrazole. Molecular weight at $m/z = 457.17$ of mass spectra is evidence of compound 4a.

2.2. Biological Activity. **2.2.1. Discussion on Antifungal Activity.** The main scaffold of newly synthesized molecules (4a–4o) consists of the pyrido[2,3-*d*]pyrimidine-2,4(1*H*,3*H*)-dione moiety. There are many reports wherein pyrido/pyrimido-pyrimidines were tested for antifungal activity. Abdelhameed et al. have reported arylidene hydrazinylpyrido[2,3-*d*]pyrimidin-4-ones as potent anti-microbial agents targeting *C. albicans*.²³ Pyrimido[4,5-*d*][1,2,4]triazolo[4,3-*a*]pyrimidines were studied for their antifungal activity by Mangoud et al. against *C. albicans*.²⁴ Sharma et al. have synthesized a pyrimido[4,5-*d*]pyrimidine-2,5-dione derivative that showed 8–100 ppm MIC values against *C. albicans*.²⁵ Taking a cue from these literature reports, as well as our previous work²⁶ wherein we have studied fused pyridine–pyrimidine hybrids for their antifungal activity, we have evaluated novel synthesized 4a–4o for *in vitro* antifungal activity using the Broth dilution method. The results of antifungal activity revealed that some of the compounds have inhibition effects toward screened fungal strains. Antifungal activity data revealed that compound 4n showed to be most potent against *C. albicans* having an MIC value of 200 $\mu\text{g}/\text{mL}$. Moreover, compound 4k showed activity against *C. albicans* having an MIC value of 250 $\mu\text{g}/\text{mL}$. Compounds 4n and 4k exhibited significant inhibition than the standard drug griseofulvin. Additionally, compounds 4c, 4e, 4h, 4i, 4j, and 4l showed comparable potency with reference drug griseofulvin

against *C. albicans* with MIC 500 $\mu\text{g}/\text{mL}$. The *in vitro* antifungal activity data of synthesized compounds 4a–4o are listed in Table 1.

2.3. Computational Study. **2.3.1. Molecular Docking Study.** The molecular interaction between the receptor and the ligand is vital information obtained from docking studies. Griseofulvin, which is used as the standard for antifungal activity, is considered to be a tubulin polymerization inhibitor.^{27,28} β -Tubulin was used as a receptor for molecular docking studies. Since the experimental three-dimensional structure of *Candida albicans* tubulin is not available in the Protein Data Bank, the structure was modeled by homology modeling using Yeast tubulin (PDB ID: 5w3f) as a template protein. The sequence coverage was around 96% with 87% identities and 96% positives. The modeled protein structure was verified by verifying 3D, Errat, and Procheck using protein structure validation server SAVES v6.0 of UCLA-DOE Lab and ProSA-web (Z-score: -9.84).

The modeled protein was prepared and used for molecular docking studies. Rathinasamy et al.²⁸ have reported that griseofulvin binds at a site distinct from the colchicine-binding site in tubulin; hence, docking studies were carried out at a site comprising amino acid residues Glu 27, Leu 215, Gln 224, Asn 227, Leu 228, Ser 231, Tyr 270, Leu 273, Thr 274, Arg 318, Lys 359, Asp 360, and Leu 361. Compound 4g showed the highest XP dock score of -5.933 kcal/mol, and the standard compound griseofulvin showed a dock score of -4.064 kcal/mol. All the compounds showed similar dock scores within a range of -5.993 to -3.982 kcal/mol. The prime MM-GBSA binding energy (MMGBSA $-\Delta G_{\text{bind}}$) values of all the compounds were better than those of griseofulvin (Table 2). The dock scores and ΔG_{bind} values showed a similar trend as the experimental antifungal activity.

Table 1. Antifungal Screening of Compounds 4a–4o Having Different Fungal Strains^a

entry	pyrazole containing fused pyridine-pyrimidine derivatives – structural formula	minimum inhibition concentrations (MIC) in $\mu\text{g/mL} \pm \text{SD}$		
		<i>C. a.</i>	<i>A. n.</i>	<i>A. c.</i>
4a		1000 \pm 3.56*	500 \pm 3.15*	500 \pm 3.51*
4b		1000 \pm 4.14*	1000 \pm 3.50*	1000 \pm 3.54*
4c		500 \pm 4.04*	1000 \pm 3.52*	1000 \pm 3.25*
4d		1000 \pm 4.02*	1000 \pm 4.25*	1000 \pm 4.14 *
4e		500 \pm 3.51 *	1000 \pm 3.50*	>1000 \pm 4.30*
4f		1000 \pm 3.54*	500 \pm 4.57*	1000 \pm 4.12*
4g		1000 \pm 3.16*	1000 \pm 4.12*	1000 \pm 4.15*
4h		500 \pm 3.52*	1000 \pm 3.45*	1000 \pm 3.45*
4i		500 \pm 3.12*	500 \pm 4.17*	500 \pm 4.15*
4j		500 \pm 3.17*	500 \pm 3.50*	500 \pm 3.15*
4k		250 \pm 4.30*	500 \pm 4.12*	1000 \pm 3.50*
4l		500 \pm 3.15*	1000 \pm 4.17*	1000 \pm 3.18*
4m		1000 \pm 3.18*	1000 \pm 3.51*	1000 \pm 3.54*
4n		200 \pm 4.35*	1000 \pm 3.55*	1000 \pm 3.45*
4o		1000 \pm 3.54*	1000 \pm 4.20*	1000 \pm 3.26*
	griseofulvin	500 \pm 2.57*	100 \pm 1.26*	100 \pm 3.15*

^aSD = standard deviation; C. A. = *Candida albicans*; A. n. = *Aspergillus niger*; A. c. = *Aspergillus clavatus*. * $p \leq 0.0001$.

A dock pose analysis of all the compounds was carried out, and the molecules showed a majorly hydrophobic interaction with protein amino acid residues. Griseofulvin showed a hydrogen bond interaction with Ser 231 and Tyr 270 (Figure 3). Molecules 4k and 4n showed better antifungal activity having similar binding energy values, but they had an additional hydrogen bond interaction with Arg 318 and Lys 359. Molecule 4k showed an additional hydrophobic π - π interaction with Tyr

270 and a π -cation interaction with Lys 359, which corresponds to the higher binding value and better antifungal activity when compared to griseofulvin (Figure 4). Molecule 4n also showed a similar hydrogen bond interaction to molecule 4k along with an additional halogen interaction with Lys 359 (Figure 5).

2.3.1.1. Evaluation of ADME-T Properties. ADME properties and toxicity were calculated for synthesized molecules to evaluate their drug-like properties. A heat map table of the

Table 2. Glide XP Dock Scores and Prime MMGBSA- ΔG_{bind} Scores

entry	XP GScore (kcal/mol)	MMGBSA ΔG_{bind} (kcal/mol)
4a	-4.340	-45.07
4b	-4.838	-62.30
4c	-4.128	-38.55
4d	-4.699	-42.64
4e	-4.169	-42.75
4f	-3.997	-41.98
4g	-5.933	-68.57
4h	-4.628	-42.99
4i	-4.151	-43.61
4j	-5.052	-48.61
4k	-4.427	-63.31
4l	-4.295	-45.27
4m	-3.982	-47.09
4n	-4.079	-44.75
4o	-4.574	-45.36
griseofulvin	-4.064	-41.41

ADME properties and toxicity parameters is provided in Table 3. Physicochemical properties like partition coefficient (log-

$P_{\text{octanol/water}}$), cell membrane permeability (P_{Caco} and P_{MDCK}), and oral absorption of all molecules are in the acceptable range. Only molecule 4l showed lower MDCK cell permeability and oral absorption value than the acceptable range. However, all the molecules showed lesser water solubility (LogS) compared to standard griseofulvin. Furthermore, the toxicity profiling using the pkCSM server revealed that all the molecules were showing a nontoxic nature.

2.3.2. Vibrational Analysis. The structure of molecule 4a was optimized with C_1 symmetry; the predicted IR spectra for molecule 4a at the B3LYP/6-31G* level in the gas phase is provided in the supplementary figures, and the comparison with experimental IR data is provided in Table 4 along with the assignment of wavenumbers to corresponding stretching modes. The calculations predicted the CH stretching modes corresponding to the phenyl rings in the expected region of $\sim 3200 \text{ cm}^{-1}$; $>C=O$ stretching was predicted at $\sim 1700 \text{ cm}^{-1}$, and $>C=N$ stretching was predicted at $\sim 1400 \text{ cm}^{-1}$.

2.3.3. Ergosterol Inhibition Activity. The antifungal mechanism of action of azole drugs is to disrupt the sterol biosynthetic pathway that leads to ergosterol biosynthesis inhibition.²⁹ Ergosterol is one of the components of the fungal plasma membrane. Inhibition of synthesis of this major sterol is one of

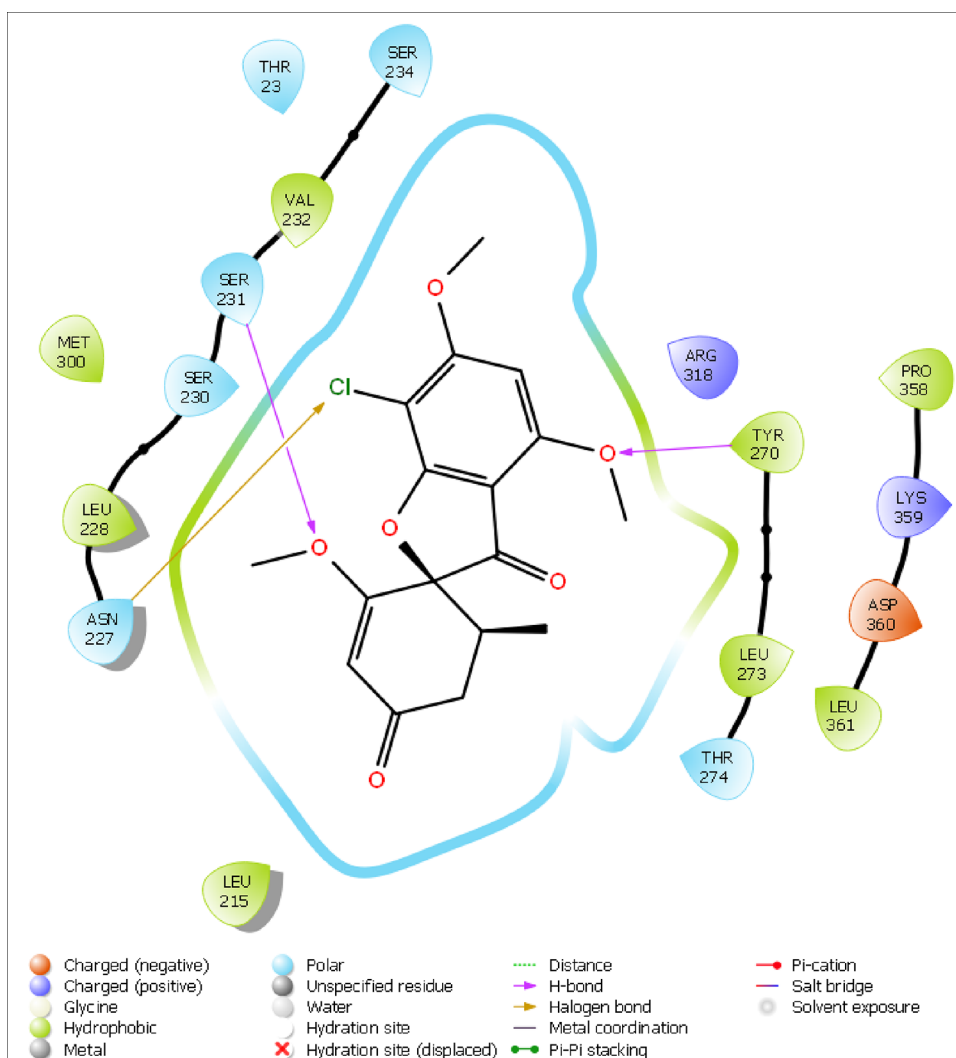


Figure 3. Ligand interaction diagram griseofulvin docked into tubulin showing a hydrogen bond interaction with residues Ser 231 and Tyr 270, and a halogen bond with Asn 227.

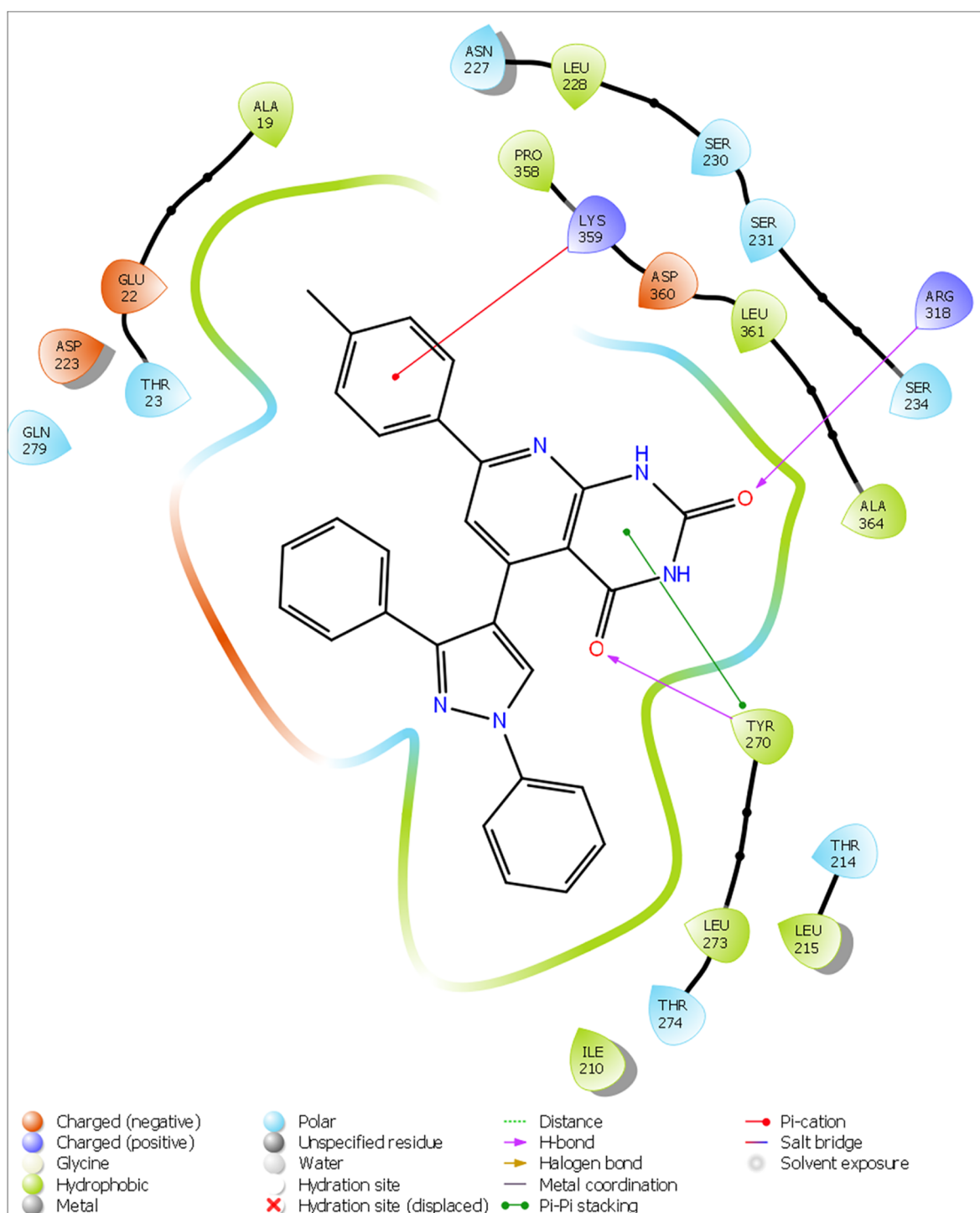


Figure 4. Ligand interaction diagram **4k** docked into tubulin showing a hydrogen bond interaction with residues Tyr 270 and Arg 318, a hydrophobic π - π interaction with Tyr 270, and a π -cation interaction with Lys 359.

the common modes of action for many antifungal drugs. The quantitative estimation of ergosterol biosynthesis was done using the protocol of Breivik et al. in 1957.^{30,31} Ergosterol quantitation was represented in terms of % ergosterol/g wet weight (Figure 6). From the ergosterol biosynthesis inhibition activity, it is clear from the graph that the % of ergosterol was decreased in compounds **4h**, **4j**, **4k**, and **4n** as compared to DMSO control in all conditions except compound **4l** (Figure 6). This clearly indicates that the plausible mode of action of these

compounds is ergosterol biosynthesis inhibition and Hasan antifungal potential.

The ergosterol inhibition activity represented by compounds **4h**, **4j**, **4k**, and **4n** was further confirmed by molecular docking studies. Table 5 shows molecular docking data represented in terms of binding energy (ΔG) in kcal/mol for sterol 14- α demethylase (PDB ID: 5v5z), the key enzyme from the ergosterol biosynthetic pathway from *C. albicans* (5v5z).³² The results indicate that all compounds bind to this enzyme with less binding energy indicating stable interactions,

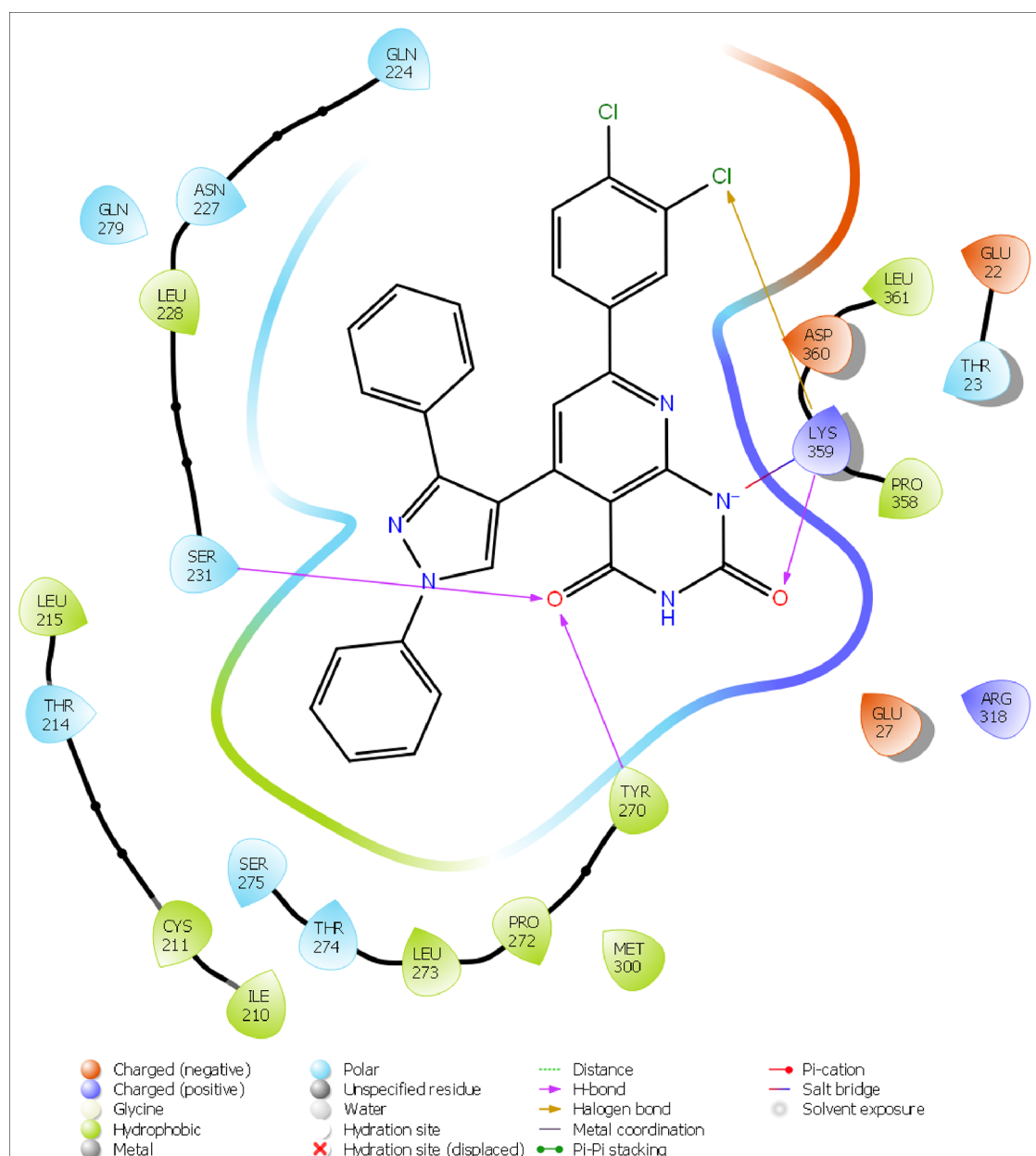


Figure 5. Ligand interaction diagram **4n** docked into tubulin showing a hydrogen bond interaction with residues Ser 231 Tyr 270 and Lys 359 and a halogen bond with Lys 359.

as shown in Table 5. Compound **4l** shows the lowest binding energy of -12.8 kcal/mol. It is observed that all five compounds bind in the active site of the enzyme, which is represented in Figure 7. Figure 7A shows the active site of SvsZ, and Figure 7B represents the binding poses of ligands **4h**, **4j**, **4k**, **4l**, and **4n**. The docking analysis of these poses was performed to check the interaction of these compounds with amino acids present in the active site (Figure 8). It was observed that all compounds were interacting with active site amino acids, suggesting its strong and stable interaction with the enzyme. These newly synthesized derivatives were subjected to the ADME study³³ for pharmacological analysis, and the data is represented in Table 5. Lipinski's rule gives us a good approach to predicting drug-likeness. All five compounds are following Lipinski's rule and therefore have the potential to be developed as lead candidates.

2.3.4. Anti-Biofilm Activity. Bacterial biofilms are highly organized bacterial complexes enclosed in a self-produced

matrix (extracellular polymeric substance, EPS) protected from the host defense. EPS plays an important role in bacterial cell aggregation, cell adhesion, and biofilm formation and protects bacterial cells from a hostile environment. Biofilms are known to have high adaptive resistance to antibiotics and other disinfectants, which makes it very difficult to treat infections.³⁴ The bacterial isolates used in the present study are of marine origin and known for biofilm formation.³⁵ The results of the biofilm inhibition of different compounds are presented in Figure 9. The inhibitory actions for biofilm formation activities were shown by all compounds at varying levels. In general, compounds highly inhibited the biofilm formation by Gram-negative bacteria (GS-5 and GM-17) compared to Gram-positive (BM-14)). This may be due to the differences in the structure between Gram-positive and Gram-negative cell walls and showed different modes of action of the tested compounds.³⁶ The tested compounds showed 14.38% (**4e**) to

Table 3. ADME-T Properties of Synthesized Molecules 4a–4o^a

molecule	MW	logPo/w	logS	P_{Caco}	P_{MDCK}	#metab	% human oral absorption	AMES toxicity	max. tolerated dose (human)	Minnow toxicity
4a	457.49	5.173	-7.847	422.086	194.739	0	91.265	no	0.458	1.452
4b	536.386	5.704	-8.619	387.788	469.626	0	80.758	no	0.446	0.919
4c	536.386	5.696	-8.551	431.878	530.023	0	81.551	no	0.46	0.875
4d	491.935	5.554	-8.237	396.664	377.95	0	93.012	no	0.457	1.138
4e	491.935	5.61	-8.483	436.948	499.41	0	94.092	no	0.46	1.021
4f	475.481	5.37	-8.035	445.758	373.901	0	92.841	no	0.456	1.765
4g	473.49	4.469	-7.055	213.81	93.366	1	94.818	no	0.438	1.761
4h	473.49	4.354	-7.275	147.736	62.613	1	91.268	no	0.442	2.007
4i	487.517	5.179	-7.59	447.053	207.219	1	91.75	no	0.456	1.472
4j	487.517	5.177	-7.785	411.643	189.536	1	91.096	no	0.457	1.671
4k	471.517	5.437	-8.312	420.486	193.941	1	92.783	no	0.447	1.093
4l	502.488	4.432	-7.946	54.14	21.156	1	70.965	no	0.443	0.504
4m	526.381	5.835	-8.457	430.176	768.652	0	82.331	no	0.464	0.503
4n	526.381	6.056	-9.04	446.201	1057.187	0	83.911	no	0.462	0.67
4o	493.471	5.57	-8.418	415.555	518.208	0	93.472	no	0.453	1.541
griseofulvin	352.771	2.185	-4.759	1553.757	1853.409	3	96.857	no	1.223	-0.274

^aAcceptable range for properties: $350 \leq MW \leq 650$, $-2.0 \leq \log Po/w \leq 6.5$, $-6.5 \leq \log S \leq 0.5$. P_{Caco} and P_{MDCK} cell permeability: <25 poor, >50 good. Percent human oral absorption >80. #metab: 1 to 8. Max. tolerated dose (MRTD) ≥ 0.47 . Minnow toxicity: $\log LC_{50} > -0.3$.

Table 4. Experimental, Calculated IR Frequency and Assignment for Molecule 4a

IR experimental frequency (wavenumber cm^{-1})	IR calculated (B3LYP/6-31G*) (wavenumber cm^{-1})	assignment
519.65	527.86	ν C–H
632.55	642.54	ν C–H
706.39	718.36	ν N–H
802.65	807.04	ν C–H
904.65	947.40	ν C–H
1276.49	1274.85	ν C–H
1432.54	1422.39	ν C=N
1479.98	1496.53	ν C–H
1543.76	1566.16	ν C–N
1609.18	1599.23	ν C=C
1659.98	1636.42	ν C=C
1689.78	1717.16	ν C=O
1755.21	1798.15	ν C=O
3001.34	3213.37	ν C–H aromatic
3178.76	3220.99	ν C–H aromatic

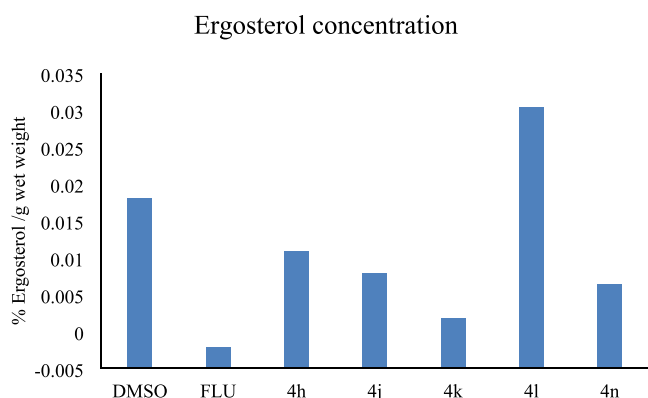


Figure 6. Ergosterol quantitation in terms of % ergosterol/g wet weight.

39.45% (4k) inhibition against *Vibrio* sp. (GS-5), similarly 36.30% (4e) to 58.59% (4k) against *Gallaecimonas* sp. (GM-17). Contrarily, these compounds showed less inhibitory action

Table 5. ADME Analysis of Active Compounds

	4h	4j	4k	4l	4n
molecular weight	473.49	487.52	471.52	502.49	526.38
A log P	4.5	4.81	5.11	4.71	6.1
H-bond acceptor	6	6	5	7	5
H-bond donor	3	2	2	2	2
rotatable bonds	4	5	4	5	4

for biofilm formation in *Staphylococcus* sp. (BM-14), which varied from 1.05% (4l) to 25.02% (4n). The reduced inhibition of biofilm development exhibited that the bacterial cell in a biofilm is more resistant to antimicrobial agents.³⁷ Altogether, our study demonstrated that the compounds inhibit biofilm formation and had the potential for anti-biofilm activity. The disc diffusion method was used to enumerate the antibacterial activities of the synthesized compounds; no zones of inhibition were observed around the discs impregnated with chemical compounds (Figure S1). The results indicated that the inhibition of biofilm formation by bacteria is not due to the toxic effect of the chemical compounds but because chemical compounds inhibited the initial attachment of bacterial cells to the surface as well as reducing cell-to-cell surface interactions.

2.3.5. The Effect of Various-Synthesized Derivatization on Antifungal Activity. The hybrids of pyrazole-bearing pyridine–pyrimidine (4a–4o) were synthesized and evaluated for their antifungal potency. The effect of various functional groups on antifungal activity is shown in Figure 10. Compound 4n having an electron-withdrawing chloro group at the third and fourth positions possessed MIC of 200 $\mu g/mL$, while compound 4k having an electron-donating methyl group at the fourth position possessed MIC of 250 $\mu g/mL$ against *C. albicans*. Compounds 4c, 4e, and 4l having electron-withdrawing bromo, chloro, and nitro groups on the fourth position possessed comparable antifungal activity against *C. albicans*. Compounds with electron-donating functional groups on the second (4i-2-OCH₃), third (4j-3-OCH₃), and fourth (4h-4-OH) positions were also found to have similar MIC values in comparison to griseofulvin.

The effect of various functional groups on ergosterol biosynthesis is shown in Figure 11. Compound 4k having an electron-donating methyl group at the fourth position on the

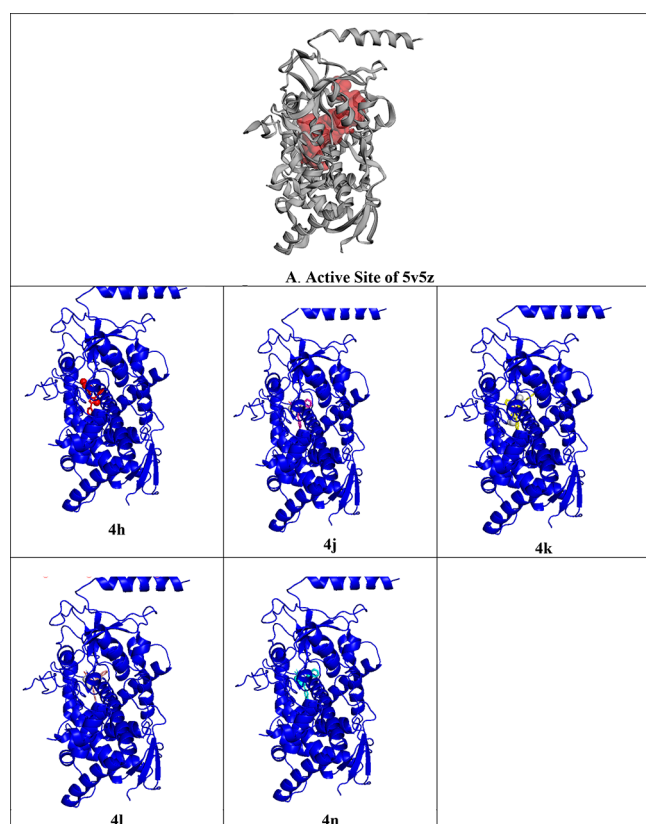


Figure 7. (A) Active site of 5v5z. (B) The two-dimensional interacting mode of the hybrid derivative in the active region of sterol 14- α demethylase from *Candida albicans* (5v5z).

phenyl ring possesses an ergosterol content of 0.0018%. Similarly, compound **4h** having an electron-donating hydroxyl group at the fourth position on the phenyl ring possesses an ergosterol content of 0.0108%. Compound **4n** having an electron-withdrawing chloro group at the third and fourth positions on the phenyl ring possesses an ergosterol content of 0.0064%, and **4j** with an electron-donating methoxy group at the third position on the phenyl ring has ergosterol content 0.0079%. Compounds having different electron-donating and -withdrawing functional groups on the third and fourth positions on the phenyl ring inhibited ergosterol biosynthesis in comparison to DMSO (0.0182%).

The effect of various functional groups on biofilm formation by Gram-positive and Gram-negative bacteria is shown in Figure 12. The potent antifungal compounds having various electron-donating and -withdrawing functional groups on the phenyl ring highly inhibited biofilm formation by Gram-negative bacteria (GS-5 and GM-17). The tested compounds showed 14.38 to 39.45% inhibition against Gram-negative *Vibrio* sp. (GS-5) and 36.30 to 58.59% against Gram-negative *Gallaecimonas* sp. (GM-17). Compounds having various electron-donating functional groups on the second (**4i-2-OCH₃**), third (**4j-3-OCH₃**), and fourth (**4k-4-CH₃**) positions on the phenyl ring inhibited biofilm formation by Gram-positive bacteria (BM-14) with 10.17 to 20.12%. Compound **4k** with the electron-negative chloro group on the third and fourth positions inhibited biofilm formation by Gram-positive bacteria (BM-14) with 25.02%.

3. CONCLUSIONS

The aldol condensation of 1,3-diphenyl-1H-pyrazole-4-carbaldehyde and appropriate acetophenone derivatives in methanolic KOH afforded the chalcone derivatives. The reaction of substituted chalcones with 6-aminouracil in the presence of acetic acid yielded the corresponding pyrazole-containing pyridine–pyrimidine derivatives. The synthesized compounds (**4a–4o**) were screened for their *in vitro* antifungal activity against three fungi. The –3,4-Cl₂ group in compound **4n** (MIC 200 μ g/mL) and the 4-CH₃ group in compound **4k** (MIC 250 μ g/mL) increased the potency for *C. albicans*, making them more active than the standard drug griseofulvin. The antifungal active derivatives were screened for ergosterol biosynthesis inhibition activity, and the compounds **4h**, **4j**, **4k**, and **4n** showed ergosterol biosynthesis inhibition. The molecular docking analysis showed that the active compounds **4h**, **4j**, **4k**, **4l**, and **4n** interacted with the active site amino acid of sterol 14- α demethylase (PDB ID: 5v5z), indicating that one of the possible modes of action is ergosterol biosynthesis inhibition. The potent antifungal agents were highly inhibitory to the biofilm formation by Gram-negative bacteria (GS-5 and GM-17) and Gram-positive bacteria (BM-14) and possessed antibiofilm activity. A stable homologous model of β -tubulin was constructed for molecular docking studies. Molecules with better antifungal activity than standard griseofulvin showed higher binding affinity, which may be attributed to an additional hydrogen bond interaction with Arg 318 and Lys 359.

4. EXPERIMENTAL PROCEDURE

4.1. Materials and Instruments. “Completion of the reaction and purity of compounds was checked on Aluminium coated TLC plates [60 F₂₅₄ (E. Merck)]. n-hexane: ethyl acetate (3:2 V/V) was used as a mobile phase and was visualized in an iodine chamber. An electrothermal melting point apparatus was used to determine melting points and was uncorrected. Elemental analysis (% C, H, N) was confirmed by a Perkin-Elmer 2400 CHN analyzer. A Perkin-Elmer FT-IR spectrophotometer was used to record IR spectra by using KBr. ¹H NMR and ¹³C NMR spectra were recorded on a Bruker Avance III 400 MHz in CDCl₃ as a solvent and tetramethylsilane (TMS) as an internal standard using a 5 mm tube. Mass spectra were obtained on SHIMADZU MS 2010 spectrometer.”³⁸

4.1.1. Synthesis of (3-(1,3-Diphenyl-1H-pyrazol-4-yl)-1-arylprop-2-en-1-ones (3a–3o). (3-(1,3-Diphenyl-1H-pyrazol-4-yl)-1-arylprop-2-en-1-ones (**3a–3o**) were prepared according to the method as per the literature method.³⁹

4.1.2. Synthesis of 5-(1,3-Diphenyl-1H-Pyrazol-4-yl)-7-arylpyrido[2,3-d]pyrimidine-2,4(1H,3H)-diones (4a–4o). A mixture of chalcone derivatives (**3a–3o**) (10 mmol), EtOH (40 mL), and 6-aminouracil (10 mmol) was refluxed in a round-bottom flask fitted with a reflux condenser. The refluxing was continued for 4 h and poured into ice-cold water. The solid precipitate was filtered and washed with hot water and dried overnight in a vacuum oven. The crude product was recrystallized from ethanol.

4.1.3. 5-(1,3-Diphenyl-1H-pyrazol-4-yl)-7-phenylpyrido[2,3-d]pyrimidine-2,4(1H,3H)-dione (4a). Yield: 75%; solid; M.P. 211–213 °C; IR (KBr, cm⁻¹): 3178, 2811 (C–H, –CH=CH–), 1755 (>C=O), 1689, 1659, 1276 (C=N, C=C, C–N); ¹H NMR (400 MHz, CDCl₃): δ = 9.45 (s, 1H, Ar-NH), 9.24 (s, 1H, Ar-NH), 8.27 (s, 1H, –CH-CN), 7.83–7.76 (m, 5H, Ar-H), 7.72–7.57 (m, 5H, Ar-H), 7.30–7.24 (m, 6H, Ar-H); ¹³C

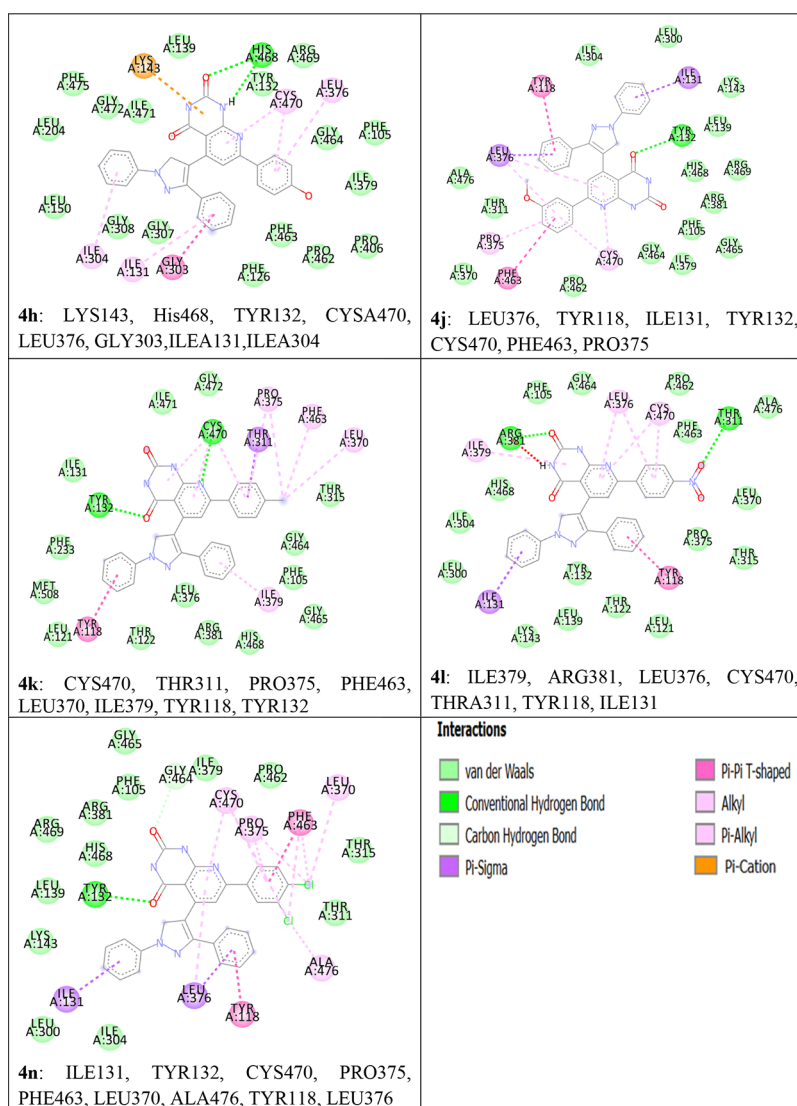


Figure 8. The two-dimensional interacting mode of the hybrid derivative in the active region of sterol 14- α demethylase from *Candida albicans* (5v5z).

NMR (100 MHz, CDCl_3): δ = 163.0 ($>\text{C}=\text{O}$), 160.6 ($>\text{C}=\text{O}$), 155.0 (Ar-C), 151.3 (Ar-C), 150.8 (Ar-C), 140.5 (C_3 of pyrazole), 139.2 (Ar-C), 137.9 (Ar-C), 135.3 (Ar-C), 133.6 (Ar-C), 132.5 (Ar-C), 131.2 (C_5 of pyrazole), 130.9 (Ar-C), 130.4 (Ar-C), 130.1 (Ar-C), 129.4 (Ar-C), 128.7 (Ar-C), 128.5 (Ar-C), 127.1 (Ar-C), 126.5 (Ar-C), 126.3 (Ar-C), 126.0 (Ar-C), 124.9 (Ar-C), 120.5 (Ar-C), 118.9 (Ar-C), 118.3 (Ar-C), 111.4 (Ar-C), 105.1 (C_4 of pyrazole); MS (m/z): 457.17 (M^+); element analysis calculated (%) for $\text{C}_{28}\text{H}_{19}\text{N}_5\text{O}_2$: C 73.51, H 4.19, N 15.31; found: C 73.57, H 4.29, N 15.42.

4.1.4. 7-(3-Bromophenyl)-5-(1,3-diphenyl-1H-pyrazol-4-yl)pyrido[2,3-d]pyrimidine-2,4(1H,3H)-dione (4b). Yield: 69%; solid; M.P. 225–227 $^\circ\text{C}$; IR (KBr, cm^{-1}): 3184, 2845 (C–H, $-\text{CH}=\text{CH}-$), 1746 ($>\text{C}=\text{O}$), 1679, 1661, 1267 (C=N, C=C, C–N), 679 (C–Br); ^1H NMR (400 MHz, CDCl_3): δ = 9.56 (s, 1H, Ar–NH), 9.47 (s, 1H, Ar–NH), 8.03 (s, 1H), 7.99–7.88 (m, 4H), 7.73–7.64 (m, 4H), 7.26–7.21 (m, 7H); ^{13}C NMR (100 MHz, CDCl_3): δ = 164.5 ($>\text{C}=\text{O}$), 160.4 ($>\text{C}=\text{O}$), 154.5 (Ar-C), 149.5 (Ar-C), 149.1 (Ar-C), 146.5 (C_3 of pyrazole), 140.3 (Ar-C), 136.9 (Ar-C), 135.4 (Ar-C), 134.4 (Ar-C), 134.1 (Ar-C), 132.9 (C_5 of pyrazole), 130.4 (Ar-C), 129.9 (Ar-C), 129.6 (Ar-C), 128.6 (Ar-C), 128.4 (Ar-C), 127.5 (Ar-

C), 125.8 (Ar-C), 122.3 (Ar-C), 119.5 (Ar-C), 117.4 (Ar-C), 115.5 (Ar-C), 110.8 (Ar-C), 102.4 (C_4 of pyrazole); MS (m/z): 535.06 (M^+); element analysis calculated (%) for $\text{C}_{28}\text{H}_{18}\text{BrN}_5\text{O}_2$: C 62.70, H 3.38, N 13.06; found: C 62.87, H 3.42, N 13.15.

4.1.5. 7-(4-Bromophenyl)-5-(1,3-diphenyl-1H-pyrazol-4-yl)pyrido[2,3-d]pyrimidine-2,4(1H,3H)-dione (4c). Yield: 67%; solid; M.P. 228–230 $^\circ\text{C}$; IR (KBr, cm^{-1}): 3173, 2831 (C–H, $-\text{CH}=\text{CH}-$), 1744 ($>\text{C}=\text{O}$), 1689, 1676, 1254 (C=N, C=C, C–N), 689 (C–Br); ^1H NMR (400 MHz, CDCl_3): δ = 9.53 (s, 1H, Ar–NH), 9.41 (s, 1H, Ar–NH), 8.06 (s, 1H), 7.94–7.86 (m, 4H), 7.71–7.60 (m, 4H), 7.28–7.22 (m, 7H); ^{13}C NMR (100 MHz, CDCl_3): δ = 164.7 ($>\text{C}=\text{O}$), 160.0 ($>\text{C}=\text{O}$), 154.8 (Ar-C), 149.8 (Ar-C), 149.0 (Ar-C), 146.8 (C_3 of pyrazole), 140.6 (Ar-C), 136.7 (Ar-C), 135.3 (Ar-C), 134.5 (Ar-C), 134.2 (Ar-C), 132.5 (C_5 of pyrazole), 130.1 (Ar-C), 129.8 (Ar-C), 129.3 (Ar-C), 128.7 (Ar-C), 128.2 (Ar-C), 127.6 (Ar-C), 125.2 (Ar-C), 122.8 (Ar-C), 119.7 (Ar-C), 117.3 (Ar-C), 115.0 (Ar-C), 110.6 (Ar-C), 102.1 (C_4 of pyrazole); MS (m/z): 535.06 (M^+); element analysis calculated (%) for $\text{C}_{28}\text{H}_{18}\text{BrN}_5\text{O}_2$: C 62.70, H 3.38, N 13.06; found: C 62.85, H 3.47, N 13.16.

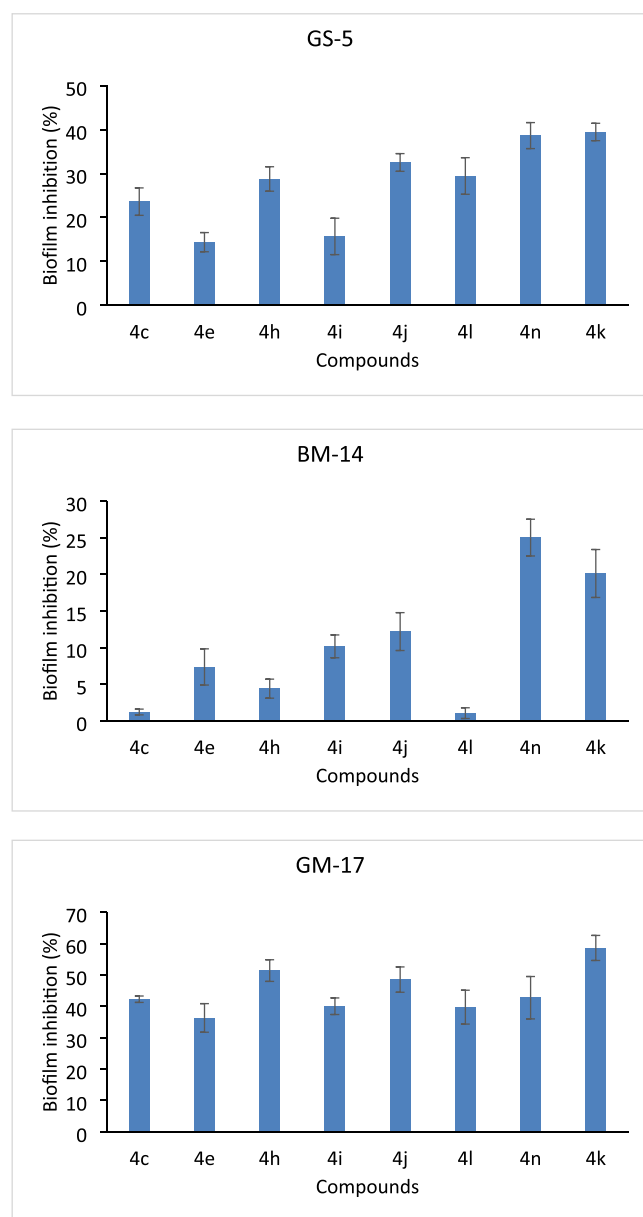


Figure 9. Biofilm formation inhibitory effects of different compounds on bacterial isolates (GS-5, *Vibrio* sp.; BM-14, *Staphylococcus* sp.; and GM-17, *Gallaecimonas* sp.).

4.1.6. 7-(2-Chlorophenyl)-5-(1,3-diphenyl-1H-pyrazol-4-yl)pyrido[2,3-d]pyrimidine-2,4(1H,3H)-dione (4d). Yield: 65%; solid; M.P. 192–193 °C; IR (KBr, cm^{-1}): 3185, 2817 (C–H, –CH=CH–), 1759 (>C=O), 1689, 1662, 1282 (C=N, C=C, C–N), 743 (C–Cl); ^1H NMR (400 MHz, CDCl_3): δ = 9.34 (s, 1H, Ar-NH), 9.21 (s, 1H, Ar-NH), 8.46 (s, 1H, Ar-H), 7.89–7.69 (m, 5H, Ar-H), 7.58–7.51 (m, 5H, Ar-H), 7.23–7.16 (m, 5H, Ar-H); ^{13}C NMR (100 MHz, CDCl_3): δ = 163.6 (>C=O), 158.3 (>C=O), 152.6 (Ar-C), 149.4 (Ar-C), 147.4 (C_3 of pyrazole), 140.8 (Ar-C), 139.5 (Ar-C), 136.3 (Ar-C), 135.7 (Ar-C), 134.4 (Ar-C), 132.8 (Ar-C), 131.6 (Ar-C), 131.4 (Ar-C), 131.1 (C_5 of pyrazole), 129.6 (Ar-C), 129.4 (Ar-C), 129.1 (Ar-C), 128.4 (Ar-C), 128.2 (Ar-C), 127.6 (Ar-C), 126.3 (Ar-C), 122.4 (Ar-C), 119.7 (Ar-C), 118.5 (Ar-C), 112.7 (Ar-C), 106.4 (C_4 of pyrazole); MS (m/z): 491.11 (M^+); element analysis calculated (%) for $\text{C}_{28}\text{H}_{18}\text{ClN}_5\text{O}_2$: C 68.36, H 3.69, N 14.24; found: C 68.43, H 3.75, N 14.32.

4.1.7. 7-(4-Chlorophenyl)-5-(1,3-diphenyl-1H-pyrazol-4-yl)pyrido[2,3-d]pyrimidine-2,4(1H,3H)-dione (4e). Yield: 69%; solid; M.P. 202–204 °C; IR (KBr, cm^{-1}): 3182, 2837 (C–H, –CH=CH–), 1754 (>C=O), 1694, 1689, 1265 (C=N, C=C, C–N), 753 (C–Cl); ^1H NMR (400 MHz, CDCl_3): δ = 9.24 (s, 1H, Ar-NH), 9.18 (s, 1H, Ar-NH), 8.40 (s, 1H, Ar-H), 7.86–7.78 (m, 5H, Ar-H), 7.55–7.50 (m, 5H, Ar-H), 7.28–7.20 (m, 5H, Ar-H); ^{13}C NMR (100 MHz, CDCl_3): δ = 163.3 (>C=O), 158.6 (>C=O), 152.3 (Ar-C), 149.8 (Ar-C), 147.4 (C_3 of pyrazole), 140.5 (Ar-C), 139.7 (Ar-C), 136.7 (Ar-C), 135.3 (Ar-C), 134.8 (Ar-C), 132.5 (Ar-C), 131.7 (Ar-C), 131.5 (Ar-C), 131.2 (C_5 of pyrazole), 129.8 (Ar-C), 129.2 (Ar-C), 129.0 (Ar-C), 128.8 (Ar-C), 128.5 (Ar-C), 127.1 (Ar-C), 126.0 (Ar-C), 122.4 (Ar-C), 119.4 (Ar-C), 118.3 (Ar-C), 112.0 (Ar-C), 106.5 (C_4 of pyrazole); MS (m/z): 491.11 (M^+); element analysis calculated (%) for $\text{C}_{28}\text{H}_{18}\text{ClN}_5\text{O}_2$: C 68.36, H 3.69, N 14.24; found: C 68.49, H 3.78, N 14.34.

4.1.8. 5-(1,3-Diphenyl-1H-pyrazol-4-yl)-7-(4-fluorophenyl)pyrido[2,3-d]pyrimidine-2,4(1H,3H)-dione (4f). Yield: 65%; solid; M.P. 199–201 °C; IR (KBr, cm^{-1}): 3188, 2816 (C–H, –CH=CH–), 1759 (>C=O), 1679, 1652, 1275 (C=N, C=C, C–N), 1269 (C–F); ^1H NMR (400 MHz, CDCl_3): δ = 9.29 (s, 1H, Ar-NH), 9.22 (s, 1H, Ar-NH), 8.37 (s, 1H, Ar-H), 7.84–7.76 (m, 5H, Ar-H), 7.51–7.46 (m, 5H, Ar-H), 7.32–7.27 (m, 5H, Ar-H); ^{13}C NMR (100 MHz, CDCl_3): δ = 164.3 (>C=O), 158.3 (>C=O), 152.7 (Ar-C), 149.3 (Ar-C), 147.7 (C_3 of pyrazole), 140.3 (Ar-C), 139.8 (Ar-C), 136.3 (Ar-C), 135.5 (Ar-C), 134.5 (Ar-C), 132.4 (Ar-C), 131.4 (Ar-C), 131.2 (Ar-C), 131.1 (C_5 of pyrazole), 129.5 (Ar-C), 129.3 (Ar-C), 129.1 (Ar-C), 128.9 (Ar-C), 128.4 (Ar-C), 127.5 (Ar-C), 126.2 (Ar-C), 122.7 (Ar-C), 119.2 (Ar-C), 118.8 (Ar-C), 112.4 (Ar-C), 106.3 (C_4 of pyrazole); MS (m/z): 475.14 (M^+); element analysis calculated (%) for $\text{C}_{28}\text{H}_{18}\text{FN}_5\text{O}_2$: C 70.73, H 3.82, N 14.73; found: C 70.85, H 3.96, N 14.79.

4.1.9. 5-(1,3-Diphenyl-1H-pyrazol-4-yl)-7-(2-hydroxyphenyl)pyrido[2,3-d]pyrimidine-2,4(1H,3H)-dione (4g). Yield: 73%; solid; M.P. 206–208 °C; IR (KBr, cm^{-1}): 3545 (–OH), 3181, 2819 (C–H, –CH=CH–), 1752 (>C=O), 1685, 1665, 1279 (C=N, C=C, C–N); ^1H NMR (400 MHz, CDCl_3): δ 9.41 (s, 1H, Ar-NH), 9.12 (s, 1H, Ar-NH), 8.53 (s, 1H, Ar-H), 8.11–7.89 (m, 4H, Ar-H), 7.75–7.60 (m, 3H, Ar-H), 7.51–7.42 (m, 4H, Ar-H), 7.26–7.17 (m, 4H, Ar-H), 7.04 (s, 1H, Ar-OH); ^{13}C NMR (100 MHz, CDCl_3): δ = 165.8 (>C=O), 161.3 (>C=O), 156.4 (Ar-C), 153.6 (Ar-C), 151.7 (Ar-C), 149.3 (Ar-C), 142.6 (C_3 of pyrazole), 140.3 (Ar-C), 135.7 (Ar-C), 133.4 (Ar-C), 132.4 (Ar-C), 131.6 (C_5 of pyrazole), 129.6 (Ar-C), 129.5 (Ar-C), 129.2 (Ar-C), 129.1 (Ar-C), 128.5 (Ar-C), 126.7 (Ar-C), 126.8 (Ar-C), 122.2 (Ar-C), 122.1 (Ar-C), 120.7 (Ar-C), 118.3 (Ar-C), 116.6 (Ar-C), 113.6 (Ar-C), 107.4 (C_4 of pyrazole); MS (m/z): 473.15 (M^+); element analysis calculated (%) for $\text{C}_{28}\text{H}_{19}\text{N}_5\text{O}_3$: C 71.03, H 4.04, N 14.79; found: C 71.2, H 4.23, N 14.98.

4.1.10. 5-(1,3-Diphenyl-1H-pyrazol-4-yl)-7-(4-hydroxyphenyl)pyrido[2,3-d]pyrimidine-2,4(1H,3H)-dione (4h). Yield: 77%; solid; M.P. 201–203 °C; IR (KBr, cm^{-1}): 3554 (–OH), 3179, 2814 (C–H, –CH=CH–), 1758 (>C=O), 1679, 1661, 1277 (C=N, C=C, C–N); ^1H NMR (400 MHz, CDCl_3): δ = 9.34 (s, 1H, Ar-NH), 9.09 (s, 1H, Ar-NH), 8.44 (s, 1H, Ar-H), 8.05–7.99 (m, 4H, Ar-H), 7.70–7.65 (m, 3H, Ar-H), 7.56–7.44 (m, 4H, Ar-H), 7.27–7.20 (m, 4H, Ar-H), 7.08 (s, 1H, Ar-OH); ^{13}C NMR (100 MHz, CDCl_3): δ = 163.3 (>C=O), 161.0 (>C=O), 156.7 (Ar-C), 153.6 (Ar-C), 151.3 (Ar-C), 149.1 (Ar-C), 142.5 (C_3 of pyrazole), 140.3 (Ar-

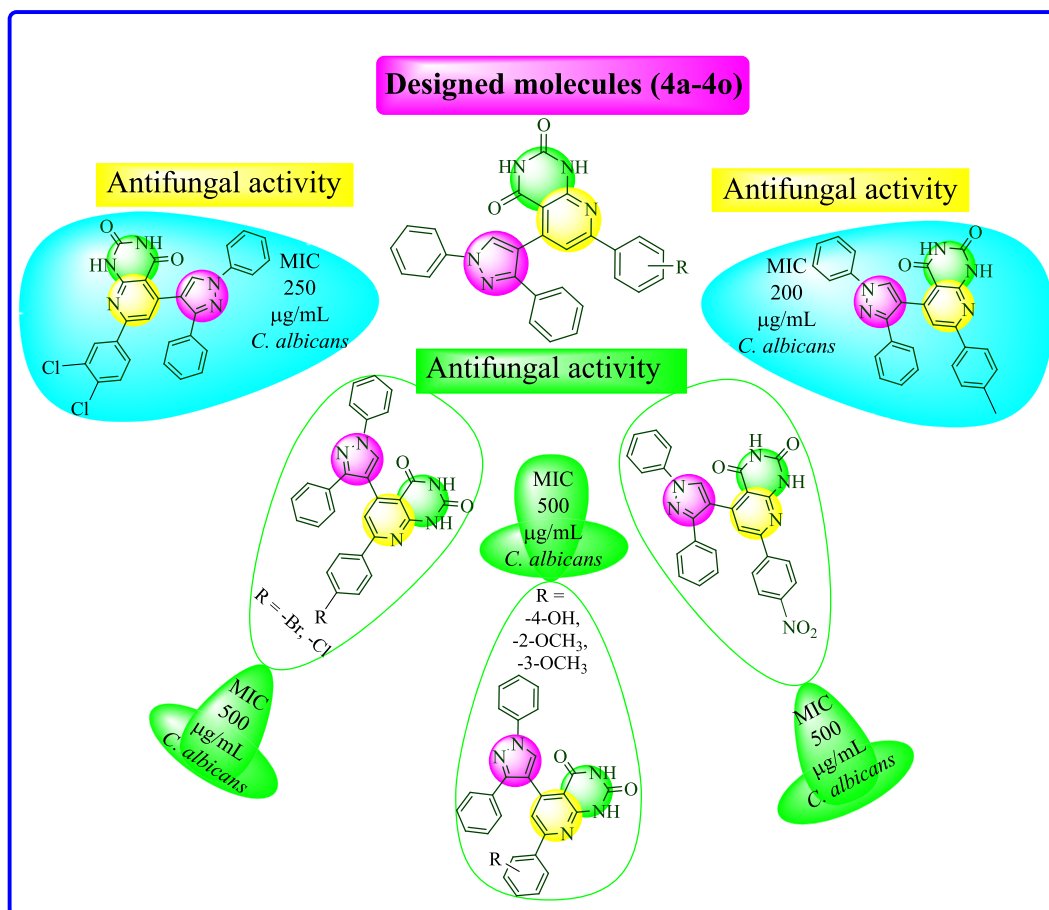


Figure 10. The effect of various functional groups on antifungal activity of pyrazole bearing pyrido[2,3-*d*]pyrimidine-2,4-dione analogues.

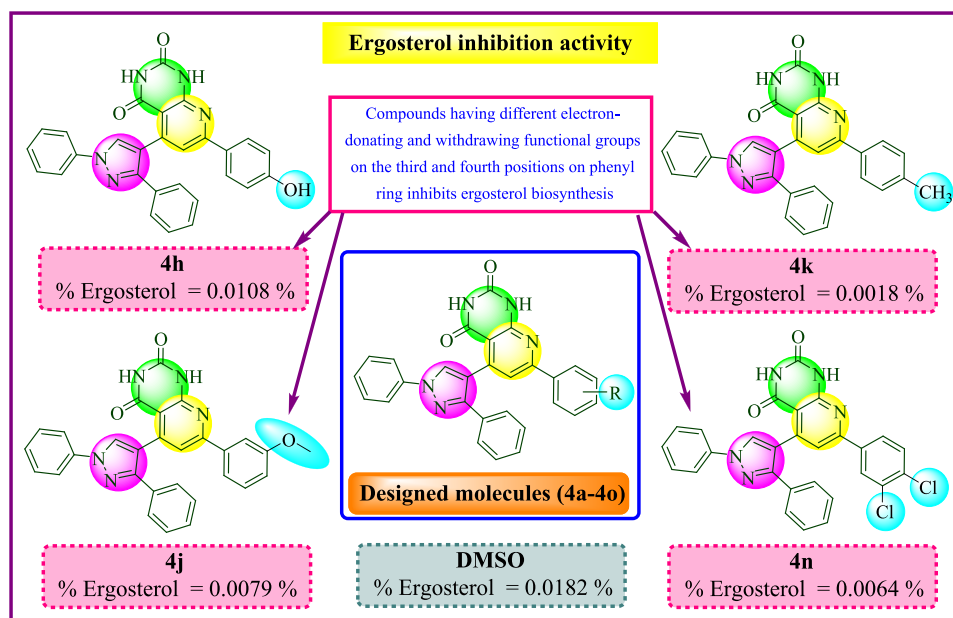


Figure 11. The effect of compound derivatization on antifungal activity of synthetic hybrids against ergosterol biosynthesis of compounds 4h, 4j, 4k, and 4n.

C), 135.2 (Ar-C), 133.7 (Ar-C), 132.7 (Ar-C), 131.2 (C₅ of pyrazole), 129.8 (Ar-C), 129.7 (Ar-C), 129.5 (Ar-C), 129.2 (Ar-C), 128.5 (Ar-C), 126.6 (Ar-C), 126.4 (Ar-C), 122.8 (Ar-C), 122.3 (Ar-C), 120.5 (Ar-C), 118.6 (Ar-C), 116.3 (Ar-C), 113.3

(Ar-C), 107.9 (C₄ of pyrazole); MS (*m/z*): 473.15 (M⁺); element analysis calculated (%) for C₂₈H₁₉N₅O₃: C 71.03, H 4.04, N 14.79; found: C 71.19, H 4.22, N 14.96.

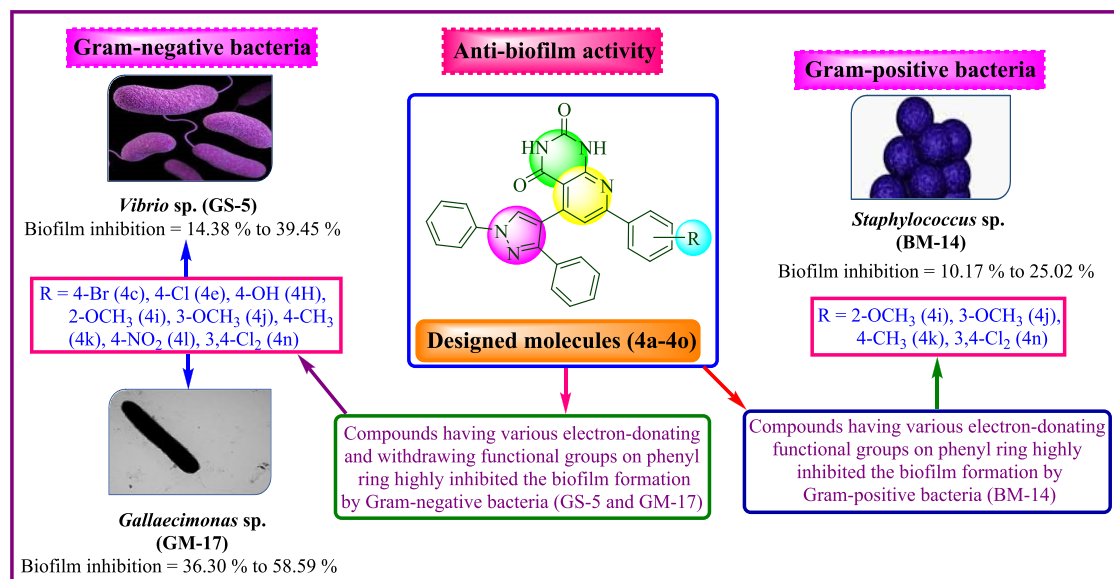


Figure 12. The effect of various functional groups on biofilm formation by Gram-positive and Gram-negative bacteria.

4.1.11. 5-(1,3-Diphenyl-1H-pyrazol-4-yl)-7-(2-methoxyphenyl)pyrido[2,3-d]pyrimidine-2,4(1H,3H)-dione (**4i**). Yield: 81%; solid; M.P. 207–209 °C; IR (KBr, cm^{-1}): 3182, 2816 (C–H, –CH=CH–), 1754 (>C=O), 1686, 1651, 1275 (C–N, C=C, C–N), 1141 (C–OCH₃); ¹H NMR (400 MHz, CDCl₃): δ = 9.66 (s, 1H, Ar-NH), 9.42 (s, 1H, Ar-NH), 8.74 (s, 1H, Ar-H), 7.70–7.67 (m, 4H, Ar-H), 7.49–7.44 (m, 1H, Ar-H), 7.51–7.39 (m, 6H, Ar-H), 7.34–7.26 (m, 4H, Ar-H), 3.88 (s, 3H, Ar-OCH₃); ¹³C NMR (100 MHz, CDCl₃): δ = 163.4 (>C=O), 162.4 (>C=O), 158.6 (Ar-C), 154.8 (Ar-C), 153.6 (Ar-C), 149.7 (Ar-C), 143.4 (C₃ of pyrazole), 140.5 (Ar-C), 139.6 (Ar-C), 138.3 (Ar-C), 132.7 (Ar-C), 132.4 (Ar-C), 131.4 (Ar-C), 131.1 (C₅ of pyrazole), 129.9 (Ar-C), 129.7 (Ar-C), 129.1 (Ar-C), 127.6 (Ar-C), 126.3 (Ar-C), 125.2 (Ar-C), 121.7 (Ar-C), 120.4 (Ar-C), 118.4 (Ar-C), 111.6 (Ar-C), 110.3 (Ar-C), 105.6 (C₄ of pyrazole), 50.9 (Ar-OCH₃); 487.16 (M⁺); element analysis calculated (%) for C₂₉H₂₁N₅O₃: C 71.45, H 4.34, N 14.37; found: C 71.55, H 4.49, N 14.47.

4.1.12. 5-(1,3-Diphenyl-1H-pyrazol-4-yl)-7-(3-methoxyphenyl)pyrido[2,3-d]pyrimidine-2,4(1H,3H)-dione (**4j**). Yield: 75%; solid; M.P. 210–212 °C; IR (KBr, cm^{-1}): 3182, 2818 (C–H, –CH=CH–), 1756 (>C=O), 1684, 1658, 1278 (C=N, C=C, C–N), 1148 (C–OCH₃); ¹H NMR (400 MHz, CDCl₃): δ = 9.50 (s, 1H, Ar-NH), 9.37 (s, 1H, Ar-NH), 8.71 (s, 1H, Ar-H), 7.76–7.70 (m, 4H, Ar-H), 7.57–7.53 (m, 1H, Ar-H), 7.51–7.44 (m, 6H, Ar-H), 7.28–7.23 (m, 4H, Ar-H), 3.91 (s, 3H, Ar-OCH₃); ¹³C NMR (100 MHz, CDCl₃): δ = 163.9 (>C=O), 162.8 (>C=O), 158.9 (Ar-C), 154.5 (Ar-C), 153.7 (Ar-C), 149.6 (Ar-C), 143.8 (C₃ of pyrazole), 140.3 (Ar-C), 139.3 (Ar-C), 138.9 (Ar-C), 132.8 (Ar-C), 132.3 (Ar-C), 131.7 (Ar-C), 131.2 (C₅ of pyrazole), 129.8 (Ar-C), 129.6 (Ar-C), 129.3 (Ar-C), 127.7 (Ar-C), 126.8 (Ar-C), 125.4 (Ar-C), 121.1 (Ar-C), 120.5 (Ar-C), 118.3 (Ar-C), 111.2 (Ar-C), 110.5 (Ar-C), 105.7 (C₄ of pyrazole), 50.1 (Ar-OCH₃); MS (*m/z*): 487.16 (M⁺); element analysis calculated (%) for C₂₉H₂₁N₅O₃: C 71.45, H 4.34, N 14.37; found: C 71.61, H 4.51, N 14.54.

4.1.13. 5-(1,3-Diphenyl-1H-pyrazol-4-yl)-7-(*p*-tolyl)pyrido[2,3-d]pyrimidine-2,4(1H,3H)-dione (**4k**). Yield: 77%; solid; M.P. 198–200 °C; IR (KBr, cm^{-1}): 3179, 2819 (C–H, –CH=CH–), 1758 (>C=O), 1685, 1667, 1278 (C=N, C=C, C–N);

¹H NMR (400 MHz, CDCl₃): δ = 9.02 (s, 1H, Ar-NH), 8.94 (s, 1H, Ar-NH), 8.17 (s, 1H, Ar-H), 7.75–7.68 (m, 6H, Ar-H), 7.56–7.39 (m, 6H, Ar-H), 7.28–7.22 (m, 3H, Ar-H), 2.53 (s, 3H, Ar-CH₃); ¹³C NMR (100 MHz, CDCl₃): δ = 158.5 (>C=O), 154.7 (>C=O), 149.7 (Ar-C), 148.4 (Ar-C), 147.7 (Ar-C), 142.9 (C₃ of pyrazole), 139.2 (Ar-C), 138.0 (Ar-C), 137.6 (Ar-C), 133.5 (Ar-C), 133.3 (Ar-C), 131.5 (C₅ of pyrazole), 131.0 (Ar-C), 129.8 (Ar-C), 129.3 (Ar-C), 129.2 (Ar-C), 128.5 (Ar-C), 127.7 (Ar-C), 127.3 (Ar-C), 126.2 (Ar-C), 124.2 (Ar-C), 121.7 (Ar-C), 118.7 (Ar-C), 115.1 (Ar-C), 113.1 (Ar-C), 106.3 (C₄ of pyrazole), 25.3 (Ar-CH₃); MS (*m/z*): 471.17 (M⁺); element analysis calculated (%) for C₂₉H₂₁N₅O₂: C 73.87, H 4.49, N 14.85; found: C 73.97, H 4.57, N 14.92.

4.1.14. 5-(1,3-Diphenyl-1H-pyrazol-4-yl)-7-(4-nitrophenyl)pyrido[2,3-d]pyrimidine-2,4(1H,3H)-dione (**4l**). Yield: 67%; solid; M.P. 228–230 °C; IR (KBr, cm^{-1}): 3180, 2824 (C–H, –CH=CH–), 1761 (>C=O), 1679, 1669, 1545 (C–NO₂), 1279 (C=N, C=C, C–N); ¹H NMR (400 MHz, CDCl₃): δ = 9.34 (s, 1H, Ar-NH), 9.27 (s, 1H, Ar-NH), 8.34 (s, 1H, Ar-H), 7.83–7.78 (m, 5H, Ar-H), 7.56–7.48 (m, 5H, Ar-H), 7.33–7.25 (m, 5H, Ar-H); ¹³C NMR (100 MHz, CDCl₃): δ = 164.5 (>C=O), 158.3 (>C=O), 152.4 (Ar-C), 149.3 (Ar-C), 147.7 (C₃ of pyrazole), 140.6 (Ar-C), 139.3 (Ar-C), 136.4 (Ar-C), 135.8 (Ar-C), 134.4 (Ar-C), 132.2 (Ar-C), 131.8 (Ar-C), 131.4 (Ar-C), 131.3 (C₅ of pyrazole), 129.7 (Ar-C), 129.3 (Ar-C), 129.2 (Ar-C), 128.6 (Ar-C), 128.3 (Ar-C), 127.5 (Ar-C), 126.3 (Ar-C), 122.8 (Ar-C), 119.6 (Ar-C), 118.2 (Ar-C), 112.6 (Ar-C), 106.3 (C₄ of pyrazole); MS (*m/z*): 502.14 (M⁺); element analysis calculated (%) for C₂₈H₁₈N₆O₄: C 66.93, H 3.61, N 16.73; found: C 67.12, H 3.75, N 16.77.

4.1.15. 7-(2,4-Dichlorophenyl)-5-(1,3-diphenyl-1H-pyrazol-4-yl)pyrido[2,3-d]pyrimidine-2,4(1H,3H)-dione (**4m**). Yield: 77%; solid; M.P. 213–215 °C; IR (KBr, cm^{-1}): 3182, 2818 (C–H, –CH=CH–), 1759 (>C=O), 1684, 1657, 1271 (C=N, C=C, C–N), 758 (C–Cl); ¹H NMR (400 MHz, CDCl₃): δ = 9.46 (s, 1H, Ar-NH), 9.37 (s, 1H, Ar-NH), 8.62 (s, 1H, Ar-H), 7.89–7.86 (m, 4H, Ar-H), 7.64–7.57 (m, 8H, Ar-H), 7.30–7.19 (m, 2H, Ar-H); ¹³C NMR (100 MHz, CDCl₃): δ = 163.4 (>C=O), 161.3 (>C=O), 157.8 (Ar-C), 155.3 (Ar-C), 151.8 (Ar-C), 144.4 (Ar-C), 141.2 (C₃ of pyrazole), 138.7 (Ar-

C), 137.2 (Ar-C), 135.6 (Ar-C), 134.2 (Ar-C), 132.9 (Ar-C), 132.5 (C₅ of pyrazole), 130.3 (Ar-C), 129.7 (Ar-C), 129.5 (Ar-C), 128.8 (Ar-C), 127.3 (Ar-C), 127.4 (Ar-C), 126.3 (Ar-C), 123.5 (Ar-C), 118.3 (Ar-C), 116.7 (Ar-C), 112.3 (Ar-C), 101.9 (C₄ of pyrazole); MS (*m/z*): 525.08 (M⁺); element analysis calculated (%) for C₂₈H₁₇C₁₂N₅O₂: C 63.89, H 3.26, N 13.31; found: C 63.98, H 3.34, N 13.45.

4.1.16. 7-(3,4-Dichlorophenyl)-5-(1,3-diphenyl-1H-pyrazol-4-yl)pyrido[2,3-d]pyrimidine-2,4(1H,3H)-dione (4n). Yield: 77%; solid; M.P. 213–215 °C; IR (KBr, cm⁻¹): 3182, 2818 (C–H, –CH=CH–), 1759 (>C=O), 1684, 1657, 1271 (C=N, C=C, C–N), 758 (C–Cl); ¹H NMR (400 MHz, CDCl₃): δ = 9.39 (s, 1H, Ar-NH), 9.32 (s, 1H, Ar-NH), 8.67 (s, 1H, Ar-H), 7.98–7.90 (m, 4H, Ar-H), 7.57–7.42 (m, 8H, Ar-H), 7.32–7.22 (m, 2H, Ar-H); ¹³C NMR (100 MHz, CDCl₃): δ = 163.9 (>C=O), 161.7 (>C=O), 157.5 (Ar-C), 155.6 (Ar-C), 151.5 (Ar-C), 144.7 (Ar-C), 141.9 (C₃ of pyrazole), 138.6 (Ar-C), 137.5 (Ar-C), 135.0 (Ar-C), 134.7 (Ar-C), 132.8 (Ar-C), 132.0 (C₅ of pyrazole), 130.4 (Ar-C), 129.9 (Ar-C), 129.3 (Ar-C), 128.2 (Ar-C), 127.9 (Ar-C), 127.7 (Ar-C), 126.8 (Ar-C), 123.5 (Ar-C), 118.3 (Ar-C), 116.1 (Ar-C), 112.5 (Ar-C), 101.6 (C₄ of pyrazole); MS (*m/z*): 525.08 (M⁺); element analysis calculated (%) for C₂₈H₁₇Cl₂N₅O₂: C 63.89, H 3.26, N 13.31; found: C 63.98, H 3.34, N 13.45.

4.1.17. 7-(2,4-Difluorophenyl)-5-(1,3-diphenyl-1H-pyrazol-4-yl)pyrido[2,3-d]pyrimidine-2,4(1H,3H)-dione (4o). Yield: 75%; solid; M.P. 204–206 °C; IR (KBr, cm⁻¹): 3174, 2814 (C–H, –CH=CH–), 1751 (>C=O), 1684, 1664, 1277 (C=N, C=C, C–N), 1287 (C–F); ¹H NMR (400 MHz, CDCl₃): δ = 9.43 (s, 1H, Ar-NH), 9.40 (s, 1H, Ar-NH), 8.76 (s, 1H, Ar-H), 7.87–7.76 (m, 4H, Ar-H), 7.49–7.39 (m, 8H, Ar-H), 7.27–7.15 (m, 2H, Ar-H); ¹³C NMR (100 MHz, CDCl₃): δ = 165.2 (>C=O), 161.5 (>C=O), 157.6 (Ar-C), 155.3 (Ar-C), 151.8 (Ar-C), 144.5 (Ar-C), 141.6 (C₃ of pyrazole), 138.3 (Ar-C), 137.7 (Ar-C), 135.4 (Ar-C), 134.8 (Ar-C), 132.4 (Ar-C), 132.1 (C₅ of pyrazole), 130.7 (Ar-C), 129.8 (Ar-C), 129.5 (Ar-C), 128.1 (Ar-C), 127.6 (Ar-C), 127.3 (Ar-C), 126.3 (Ar-C), 123.7 (Ar-C), 118.4 (Ar-C), 116.6 (Ar-C), 112.3 (Ar-C), 101.6 (C₄ of pyrazole); MS (*m/z*): 493.14 (M⁺); element analysis calculated (%) for C₂₈H₁₇F₂N₅O₂: C 68.15, H 3.47, N 14.19; found: C 68.45, H 3.54, N 14.56.

5. MATERIALS AND METHODS

5.1. Antifungal Bioassay. “The synthesized compounds (4a–4o) were screened for their antifungal activity against *Candida albicans* (MTCC-227), *Aspergillus niger* (MTCC-282), *Aspergillus clavatus* (MTCC-1323) at concentrations of 1000, 500, and 250 μg/mL. The experiments were conducted in triplicates. The active compounds obtained as above in primary screening were similarly diluted to obtain 200, 125, 100, 62.5, 50, 25, and 12.5 μg/mL concentrations for secondary screening to test in the second set of dilutions against all microorganisms. ‘Griseofulvin’ was used as a standard drug for antifungal activity and was considered a positive control. For the growth of fungi, Sabouraud dextrose broth was used and incubated at 28 °C in aerobic conditions for 48 h. DMSO and sterilized distilled water were used as negative controls.”⁴⁰ (Figures S2–S4).

5.2. Molecular Docking and Computational Methodology for Target β-Tubulin. The β-tubulin protein sequence of *C. albicans* was retrieved from the NCBI protein database. A protein–protein blast (BLASTP)^{41,42} was carried out to obtain homologous proteins having an experimental structure retrieved from the Protein Data Bank (PDB). The sequences of target and

template proteins were aligned using clustalX, and the protein structure of *C. albicans* β-tubulin protein was constructed using the template protein by homology modeling using Modeller 10.^{43–45} The protein structure was validated by protein structure validation server SAVES v 6.0 of UCLA-DOE Lab.^{46–48} The constructed protein of *C. albicans* β-tubulin was prepared using the protein preparation wizard in Schrödinger Suite using the default parameter. A grid was constructed at the Taxol binding site, and molecules 4a–4o were docked into the Taxol binding site by the GLIDE module using the extra precision (XP) mode. The docked complexes were further subjected to binding energy calculation using PRIME-MMGBSA. Molecule 4a was optimized, and vibrational frequency calculations were performed in Gaussian applying the DFT method using B3LYP/6-31G* level theory. ADME properties were calculated using the QikProp module of Schrödinger suite, and toxicity parameters were calculated by the online pkCSM server.⁴⁹

5.3. Ergosterol Inhibition Activity. **5.3.1. Materials and Methods.** Ergosterols were extracted by using the standard protocol.⁵⁰ The sterols were quantitated by Breivik and Owades in 1957.³⁰ Overnight-grown *Candida* culture was inoculated in 50 mL of the potato dextrose broth containing the MIC concentration of compounds. All the solutions were prepared in 2 mL of DMSO according to their MIC (500 μg/mL for compounds 4h, 4j, and 4l; 250 μg/mL for compounds 4k; and 200 μg/mL for compounds 4n), 50 μg/mL concentration of fluconazole, and 2 mL concentration of DMSO. The cultures were incubated on a shaker at 37 °C for 72 h. The cultures were centrifuged at 5000 rpm for 5 min and washed once with sterile distilled water for harvesting the fungal cells. The net wet weight of the cell pellet was measured. 3 mL of 25% alcoholic KOH was added to each pellet and mixed by vortexing for 1 min. This was incubated at 85 °C water bath for 1 h. A mixture of 1 mL of sterile distilled water and 3 mL of *n*-heptane was added to each tube after cooling for extraction of sterols. The upper heptane layer was transferred to a clean tube and diluted 10 times with *n*-heptane:ethanol (1:1) and canned spectrophotometrically from 240 and 300 nm. Ergosterol content was calculated as a percentage of the wet weight of the cell by the following equations:

$$\% \text{ergosterol} = (\% \text{ergosterol} + \% 24(28)\text{DHE}) - (\% 24(28)\text{DHE}) \quad (1)$$

$$(\% \text{ergosterol} + \% 24(28)\text{DHE}) = \left[\left(\frac{A_{281.5}}{290} \right) \times F \right] / \text{pellet weight} \quad (2)$$

$$\% 24(28)\text{DHE} = \left(\frac{A_{230}}{518} \right) \times F / \text{pellet weight} \quad (3)$$

$$\left[\left(\frac{A_{281.5}}{290} \right) 3 F \right] / \text{pellet weight}, \% 24(28)\text{DHE} \quad (4)$$

where *F* is the factor for dilution in ethanol and 290 and 518 are the *E* values (in percentages per centimeter) determined for crystalline ergosterol and 24(28)DHE, respectively.

5.3.2. Molecular Docking Analysis of Target Sterol 14-Alpha Demethylase Proteins/Macromolecules. Sterol 14-demethylase is one of the important enzymes in ergosterol biosynthesis in fungi and is used as a target for many antifungal agents.^{32,33} This enzyme was selected to find the mechanism of action of these compounds as an antifungal candidate. The 3D crystal structure of this enzyme from *C. albicans* (PDB ID: 5v5z)

was downloaded from the Protein Data Bank (<https://www.rcsb.org/>), in .pdb format.⁵¹ The protein molecule was cleaned by removing bound complexes. The protein structure was prepared by removing water molecules using PyMOL version 2.3.3.

5.3.3. Preparation of Ligands. The 3D structures of the ligands were drawn, and energy was minimized by Open Babel, an open-source platform.

5.3.4. Determination of Active Sites. The Computed Atlas for Surface Topography of Proteins (CASTp)⁵² and BIOVIA Discovery Studio 4.5⁵³ were used to determine the amino acids in the active site of a protein (<http://sts.bioe.uic.edu/castp/index.html?2011>).

5.3.5. Molecular Docking. Ligand and protein optimization was done using PyMOL version 2.3.3 [The PyMOL Molecular Graphics System, Version 2.0 Schrödinger LLC]. For ligand optimization, the geometry of ligands was cleaned, whereas, for protein, the water was removed. The docking was performed by using PyRx 0.8.⁵⁴ The docking analyses were performed using both PyMOL as well as BIOVIA Discovery Studio 4.5.⁵¹

5.4. Anti-Biofilm Activity. **5.4.1. Materials and Methods.** The ability of the compounds (**4c**, **4e**, **4h**, **4i**, **4j**, **4l**, **4n**, and **4k**) to inhibit biofilm formation was measured using the microplate-based assay, as described in ref 34 with three different marine biofilm-forming bacterial strains (GS-5, *Vibrio* sp. (Gram-negative); BM-14, *Staphylococcus* sp. (Gram-positive); and GM-17, *Gallaecimonas* sp. (Gram-negative)).³⁵ Bacterial isolates were cultured in ZMB (Zobell Marine Broth) overnight and diluted in sterile ZMB broth containing 1% glucose in 1:10 ratio. 10 μ L of each chemical compound in the concentration of 400 μ g/mL dissolved in DMSO was transferred into the wells of a microtiter plate containing 90 μ L of diluted bacterial culture and replicated six times. Thereafter, the plate was kept on a stirrer for proper mixing of culture and compounds and kept in a static incubator at 30 °C for 24 h. After incubation, the wells were washed three times with distilled water and dripped with 1% crystal violet solution (100 μ L), incubated at room temperature for 30 min, and again, washed with distilled water three times and dried at room temperature. Next, 100 μ L of 70% ethanol was added to each well, and optical density was measured at 590 nm using a microplate spectrophotometer. Bacterial culture without compound was used as control. The percentage biofilm inhibition was calculated with the following formula:

$$\text{biofilm inhibition (\%)} = \frac{\text{OD (control)} - \text{OD (experimental)}}{\text{OD (control)}} \times 100 \quad (5)$$

For the antimicrobial susceptibility assay, the disc diffusion method was used. With the help of a sterile cell spreader, the bacterial colony adjusted to 0.6 OD, spread on Zobell marine agar, and dried for 15 min. Discs were impregnated in the chemical compounds (400 μ g/mL) and then placed on a Zobell Marine Agar plate. Each plate contained five disc, four chemical-compound-treated discs, and one as a control followed by incubation at 37 °C for 48 h, and the zone of inhibition was examined.

■ ASSOCIATED CONTENT

SI Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acsomega.3c01722>.

Antibacterial activities of chemical compounds; antifungal activity against *C. albicans*; antifungal activity against *A. niger*; antifungal activity against *A. clavatus*; Procheck–Ramachandran plot; Verify3D plot (Verify3D—Pass); Errat—overall quality factor 77.6722; ProSa; ligand interaction diagram of **4a**; ligand interaction diagram of **4k**; ligand interaction diagram of **4n**; ligand interaction diagram of griseofulvin; IR frequency data obtained from DFT; IR frequency data; ¹H NMR spectra of compound **4a**; ¹³C NMR spectra of compound **4a**; mass spectra of compound **4a**; IR spectra of compound **4a**; ¹H NMR spectra of compound **4c**; ¹³C NMR spectra of compound **4c**; ¹H NMR spectra of compound **4e**; ¹³C NMR spectra of compound **4e**; mass spectra of compound **4e**; ¹H NMR spectra of compound **4h**; ¹³C NMR spectra of compound **4h**; mass spectra of compound **4h**; ¹H NMR spectra of compound **4j**; ¹³C NMR spectra of compound **4j**; mass spectra of compound **4j**; ¹H NMR spectra of compound **4k**; ¹³C NMR spectra of compound **4k**; mass spectra of compound **4k**; ¹H NMR spectra of compound **4n**; ¹³C NMR spectra of compound **4n**; mass spectra of compound **4n** (PDF)

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Notes

The authors declare no competing financial interest.

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