






Genome Sequence of Genotype 1A Hepatovirus A Isolated from Plasma from a Haitian Child

M. Mahbulul Alam,^{a,b}  Maha A. Elbadry,^{a,b*} Julia C. Loeb,^{a,b} Caroline J. Stephenson,^{a,b} Rigan Louis,^{a,c} Carla Mavian,^{a,d} Remi Charrel,^{a,e} Sabita Rezwana Rahman,^f  J. Glenn Morris, Jr.,^{a,g}  John A. Lednicky^{a,b}

^aEmerging Pathogens Institute, University of Florida, Gainesville, Florida, USA

^bDepartment of Environmental and Global Health, College of Public Health and Health Professions, University of Florida, Gainesville, Florida, USA

^cState University of Haiti Faculty of Medicine and Pharmacy, Port-au-Prince, Haiti

^dDepartment of Pathology, Immunology, and Laboratory Medicine, College of Medicine, University of Florida, Gainesville, Florida, USA

^eUnite des Virus Emergents, Aix Marseille Université, IRD 190, INSERM U1207, Marseille, France

^fUniversity of Dhaka, Dhaka, Bangladesh

^gDepartment of Medicine, College of Medicine, University of Florida, Gainesville, Florida, USA

ABSTRACT Genotype 1A hepatovirus A was identified by quantitative reverse transcription-PCR and isolated from plasma from a Haitian child with acute undifferentiated febrile illness and malaise. The strain was most closely related to Brazilian strains, consistent with recognized patterns of virus movement in the Caribbean region.

Hepatovirus A (HAV) is a single-stranded nonenveloped RNA virus belonging to the genus *Hepatovirus* of the family *Picornaviridae*. In low- and middle-income countries, it has been estimated that 90% of children are infected with the virus before the age of 10 years (1). As reported previously, our group monitored the occurrence of acute undifferentiated febrile illness in a cohort of children in Gressier, Haiti, from 2014 to 2019 (2–4). Studies were approved by the University of Florida and Haitian national institutional review boards, and signed informed consent forms from parents and assent from students were obtained before enrollment in the study.

HAV genomic RNA was detected by quantitative reverse transcription-PCR (qRT-PCR) in 2 (0.3%) of 677 plasma samples collected from febrile children as part of this study, by using the primer system described by Jothikumar et al. (5) and a modified HAV probe, namely, 5'-6-carboxyfluorescein (FAM)-CTTARGCTARTACTTCTATGAAGAGATGC-black hole quencher 1 (BHQ1)-3', in which two R degeneracies (underlined) were inserted to replace a G at position 417 and an A at position 422. Primers and probe were combined and freeze-dried in a single glass vial (6). The quantification cycle (C_q) values for sample 15-1-1251, which was collected in January 2015, and sample 18-1-2097, which was collected in November 2016, were 33.37 and 40.27, respectively. Attempts were made to isolate the virus in MRC-5 cells (7) and were successful only for sample 18-1-2097. Sample 18-1-2097 was from a 6-year-old child who presented with a temperature of 38.1°C and complaints of malaise and mouth sores; no jaundice was noted.

The genomic sequence of sample 18-1-2097 was obtained from virus RNA that had been extracted from plasma (8) and Sanger sequenced with a gene-walking approach (9, 10) using nonoverlapping primers. Briefly, viral RNA was extracted from 140 μ l of plasma using a QIAamp viral RNA extraction kit (Qiagen, Valencia, CA), RT was performed using the AccuScript high fidelity first strand cDNA kit (Agilent Technologies, Santa Clara, CA) in the presence of SUPERase-In RNase inhibitor (Ambion, Austin, TX), and PCR was sequentially performed using Q5 high fidelity DNA polymerase (New England Biolabs, Inc., Ipswich, MA) and the primers identified in Table 1. To determine the sequence of the 5' end of the virus genome, 20 μ L of

Editor Kenneth M. Stedman, Portland State University

Copyright © 2022 Alam et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to John A. Lednicky, jllednicky@phhp.ufl.edu, or J. Glenn Morris, Jr., jgmorris@epi.ufl.edu.

*Present address: Maha A. Elbadry, Procter and Gamble, Cincinnati, Ohio, USA.

The authors declare no conflict of interest.

Received 20 May 2022

Accepted 24 July 2022

Published 11 August 2022

TABLE 1 Primers for sequencing HAV

Primer	Sequence (5' to 3')	Nucleotide positions in GenBank accession no. OK625565.1	Reference	Nucleotide positions in GenBank accession no. MG049743.1	Amplicon size (bp)
5'-RACE-R	AACAACCTACCAATATCCGC	480–461	2		
HAV for	GGTAGGCTACGGGTGAAAC	392–410	2		88
HAV rev	AACAACCTACCAATATCCGC	480–461	2		
HAV For4	TACCTCACCGCCGTTTGCTAGGC	64–87	1		417
HAV rev	AACAACCTACCAATATCCGC	480–461	2		
HepA for1	CTTAAGCTATTACTTCTATGAAGAGATGC	413–441	This work		648
HepA rev1	CAGTTCCACCACATCCAATTTGCAACTTC	1060–1031	This work	1030–1001	
HepA for 2	GGCTCACTACACATGCTCTCTTTCATG	1005–1031	This work		853
HepA rev 2	CTACCTGAATGATATTTGGTTGGAAAAACC	1857–1828	This work	1827–1798	
HepA for 3	GGCTTCTATCTGTCAAATGTTTTG	1771–1794	This work		810
HepA rev 3	GATGGTAAACCATGCGGAGGATTTGAAG	2580–2553	This work	2550–2523	
HepA for 4	GGGAAGGTCTCACTTTTTGTG	2473–2493	This work		847
HepA rev 4	CCTAGTATCAGCAGTTACTCTCTCC	3319–3294	This work	3289–3264	
HepA for 5	GTGCTCCACCTCCTAGGAAAATGAAG	3200–3226	This work		952
HepA rev 5	GCTGGTATCTTTAAGAATGTTAAG	4151–4127	This work	4121–4097	
HepA for 6	GGTTATATAACAAATGAAGGAT	4068–4090	This work		816
HepA rev 6	CACAAGACATGTCCTTGATTGCATC	4883–4859	This work	4853–4829	
HepA for 7	GATCGTAGACTTCATTTAAGGTTGAAG	4763–4790	This work		843
HepA rev 7	CTTGATAAAAATGTTAGTAATATCTCT	5605–5579	This work	5575–5549	
HepA for 8	GGGATTTCAAGATGTTGTTCTAATG	5530–5554	This work		844
HepA rev 8	GGCCAGTCGTGGATGAACCTAACAG	6373–6347	This work	6343–6317	
HepA for 9	GGCTCCAGGCATTGATGCTATTAATA	6241–6266	This work		773
HepA rev 9	GAACATCTCTGGAAAAGACTAT	7013–6992	This work	6983–6962	
HepA for 10	GAGGATCTTTGTTACGGAGATG	6961–6983	This work		542
HepA rev 10	TTTTTTTTTTTTTTTTTTTTTTT	7502–7477	This work		

purified RNA was treated with DNase- and RNase-free proteinase K (New England Biolabs) to remove the 5' VPg (11–13), followed by 5' rapid amplification of cDNA ends (RACE) using a FirstChoice RLM-RACE kit (Thermo Fischer Scientific) following the manufacturer's instructions. Quality scores for the Sanger sequences ranged from 52 to 67 (11–13). Excluding the poly(A) tail, the virus genome length is 7,477 ribonucleotides (rnt) (A, 2,181 rnt; U, 2,449 rnt; G, 1,638 rnt; C, 1,209 rnt), with a G+C content of 38.1%.

The maximum likelihood phylogeny was constructed by using IQ-TREE (14–19) with all available complete genomes for HAV strains from humans available in GenBank; genotyping followed the methods described by Ramachandran et al. (20). The phylogenetic analysis shows that the genome sequence of the Haitian HAV isolate belongs to a well-supported monophyletic clade in genotype 1A that includes HAV genomes from the Americas, including Brazil, Mexico, and the United States, between 2009 and 2018 (Fig. 1). In particular, the genome sequence of the Haitian isolate clusters near a Brazilian HAV genome from 2017; the phylogenetic proximity and the short branches separating the Haitian genome sequence from the Brazilian sequence, which was from an HAV case that occurred 1 year after the collection of the virus in Haiti, are consistent with a recent common source or exchange of viruses between these two countries.

Data availability. The virus was designated hepatovirus A/0789/Haiti/2016, and its sequence was deposited in GenBank under accession number [OK625565.1](#).

ACKNOWLEDGMENTS

This work was supported in part by the European Commission (European Virus Archive Global [EVA GLOBAL] project grant 871029 of the Horizon 2020 Research and Innovation Programme awarded to J.A.L. and R.C.) and by the National Institute of Allergy and Infectious Diseases (NIH grant R01 AI123657S awarded to J.G.M.). The material was provided by the European Virus Archive Marseille (EVAM) under the label technological platforms of Aix-Marseille Université.

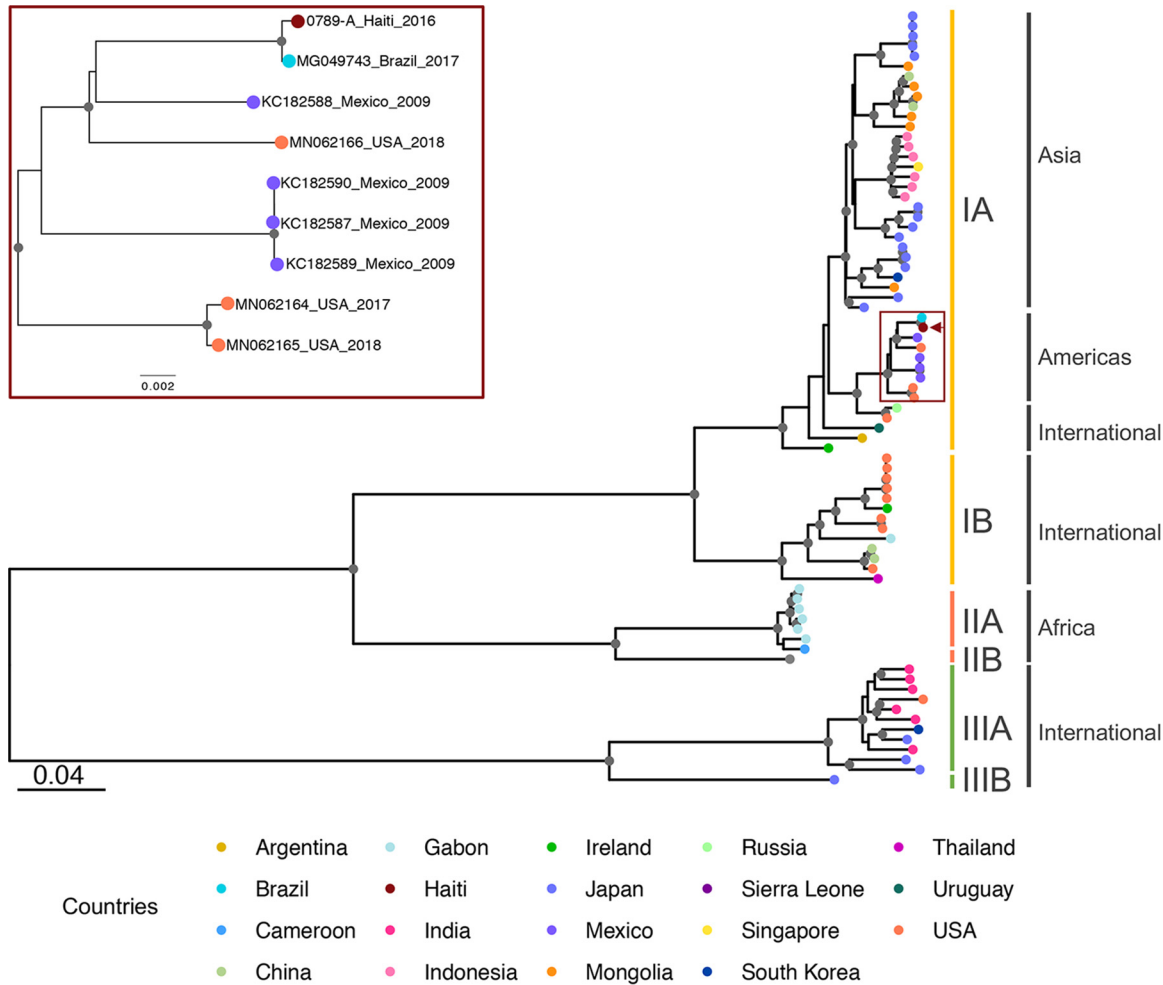


FIG 1 Phylogenetic inference of the HAV strains from a human source. The maximum likelihood phylogenetic tree of HAV strains from humans is based on the complete genome and was inferred using IQ-TREE. Gray circles at internal nodes represent >90% bootstrap support. Colored circles at the tips show the collection locations based on the legend at the bottom. The red rectangle shows the magnification of the subtree based on the American subclade containing the isolates from Haiti, Brazil, Mexico, and the United States with GenBank accession numbers (rectangle in the full phylogenetic tree). The Haitian isolate is indicated with a red arrow. Genotype classification is shown to the right of the tree.

We thank Laurence Thirion and Gregory Molle for excellent technical assistance.

REFERENCES

- World Health Organization. 27 July 2021. Hepatitis A. <https://www.who.int/news-room/fact-sheets/detail/hepatitis-a>. Accessed 4 November 2021.
- Ball JD, Elbadry MA, Telisma T, White SK, Chavannes S, Anilis MG, Prospero M, Cummings DAT, Lednicky J, Morris JG, Beau de Rochars VM. 2019. Clinical and epidemiological patterns of *Chikungunya virus* infection and coincident arboviral disease in a school cohort in Haiti, 2014/2015. *Clin Infect Dis* 68:919–926. <https://doi.org/10.1093/cid/ciy582>.
- Lednicky JA, Tagliamonte MS, White SK, Elbadry MA, Alam MM, Stephenson CJ, Bonny TA, Loeb JC, Telisma T, Chavannes S, Ostrov DA, Mavian C, Beau De Rochars VM, Salemi M, Morris JG. 2021. Independent infections of porcine deltacoronavirus among Haitian children. *Nature* 600:133–137. <https://doi.org/10.1038/s41586-021-04111-z>.
- Elbadry MA, Carvalho RD, Blohm G, Stephenson C, Loeb J, White S, Telisma T, Chavannes S, Beau De Rochars VM, Salemi M, Morris JG, Lednicky JA. 2021. Orthobunyaviruses in the Caribbean: Malao and Oropouche virus infections in school children in Haiti in 2014. *PLoS Negl Trop Dis* 15:e0009494. <https://doi.org/10.1371/journal.pntd.0009494>.
- Jothikumar T, Cromeans L, Sobsey D, Robertson BH. 2005. Development and evaluation of a broadly reactive Taqman assay for rapid detection of hepatitis A virus. *Appl Environ Microbiol* 71:3359–3363. <https://doi.org/10.1128/AEM.71.6.3359-3363.2005>.
- Thirion L, Dubot-Peres A, Pezzi L, Corcostegui I, Touinssi M, de Lamballerie X, Charrel RN. 2020. Lyophilized matrix containing ready-to-use primers and probe solution for standardization of real-time PCR and RT-qPCR diagnostics in virology. *Viruses* 12:159. <https://doi.org/10.3390/v12020159>.
- De Chastonay J, Siegl G. 1987. Replicative events in hepatitis A virus-infected MRC-5 cells. *Virology* 157:268–275. [https://doi.org/10.1016/0042-6822\(87\)90269-8](https://doi.org/10.1016/0042-6822(87)90269-8).
- Bruni R, Taffon S, Equestre M, Chionne P, Madonna E, Rizzo C, Tosti ME, Alfonsi V, Ricotta L, De Medici D, Di Pasquale S, Scavia G, Pavoni E, Losio MN, Romanò L, Zanetti AR, Morea A, Pacenti M, Palù G, Capobianchi MR, Chironna M, Pompa MG, Ciccaglione AR, Italian National Task Force on Hepatitis A. 2016. Key role of sequencing to trace hepatitis A viruses circulating in Italy during a large multi-country European foodborne outbreak in 2013. *PLoS One* 11: e0149642. <https://doi.org/10.1371/journal.pone.0149642>.
- Lednicky J, Beau De Rochars VM, El Badry M, Loeb J, Telisma T, Chavannes S, Anilis G, Cella E, Ciccozzi M, Rashid M, Okech B, Salemi M, Morris JG. 2016. Zika

- virus outbreak in Haiti in 2014: molecular and clinical data. *PLoS Negl Trop Dis* 10:e0004687. <https://doi.org/10.1371/journal.pntd.0004687>.
10. Lednicky JA, Lauzard M, Fan ZH, Jutla A, Tilly TB, Gangwar M, Usmani M, Shankar SN, Mohamed K, Eiguren-Fernandez A, Stephenson CJ, Alam MM, Elbadry MA, Loeb JC, Subramaniam K, Waltzek TB, Cherabuddi K, Morris JG, Wu C-Y. 2020. Viable SARS-CoV-2 in the air of a hospital room with COVID-19 patients. *Int J Infect Dis* 100:476–482. <https://doi.org/10.1016/j.ijid.2020.09.025>.
 11. Perez Bercoff R, Gander M. 1978. In vitro translation of mengovirus RNA deprived of the terminally-linked (capping?) protein. *FEBS Lett* 96: 306–312. [https://doi.org/10.1016/0014-5793\(78\)80424-4](https://doi.org/10.1016/0014-5793(78)80424-4).
 12. Langereis MA, Feng Q, Nelissen FH, Virgen-Slane R, van der Heden van Noort GJ, Maciejewski S, Filippov DV, Semler BL, van Delft FL, van Kuppeveld FJ. 2014. Modification of picornavirus genomic RNA using 'click' chemistry shows that unlinking of the VPg peptide is dispensable for translation and replication of the incoming viral RNA. *Nucleic Acids Res* 42:2473–2482. <https://doi.org/10.1093/nar/gkt1162>.
 13. Flanagan JB, Petterson RF, Ambros V, Hewlett NJ, Baltimore D. 1977. Covalent linkage of a protein to a defined nucleotide sequence at the 5'-terminus of virion and replicative intermediate RNAs of poliovirus. *Proc Natl Acad Sci U S A* 74:961–965. <https://doi.org/10.1073/pnas.74.3.961>.
 14. Katoh K, Standley DM. 2016. A simple method to control over-alignment in the MAFFT multiple sequence alignment program. *Bioinformatics* 32: 1933–1942. <https://doi.org/10.1093/bioinformatics/btw108>.
 15. Nguyen LT, Schmidt HA, von Haeseler A, Minh BQ. 2015. IQ-TREE: a fast and effective stochastic algorithm for estimating maximum-likelihood phylogenies. *Mol Biol Evol* 32:268–274. <https://doi.org/10.1093/molbev/msu300>.
 16. Schmidt HA, Strimmer K, Vingron M, von Haeseler A. 2002. TREE-PUZZLE: maximum likelihood phylogenetic analysis using quartets and parallel computing. *Bioinformatics* 18:502–504. <https://doi.org/10.1093/bioinformatics/18.3.502>.
 17. Strimmer K, von Haeseler A. 1997. Likelihood-mapping: a simple method to visualize phylogenetic content of a sequence alignment. *Proc Natl Acad Sci U S A* 94:6815–6819. <https://doi.org/10.1073/pnas.94.13.6815>.
 18. Kalyanamoorthy S, Minh BQ, Wong TKF, von Haeseler A, Jermini LS. 2017. ModelFinder: fast model selection for accurate phylogenetic estimates. *Nat Methods* 14:587–589. <https://doi.org/10.1038/nmeth.4285>.
 19. Hoang DT, Chernomor O, von Haeseler A, Minh BQ, Vinh LS. 2018. UFBoot2: improving the ultrafast bootstrap approximation. *Mol Biol Evol* 35:518–522. <https://doi.org/10.1093/molbev/msx281>.
 20. Ramachandran S, Xia G-L, Dimitrova Z, Lin Y, Montgomery M, Augustine R, Kamili S, Khudyakov Y. 2021. Changing molecular epidemiology of hepatitis A virus infection, United States, 1996–2019. *Emerg Infect Dis* 27: 1742–1745. <https://doi.org/10.3201/eid2706.203036>.