Clinical pharmacokinetics and dose recommendations for posaconazole gastroresistant tablets in children with cystic fibrosis

Siân Bentley 💿 ^{1,2}*, Jane C. Davies^{2,3}, Silke Gastine⁴, Jackie Donovan⁵ and Joseph F. Standing 💿 ^{4,6}

¹Pharmacy Department, Royal Brompton Hospital, London, UK; ²National Heart and Lung Institute, Imperial College London, London, UK; ³Paediatric Respiratory Medicine Department, Royal Brompton Hospital, London, UK; ⁴Infection, Immunity and Inflammation, Great Ormond Street Institute of Child Health, University College London, London, UK; ⁵Clinical Biochemistry Department, Royal Brompton Hospital, London, UK; ⁶Pharmacy Department, Great Ormond Street Hospital, London, UK; ⁶Dundation Trust, London, UK

*Corresponding author. E-mail: s.bentley@rbht.nhs.uk

Received 1 April 2021; accepted 21 July 2021

Objectives: To investigate the population pharmacokinetics of posaconazole gastroresistant tablets in children with cystic fibrosis (CF) and perform simulations to recommend optimal doses.

Patients and methods: Children from a paediatric CF centre who had received posaconazole tablets and underwent therapeutic drug monitoring were identified from pharmacy records. Relevant clinical data were collated from case notes and electronic patient records and used to develop an allometrically scaled population pharmacokinetic model. A stepwise covariate model-building exercise evaluated the influence of interacting medicines and liver function.

Results: One hundred posaconazole serum concentrations were collected from 37 children with a median age of 14 years (range 7–17). Posaconazole pharmacokinetics were adequately described by a one-compartment model with inter-individual variability on clearance. Dose simulations demonstrated a 77%–83% probability of attaining a trough target of 1 mg/L with a dose of 300 mg every 12 h for two doses then 300 mg once daily (OD) in children aged 6–11 years; and 86%–88% with a dose of 400 mg every 12 h for two doses then 400 mg OD in adolescents aged 12–17 years. This dose scheme also yielded a 90% probability of achieving an AUC of 30 mg·h/L. AUC and trough concentration were highly correlated (r^2 = 0.98). Simulations showed that trough concentrations of >0.75 mg/L would exceed an AUC of 30 mg·h/L in 90% of patients.

Conclusions: A starting dose of 300 mg OD in those aged 6–11 years and 400 mg OD in those aged 12–17 years (following loading doses) yields a 90% probability of attaining an AUC of 30 mg·h/L.

Introduction

Cystic fibrosis (CF) is a common autosomal recessive genetic disorder, resulting from the defective CF transmembrane conductance regulator (CFTR) gene, which causes abnormal mucous secretions in multiple organs, notably the lungs, liver and pancreas. Lung disease is characterized by chronic pulmonary infection by opportunistic bacteria and fungi, leading to inflammation and a progressive decline in lung function. *Aspergillus fumigatus* is one of the most commonly isolated fungi in sputum from CF patients.¹ Reported prevalence in children varies between 16% and 42%.^{2,3} In 2019, UK CF Registry data reported *Aspergillus* grown in 9.4% of children (aged 0–15 years) in the 12 months prior to the 2019 annual review. This is proportionally higher in older age groups, with positive sputum samples found in 10.1% of children aged 8– 11 years and 18.9% in those aged 12–15 years.⁴ Other fungi commonly isolated in CF include *Scedosporium* species and *Exophiala dermatitidis*.¹

Triazole antifungal medicines, such as posaconazole, are used for the treatment of lung disease caused by these fungi in children with CF. Although there are no national or international guidelines specific to CF, this is in line with international guidelines for the wider population.⁵⁻⁷ Posaconazole is not licensed in children <18 years old in the UK and EU, or those <13 years old in the USA; however, it is increasingly being used in children with CF due to the poor tolerability, toxicity and high inter-individual variability (IIV) in drug levels of the alternative azoles, itraconazole and voriconazole.^{8,9} Due to the lack of data supporting posaconazole dosing regimens in children, as well as the aforementioned IIV exhibited in other azoles, therapeutic drug monitoring (TDM) is routinely carried out. The ESCMID-ECMM-ERS guidelines for the diagnosis and management of *Aspergillus* diseases and British Society of Medical

© The Author(s) 2021. Published by Oxford University Press on behalf of the British Society for Antimicrobial Chemotherapy. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/ by/4.0/), which permits unrestricted reuse, distribution, and reproduction in any medium, provided the original work is properly cited. Mycology antifungal TDM guidelines suggest a posaconazole treatment target trough concentration of >1 mg/L, with no upper limit yet shown to be associated with toxicity.^{7,10}

People with CF are considered to be a unique population, displaying altered pharmacokinetics for some drugs, most likely as a result of comorbidities affecting drug absorption and disposition.¹¹ This has been borne out in several studies; in a small prospective pharmacokinetic study of itraconazole in paediatric CF patients, many failed to achieve adequate serum concentrations with standard doses.¹² More recently, in a retrospective observational study evaluating posaconazole plasma concentrations in lung transplant recipients, C_{min} for the tablet formulation was found to be 48% lower in patients with CF.¹³ Together with a paucity of paediatric pharmacokinetic data for medicines new to the market on which to base dosing regimens, the use of inappropriate dosing regimens may reduce the effectiveness of posaconazole in this cohort of patients, increase the risk of adverse effects and potentially drive resistance; in a tertiary institution, 16.2% of A. fumigatus isolates from CF patients were found to be resistant to azole antifungals.14

A recently published population pharmacokinetic model provided dose recommendations for a liquid formulation of posaconazole in immunocompromised children.¹⁵ Given the potential differences in the pharmacokinetics of some medicines in people with CF, and extensive use of the tablet formulation of posaconazole in this population, with known pharmacokinetic differences to the liquid formulation,¹⁶ our study aimed to develop a population pharmacokinetic model of posaconazole gastroresistant tablets in a cohort of paediatric CF patients. This was used to evaluate the influence of patient characteristics, including interacting medicines, on posaconazole exposure. We then performed simulations to produce dosing recommendations in this unique population.

Patients and methods

Study design

Retrospective patient data collected during the routine care of children aged between 5 and 17 years (both inpatients and outpatients) within the paediatric CF clinic at a tertiary centre, who had received posaconazole gastroresistant tablets for treatment of CF-related fungal lung disease between 2014 and 2020, were collated. The study and use of anonymized clinical data was approved by the Trust's Research and Development Office and Health Research Authority, reference number 20/HRA/3760. As the study involved the retrospective use of anonymized data, formal participation/ parental consent was not required. Patients were identified by matching pharmacy and clinical biochemistry records by pharmacy staff who routinely care for children with CF. Patients who had at least one posaconazole TDM sample taken were included. The time and date of the posaconazole TDM sample, along with the reported concentration, were extracted from electronic patient records. Samples that were below the limit of quantification in the analysis were recorded in the dataset as LOQ/2. For inpatients, dosing history was taken from the electronic nursing administration record, capturing doses given in the preceding 48 h before sampling. For outpatients, the time of the last dose, if not recorded, was presumed to be at 8 am on the day of the sample and assumed to be at steady state based on the patient's drug history. The model was run with and without these samples to check if this assumption led to biased parameter estimates. Additional data collected from electronic patient records were basic demographic information, liver function at the time of sampling, concomitant

Analytical method

Posaconazole analysis was carried out using 2D TurboFlowTM HPLC tandem MS (2D HPLC-MS/MS). This was on a Thermo Fisher Scientific TranscendTM prior to 2017 and a Prelude SPLCTM HPLC system coupled with a TSQ EnduraTM triple quadruple mass spectrometer (Thermo Fisher Scientific, CA, USA) thereafter.

Analysis was carried out by Trust laboratories using in-house methodology, developed in line with FDA and EMA guidelines.^{17,18} The intra- and inter-assay precision across the reporting range of 0.2 to 8 mg/L were below 5.2%.

Samples were stored at -20° C prior to analysis. Posaconazole was extracted from serum via protein precipitation with acetonitrile. Posaconazole-d4 was used as the corresponding internal standard. Chromatography mobile phases were 10 nM ammonium acetate in ultrapure water (UP-H₂O) and 10 nM ammonium acetate in methanol and posaconazole was detected using atmospheric chemical ionization in positive ion mode.

Population pharmacokinetic model building

Parameters for a one-compartment model assuming first-order absorption and elimination were estimated using non-linear mixed-effects modelling with NONMEM (Version 7.4; ICON Development Solutions, Ellicott City, MD, USA),¹⁹ using the FOCE with interaction algorithm. IIV was tested for clearance assuming a log-normal distribution. The residual error included additive and proportional terms.

Allometric size scaling of clearance and volume terms were added a priori¹³ in the following form:

$$p_i = \theta_p \left(\frac{c_i}{\overline{c}}\right)^{\theta_c}$$

where p_i is the individual parameter of interest, c_i is the individual value of the covariate and \overline{c} is the typical value of the covariate in the population. In the fixed allometric weight scaling, c_i was the individual body weight, \overline{c} was set to 70 kg and θ_c was 0.75 for clearance and 1 for central volume.

For nested models, significant model improvement through additional parameters was evaluated with the likelihood ratio test, the difference in $-2 \log$ -likelihood (objective function value, OFV, in NONMEM) of the models being asymptotically χ_1^2 distributed. Covariates were added if the likelihood ratio test indicated a significant improvement in fit at the level of P < 0.01. A stepwise covariate model (SCM)-building exercise with Perl Speaks NONMEM (PsN)²⁰ considered forward inclusion with a *P* value of ≤ 0.05 and backwards elimination with a *P* value of ≤ 0.01 . The following covariates were tested on clearance: liver function abnormality (if any of ALT, alkaline phosphatase, AST, GGT or bilirubin were outside of the normal reference ranges); and potential interacting medicines that were identified in patients included in the dataset: Orkambi[®] (lumacaftor/ivacaftor), rifampicin, rifabutin, clarithromycin, histamine H₂-receptor antagonists and proton pump inhibitors.

Dose simulations

Dosing simulations were performed in R.²¹ The multivariate mean, variance and covariance between age and weight were estimated from the patient demographics and then simulations performed to generate 1000 age-weight pairs for each simulation scenario. Using

both this dataset and the final model, simulations were generated to assess the probability of attaining a trough target concentration of $1 \text{ mg/L}^{7,10}$ and an alternative target of an AUC of 30 mg·h/L.^{22,23}

Model evaluation

Final model evaluation consisted of plotting predictions versus observations, and conditional weighted residuals versus time and predictions. A visual predictive check with 1000 samples was performed to check predictive performance, and a non-parametric bootstrap (1000 samples) was conducted to evaluate parameter robustness.

Results

Study population

The initial dataset included 109 posaconazole serum concentrations from 41 children, obtained during 65 treatment courses. From this, nine serum concentrations were excluded (three where we were unable to ascertain the timing of the sample; three due to incomplete dose history, including uncertainty on whether doses had been administered; two below the limit of quantification in a patient known to have been non-adherent; and one where it was identified that the patient had missed a number of the preceding doses). The final dataset therefore included 100 levels from 37 children, with a median age of 14 years (range 7–17). There were 29 samples taken from children aged 7–11 years, with median weight of 31.5 kg (range 25–58). The median posaconazole serum concentration for this age group was 3.08 mg/L (range 0.2–8.91) from a median dose of 300 mg (range 100–300). Of 71 samples taken from children aged 12-17 years, with median weight of 50 kg (range 34.7–82.8), the median posaconazole serum concentration was 2.07 mg/L (range 0.2–4.96) from a median dose of 300 mg (range 200-600). There were 47 samples from outpatients where the time of the last dose was presumed to be at 8 am on the day of the sample. Study population characteristics are detailed in Table 1.

Population pharmacokinetics

The pharmacokinetics of posaconazole in children with CF were adequately described by a one-compartment model with IIV on clearance and additive and proportional residual error. Goodnessof-fit plots, and a visual predictive check indicating an acceptable model fit, are given in Figure 1. Parameter estimates and bootstrap results from the final model are included in Table 2. Of the 1000 bootstrap samples, 5 were omitted due to minimization terminating and 107 with final estimate zero gradients. No covariates were selected during the stepwise covariate modelling. The absorption rate constant (K_{a}) was $0.16 h^{-1}$, apparent clearance (CL/F) was 8.43 L/h and apparent volume (V/F) was 186 L, standardized to a 70 kg individual. The corresponding estimates for the reduced dataset (including only those samples with known dose times) were: $0.13 h^{-1}$, 7.93 L/h and 108 L for K_a, CL/F and V/F, respectively, and hence there was no apparent influence of assuming dose time so all samples were included.

 Table 1. Demographics of patients included in pharmacokinetic analysis

| Variable | Included patients (<i>n</i> = 37) |
|--|------------------------------------|
| TDM samples, <i>n</i> | 100 |
| Samples/patient, <i>n</i> , median (range) | 2 (1-9) |
| Organism/condition being treated at the time | 53/17/26/4 |
| of the sample: ABPA/Aspergillus/ | |
| Scedosporium/Exophiala | |
| Age, years (range) | 14 (7–17) |
| Weight, kg (range) | 45.55 (25-82.8) |
| Age 6–11 years | 31.5 (25–58) |
| Age 12–17 years | 50 (34.7-82.8) |
| Sex, male/female, <i>n</i> | 13/24 |
| Baseline (prior to each course) FEV ₁ predicted, % (range) | 76 (42–105) |
| Age 6–11 years | 87 (47–105) |
| Age 12–17 years | 72 (42–97) |
| Baseline (prior to each course) BMI, kg/m ² (range) | 18.55 (14.55–25.38) |
| Age 6–11 years | 16.61 (14.55–25.38) |
| Age 12–17 years | 19.41 (16.06-23.66) |
| Dose, mg per day (range) | 300 (100–600) |
| Age 6–11 years | 300 (100-300) |
| Age 12–17 years | 300 (200–600) |
| Dose, mg/kg per day (range) | 6.48 (3.45-14.05) |
| Age 6–11 years | 8.33 (3.45-12) |
| Age 12–17 years | 6.06 (4.11-14.05) |
| Concentration, mg/L (range) | 2.41 (0.2-8.91) |
| Age 6–11 years | 3.08 (0.2-8.91) |
| Age 12–17 years | 2.07 (0.2-4.96) |
| Sample time after dose, h (range) | 1-31 |
| Samples when patient taking PPI or H_2 -receptor antagonist, % | 76 |
| Samples when patient taking clarithromycin, % | 2 |
| Samples when patient taking rifampicin, % | 2 |
| Samples when patient taking rifabutin, % | 11 |
| Samples when patient taking Orkambi [®] , % | 3 |
| Samples where liver function tests normal, % | 40 |

ABPA, allergic bronchopulmonary aspergillosis; Aspergillus, A. fumigatus; Scedosporium, S. apiospermum and L. prolificans; Exophiala, E. dermatitidis; H₂-receptor antagonist, histamine H₂-receptor antagonist; PPI, proton pump inhibitor.

Dose simulations

The simulations using the derived pharmacokinetic parameters from the model demonstrated that by using the tablet formulation the probability of attaining a trough target concentration of 1 mg/L with the standard adult starting dose of 300 mg every 12 h for two doses (loading) followed by 300 mg once daily (OD) was 77% in children aged 9–11 years and 83% in children aged 6–8 years; using a starting dose of 400 mg every 12 h for two doses followed by 400 mg OD, the probability of attaining a trough target of 1 mg/L was 88% in adolescents aged 12–14 years and 86% in adolescents aged 15–17 years. However, simulations using an alternative AUC target showed that with these dosing regimens

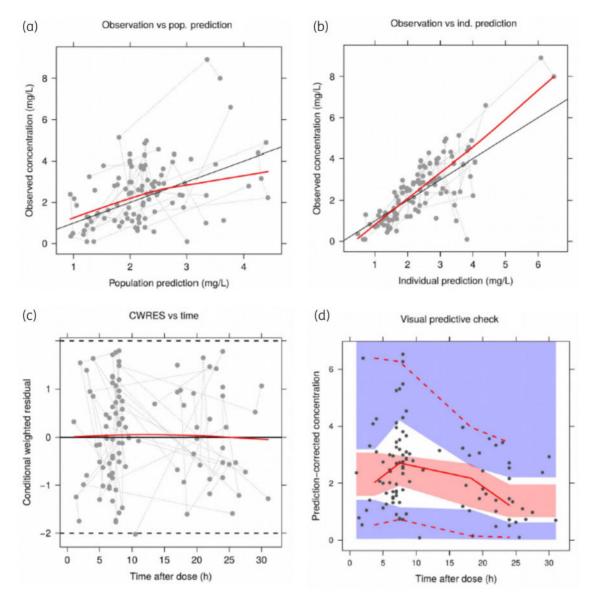


Figure 1. Goodness-of-fit plots for the final model. (a) Population (pop.) predictions versus observations; (b) individual (ind.) predictions versus observations; (c) conditional weighted residuals (CWRES) versus time after dose (time); and (d) visual predictive check showing model-simulated 95% CIs for the simulated 2.5th, 50th and 97.5th percentiles (shaded areas) compared with the observed percentiles (lines). This figure appears in colour in the online version of *JAC* and in black and white in the print version of *JAC*.

| Parameter | Estimate (%RSE) | IIV %CV (%RSE) | Bootstrap median (95% CI) | Bootstrap IIV %CV (95% CI) |
|----------------------------|-----------------|----------------|---------------------------|----------------------------|
| CL/F (L/h) ^a | 8.43 (8.7) | 38 (36.8) | 8.38 (7.11-10.07) | 37 (20–52) |
| V/F (L) ^a | 186 (55.4) | _ | 197 (107.96–413.55) | _ |
| K_{a} (h ⁻¹) | 0.16 (55.5) | _ | 0.15 (0.05-2.09) | _ |
| Proportional error (%CV) | 36 (23.2) | _ | 35 (24–44) | _ |
| Additive error (mg/L) | 0.15 (48.4) | _ | 0.15 (0.02–0.42) | _ |

Table 2. Parameter estimates from the final model

%CV, coefficient of variation; RSE, relative standard error.

The NONMEM model code is given in the Supplementary data available at JAC Online.

^aCL/F and V/F allometrically scaled.

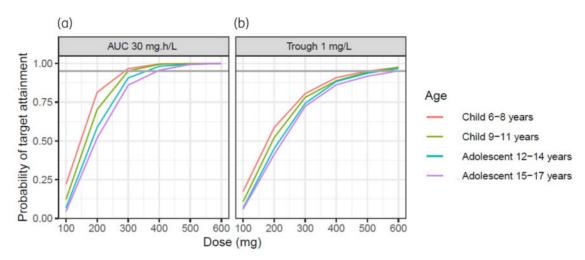


Figure 2. Simulated probability of AUC being 30 mg·h/L (a) or trough concentration being 1 mg/L (b) for OD dosing split by age group. The grey horizontal line represents a 90% PTA. This figure appears in colour in the online version of *JAC* and in black and white in the print version of *JAC*.

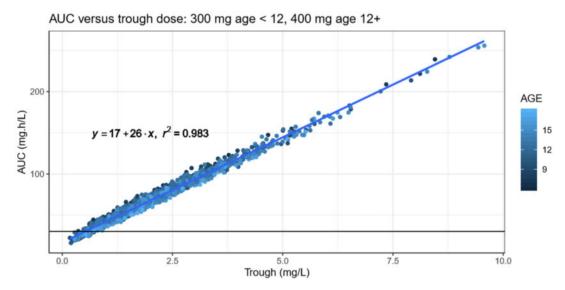


Figure 3. Simulation of AUC versus trough concentration using a dose of 300 mg every 12 h for two doses then 300 mg OD in children <12 years old and 400 mg every 12 h for two doses then 400 mg OD in children aged 12 years and above. Black horizontal line represents an AUC of 30 mg·h/L. This figure appears in colour in the online version of *JAC* and in black and white in the print version of *JAC*.

there was a greater probability (more than 90% in both age groups) of attaining an AUC of 30 mg·h/L. Plots of simulated target attainment versus dose are shown in Figure 2. AUC and trough concentration were highly correlated ($r^2 = 0.98$) (see Figure 3) and simulations showed that trough concentrations of >0.75 mg/L would exceed an AUC of 30 mg·h/L in 90% of patients.

Discussion

Here we report the first population pharmacokinetic analysis of posaconazole gastroresistant tablets in 37 children with CF, 14 of whom were less than 12 years old, who received posaconazole to treat CF-related fungal lung disease. A one-compartment model adequately described the data (Figure 1) and no significant covariates were found. We have made simulations from the model that

have suggested an initial starting dose of 300 mg every 12 h for two doses then 300 mg OD in children aged 6–11 years and 400 mg every 12 h for two doses then 400 mg OD in children aged 12–17 years.

Posaconazole has been prospectively studied in 14 children with CF. The study⁸ reported that posaconazole was well tolerated and improvements in lung function were observed. Posaconazole plasma trough concentrations of >1 mg/L were achieved in 100% of children taking the tablet formulation, all of whom were aged 12 years and older and receiving a dose of 300 mg OD. A case study of a 13-year-old male patient with CF treated with posaconazole tablets for *A. fumigatus* infection also demonstrated that plasma trough concentrations of >1 mg/L were attained with a dose of 300 mg, along with improved lung function and eradication of the organism.²⁴ However, despite these positive experiences thus far,

there have been no published pharmacokinetic data to support dosing specific to children with CF, including younger children, and especially in the context of altered pharmacokinetics in this population.

The population pharmacokinetics of posaconazole in children with CF from our model are broadly in line with the parameters ascertained by Boonsathorn et al.¹⁵ in children without CF who estimated CL/F to be 14.95 L/h and V/F to be 201.7 L. Although the CIs of the CL/F value derived from our model (7.11-10.07) are within the stated 95% CIs for CL/F derived by Boonsathorn (6.3-34.1), the lower value in our model may be explained by the difference in formulations used by the patients in each of the models, with lower bioavailability of the suspension used in the model in children without CF leading to a higher estimate of CL/F.²⁵ However, our value is similar to the CL/F of the tablet formulation (7.3 L/h) derived from the population pharmacokinetic model described by Petitcollin et al.²⁶ in adult haematological patients. We had limited data in the absorption phase and our absorption rate constant was somewhat lower than that reported by others (0.16 versus $0.588 \, h^{-1}$).²⁶ Fixing K_a to 0.588 h⁻¹ had limited impact on model fit and other parameter estimates ($\Delta OFV + 2.64$ points when fixing K_a), indicating inferences on AUC or trough concentration would not be affected.

Posaconazole is metabolized via UDP glucuronidation (UDPglucuronosyltransferase, UGT) and is a substrate for p-glycoprotein (P-qp) efflux.²⁷ Children with CF often require antibiotics to treat pulmonary infections caused by Staphylococcus aureus and non-tuberculous mycobacteria including Mycobacterium avium complex. Recommended treatment for these includes rifampicin and rifabutin (inducers of these pathways) and clarithromycin (an inhibitor), which respectively have the potential to either reduce or increase serum concentrations of posaconazole if given concomitantly. Many patients with CF are now treated with CFTR modulators, which target the basic CFTR channel defect. One of these, Orkambi®, a combination of ivacaftor and lumacaftor, can reduce posaconazole serum concentrations by induction of UGT via the lumacaftor component, and concomitant use with posaconazole is not recommended.²⁷ Posaconazole can also increase exposure of the ivacaftor component of Orkambi[®], though due to the induction effect of the lumacaftor component of Orkambi® on CYP3A enzymes at steady state the overall exposure of ivacaftor with concomitant posaconazole does not exceed that achieved in patients receiving ivacaftor monotherapy.²⁸ Therefore it is not necessary to adjust the dose of Orkambi[®] if initiating posaconazole in patients already commenced and stabilized on this medicine, as was the case in our cohort. We tested these interacting drugs as covariates in the analysis as it was hypothesized that these may be associated with changes in posaconazole CL/ F. However, none were statistically significant. It is likely that the small percentage of samples included within the analysis was insufficient to detect any effects. We also tested the effect of concurrent use of proton pump inhibitors and histamine H₂receptor antagonists as a covariate, and no effect on the CL/F of the tablet formulation was seen, confirming the findings already seen in healthy subjects.²⁹ Similarly, liver function abnormalities were not found to have an effect on CL/F when tested as a covariate within the analysis. This concurs with findings by Petitcollin et al.²⁶ who suggested that although baseline ALT

3252

levels were negatively correlated with posaconazole clearance, the effect was small, with a 5-fold increase in ALT levels resulting in only a 25% reduction in clearance.

Dosing simulations were carried out against a target trough concentration of 1 mg/L, as per established guidelines,^{7,10} and an alternative target of an AUC of 30 mg·h/L. The latter target was chosen from the EUCAST Rationale Document for Clinical Breakpoints for Aspergillus, which states this value as it is associated with a 75% response rate in patients receiving posaconazole as salvage therapy for invasive aspergillosis and is equivalent to a C_{ava} of 1.25 mg/L.^{22,30} Although neither of these targets is well validated in larger cohorts including those with CF, Gastine et al.²³ confirmed that in a rabbit model of A. fumigatus invasive pulmonary aspergillosis, an AUC greater than 30 mg h/L was associated with adequate resolution of the galactomannan index—a surrogate marker of antifungal response against Aspergillus species. For azole antifungals, the pharmacodynamics and thus response to treatment are ultimately related to the MIC of an organism, and specifically the AUC_{0-24}/MIC ratio, which is the pharmacodynamic index for posaconazole consistently shown to be most closely correlated with efficacy. Gastine et al.²³ utilized an A. fumigatus strain with an MIC of 0.25 mg/L in line with the EUCAST epidemiological cutoff value for this organism, resulting in an AUC₀₋₂₄/MIC ratio of 120. Recommendations for the AUC_{0-24}/MIC target range between 100 and 200 for treatment of A.fumigatus.^{22,27,31}

Our simulations showed that an AUC of 30 mg h/L was easier to attain with 'standard' dosing, that is doses in a similar range to those licensed in adults, than a trough concentration of 1 mg/L. With the number of samples required to determine AUC accurately being impractical in clinical use, our simulations showed that with AUC and trough concentrations highly correlated, trough concentrations of >0.75 ma/L (lower than the recommended 1 ma/L target trough concentration for treatment of susceptible fungal diseases) would exceed an AUC of 30 mg·h/L in 90% of patients. Future work is required to evaluate whether a trough concentration of 0.75 mg/L, as a surrogate for AUC, is sufficient to optimize outcome in this cohort for the treatment of Asperaillus-related lung disease, and to evaluate response for less susceptible strains of A. fumigatus where we would expect to increase the AUC attained to maintain an AUC₀₋₂₄/MIC ratio of >100. Our study also included children with lung disease associated with Scedosporium species (Lomentospora prolificans and Scedosporium apiospermum) and E. dermatitidis. Unlike Aspergillus, pharmacodynamic targets have not been elucidated for these organisms and there is a need to identify these and produce recommendations on the optimal use of posaconazole in children with CF with these organisms.

This study is limited by its retrospective nature and ideally a prospective study with optimal sample timing should be carried out. However, given the difficulty in recruiting to studies requiring multiple blood samples in children, it is likely that our study included a larger cohort than might otherwise have been possible. Our simulations used recommended, though unvalidated, targets in our cohort of patients. There is a need to refine and validate these according to the organism, site of action and disease state, and most importantly assess the clinical outcome in children with CF.

Conclusions

Our study confirms that the population pharmacokinetics of posaconazole in children with CF are in line with data published in other paediatric populations. Our dose simulations suggest that a starting dose of 300 mg every 12 h for two doses (loading) then 300 mg OD in children aged 6–11 years and 400 mg every 12 h for two doses then 400 mg OD in adolescents aged 12–17 years will ensure a 90% probability of attaining an AUC of 30 mg·h/L, a target which is known in other patient populations to be associated with a favourable outcome when treating *Aspergillus* infections.^{22,30} If aiming for trough concentration targets of >1 mg/L, 12%–23% of patients are predicted to require a higher dose.

There is a need to establish pharmacodynamic targets for other fungi commonly found in children with CF and to evaluate clinical outcomes using both these targets, and those already recommended for *A. fumigatus*, including whether a trough concentration of 0.75 mg/L is sufficient to optimize outcome for the treatment of *Aspergillus*-related lung disease.

Funding

This study was supported by the National Institute of Health Research (NIHR) through a Health Education England/NIHR Pre-doctoral Clinical Academic Fellowship (S.B., NIHR300407), a Senior Investigator Award (J.C.D., NIHR201402) and the Imperial Biomedical Research Centre. J.F.S. and S.G. were supported at an institution level by the NIHR Great Ormond Street Biomedical Research Centre and J.F.S. received a UK Medical Research Council fellowship (MR/M008665/1).

Transparency declarations

None to declare.

Supplementary data

The NONMEM model code is available as Supplementary data at JAC Online.

References

1 Renner S, Nachbaur E, Jaksch P *et al.* Update on respiratory fungal infections in cystic fibrosis lung disease and after lung transplantation. *J Fungi* (*Basel*) 2020; **6**: 381.

2 Amin R, Dupuis A, Aaron SD *et al.* The effect of chronic infection with *Aspergillus fumigatus* on lung function and hospitalization in patients with cystic fibrosis. *Chest* 2010; **137**: 171–6.

3 Saunders RV, Modha DE, Claydon A *et al.* Chronic Aspergillus fumigatus colonization of the pediatric cystic fibrosis airway is common and may be associated with a more rapid decline in lung function. *Med Mycol* 2016; **54**: 537–43.

4 Cystic Fibrosis Trust. UK Cystic Fibrosis Registry: Annual Data Report 2019. 2020. https://www.cysticfibrosis.org.uk/sites/default/files/2020-12/2019%20 Registry%20Annual%20Data%20report_Sep%202020.pdf.

5 Denning DW, Cadranel J, Beigelman-Aubry C *et al*. Chronic pulmonary aspergillosis: rationale and clinical guidelines for diagnosis and management. *Eur Respir J* 2016; **47**: 45–68.

6 Tortorano AM, Richardson M, Roilides E *et al.* ESCMID and ECMM joint guidelines on diagnosis and management of hyalohyphomycosis: *Fusarium* spp., *Scedosporium* spp. and others. *Clin Microbiol Infect* 2014; **20** Suppl 3: 27-46.

7 Ullmann AJ, Aguado JM, Arikan-Akdagli S *et al.* Diagnosis and management of *Aspergillus* diseases: executive summary of the 2017 ESCMID-ECMM-ERS quideline. *Clin Microbiol Infect* 2018; **24** Suppl 1: e1–38.

8 Patel D, Popple S, Claydon A *et al.* Posaconazole therapy in children with cystic fibrosis and *Aspergillus*-related lung disease. *Med Mycol* 2020; **58**: 11–21.

9 Bentley S, Gupta IM, Balfour-Lynn IM. Subtherapeutic itraconazole and voriconazole levels in children with cystic fibrosis. *J Cyst Fibros* 2013; **12**: 418–9.

10 Ashbee RH, Barnes RA, Johnson EM *et al.* Therapeutic drug monitoring (TDM) of antifungal agents: guidelines from the British Society for Medical Mycology. *J Antimicrob Chemother* 2014; **69**: 1162–76.

11 De Sutter P, Gasthuys E, Van Braeckel E *et al.* Pharmacokinetics in patients with cystic fibrosis: a systematic review of data published between 1999 and 2019. *Clin Pharmacokinet* 2020; **59**: 1551–73.

12 Conway SP, Etherington C, Peckham DG *et al.* Pharmacokinetics and safety of itraconazole in patients with cystic fibrosis. *J Antimicrob Chemother* 2004; **53**: 841–7.

13 Jeong W, Snell GI, Levvey BJ *et al.* Single-centre study of therapeutic drug monitoring of posaconazole in lung transplant recipients: factors affecting trough plasma concentrations. *J Antimicrob Chemother* 2018; **73**: 748–56.

14 Abdolrasouli A, Scourfield A, Rhodes J *et al*. High prevalence of triazole resistance in clinical *Aspergillus fumigatus* isolates in a specialist cardiothoracic centre. *Int J Antimicrob Agents* 2018; **52**: 637–42.

15 Boonsathorn S, Cheng I, Kloprogge F *et al.* Clinical pharmacokinetics and dose recommendations for posaconazole in infants and children. *Clin Pharmacokinet* 2019; **58**: 53–61.

16 Durani U, Tosh PK, Barreto JN *et al.* Retrospective comparison of posaconazole levels in patients taking the delayed-release tablet versus the oral suspension. *Antimicrob Agents Chemother* 2015; **59**: 4914–8.

17 US FDA. Guidance for Industry: Bioanalytical Method Validation. 2018. https://www.fda.gov/media/70858/download.

18 EMA. Guideline on Bioanalytical Method Validation. Committee for Medicinal Products for Human Use. EMEA/CHMP/EWP/192217/2009. 2011. http://www.ema.europa.eu/docs/en_GB/document_library/Scientific_guide line/2011/08/WC500109686.pdf.

19 Beal S, Sheiner LB, Boeckmann A *et al. NONMEM User's Guides (1989–2013).* Icon Development Solutions, 2013.

20 Lindbom L, Pihlgren P, Jonsson EN. PsN-Toolkit—a collection of computer intensive statistical methods for non-linear mixed effect modeling using NONMEM. *Comput Methods Programs Biomed* 2005; **79**: 241–57.

21 R Core Team. R: A language and environment for statistical computing. R Foundation for Statistical Computing. 2020. https://www.R-project.org/.

22 EUCAST. Rationale document for EUCAST clinical breakpoints: posaconazole, version 3.0. 2020. https://www.eucast.org/fileadmin/src/media/PDFs/ EUCAST_files/Rationale_documents/Posaconazole_RD_v3.0_final_final_18_ 02.pdf.

23 Gastine S, Hope W, Hempel G *et al.* Pharmacodynamics of posaconazole in experimental invasive pulmonary aspergillosis: utility of serum galactomannan as a dynamic endpoint of antifungal efficacy. *Antimicrob Agents Chemother* 2021; **65**: e01574-20.

24 Shearin S, Bell T. Treatment of *Aspergillus fumigatus* infection with posaconazole delayed-release tablets. *Am J Health Syst Pharm* 2018; **75**: 958–61.

25 Chen L, Krekels EHJ, Verweij PE *et al.* Pharmacokinetics and pharmacodynamics of posaconazole. *Drugs* 2020; **80**: 671–95.

26 Petitcollin A, Boglione-Kerrien C, Tron C *et al.* Population pharmacokinetics of posaconazole tablets and Monte Carlo simulations to determine whether all patients should receive the same dose. *Antimicrob Agents Chemother* 2017; **61**: e01166-17.

27 Summary of Product Characteristics – Posaconazole (Noxafil). Merck Sharp & Dohme (UK) Ltd. https://www.medicines.org.uk/emc/product/5388/ smpc.

28 Summary of Product Characteristics – Orkambi. Vertex Pharmaceuticals (Europe) Ltd. https://www.medicines.org.uk/emc/product/8952/smpc.

29 Kraft WK, Chang PS, van Iersel ML *et al.* Posaconazole tablet pharmacokinetics: lack of effect of concomitant medications altering gastric pH and gastric motility in healthy subjects. *Antimicrob Agents Chemother* 2014; **58**: 4020–5. **30** Walsh TJ, Raad I, Patterson TF *et al.* Treatment of invasive aspergillosis with posaconazole in patients who are refractory to or intolerant of conventional therapy: an externally controlled trial. *Clin Infect Dis* 2007; **44**: 2–12.

31 Lewis RE, Albert ND, Kontoyiannis DP. Comparative pharmacodynamics of posaconazole in neutropenic murine models of invasive pulmonary aspergillosis and mucormycosis. *Antimicrob Agents Chemother* 2014; **58**: 6767–72.