





Citation: Radin JM, Hawksworth AW, Ortiguerra RG, Brice GT (2015) Seroprotective Antibodies to 2011 Variant Influenza A(H3N2v) and Seasonal Influenza A (H3N2) among Three Age Groups of US Department of Defense Service Members. PLoS ONE 10(3): e0121037. doi:10.1371/journal.pone.0121037

Academic Editor: Suryaprakash Sambhara, Centers for Disease Control and Prevention, UNITED STATES

Received: August 13, 2014

Accepted: January 28, 2015

Published: March 27, 2015

Copyright: This is an open access article, free of all copyright, and may be freely reproduced, distributed, transmitted, modified, built upon, or otherwise used by anyone for any lawful purpose. The work is made available under the Creative Commons CC0 public domain dedication.

Data Availability Statement: All relevant data are within the paper and its Supporting Information files.

Funding: Report number 14-29, supported by Department of Defense Global Emerging Infections Surveillance and Response System under Work Unit No. 60805. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript. Dr. Jennifer Radin, Mr. Anthony Hawksworth, and Mr. Ryan Ortiguerra are employed by The Henry M. Jackson Foundation for the Advancement of Military Medicine, Inc. and are

RESEARCH ARTICLE

Seroprotective Antibodies to 2011 Variant Influenza A(H3N2v) and Seasonal Influenza A(H3N2) among Three Age Groups of US Department of Defense Service Members

Jennifer M. Radin*, Anthony W. Hawksworth, Ryan G. Ortiguerra, Gary T. Brice

Operational Infectious Diseases Department, Naval Health Research Center, San Diego, California, United States of America

* jennifer.radin@med.navy.mil

Abstract

Background

In 2011, a new variant of influenza A(H3N2) emerged that contained a recombination of genes from swine H3N2 viruses and the matrix (M) gene of influenza A(H1N1)pdm09 virus. New combinations and variants of pre-existing influenza viruses are worrisome if there is low or nonexistent immunity in a population, which increases chances for an outbreak or pandemic.

Methods

Sera collected in 2011 were obtained from US Department of Defense service members in three age groups: 19–21 years, 32–33 years, and 47–48 years. Pre- and post-vaccination samples were available for the youngest age group, and postvaccination samples for the two older groups. Specimens were tested using microneutralization assays for antibody titers against H3N2v (A/Indiana/10/2011) and seasonal H3N2 virus (A/Perth/16/2009).

Results

The youngest age group had significantly (p<0.05) higher geometric mean titers for H3N2v with 165 (95% confidence interval [CI]: 105-225) compared with the two older groups, aged 32–33 and 47–48 years, who had geometric mean titers of 68 (95% CI: 55-82) and 46 (95% CI: 24-65), respectively. Similarly, the youngest age group also had the highest geometric mean titers for seasonal H3N2. In the youngest age group, the proportion of patients who seroconverted after vaccination was 12% for H3N2v and 27% for seasonal H3N2.

Discussion

Our results were similar to previous studies that found highest seroprotection among young adults and decreasing titers among older adults. The proportion of 19- to 21-year-olds who seroconverted after seasonal vaccination was low and similar to previous findings.



funded to do this work by the US Government. CDR Gary Brice is a military service member. The views expressed in this article are those of the authors and do not necessarily reflect the official policy or position of the Department of the Navy, Department of Defense, or the US Government. Approved for public release; distribution is unlimited. US Government Work (17 USC 105). Not copyrighted in the US. This research has been conducted in compliance with all applicable federal regulations governing the protection of human subjects in research (Protocol NHRC.2013.0025).

Competing Interests: The authors have declared that no competing interets exist.

Improving our understanding of H3N2v immunity among different age groups in the United States can help inform vaccination plans if H3N2v becomes more transmissible in the future.

Introduction

A new influenza A(H3N2) variant virus emerged in 2011, referred to as H3N2v, which contained a recombination of genes from swine H3N2 viruses and the M or matrix protein from the influenza A(H1N1)pdm09 (pH1N1) virus [1]. From 2005 to 2011, human infections with swine influenza viruses were relatively rare in the United States, with only 35 known cases [2]; however, the frequency of human cases increased dramatically in 2011 and 2012, when over 300 cases of H3N2v were identified [3,4]. These cases occurred predominately among young children exposed to swine at agricultural fairs [4]. Although there have only been a few suspected human-to-human transmitted cases [3,4], there is concern that this virus could mutate, improving its ability to spread between humans, causing new outbreaks or even a pandemic in the future. This is especially worrisome because H3N2 seasonal viruses have historically resulted in more severe illness, such as hospitalization and death [5,6].

Several studies have found that protective antibodies against H3N2v generally tend to increase among older age groups of children, peak among young adults, and then decrease among older age groups of adults [7–9]. These studies have also shown little to no immunity among children younger than age 10 years, which coincides with the ages of the H3N2v cases [7–9]. Additionally, several studies have found that vaccination with seasonal trivalent inactivated influenza vaccine (TIV) results in only small improvements in cross-protective immunity to H3N2v [7,9,10]. However, incongruous age categories, vaccine type used, different populations, small sample sizes, and different testing methods make comparisons between existing studies challenging.

To date, there have only been a few serologic studies assessing cross-protective immunity and seroconversion against H3N2v, and the one study completed in the United States used a broad age category for adults, which ranged from 18 to 49 years. Our study aims to add to the existing literature by evaluating the cross-protective antibodies among more-specific adult age categories of US Department of Defense (DoD) service members in 2011. Military-specific populations are especially important because they are often at increased risk of respiratory infections due to high-density living quarters and physical demands. Gaining a better understanding of pre- and post- vaccine immunity will help inform targeted immunization or treatment of high-risk groups if an H3N2v outbreak or pandemic were to emerge.

Materials and Methods

Participants and procedures

Three age groups of active-duty DoD service members were used for this study: 19–21 years, 32–33 years, and 47–48 years. Pre- and post-vaccination sera were obtained from the youngest group (all were US military basic trainees at either Coast Guard Training Center, Cape May, New Jersey; Fort Jackson, South Carolina; or Marine Corps Recruit Depot Parris Island, South Carolina) during a public health investigation, and sera for the other two groups were obtained from the DoD Serum Repository. All serum samples were de-identified for this study. Fortynine paired sera samples were tested for the youngest age group, and 50 single serum samples were tested for each of the two older age groups.



Pre-vaccination sera for the 19–21 year olds were collected between August 2009 and January 2011 and post-sera were collected in March 2011. More than 90% of DoD service members receive annual seasonal influenza vaccines, therefore the two oldest age groups were considered vaccinated. Post-vaccination sera for the older two groups were collected in January 2011, about 2 to 4 months after vaccination. All sera were collected prior to the first cases of H3N2v identified in the US in August 2011 [3]. Both TIV and LAIV vaccines were administered and for the 2010–2011 influenza season, the influenza vaccine composition consisted of an A/California/7/2009 (H1N1)-like virus; an A/Perth/16/2009 (H3N2)-like virus; and a B/Brisbane/60/2008-like virus [11].

Specimens were tested using microneutralization (MN) assays for antibody titers against H3N2v (A/Indiana/10/2011) and seasonal H3N2 virus (A/Perth/16/2009). Viruses used were grown in Madin-Darby canine kidney cells, and adjusted to a consistent concentration throughout the MN assays. Each specimen was tested in duplicate, and the geometric mean of both was used. An MN titer of \geq 80 was considered seropositive [7,9], and a 4-fold or greater rise in titer from pre- to post-vaccination with a postvaccination MN titer of \geq 80 was considered seroconversion.

Statistical analysis

The mean of the geometric mean, as well as the percentage of specimens with MN titers \geq 80 were calculated for each age group. An analysis of variance (ANOVA) was used to identify if any differences in the means existed across the age groups, and Tukey's honestly significant difference post hoc test was used to identify where the differences existed. A chi-squared test was used to identify any differences in the percentage of MN titers \geq 80 across age groups.

All statistical analyses were conducted using SAS software, version 9.3 (SAS Institute Inc., Cary, North Carolina). PROC FREQ and PROC MEAN were used for frequencies and means, respectively, and PROC GLM was used for the ANOVA and post hoc tests. Additionally PROC CORR was used to calculate the Pearson correlation coefficient comparing the geometric mean titers of H3Nv and seasonal H3N2.

Ethics statement

This research was conducted in compliance with all applicable federal and international regulations governing the protection of human subjects in research (Protocol NHRC.2013.0025). Authors were involved in collection of sera samples from the youngest age group and had access to personally identifiable information (PII) prior to its anonymization for the study. For the older two age groups, authors were not involved with sera collection and never had access to PII, although it is stored in a DoD sera repository. Since all specimens in this study were collected previously and were de-identified for the purposes of this study, the Naval Health Research Center institutional review board committee classified this study as minimal risk, exempt from full committee review.

Results

Postvaccination MN titers against H3N2v differed across the three age groups. The youngest age group (19-to-21-year-olds) had the highest geometric mean titer of 165 (95% confidence interval [CI]: 105–225), and the two older age groups, aged 32–33 and 47–48 years, had the lowest geometric mean titers, with 68 (95% CI: 55–82) and 46 (95% CI: 24–65), respectively. The youngest age group was statistically higher than the older two (p<0.05) in terms of geometric mean titers, but the two older groups were not significantly different from each other. The proportion with mean postvaccination titers \geq 80 for H3N2v was 69% for those aged 19–



Table 1. Geometric mean titer, percentage with microneutralization titer \geq 80 or hemagglutination inhibition titer \geq 40, and seroconversion, by age group and selected summary of previous studies with similar age groups.

Study population	Age group (years)	Birth years	n	Vaccination: LAIV, TIV, Both (% LAIV), or cross-sectional sample	-	Test type	Geometric mean MN or HI titer (95% CI)		% with MN titer \geq 80 or HI titer \geq 40		Sero- conversion
							Pre-vaccination	Post-vaccination	Pre-vaccination	Post-vaccination	(%)
US (DoD)	19–21	1990– 1992	49	Both (77%)	H3N2v ^b	MN	124 (86–162)	165 (105–225)	55 (41–69)	69 (56–82)	12 (3–21) ^a
					H3N2 ^c		69 (39–100)	175 (97–253)	29 (16–42)	51 (37–65)	27 (15–39) ^a
	32–33	1977– 1978	50	Both (38%)	H3N2v ^b	MN		68 (55–82)		44 (30–58)	
					H3N2 ^c			38 (22-54)		16 (6–26)	
	47–48	1962– 1963	50	Both (45%)	H3N2v ^b	MN		46 (24–68)		16 (6–26)	
					H3N2 ^c			52 (28–76)		24 (12–36)	
US (NHANES & a 2010-11 TIV study)	18–49		30	TIV	H3N2v ^d	MN	55 (31–98)	95 (51–177)	43	63	13
					H3N2 ^c		31 (16–61)	172 (94–316)	27	70	50
Canada	20-59		65	TIV	H3N2v ^b	HI	14 (10–18)	22 (16–29)	26 (15–37)	38 (26–50)	11 (3–19)
					H3N2 ^e		22 (16–29)	128 (94–173)	35 (23–47)	89 (81–97)	65 (53–77)
							Cross-sec	tional sample	Cross-sect	ional sample	
	20–29		98	Cross-sectional	H3N2v ^b	HI	43 (34–54)		59 (49–69)		
					H3N2 ^f		16 (12–22)	28 (19–36)	
	30–39		100	Cross-sectional	H3N2v ^b	HI	22 (18–26)	35 (2	26–44)	
					H3N2 ^f		15 (11–18)	31 (2	22–40)	
	40–49		100	Cross-sectional	H3N2v ^b	HI	9 (8–10)	7 (2	2–12)	
					H3N2 ^f		12	(9–15)	25 (17–34)	
Norway	18–24	1993– 1987	28	Cross-sectional	H3N2v ^g	HI	37 (27–52)	71 (53–86)	
	25–34	1986– 1977	45		H3N2v ^g		41 (31–53)	71 (57–83)	
	35–44	1976– 1967	27		H3N2v ^g		27 (20–35)	48 (3	30–67)	
	45–54	1966– 1957	22		H3N2v ^g		11 ((8–16)	14 (4–33)	

DoD, Department of Defense; HI, hemagglutination inhibition; LAIV, live attenuated influenza vaccine; MN, microneutralization; NHANES, National Health and Nutrition Examination Survey; TIV, trivalent influenza vaccine.

doi:10.1371/journal.pone.0121037.t001

21 years, and declined to 44% and 16% among those aged 32–33 and 47–48 years, respectively. All age groups were significantly different from each other for the proportion with mean titers \geq 80 (p<0.05). Additionally, as the ages increased, there was a steady decline in the number of participants with titers \geq 160 and a steady increase in the number of participants with titers \leq 20 (<u>Table 1</u> and <u>Fig. 1</u>).

For seasonal H3N2 titers, the youngest age group also had the highest geometric mean titer with 175 (95% CI: 97–253), followed by the oldest age group with 52 (95% CI: 28–76). The lowest titers were found in the middle age group with 38 (95% CI: 22–54), although they were not

 $^{^{\}mathrm{a}}4\text{-fold}$ or greater increase, with postvaccination MN titers \geq 80.

^bA/Indiana/10/2011.

^cA/Perth/16/2009.

^dA/Minnesota/11/2010.

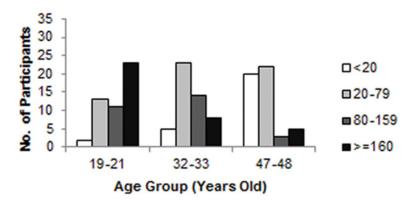
eA/Wisconsin/15/2009.

fA/Brisbane/10/2007.

gA/Indiana/08/2011.



A. A/Indiana/10/2011



B. A/Perth/16/2009

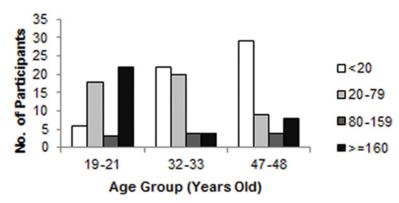


Fig 1. Postvaccination microneutralization titers by three age groups of US Department of Defense service members. (A) H3N2v (A/Indiana/10/2011) and (B) seasonal H3N2 (A/Perth/16/2009).

doi:10.1371/journal.pone.0121037.g001

significantly different from the oldest age group. Similarly, the proportion with postvaccination MN titers \geq 80 for seasonal H3N2 was 51% for those aged 19–21 years, 16% for 32–33 years, and 24% for 47–48 years. Interestingly, the titers for those aged 32–33 years were significantly lower for H3N2 than for H3N2v. The titer trends for seasonal H3N2 were more variable, with the youngest having very few intermediate titers of 80–159 (see Table 1 and Fig. 1).

In the youngest age group, 12% seroconverted to H3N2v and 27% seroconverted to seasonal H3N2 following vaccination (<u>Table 1</u>).

The Pearson correlation between H3N2v and seasonal H3N2 geometric mean titers was 0.65 (p<0.0001) for pre-vaccination among the 19- to 21-year olds and 0.59 (p<0.0001) for post-vaccination among all age groups.

Discussion

H3N2v first emerged in 2011 with 13 cases identified between August 2011 and April 2012 [12], and it quickly grew to 305 between July 9 and September 7, 2012 [4]. However, modeling suggests that the actual number was much higher: approximately 2,000 cases during the first time period due to lack of identification by existing surveillance systems [12]. Among the 13



initial cases, 12 were children, with most under age 10 years, and 54% had known swine exposure [3]. Among the 305 cases, 92% were younger than 18 years, and 95% had direct or indirect exposure to swine [4]. Although most cases had swine exposure, there were 15 possible human-to-human transmitted cases [4]. The possibility of future human-to-human H3N2v transmission is a concern considering the high susceptibility among children and older adults.

In this study we found that young adults aged 19–21 years had significantly higher geometric mean titers (p<0.05) mean antibody titers against H3N2v and seasonal H3N2 compared with the older adults, aged 32–33 and 47–48 years. We also found that with increased age came decreased antibodies to H3N2v, although the geometric mean titers of the two older age groups in our study were not significantly different. Seropositivity to influenza may vary by age as a result of different exposures to circulating influenza viruses during childhood and different exposures in adulthood that would have boosted antibodies, thus leading to cohort effects [7]. Regional differences in circulating strains across countries may also influence seropositivity and explain some of the variation seen across study populations.

Due to the characteristics of our study population, it is unlikely that the participants were exposed to H3N2v directly. However, it is possible that this population, especially the youngest age group, had recent natural exposure to seasonal H3N2. Natural infection has been found to result in stronger immunologic responses than vaccination, especially in younger adults [13], which may explain the large proportion of participants in the youngest age group with seasonal H3N2 titers \geq 160 (Fig. 1).

Similar seropositive proportions to H3N2v by age group have been found in other studies in the United States, Norway, and Canada [7–9]. The Canadian study used hemagglutination inhibition (HI) assay titers and found that 0% of children <5 years of age had seropositive antibodies for H3N2v, <20% of individuals aged \le 14 and \ge 40 years, and approximately 50% for individuals aged 15–39 years [7]. The Norwegian study found 71% seropositivity among 18- to 34-year-olds, and the lowest seropositivity among children younger than age 12 (0%) and 45- to 54-year-olds (14%) [8]. The previous US study found no seropositivity among children aged 6–35 months, 33% among adults aged 18–49 years, and 17% among adults aged 65 years and older [9] (8). Overall, these studies agreed with our results, showing the highest seropositivity among young adults and lowest among 40- to 54-year-olds.

Our study found H3N2v seroconversion rates similar to other studies, with 12% seroconverting in the 19–21 year old age group (the only group for which we had pre-vaccination sera) following vaccination. The study in Canada found seroconversion rates were <15% in all age groups and across all vaccine groups [7]. A study by the Centers for Disease Control and Prevention found similar increases against H3N2v among adults who received TIV, with 13–17% experiencing seroconversion. However, they did not find seroconversion in children aged 6–35 months [9]. Similarly, studies among ferrets have also found limited to no protection to H3N2v among individuals vaccinated with TIV [10].

Our study also had rates of seroconversion to seasonal H3N2 similar to those of previous studies that used live attenuated influenza virus (LAIV) [14]. However, our study had much lower seroconversion for seasonal H3N2 compared with the Canadian study that used TIV. Additional seroconversion studies in other age groups will be important for understanding the full benefit of seasonal vaccine protection for H3N2v and seasonal H3N2 viruses.

This study consisted of US DoD service members who are a highly vaccinated and an overall active and healthy population. Consequently, this study population may be less generalizable to the general public who may have lower immune responses to vaccination due to comorbidities. Additionally, a high proportion of the 19- to 21-year-olds in our study received LAIV (77%). Previous studies have found higher efficacy and effectiveness with LAIV compared to TIV among children, but mixed results among adults [15,16]. However, a previous study



comparing seroconversion rates among military recruits found that they were higher for TIV [17]. This may be a result of higher mucosal immune response and lower systemic immune response with the intranasal LAIV vaccine, resulting in low seroconversion [18]. Similarly among the 19- to 21-year-olds in our study, the mean geometric mean titers for H3N2v among the LAIV vaccinated group in our study were smaller (130, 95% CI: 86–174) than the TIV vaccinated group (282, 95% CI: 45–519). This could explain why the seroconversion rate we observed was similar to previous studies that examined response to LAIV.

The two older age groups in this study were presumably vaccinated routinely for many years, whereas the younger group likely had low vaccination rates similar to the general population in the years prior to the study. It is postulated that once a person is exposed to an influenza vaccine, he or she may reach an "antibody ceiling," and his or her antibody titers will not increase in response to infection [14]. However, it has also been found that prior influenza vaccination may negatively impact future influenza immunological responses [19–23], especially previous vaccination with TIV [23]. This may explain, in part, why we saw very low titers against H3N2 (Perth) in the two older age groups compared with the previous US study [9], which likely had lower vaccination rates in prior seasons.

Although serologic testing is often used to identify subclinical infections and infections that would not be identified by real-time polymerase chain reaction, it can be nonspecific, with serologic testing sometimes picking up cross-reacting antibodies from similar influenza strains [24]. Our study found a moderate but significant correlation between H3N2v and seasonal H3N2v geometric mean titers, both before and after influenza vaccination, which may reflect some cross protective immunity between the two virus strains.

Specificity of diagnostic tests can also vary across age groups; one study found lower MN specificity for pH1N1 in older age groups and 80% sensitivity when the \geq 80 cutoff was used [25]. Another study found that an HI titer of 110 instead of 40 corresponded to 50% clinical protection among children, therefore different cutoffs should be used for different age groups [26]. Consequently, seroprotection may be overestimated in our young adult group. Additionally, antibody titers decrease over time since vaccination, with faster declines seen among older age groups [27]. These declines were seen as early as 6 months [27] and could have also played a role in the lower titers seen among the two older groups who were sampled on average further from the time of vaccination compared with the youngest group.

Despite these concerns, MN titers measure the number of neutralizing antibodies and may have greater sensitivity than HI antibody titers, especially with novel influenza viruses [25]. Although, the 50% seroprotective level for MN is not known for H3N2v specifically, a previous study found that MN titers were usually double that of HI titers for pH1N1 [25], and previous H3N2v serology studies have used MN and HI cutoffs of \geq 80 or \geq 40, respectively, to represent a 50% seroprotective level [7,9]. However, the US National Health and Nutrition Examination Survey study, which used both HI and MN titers on the same samples, showed that the percentage with seroprotective titers was consistently higher for MN than HI when the \geq 80 and \geq 40 cutoffs were used [9]. The variation among and between different serology tests should be taken into consideration when comparing serological results.

Improving our knowledge of H3N2v cross-protective antibodies among specific age groups is important in case this strain becomes more easily transmissible from human to human. The main differences across age groups are likely a result of different influenza exposures to circulating strains during childhood, during which time the strongest immune response is mounted, and differences in exposures later in life that boost antibodies [28]. Identifying groups who are at highest risk is important since there are often shortages of vaccines at the beginning of an outbreak or pandemic. Age groups with lower cross-protective antibodies could be targeted to receive priority vaccination, which could potentially reduce the spread of the virus.



Supporting Information

S1 IRB. Institutional Review Board approval.

(PDF)

S2 IRB. Institutional Review Board completion.

(PDF)

S1 Dataset.

(XLSX)

Acknowledgments

We thank Patrick Blair, Daisy Cabrera, Christopher Clagett, Nakia Clemmons, Angelia Cost, Shawn Garcia, Kris Legge, Michelle LeWark, Chris Myers, Laura Pacha, and Damaris Padin.

Author Contributions

Conceived and designed the experiments: AWH JMR. Performed the experiments: RO. Analyzed the data: JMR AWH. Wrote the paper: JMR AWH RO GTB.

References

- Lindstrom S, Garten R, Balish A, Shu B, Emery S, Berman L, et al. Human infections with novel reassortant influenza A(H3N2)v viruses, United States, 2011. Emerg Infect Dis. 2012; 18: 834–837. doi: 10.3201/eid1805.111922 PMID: 22516540
- Centers for Disease Control and Prevention (CDC). Update: Influenza A(H3N2)v transmission and guidelines—five states, 2011. MMWR Morb Mortal Wkly Rep. 2012; 60: 1741–1744. PMID: 22217624
- Epperson S, Jhung M, Richards S, Quinlisk P, Ball L, Moll M, et al. Human infections with influenza A (H3N2) variant virus in the United States, 2011–12. Clin Infect Dis. 2013; 57: S4–S11. doi: 10.1093/cid/cit272 PMID: 23794729
- Jhung MA, Epperson S, Biggerstaff M, Allen D, Balish A, Barnes N, et al. Outbreak of variant influenza A(H3N2v) virus in the United States. Clin Infect Dis. 2013; 57: 1703–1712. doi: 10.1093/cid/cit649 PMID: 24065322
- Thompson WW, Shay DK, Weintraub E, Brammer L, Bridges CB, Cox NJ, et al. Influenza-associated hospitalizations in the United States. JAMA 2004; 292: 1333–1340. PMID: 15367555
- Thompson WW, Shay DK, Weintraub E, Brammer L, Cox N, Anderson L, et al. Mortality associated with influenza and respiratory syncytial virus in the United States. JAMA 2003; 289: 179–186. PMID: 12517228
- Skowronski DM, Janjua NZ, Serres GD, Purych D, Gilca V, Scheifele DW, et al. Cross-reactive and vaccine-induced antibody to an emerging swine-origin variant of influenza A virus subtype H3N2 (H3N2v). J Infect Dis. 2012; 206: 1852–1861. doi: 10.1093/infdis/jis500 PMID: 22872731
- **8.** Waalen K, Kilander A, Dudman SG, Ramos-Ocao R, Hungnes O. Age-dependent prevalence of antibodies cross-reactive to the influenza A(H3N2) variant virus in sera collected in Norway in 2011. Euro Surveill. 2012; 17: 1–5.
- Centers for Disease Control and Prevention (CDC). Antibodies cross-reactive to influenza A (H3N2)
 variant virus and impact of 2010–11 seasonal influenza vaccine on cross-reactive antibodies—United
 States. MMWR Morb Mortal Wkly Rep. 2012; 16: 237–241.
- Houser KV, Katz JM, Tumpey TM. Seasonal trivalent inactivated influenza vaccine does not protect against newly emerging variants of influenza A (H3N2v) virus in ferrets. J Virol. 2013; 87: 1261–1263. doi: 10.1128/JVI.02625-12 PMID: 23115290
- World Health Organization (WHO). Recommended viruses for influenza vaccines for use in the 2010– 2011 northern hemisphere influenza season. Available at: http://www.who.int/influenza/vaccines/virus/2010_11north/en/. Accessed 20 November 2014.
- 12. Biggerstaff M, Reed C, Epperson S, Jhung MA, Gambhir M, Bresee JS, et al. Estimates of the number of human infections with influenza A (H3N2) variant virus, United States, August 2011–April 2012. Clin Infect Dis. 2013; 57: S12–S15. doi: 10.1093/cid/cit273 PMID: 23794726



- Chan KH, To KK, Hung IF, Zhang AJ, Chan JF, Cheng VC, et al. Differences in antibody responses of individuals with natural infection and those vaccinated against pandemic H1N1 2009 influenza. Clin Vaccine Immunol. 2011; 18: 867–873. doi: 10.1128/CVI.00555-10 PMID: 21411604
- Petrie JG, Ohmit SE, Johnson E, Cross RT, Monto AS. Efficacy studies of influenza vaccines: effect of end points used and characteristics of vaccine failures. J Infect Dis. 2011; 203: 1309–1315. doi: 1093/infdis/jir015 PMID: 21378375
- Ambrose CS, Levin MJ, Belshe RB. The relative efficacy of trivalent live attenuated and inactivated influenza vaccines in children and adults. Influenza Other Respir Viruses. 2011; 5: 67–75. doi: 10.1111/j. 1750-2659.2010.00183.x PMID: 21306569
- Phillips CJ, Woolpert T, Sevick C, Faix D, Blair PJ, Crum-Cianflone NF. Comparison of the effectiveness of trivalent inactivated influenza vaccine and live, attenuated influenza vaccine in preventing influenza-like illness among US military service members, 2006–2009. Clin Infect Dis. 2012; 56: 11–19. doi: 10.1093/cid/cis860 PMID: 23183869
- Faix DJ, Hawksworth AW, Myers CA, Hansen CJ, Ortiguerra RG, Halpin R, et al. Decreased serologic response in vaccinated military recruits during 2011 correspond to genetic drift in concurrent circulating pandemic A/H1N1 viruses. PLoS One 2012; 7: e3481.
- Barria MI, Garrido JL, Stein C, Scher E, Ge Y, Engel SM, et al. Localized mucosal response to intranasal live attenuated influenza vaccine in adults. J Infect Dis. 2013; 207: 115–124. doi: 10.1093/infdis/iis641 PMID: 23087433
- Song JY, Cheong HJ, Seo YB, Kim IS, Noh JY, Choi WS, et al. Long-term immunogenicity of the pandemic influenza A/H1N1 2009 vaccine among health care workers: influence of prior seasonal influenza vaccination. Clin Vaccine Immunol. 2013; 20: 513–516. doi: 10.1128/CVI.00725-12 PMID: 23365206
- Ohmit SE, Petrie JG, Malosh RE, Cowling BJ, Thompson MG, Shay DK, et al. Influenza vaccine effectiveness in the community and the household. Clin Infect Dis. 2013; 56: 1363–1369. doi: 10.1093/cid/cit060 PMID: 23413420
- Beyer WE, Palache AM, Sprenger MJ, Hendricksen E, Tukker JJ, Darioli R, et al. Effects of repeated annual influenza vaccination on vaccine sero-response in young and elderly adults. Vaccine 1996; 14: 1331–1339. PMID: 9004442
- 22. Huijskens E, Rossen J, Mulder P, van Beek R, van Vugt H, Verbakel J, et al. Immunogenicity, boostability, and sustainably of the immune response after vaccination against influenza A virus (H1N1) 2009 in a healthy population. Clin Vaccine Immunol. 2011; 18: 1401–1405. doi: 10.1128/CVI.05046-11 PMID: 21795459
- Sasaki S, He XS, Homes TH, Dekker CL, Kemble GW, Arvin AM, et al. Influence of prior influenza vaccination on antibody and B-cell responses. PLoS One 2008; 3: e2975. doi: 10.1371/journal.pone. 0002975 PMID: 18714352
- Kelly H, Peck HA, Laurie KL, Wu P, Nishiura H, Cowling BJ. The age-specific cumulative incidence of infection with pandemic influenza H1N1 2009 was similar in various countries prior to vaccination. PLoS One 2011; 6: e21828. doi: 10.1371/journal.pone.0021828 PMID: 21850217
- Veguilla V, Hancock K, Schiffer J, Garguillo P, Lu X, Aranio D, et al. Sensitivity and specificity of serologic assays for detection of human infection with 2009 pandemic H1N1 virus in U.S. populations. J Clin Microbiol. 2011; 49: 2210–2215. doi: 10.1128/JCM.00229-11 PMID: 21471339
- Black S, Nicolay U, Vesikari T, Knuf M, Del Giudice G, Della Cioppa G, et al. Hemagglutination inhibition antibody titers as a correlate of protection for inactivated influenza vaccines in children. Pediatr Infect Dis. 2011; 30: 1081–1085. doi: 10.1097/INF.0b013e3182367662 PMID: 21983214
- Song JY, Cheong HJ, Hwang IS, Choi WS, Jo YM, Park DW, et al. Long-term immunogenicity of influenza vaccine among the elderly: risk factors for poor immune response and persistence. Vaccine 2010; 28: 3929–3935. doi: 10.1016/j.vaccine.2010.03.067 PMID: 20394719
- Skowronski DM, De Serres G, Janjua NZ, Gardy JL, Gilca V, Dionne M, et al. Cross-reactive antibody
 to swine influenza A(H3N2) subtype virus in children and adults before and after immunization with
 2010/11 trivalent inactivated influenza vaccine in Canada, August to November 2010. Euro Surveill.
 2012; 17: 1–8.