



Direct Cardiac Actions of Sodium Glucose Cotransporter 2 Inhibitors Target Pathogenic Mechanisms Underlying Heart Failure in Diabetic Patients

Laween Uthman¹, Antonius Baartscheer², Cees A. Schumacher², Jan W. T. Fiolet², Marius C. Kuschma¹, Markus W. Hollmann¹, Ruben Coronel^{2,3}, Nina C. Weber¹ and Coert J. Zuurbier^{1*}

¹ Laboratory of Experimental Intensive Care and Anesthesiology, Amsterdam UMC, University of Amsterdam, Meibergdreef, Amsterdam, Netherlands, ² Clinical and Experimental Cardiology, Amsterdam UMC, University of Amsterdam, Meibergdreef, Amsterdam, Netherlands, ³ IHU Liryc, Electrophysiology and Heart Modeling Institute, Fondation Bordeaux Université, Bordeaux, France

OPEN ACCESS

Edited by:

Gaetano Santulli,
Columbia University, United States

Reviewed by:

Xiaoqiang Tang,
Sichuan University, China
Wayne Rodney Giles,
University of Calgary, Canada

*Correspondence:

Coert J. Zuurbier
c.j.zuurbier@amc.uva.nl

Specialty section:

This article was submitted to
Cardiac Electrophysiology,
a section of the journal
Frontiers in Physiology

Received: 31 July 2018

Accepted: 22 October 2018

Published: 21 November 2018

Citation:

Uthman L, Baartscheer A, Schumacher CA, Fiolet JWT, Kuschma MC, Hollmann MW, Coronel R, Weber NC and Zuurbier CJ (2018) Direct Cardiac Actions of Sodium Glucose Cotransporter 2 Inhibitors Target Pathogenic Mechanisms Underlying Heart Failure in Diabetic Patients. *Front. Physiol.* 9:1575. doi: 10.3389/fphys.2018.01575

Sodium glucose cotransporter 2 inhibitors (SGLT2i) are the first antidiabetic compounds that effectively reduce heart failure hospitalization and cardiovascular death in type 2 diabetics. Being explicitly designed to inhibit SGLT2 in the kidney, SGLT2i have lately been investigated for their off-target cardiac actions. Here, we review the direct effects of SGLT2i Empagliflozin (Empa), Dapagliflozin (Dapa), and Canagliflozin (Cana) on various cardiac cell types and cardiac function, and how these may contribute to the cardiovascular benefits observed in large clinical trials. SGLT2i impaired the Na⁺/H⁺ exchanger 1 (NHE-1), reduced cytosolic [Ca²⁺] and [Na⁺] and increased mitochondrial [Ca²⁺] in healthy cardiomyocytes. Empa, one of the best studied SGLT2i, maintained cell viability and ATP content following hypoxia/reoxygenation in cardiomyocytes and endothelial cells. SGLT2i recovered vasoreactivity of hyperglycemic and TNF- α -stimulated aortic rings and of hyperglycemic endothelial cells. Anti-inflammatory actions of Cana in IL-1 β -treated HUVEC and of Dapa in LPS-treated cardiofibroblast were mediated by AMPK activation. In isolated mouse hearts, Empa and Cana, but not Dapa, induced vasodilation. In ischemia-reperfusion studies of the isolated heart, Empa delayed contracture development during ischemia and increased mitochondrial respiration post-ischemia. Direct cardiac effects of SGLT2i target well-known drivers of diabetes and heart failure (elevated cardiac cytosolic [Ca²⁺] and [Na⁺], activated NHE-1, elevated inflammation, impaired vasorelaxation, and reduced AMPK activity). These cardiac effects may contribute to the large beneficial clinical effects of these antidiabetic drugs.

Keywords: SGLT2 inhibitors, cardiomyocyte, endothelial cell, smooth muscle cell, cardiac fibroblast, isolated heart, 2 type diabetes, heart failure

SGLT2 INHIBITORS IN DIABETES AND HEART FAILURE: EXPLORING DIRECT CARDIAC EFFECTS

Sodium glucose cotransporter 2 (SGLT2) inhibitors (SGLT2i) are kidney-targeted anti-diabetic agents that have exhibited marked reductions in cardiovascular events and mortality of type 2 diabetes (T2D) patients. In two large clinical trials, Empa, (Zinman et al., 2015) and Cana, (Neal et al., 2017) reduced the risk of heart failure related hospitalization by 35 and 33%, respectively. These trials demonstrate that SGLT2i may serve as an effective treatment strategy against heart failure in a T2D setting. SGLT2i were designed to inhibit the kidney-specific SGLT2 and to induce glycosuria. SGLT2 is part of a family of proteins facilitating glucose translocation in a variety of tissues. SGLT2 is mainly expressed in the kidney, with some expression in the pancreatic alpha cells. In the kidney, SGLT2 is located in the first part of the proximal tubule, where it enables ~90% of glucose reabsorption from the urine; the other 10% reabsorption is through SGLT1, which is located in the distal part of the proximal tubule. SGLT2i use is associated with ~50 g/day of glucose loss, which corresponds with ~200 kcal/day (Ferrannini et al., 2015). Indeed, administration of a SGLT2i on top of standard glucose lowering therapy (metformin, insulin and sulfonylureas) modestly reduced (~0.4%) glycated hemoglobin plasma levels and was associated with small decreases in body weight, plasma insulin and blood pressure (Zinman et al., 2015 Neal et al., 2017). However, it is unlikely that the small changes in these parameters can explain the large beneficial actions of SGLT2i, and thus preclinical studies were sought to explore possible underlying mechanisms.

In vivo preclinical studies have shown decreased ROS levels, inflammatory cytokines and vascular dysfunction after SGLT2i

administration in animals with diabetes (Oelze et al., 2014; Andreadou et al., 2017; Tanajak et al., 2018). Moreover, preserved cardiac function has been observed with administration of SGLT2i Empa in a non-diabetic model of chronic heart failure (Byrne et al., 2017). In microvascular coronary endothelial cells of mice with STZ-induced diabetes, Empa administration resulted in activation of AMPK, improved mitochondrial function through inhibition of Drp1-mediated fission, restoration of vascular barrier function, eNOS phosphorylation, adhesion molecules expression and reduced mitochondrial ROS levels (Zhou et al., 2018). However, the *in vivo* animal studies cannot discern whether the cardiovascular beneficial effects of Empa and other SGLT2i are due to kidney-related systemic alterations or due to direct cardiovascular effects, or both. To examine whether SGLT2i operate directly on cardiac specific pathophysiological mechanisms without interference of other mediating factors, including plasma circulating glucose and insulin, isolated cardiac cell and organ studies are required. In this review we summarize the results of these studies and discuss how these direct cardiac effects of the inhibitors may contribute to the effects seen on heart failure. We restrict ourselves to cardiac cellular studies using clinically relevant SGLT2i concentrations ($\leq 10 \mu\text{M}$).

PATHOGENESIS LINKING DIABETES AND HEART FAILURE

T2D is associated with a two to five fold higher risk of developing heart failure (Kannel et al., 1974; Cavender et al., 2015). Vice versa, chronic heart failure patients have an increased risk of developing T2D and metabolic abnormalities, including increased insulin resistance (Riehle and Abel, 2016). Thus, the presence of one disease impacts the development or progression of the other. Patients with heart failure and T2D have a worse prognosis than those with either one of the diseases (Cubbon et al., 2013).

Heart failure is a syndrome characterized by fatigue, dyspnea and the inability to exercise. It often develops after MI, valvular disease, chronic tachycardia, hypertension, diabetic cardiomyopathy, or in the setting of various genetic or acquired cardiomyopathies. The initial remodeling process associated with heart failure primarily includes left ventricular hypertrophy, fibrosis and diastolic dysfunction. Although the exact causes of T2D are still debated, hyperinsulinemia, insulin resistance, hyperglycemia and dyslipidemia are considered early contributors to the development of T2D (Jia et al., 2018). In T2D patients, insulin resistance and increased salt sensitivity lead to hypertension by increased sodium retention, activation of sympathetic nervous system and atherosclerosis (Feldstein, 2002; Lastra et al., 2010). Atherosclerosis in turn may cause MI and heart failure. Furthermore, the T2D heart is considered to be in a state of metabolic overload, with elevated glycogen and lipid stores within the heart (Varma et al., 2018). T2D is only one of several diseases that can develop into heart failure (see above). Early on in the development of heart failure, there are several key mechanisms that instigate functional and structural cardiac impairments, which are shared among heart failure and

Abbreviations: ΔG_{ATP} , Gibbs free energy of ATP hydrolysis; $[\text{Ca}^{2+}]_c$, cytoplasmic calcium; $[\text{Na}^+]_c$, cytoplasmic sodium; 2-NBDG, 2-(N-(7-Nitrobenz-2-oxa-1,3-diazol-4-yl)Amino)-2-Deoxyglucose; AGE, advanced glycation end-product(s); Akt, protein kinase B; AMP/ADP, adenosine monophosphate/adenosine diphosphate; AMPK, 5' adenosine monophosphate-activated protein kinase; ASC, apoptosis-associated speck-like protein containing a caspase recruitment domain; CABG, coronary artery bypass graft; Cana, Canagliflozin; C_{max} , maximal serum concentration; Dapa, Dapagliflozin; Drp1, dynamin-related protein; Empa, Empagliflozin; eNOS, endothelial nitric oxide synthase; ETC, electron transport chain; HAEC, human aortic endothelial cells; HbA1c, hemoglobin A1c; HFpEF, heart failure with preserved ejection fraction; HUVEC, human umbilical cord vein endothelial cells; ICAM-1, intercellular Adhesion Molecule 1; IL-1 β , interleukin 1 β ; IL-10, interleukin 10; IL-6, interleukin 6; IR, ischemia-reperfusion; IRS-1, insulin receptor substrate 1; JNK, c-Jun N-terminal kinases; LPS, lipopolysaccharides; LTCC, L-type calcium channel; MCP-1, monocyte chemoattractant protein 1; MI, myocardial infarction; Na^+/K^+ , ATPase sodium-potassium ATPase; NCX, $\text{Na}^+/\text{Ca}^{2+}$ -exchanger; NF κ B, nuclear factor-kappa B; NHE, sodium hydrogen exchanger; NLRP3, NACHT, LRR and PYD domains-containing protein 3; NO-cGMP-PKG, nitric oxide - cyclic guanosine monophosphate - protein kinase G; NOX2, NADPH oxidase 2; PA, palmitic acid; PCr/ATP, phosphocreatine/adenosine triphosphate; PTCA, percutaneous transluminal coronary angioplasty; ROS, reactive oxygen species; SGLT1, sodium glucose cotransporter 1; SGLT2, sodium glucose cotransporter 2; SGLT2i, sodium glucose cotransporter 2 inhibitor(s); SMC, smooth muscle cell(s); SMIT-1, sodium-myoinositol co-transporter 1; SR, sarcoplasmic reticulum; STAT3, signal transducer and activator of transcription 3; STZ, streptozotocin; T2D, type 2 diabetes; TNF- α , tumor necrosis factor α ; VCAM-1, vascular cell adhesion protein 1.

all these diseases, including T2D (Bugger and Abel, 2014; Packer, 2017). Therefore, targeting these early overlapping mechanisms is a promising strategy to treat heart failure. Several of these mechanisms with potential relevance to direct myocardial effects of SGLT2i are summarized in **Figure 1**.

First, elevated cytosolic $[Na^+]_c$ and $[Ca^{2+}]_c$ ($[Na^+]_c$ and $[Ca^{2+}]_c$) are observed in cardiomyocytes from failing and T2D hearts (Despa et al., 2002; Lambert et al., 2015; Despa, 2018). Elevations in $[Na^+]_c$ and $[Ca^{2+}]_c$ are coupled to reduced mitochondrial $[Ca^{2+}]_m$ ($[Ca^{2+}]_m$) in cardiomyocytes, which will decrease the energetic and redox function of mitochondria (Murphy and Eisner, 2009; Bay et al., 2013). Elevated $[Ca^{2+}]_c$ is perceived in hyperglycemia- and LPS-induced endothelial dysfunction (Wang et al., 2008; Cui et al., 2013), atrial fibrillation (Neef et al., 2010) and in myocytes during IR injury (Garcia-Dorado et al., 2012). Increases in $[Na^+]_c$ and $[Ca^{2+}]_c$ are caused by perturbed ion fluxes due to altered activity of ion channels and/or transporters, including the sodium-calcium exchanger (NCX), the sodium-hydrogen exchanger 1 (NHE-1), the ryanodine receptor regulating SR-calcium release and the sodium-potassium ATPase (Na^+/K^+ ATPase), and targeting these ion regulating transporters has been proposed to improve cardiovascular function (Baartscheer et al., 2003a; Despa et al., 2012; Karmazyn, 2013; Luo et al., 2013; Sasahara et al., 2013).

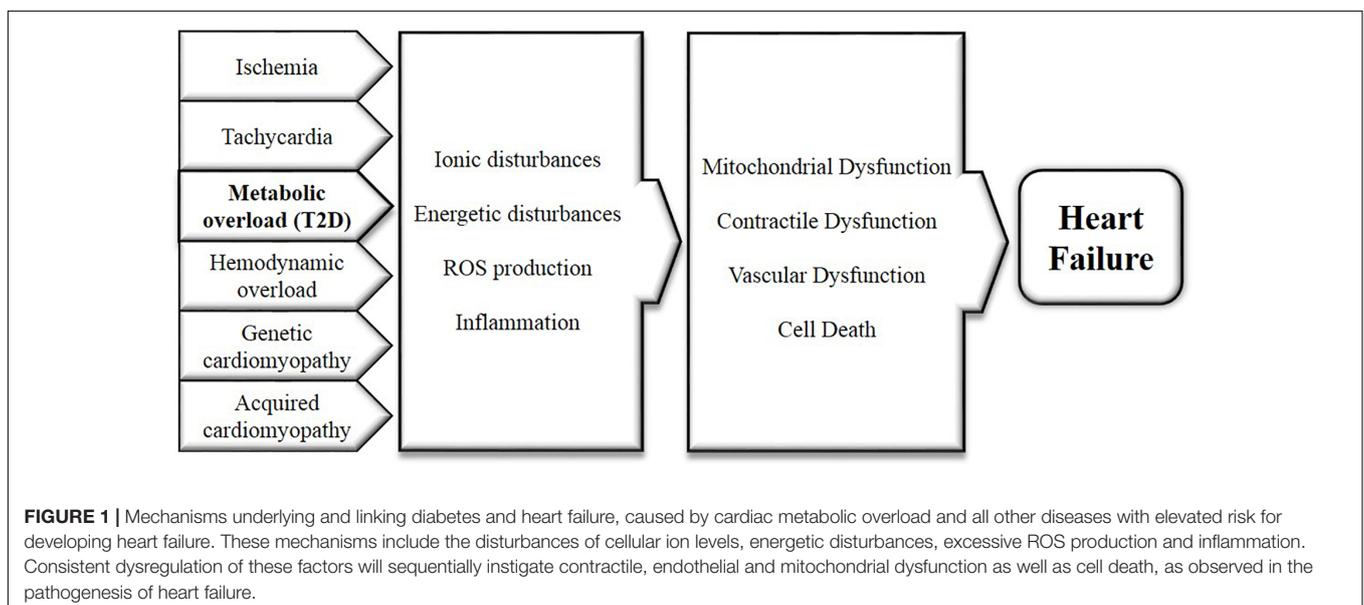
Second, the development of an energy crisis, best reflected by decreases in cardiac PCr/ATP, is present in both heart failure and diabetic cardiomyopathy. As a consequence, the cellular energy sensor AMPK is altered. AMPK activity is increased in failing myocytes (Tian et al., 2001), whereas in T2D divergent AMPK alterations have been found (Varma et al., 2018). AMPK acts as an energy sensor, and activates catabolic actions such as increasing glucose uptake and glycolysis, and impairing anabolic reactions (Beauloye et al., 2011). Targeting the AMPK pathway by increasing its activity have been shown to combat cardiac hypertrophy and heart failure (Gélinas et al., 2018). Besides,

AMPK activity is relevant in non-metabolic cellular processes, including the regulation of vascular tone and the suppression of inflammation (Beauloye et al., 2011; He et al., 2015; Cordero et al., 2018). Metformin, the first-line drug in the treatment of T2D for the last two decades, is known to increase AMPK (Foretz et al., 2014), an effect that can also be observed in the human non-diabetic heart tissue (El Messaoudi et al., 2015).

Third, early increased oxidative stress figures prominently in both heart failure (Dey et al., 2018) and T2D pathogenesis (Shah and Brownlee, 2017). The highly reactive ROS can oxidize many proteins in its vicinity, with consequently altered function of that protein. ROS is also an upstream driver of endothelial dysfunction associated with heart failure, by the increase of nitric oxide synthase (NOS) uncoupling, switching the production of NO into that of peroxynitrite (Paulus and Tschöpe, 2013; Joshi et al., 2014; Münzel et al., 2015). Reducing ROS therefore will normalize the function of many proteins of cardiac cells and may so undermine the development of heart failure.

Fourth a chronic low-grade state of inflammation is present in both heart failure and diabetes. Cellular ion dysregulation, AMPK inactivity and ROS production are relevant factors in the development of inflammation (Fрати et al., 2017). Other mechanisms that underlie the progression of inflammation in diabetes may include increased O-GlcNAcylation, formation of AGEs, activation of the renin-angiotensin-aldosterone system, damage-associated molecular patterns (DAMPs) and the production of cytokines and adipokines (Fрати et al., 2017). Inflammation may lead to fibrosis, cell death (pyroptosis) and cardiac remodeling, and its suppression may therefore be a relevant mechanism in the prevention and treatment of diabetes-associated heart failure.

These four mechanisms are not separate entities but are all intrinsically interrelated, and together may induce contractile, vascular and mitochondrial dysfunction, and cell death in the heart, which may evolve into left ventricular concentric or



eccentric hypertrophy and heart failure (Boudina et al., 2007; Van Heerebeek et al., 2008; Miki et al., 2013; Sena et al., 2013; Cassidy et al., 2015; Munasinghe et al., 2016). In the subsequent paragraphs, SGLT2i effects in the major cardiac cell types will be discussed in relation to these four early signatures (ion homeostasis, energy balance, oxidative stress and inflammation) of both heart failure development and diabetic cardiomyopathy.

SGLT2 INHIBITORS IN CARDIOMYOCYTES

Intracellular Ion Regulation

Several reports have documented that SGLT2i modify ionic homeostasis in cardiomyocytes. Hamouda et al. (2014) showed that 1–10 μM Dapa reduced the amplitudes of cell shortening and L-type Ca^{2+} current in ventricular cardiomyocytes from STZ-treated and control rats. Dapa only lowered systolic $[\text{Ca}^{2+}]_c$, but not diastolic $[\text{Ca}^{2+}]_c$, in STZ-treated rat cardiomyocytes, however, this effect was absent in control rat cardiomyocytes. These changes were observed at $t = 5$ min and were absent after 1–3 h of incubation with 1 μM Dapa (Hamouda et al., 2014). These data suggest that Dapa may confer acute negative inotropic effects in the diabetic cardiomyocyte. In support of this, Empa (0.25–1 μM) reduced $[\text{Na}^+]_c$, diastolic and systolic $[\text{Ca}^{2+}]_c$, and impaired the NHE-1 activity in healthy rabbit cardiomyocytes (Baartscheer et al., 2017). In that study, Empa also increased $[\text{Ca}^{2+}]_m$ in rat cardiomyocytes. This finding may reflect improved mitochondrial capacity to synthesize ATP and target oxidants, which would be beneficial to restore the energetic state of myocytes that is known to be decreased in heart failure. The lowering of $[\text{Na}^+]_c$ by Empa was observed at 5 mM and 11 mM glucose incubations and inhibition of NHE-1 occurred in the presence and absence of extracellular glucose, supporting the notion that glucose and SGLT's were not involved in these direct Empa effects. In addition, we recently showed that Dapa (1 μM) and Cana (3 μM) also reduced $[\text{Na}^+]_c$ and impaired NHE-1 activity in mouse cardiomyocytes (Uthman et al., 2018a). Further prove for SGLT2i interaction with NHE-1 was obtained by molecular binding studies, showing SGLT2i to exhibit high binding affinities with the extracellular Na^+ -binding site of the NHE (Uthman et al., 2018a). These data indicate that the SGLT2i exert an off-target effect on the NHE-1.

Mitochondrial Function

Empa increased cell viability and preserved ATP levels following hypoxia/reoxygenation in cultured H9C2 embryonic heart-derived cells (Andreadou et al., 2017). These effects were equally present in myocytes stimulated by AGE, and Empa did not change the expression level of RAGE (receptor for AGE), suggesting that these pro-survival mechanisms of Empa were not mediated through AGE/RAGE signaling. In mitochondria derived from cardiac muscle fibers obtained from isolated IR rat hearts, Empa increased complex II respiration, and permeabilized and uncoupled the inner mitochondrial membrane (Jespersen et al., 2017). The authors discuss that these mitochondrial changes may improve remodeling following IR conditions.

Expression of SGLT's

Although the SGLT2 has not been detected in cardiomyocytes and the heart, SGLT1 appears to be highly expressed in human, rat and mouse hearts and cardiomyocytes, and may even be upregulated in ischemic, hypertrophic, failing and diabetic hearts (Banerjee et al., 2009; Kashiwagi et al., 2015; Lambert et al., 2015; Vrhovac et al., 2015; Di Franco et al., 2017; Van Steenbergen et al., 2017). Currently, the relationship between SGLT2i and SGLT1 in failing and diabetic hearts has not been investigated.

Overall, SGLT2i directly affect cardiomyocytes by reducing $[\text{Na}^+]_c$ and $[\text{Ca}^{2+}]_c$, by inhibition of NHE-1, and by increasing mitochondrial function and cell viability (Figure 2). The exact membrane receptors/transporters mediating these effects in cardiomyocytes have not yet been identified, with the exemption of NHE-1 and LTCC.

SGLT2 INHIBITORS IN VASCULAR CELLS

Vascular function may be directly affected by SGLT2i through the changing of endothelial or vascular smooth muscle cell (SMC) function. Various reports have investigated SGLT2i effects on AMPK activity, mitochondrial function, vasodilation and on the expression of adhesion molecules in hyperglycemic and inflamed vascular cells (Figure 3).

AMPK Activity

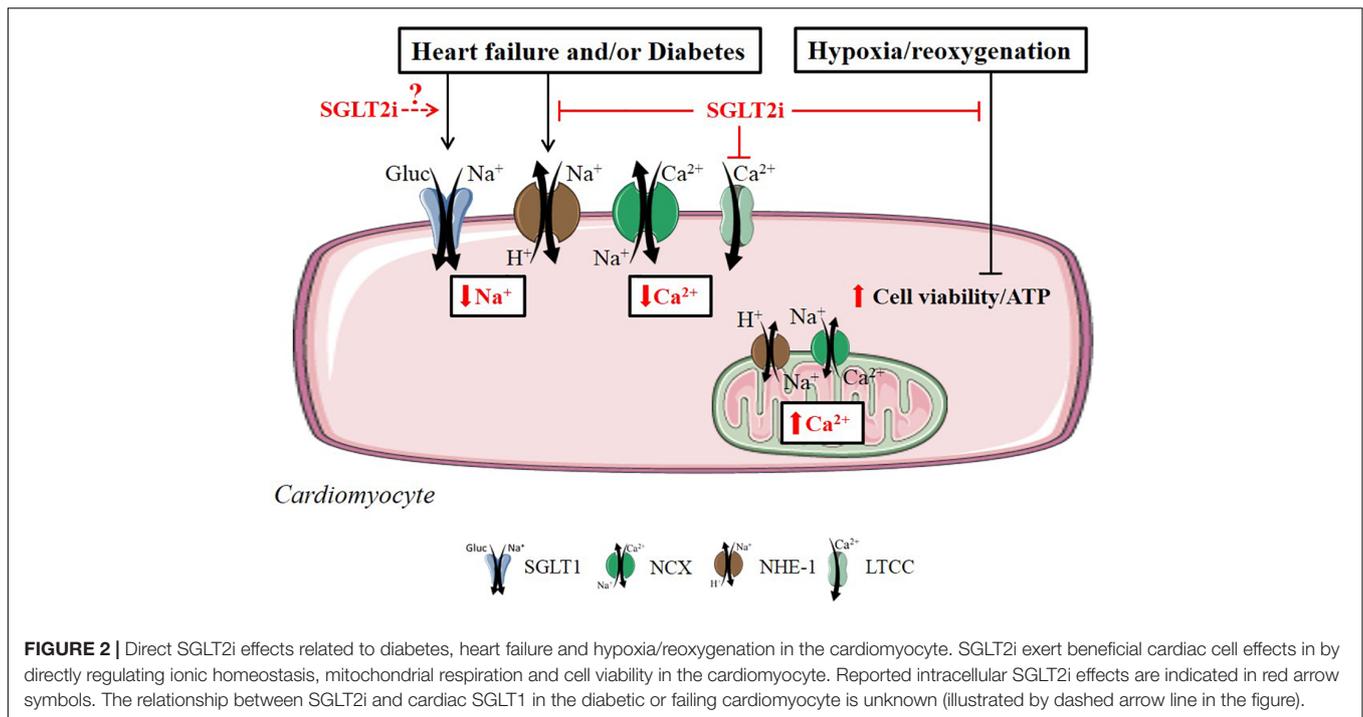
Cell studies with SGLT2i in various disease models have generally shown that SGLT2i improve vascular function and/or reduce markers of atherogenesis, by directly acting on vascular cells. Increased AMPK activity was observed in intact HUVEC and HAEC, after 15–30 min treatment with Cana (≥ 10 μM) but not with Empa or Dapa (Mancini et al., 2018). Similarly, phosphorylation of ACC as an indicator of AMPK activation was increased acutely with 10 μM Cana in cultured human aortic vascular SMC (Mancini et al., 2018).

Cell Viability and Mitochondrial Function

In HUVEC subjected to hypoxia/reoxygenation, Empa increased cell viability and preserved ATP levels in the presence and absence of AGE (Andreadou et al., 2017). In healthy cells, viability remained unaltered with 24 h Empa or Dapa treatment, but was modestly reduced with Cana treatment, which is suggested to be caused by reduced glucose uptake causing compromised ATP synthesis (Mancini et al., 2018). Empa (1 μM) did prevent mitochondrial dysfunction in hyperglycemic mouse aortic rings, i.e., increased oxygen consumption rates and reduced proton leakage (El-daly et al., 2018). It is uncertain, however, whether Empa would affect the vasculature at physiologically more relevant hyperglycemic glucose concentrations (<20 mM). Together, these data suggest that SGLT2i directly improve cell viability under conditions of high glucose, and that this may be due to mitochondrial alterations.

Vasodilation

The effects of Dapa on endothelial cell- and SMC- dependent vascular tone in various models have been studied. Acute Dapa



administration (1 nM–100 μ M) increased vasorelaxation in an endothelial cell-independent manner in healthy mouse aortic rings, while in a hyperglycemic setting (44 mM glucose) 1 h pre-incubation with 1 μ M Dapa restored relaxation without influencing SMC vasoreactivity (Gaspari et al., 2017). Acute treatment with high dose Cana (10 μ M) caused vascular relaxation in pulmonary, but not coronary artery rings from single STZ-injected mice (Han et al., 2015), indicating vascular bed specific effects of SGLT2i. Pre-treatment with 0.5–1 μ M Empa, 50 nM Dapa or 50 nM Cana for 24 h resulted in a stronger vasorelaxation response in hyperglycemic (25 mM glucose) mice aortic rings following application of acetylcholine or a proteinase-activated receptors 2 agonist (El-daly et al., 2018). Likewise, exposure of 1 μ M Empa for 3 days promoted eNOS activity (nitrite formation) in hyperglycemic HUVEC, yet cell density was not restored at this physiologically relevant Empa dosage (Steven et al., 2017), suggesting that the functional improvement by Empa did not directly correlate to increased cell viability.

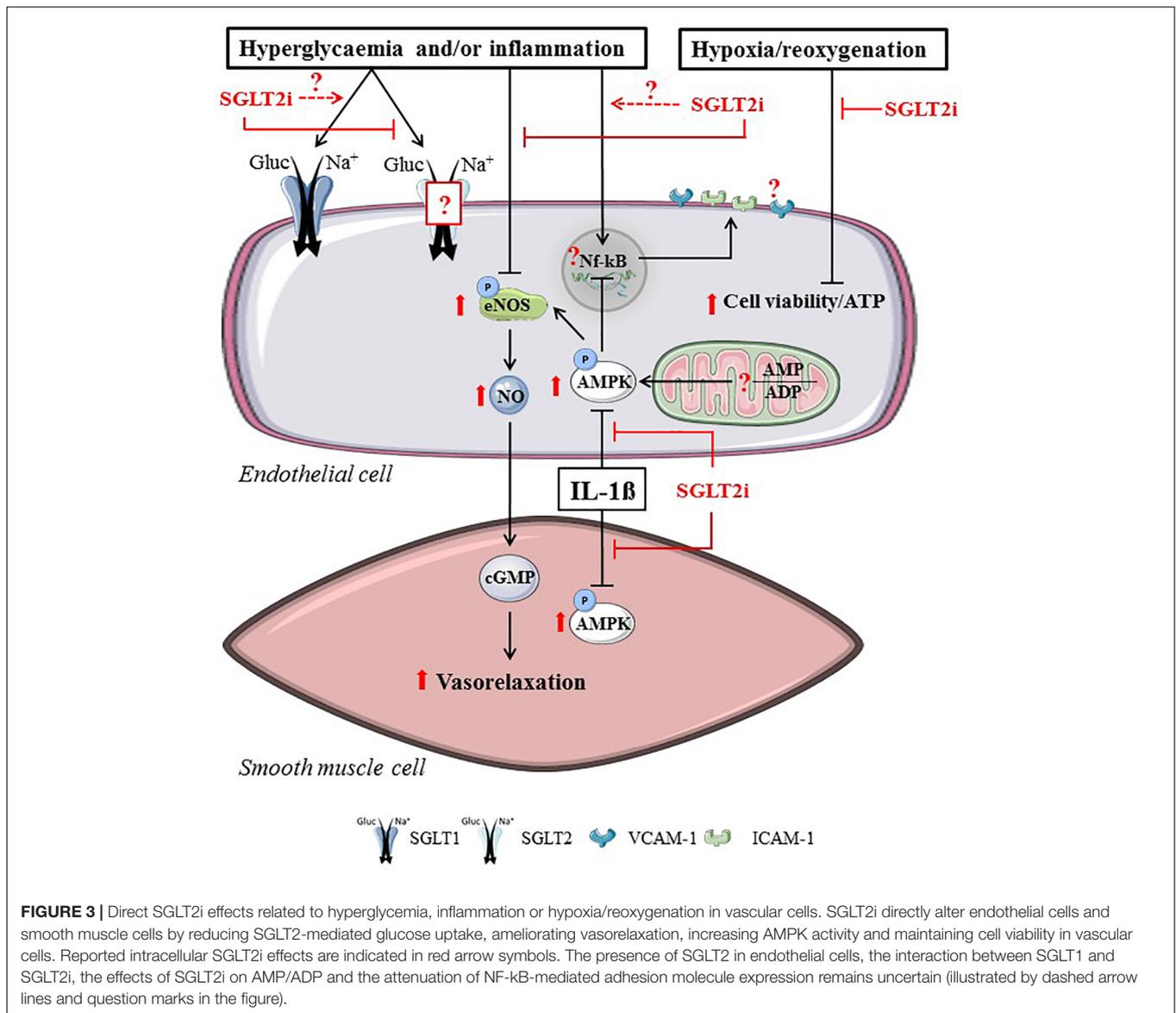
Cell Adhesion

In IL-1 β -stimulated HUVEC and HAEC, 10 μ M Cana attenuated secretion of MCP-1 and IL-6, which was at least partly associated with increased AMPK activity (Mancini et al., 2018). Dapa treatment (1–1000 nM) resulted in lower ICAM-1 and VCAM-1 protein levels and NF κ B mRNA expression in HUVEC subjected to TNF- α or to a hyperglycemic insult. However, low dose Dapa (\leq 10 nM), but not 100 nM and 1 μ M Dapa, seemed to reduce the inflammatory responses from hyperglycemia or TNF- α (Gaspari et al., 2017). Knowing that the maximal plasma concentrations of Dapa healthy volunteers taking 10 mg Dapa fluctuates between ~6–300 nM (Kasichayanula et al., 2011), it raises the possibility

of Dapa having anti-inflammatory actions in the T2D and/or heart failure patient. Contrary to Dapa application, Mancini et al. (2018) reported that adhesion molecule expression, JNK and NF κ B signaling of LPS-stimulated HAEC were not altered after Cana administration (Mancini et al., 2018), although adhesion to pro-monocytic cells was reduced. Our own results indicated that SGLT2i pre-incubation does not prevent ICAM and VCAM expression and the increase in permeability in TNF- α stimulated HUVEC and human coronary artery endothelial cells [HCAEC, (Uthman et al., 2018b)].

Expression of SGLT's

As several studies reported absence of SGLT2 expression in the heart (Han et al., 2015; Vrhovac et al., 2015; Di Franco et al., 2017; Van Steenbergen et al., 2017), SGLT2 may be excluded as a possible explanation for the direct vascular effects of SGLT2i. However, others have proposed that the SGLT2 mRNA is ubiquitously expressed in most human tissue (Zhou et al., 2003) and that the SGLT2 protein is also present in endothelial cells (El-daly et al., 2018; Li et al., 2018). In HUVEC, both SGLT1 and SGLT2 expressions were demonstrated at the protein and mRNA level and their quantities were elevated in the presence of PA, which also resulted in reduced Akt, IRS-1 and eNOS phosphorylation and nitric oxide concentration (Li et al., 2018). PA-induced endothelial dysfunction was partially restored with administration of the classic SGLT inhibitor phlorizin, and knockdown of SGLT1 or SGLT2. Additionally, Empa reduced glucose uptake in cultured endothelial cells similar to SGLT1/2 inhibitor sotagliflozin, measured by reduced glucose analog 2-NBDG uptake (El-daly et al., 2018). So far, no direct target of SGLT2i on endothelial cells has been identified, although the



presence of SGLT1 and SGLT2 in endothelial cells might be connected to the direct vascular effects summarized above.

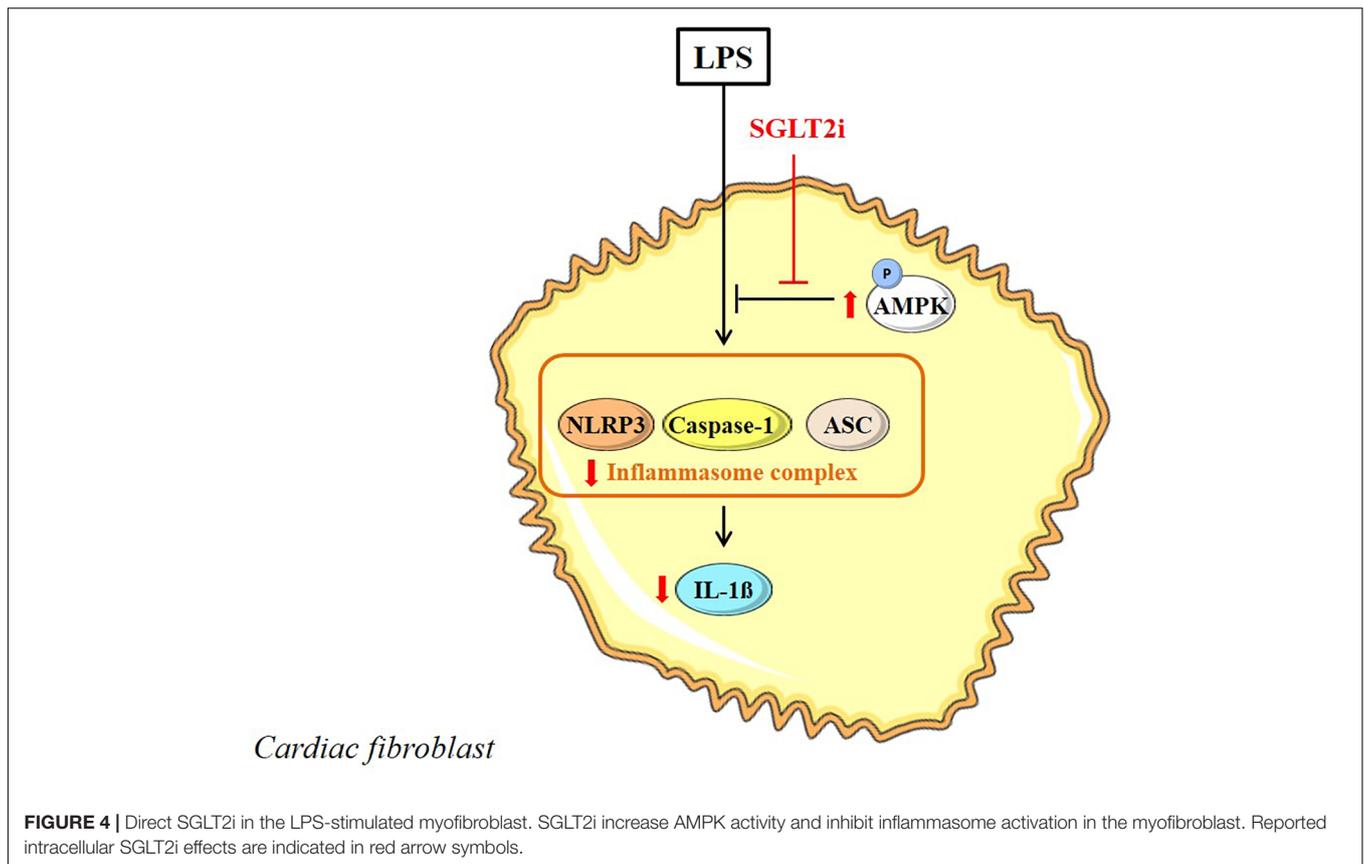
SGLT2 INHIBITORS IN CARDIAC FIBROBLASTS

Cardiac fibroblasts are valuable targets for therapeutic applications due to their role in cardiac remodeling after MI. Ye et al. (2017) studied SGLT2i effects in mouse cardiac fibroblasts stimulated with LPS. Pre-incubation with Dapa (0.3–0.5 μM) showed attenuation of LPS-induced upregulation of NLRP3, ASC, IL-1β and caspase-1 mRNA levels (Ye et al., 2017). Administration of phlorizin (100 μM) did not exhibit similar responses as Dapa and this indicates that SGLT-independent mechanisms were affected by Dapa. Effects of Dapa on NLRP3, TNF-α and caspase-1 were similar to A769662, an AMPK

activator, while co-administration of Dapa with Compound C, an AMPK inhibitor, abrogated the anti-inflammatory effects, including phosphorylation of AMPKα at Thr172. Thus, Dapa induces anti-inflammatory responses in myofibroblasts, mediated through increased AMPK activation without the involvement of SGLT (Figure 4).

SGLT2 INHIBITORS IN ISOLATED HEARTS

Data retrieved from isolated heart studies indicate that SGLT2i have direct cardiac effects at organ level, i.e., acute vasodilation, delayed ischemic contracture onset and increased post-ischemic P-STAT3 expression. The degree of IR injury and vasodilation, however, relies upon the concentration administered as well as the type of SGLT2i.



Vasodilation

Several studies have addressed the effects of SGLT2i in the isolated perfused heart. Acute coronary vasodilation occurs in intact mouse hearts following Empa (1 μM) or Cana (3 μM), but not after Dapa (1 μM) (Uthman et al., 2018a). These findings may reflect the differences in drug efficacy since Dapa did show a trend toward vasodilation, which may suggest that a higher concentration of Dapa could induce vasodilation in the healthy heart.

Ischemia-Reperfusion

Empa (4.75 μM) administration prior to ischemia did not change infarct size after 40 min ischemia and 30 min reperfusion in the isolated rat heart (Jespersen et al., 2017). However, during a 25 min ischemia and 2 h reperfusion protocol in isolated mouse hearts, Empa (1–10 μM) delayed ischemic contracture development similar to the effects of a classic NHE-1 inhibitor Cariporide. Reperfusion injury remained unaltered or even worsened for the Empa-treated hearts under these conditions (Uthman et al., 2018c). These effects may indicate an ATP-sparing effect of SGLT2i that is caused by inhibition of NHE-1 during ischemia, although NHE-1 inhibition during reperfusion may differ between Empa and Cariporide. Furthermore, the detrimental outcomes of high-dosage Empa (higher than the maximal concentration observed in clinical trials), may possibly refer to potential cell toxic effects of this drug used at a

higher concentration during IR injury. Furthermore, Empa did not change mitochondrial calcium retention capacity in mitochondria isolated from mouse hearts (Andreadou et al., 2017). This suggests that Empa does not directly affect the mitochondrial permeability transition pore, which is in line with findings reflecting that Empa does not protect the isolated heart against IR injury (Jespersen et al., 2017; Uthman et al., 2018c).

Fibrosis

Perfusion with a high dose of Dapa (10 μM) for 60 min increased STAT3 phosphorylation in the remote area of post-ischemic rat hearts (Lee et al., 2017). Increased p-STAT3 levels were associated with higher IL-10 levels in Dapa treated hearts, which is suggested to facilitate the conversion of macrophages from an inflammatory M1 to anti-inflammatory M2 phenotype and therefore inhibit myofibroblast differentiation and extracellular matrix formation.

DISCUSSION

This review provides an overview of the direct effects of SGLT2i in the heart and in the most prominent cell types present in the heart, including cardiomyocytes, endothelial cells, vascular SMC and cardiofibroblasts. **Figures 2–4** graphically summarize the described effects of SGLT2i in these cardiac cell types. They show that SGLT2i positively affect the Na^+ and Ca^{2+} homeostasis, inhibit NHE-1 activity, improve mitochondrial

respiration, and increase post-hypoxic ATP levels and cell viability in cardiomyocytes. In vascular cells, SGLT2i can directly improve vasorelaxation, increase AMPK activity, eNOS phosphorylation and, although inconsistently, adhesion molecule expression in hyperglycemic and/or inflamed conditions. In LPS-treated cardioblasts, SGLT2i exert anti-inflammatory actions similar to AMPK activation pathways. Finally, in isolated hearts SGLT2i improved vasodilation, delayed ischemic contracture and increased post-ischemic STAT3 phosphorylation. These preclinical cell/organ studies clearly show that there are direct cardiac effects of SGLT2i, offering several off-target, cardioprotective mechanisms that may contribute to the beneficial effects observed in the clinical trials.

Ionic Homeostasis in Diabetes and Heart Failure

A growing number of pre-clinical studies have demonstrated beneficial cardiovascular effects of SGLT2i independent of their systemic glucose-lowering actions (Andreadou et al., 2017; Baartscheer et al., 2017; Byrne et al., 2017; Ye et al., 2017; Uthman et al., 2018a). In light of these results, the direct lowering of $[Na^+]_c$ by SGLT2i may constitute part of the beneficial effects of SGLT2i through attenuation of inflammation, endothelial dysfunction, oxidative stress and cardiac remodeling.

To reduce the global cardiovascular risk, the World Health Organization advises to lower daily salt intake within the population to <5 g (WHO, 2012). High salt (i.e., Na^+) intake evidently predicts the development of heart failure and diabetes (He and MacGregor, 2018) as well as an increased incidence of T2D in individuals with high caloric intake (Lanaspa et al., 2018). Na^+ loading contributes to elevated oxidative stress and activation of the NLRP3 inflammasome (inflammation), consequently leading to the development of diabetes in humans (Wan et al., 2018). In endothelial cells, a mild increase of extracellular Na^+ led to elevated levels of adhesion molecules and the transmigration of peripheral blood mononuclear cells (Dmitrieva and Burg, 2015). An exact molecular mechanism of Na^+ loading that causes inflammation has not been experimentally determined, although it has been reported that the efflux of K^+ via the Na^+/K^+ -ATPase is a necessary step that triggers NLRP3 activation (Muñoz-Planillo et al., 2013). Increased Na^+/K^+ -ATPase through elevated extracellular Na^+ may therefore cause K^+ efflux and activate NLRP3. Notably, increased extracellular Na^+ also led to activation of NLRP3 independent of K^+ efflux (Muñoz-Planillo et al., 2013). Taken together, extracellular Na^+ is an important determinant of inflammation and may therefore be relevant in targeting cardiovascular disorders.

A modest increase in extracellular glucose (from 5.5 to 11 mM) acutely elevated $[Na^+]_c$ in isolated cardiomyocytes (Baartscheer et al., 2017). Chronic hyperglycemia, as occurring in T2D and heart failure, may thus at least partly be accountable for the increase in cardiac $[Na^+]_c$. Increased $[Na^+]_c$ at a steady extracellular glucose concentration was recently observed in T2D rat cardiomyocytes, which could be ascribed to elevated Na^+ -glucose cotransport through increased expression of SGLT1

(Lambert et al., 2015). Cardiac SGLT1 expression is increased in conditions of T2D and heart failure, in both animal and man (Banerjee et al., 2009; Lambert et al., 2015; Di Franco et al., 2017). Thus, $[Na^+]_c$ in cardiomyocytes can be increased due to increased extracellular glucose levels as well as increased SGLT1 expression.

Elevation of $[Na^+]_c$ results in a secondary rise of $[Ca^{2+}]_c$ via the Na^+/Ca^{2+} -exchanger (NCX) (Baartscheer et al., 2003a). Studies support the contention that increases in $[Na^+]_c$ and $[Ca^{2+}]_c$ induce signaling cascades that lead to dysregulation of mitochondrial homeostasis, including impaired energetics and elevated ROS production, and as such reduced cardiac hypertrophy and remodeling [reviewed in detail elsewhere: (Bay et al., 2013)].

The reduction in $[Ca^{2+}]_c$ by SGLT2i appears to be directly correlated to the lowering of $[Na^+]_c$. The $[Na^+]_c$ lowering effects of SGLT2i can be explained by inhibition of NHE-1. Cardiomyocytes express high amounts of NHE-1 (Yokoyama et al., 2000), which is also of paramount importance for pH regulation during pathological conditions, including diabetes, IR injury and heart failure (ten Hove et al., 2003; Packer, 2017). Preclinical studies have shown that pharmacological interventions targeting sarcolemmal sodium ion transporters proved effective in ameliorating heart failure (Kusumoto et al., 2001; Engelhardt, 2002; Baartscheer et al., 2003b, 2005, 2008; Baartscheer and Van Borren, 2008). However, these promising preclinical studies in the setting of HF on the use of NHE-1 inhibition never translated into clinical testing for HF, because of negative results obtained with the use of these inhibitors in the setting of acute cardiac ischemia. Clinical trials on the short-term use (<7 days) of NHE-1 inhibitors in the setting of acute MI in CABG and PTCA patients showed a reduction in the incident of non-fatal MI without an overall benefit on cardiovascular mortality. These clinical studies were halted due to the occurrence of an increased incidence in cerebrovascular events in these acute cardiovascular conditions following treatment with the NHE-1 inhibitors [Control vs. NHE inhibitor; EXPEDITION 3.0% vs. 5.2% ($p < 0.001$); GUARDIAN-trial 1.0% vs. 1.5%; ESCAMI-trial 0.2 vs. 2.2% (not significant)] (Théroux et al., 2000; Zeymer et al., 2001; Mentzer et al., 2008). In addition, the optimal dosage, timing and duration of the intervention and the patient populations that could most likely benefit from NHE-1 inhibitors were insufficiently investigated by phase 2 clinical trials, which may explain the divergence between the highly promising preclinical results of NHE-1 inhibitors and the ambiguity hereof in the known clinical trials. Furthermore, it is known that mild acidosis, but not severe acidosis, may be protective against ischemia in neuronal tissue (Vornov et al., 1996). Most importantly, to date no clinical trial has investigated the effects of using NHE-1 inhibitors as chronic therapy for the treatment of heart failure. Possibly, the risk of stroke may be less pronounced or even absent if NHE-1 inhibitors are used chronically, not immediately halted, used at milder dosages and in combination with platelet inhibitors. Therefore, the clinical efficacy of inhibiting NHE-1 and lowering $[Na^+]_c$ in different chronic cardiac disorders remains to be tested.

Previous experiments with Empa have shown that NHE-1 inhibition also occurred in glucose-free conditions, thus eliminating SGLT's as mediators for the lowering of $[Na^+]_c$ (Baartscheer et al., 2017). Whether SGLT2i also inhibit NHE-1 activity in diabetic cardiomyocytes has not yet been investigated, although increased activity of NHE-1, accompanied by higher $[Ca^{2+}]_c$ and hypertrophy has been observed in the diabetic myocardium (Darmellah et al., 2007). Others have reported elevated NHE-1 expression in glucose-induced hypertrophy in isolated rat cardiomyocytes, which may be prevented by treatment with Cariporide (Chen et al., 2007).

In diabetic and failing hearts, the cellular energetics (PCr/ATP) are impaired (Beer et al., 2002; Levelt et al., 2016). ATP generation is directly linked to $[Ca^{2+}]_m$ as it determines the regulation of dehydrogenases, generation of reducing equivalents for oxidative phosphorylation, ΔG_{ATP} and the inner mitochondrial membrane potential (Glancy and Balaban, 2012). It has been demonstrated that $[Ca^{2+}]_m$ is reduced when $[Na^+]_c$ is elevated from 5 to 15 mM as the exchange of $[Ca^{2+}]_m$ and $[Na^+]_c$ through the mitochondrial NCX is raised (Liu and Rourke, 2008). Conversely, Empa led to elevated $[Ca^{2+}]_m$ in myocytes (Baartscheer et al., 2017), probably because of reduced exchange of $[Na^+]_c$ for $[Ca^{2+}]_m$ due to lower $[Na^+]_c$. The increased $[Ca^{2+}]_m$ is likely to increase cardiac energetics resulting in an increased PCr/ATP ratio (Bertero and Maack, 2018). Although in healthy hearts SGLT2i were without effect on PCr/ATP levels (Uthman et al., 2018a), it cannot be excluded that SGLT2i will increase PCr/ATP levels in diabetic and failing myocytes, especially since it is already observed that ATP levels are preserved in cultured cardiomyocytes subjected to hypoxia/reoxygenation (Andreadou et al., 2017).

AMPK Activation to Improve Energetics and Prevent Inflammation

5' AMPK is a regulator of cardiac energy metabolism. Given that myocardial energetic status is severely compromised in failing (Neubauer, 2007; Ingwall, 2009) and diabetic (Levelt et al., 2016) hearts, AMPK activation could enhance cardiac energy metabolism and hence restore myocardial energy levels (Beauloye et al., 2011). Furthermore, activation of AMPK reduced inflammation and oxidative stress levels in PA-stimulated endothelial cells (Li et al., 2015). Cana in endothelial cells and Dapa in myofibroblasts activated AMPK, resulting in reduced inflammatory responses in these cells (Ye et al., 2017; Mancini et al., 2018). A possible mechanism by which SGLT2i change AMPK activity is through complex I inhibition and increased AMP/ADP ratio, which was examined in mouse hepatocytes (Hawley et al., 2016). However, this effect was only observed with high doses of Cana, and not with Empa or Dapa. One may also postulate that the inhibition of glucose uptake by SGLT2 inhibition may activate AMPK as a compensatory feedback mechanism to restore cell metabolism. Limited data are available as of yet that explains how SGLT2i could activate AMPK. Further research with, e.g., knock-down models of SGLT2 could elaborate on the cellular mechanism of SGLT2i to directly activate AMPK.

Whether the SGLT2i convey through similar AMPK activation pathways in cardiac cells needs further investigation. Based on the existing research, lower dosages of Dapa ($\leq 0.5 \mu M$) seemed to be most effective for AMPK activation and the suppression of inflammation, while higher Cana concentrations activated AMPK and reduced inflammatory responses in AMPK-dependent and -independent manners. These results imply that the different SGLT2i activate AMPK at different concentrations.

Inflammation and Oxidative Stress

Inflammation is considered an essential driving factor of cardiovascular disease in diabetes, whereas elevated levels of extracellular glucose alone is insufficient to induce a pro-atherogenic state (Sharma et al., 2018). Instead, the combination of glucotoxic and inflammatory stimuli are needed to establish an environment prone to developing atherosclerosis (Azcutia et al., 2010). Current understandings of the pathophysiology of heart failure, in particular with preserved ejection fraction (HFpEF) postulate that the presence of comorbidities cause microvascular inflammation that ultimately leads to the development of heart failure (Paulus and Tschöpe, 2013). Importantly, vascular inflammation can be induced by reduced AMPK activity (He et al., 2015) as well as high Na^+ loading, the latter resulting in elevation of fasting blood glucose levels, oxidative stress and insulin resistance (Wan et al., 2018). Endothelial inflammation then leads to perturbed NO-cGMP-PKG signaling and increased leukocyte trafficking, which subsequently induces cardiomyocyte hypertrophy and myofibroblast differentiation, and as such cardiac remodeling. Since SGLT2i have demonstrated direct anti-inflammatory actions in vascular cells, targeting this new paradigm may potentially and at least partly explain the positive results observed in the clinical trials and may identify novel therapeutic implications for SGLT2i.

A Window for SGLT2i as Anti-arrhythmic Therapy

In heart failure, cardiac arrhythmias primarily depend on triggered activity (Coronel et al., 2013). Elevated levels of diastolic $[Ca^{2+}]_c$ increase the open probability of the Ryanodine receptor resulting in spontaneous release of calcium from the SR. This in turn activates the NCX, leading to the transient inward current responsible for delayed after depolarizations (Baartscheer et al., 2003c). In addition, it has been shown that the number of after-transients measured in isolated myocytes is related to the occurrence of ventricular tachycardia *in vivo* (Janse et al., 2001). No data are available on the effects of SGLT2i on arrhythmogenesis. However, under pathologic conditions where myocardial $[Na^+]_c$ is increased, such as in heart failure, a reduction of $[Na^+]_c$ is associated with a reduction of $[Ca^{2+}]_c$, the number and amplitude of Ca^{2+} after-transients and the associated arrhythmias (Despa et al., 2002; Pogwizd et al., 2003; Bers, 2014). In the same vein of thought, we speculate that the inhibitory effect of SGLT2i on Na^+ influx during heart failure is antiarrhythmic.

Erickson et al. (2013) have documented that Ca^{2+} /calmodulin-dependent kinase II (CaMKII) is elevated in diabetes and that it is associated with increased Ca^{2+} -release events from the SR. Thus, diabetes directly impacts arrhythmogenesis. Recent work by Mustrup et al. (2018) has demonstrated that Empa reduced CaMKII activity and CaMKII-dependent SR Ca^{2+} leak. Thus, Empa is likely to directly reduce Ca^{2+} -release and arrhythmias. In addition, indirect effects of SGLT2i on arrhythmogenesis can be mediated through attenuation of the cardiac remodeling and hypertrophic phase known to occur in diabetes (Byrne et al., 2017).

The Presence of SGLT2 in Endothelial Cells?

Given that several studies have reported that SGLT2i directly reduced glucose uptake in endothelial cells, it may be possible that this occurred as a result of SGLT2 inhibition. Conflicting results regarding the expression of SGLT2 in the coronary vasculature have been reported. While SGLT2 has not been detected at all in the heart (Van Steenbergen et al., 2017), increasing evidence demonstrate the existence of SGLT2 in non-cardiac endothelial cells (El-daly et al., 2018; Li et al., 2018). Existing data that suggested the absence of SGLT2 in the endothelium had only attempted to detect SGLT2 at mRNA levels. However, more recent studies have postulated the presence of SGLT2 in endothelial cells at protein level and that its expression levels are amendable by exposure to SGLT2-specific siRNA or PA (El-daly et al., 2018; Li et al., 2018), providing further support for a novel mechanism of SGLT2 and its inhibitors in vascular cells. A major limitation for the identification of SGLT2 at protein level is the lack of knowledge on the quality and specificity of the antibodies used. Therefore, further development of these techniques are highly warranted. Whether diabetic or failing conditions may evoke SGLT2 expression in cardiac endothelial cells has so far not been investigated.

SMIT-1 as Potential Target for SGLT2i

Another possible target for SGLT2i is the sodium-myoinositol cotransporter 1 (SMIT-1), a member of the SGLT receptor family. Overexpression of SMIT-1 leads to activation of NOX2, ROS production and increased glucose sensitivity in cardiomyocytes, and its deletion triggered opposite effects (Van Steenbergen et al., 2017). The effects of increased SMIT-1 expression were not associated with increased glucose uptake with higher extracellular glucose concentration, and the authors suggest that SMIT-1 effects relate to extended glucose sensitization that could alter downward signaling events related to $[\text{Na}^+]_c$ and $[\text{Ca}^{2+}]_c$. Considering the $[\text{Na}^+]_c$ lowering effect of SGLT2i, SGLT2i targeting SMIT-1 during hyperglycemic condition to attenuate hyperglycemia-induced damage might be assumed. However, Baartscheer et al. (2017) showed Na^+ -lowering with Empa, even in the absence of glucose. Furthermore, the IC50 of Empa and Cana for SMIT-1 were estimated at 8.3 μM and 5.6 μM , respectively, which is far off the

C_{max} for Empa (0.6 μM) (Suzuki et al., 2012; Scheen, 2014).

Future Directions

Our current understanding is that cardiac $[\text{Na}^+]_c$ is raised in conditions of heart failure and diabetes, while SGLT2i cause the reduction of $[\text{Na}^+]_c$, through inhibition of NHE-1. Studies considering the impact of SGLT2i on NHE-1 activity, $[\text{Na}^+]_c$, $[\text{Ca}^{2+}]_c$, $[\text{Ca}^{2+}]_m$ and mitochondrial energetics in diabetic and failing cardiomyocytes are warranted. Furthermore, whether the reduction of inflammation observed with SGLT2i is a direct consequence of Na^+ lowering is as of yet unknown. It remains uncertain whether the SGLT2 is expressed in cardiac endothelial cells and, if so, is involved in the cardiac effects of SGLT2i. Likewise, since SGLT1 expression in cardiomyocytes is increased in diabetic, heart failure and hypertrophic conditions, the effect of SGLT2i on SGLT1 under these circumstances should be investigated. The identification of SGLT2 in the endothelium with existing techniques requires accurate validation. Moreover, SMIT-1 might be a valuable target to study in the field of heart failure and SGLT2i.

CONCLUSION

In conclusion, increased intracellular sodium concentration is an early hallmark and driver in the pathogenesis of heart failure and T2D. Besides other mechanisms, SGLT2i lower $[\text{Na}^+]_c$ in cardiomyocytes, activate AMPK in endothelial cells and cardiofibroblasts, and inhibit cellular glucose uptake in endothelial cells, to favorably interfere with $[\text{Ca}^{2+}]_c$ homeostasis, improve mitochondrial function, reduce inflammation and ROS production and restore nitric oxide formation. These effects may explain the beneficial effects of SGLT2i to prevent heart failure and other related cardiac complications in the diabetic as well as the non-diabetic heart.

AUTHOR CONTRIBUTIONS

LU conducted the literature review, drafted the article, provided critical revision of the article, and final approval of the version to be published. AB, CS, JF, MK, MH, RC, and NW provided critical revision of the article and final approval of the version to be published. CZ drafted the article, provided critical revision of the article, and final approval of the version to be published.

FUNDING

RC received funding from Leducq Transatlantic Network of Excellence, RHYTHM, 16CVD02.

REFERENCES

- Andreadou, I., Efentakis, P., Balafas, E., Togliatto, G., Davos, C. H., Varela, A., et al. (2017). Empagliflozin limits myocardial infarction in vivo and cell death in vitro: role of STAT3, mitochondria, and redox aspects. *Front. Physiol.* 8:1077. doi: 10.3389/fphys.2017.01077
- Azcutia, V., Abu-Taha, M., Romacho, T., Vázquez-Bella, M., Matesanz, N., Luscinskas, F. W., et al. (2010). Inflammation determines the pro-adhesive properties of high extracellular D-glucose in human endothelial cells in vitro and rat microvessels in vivo. *PLoS One* 5:e10091. doi: 10.1371/journal.pone.0010091
- Baatscheer, A., Hardziyenka, M., Schumacher, C. A., Belterman, C. N. W., Van Borren, M. M., Verkerk, A. O., et al. (2008). Chronic inhibition of the Na^+/H^+ -exchanger causes regression of hypertrophy, heart failure, and ionic and electrophysiological remodeling. *Br. J. Pharmacol.* 154, 1266–1275. doi: 10.1038/bjp.2008.189
- Baatscheer, A., Schumacher, C. A., Belterman, C. N. W., Coronel, R., and Fiolet, J. W. T. (2003a). $[\text{Na}^+]$ and the driving force of the $\text{Na}^+/\text{Ca}^{2+}$ -exchanger in heart. *Cardiovasc. Res.* 57, 986–995. doi: 10.1016/S0008-6363(02)00848-9
- Baatscheer, A., Schumacher, C. A., Van Belterman, M. M., Borren, C. N., Coronel, R., and Fiolet, J. W. T. (2003b). Increased Na^+/H^+ -exchange activity is the cause of increased $[\text{Na}^+]$ and underlies disturbed calcium handling in the rabbit pressure and volume overload heart failure model. *Heart Fail.* 57, 1015–1024. doi: 10.1016/S0008-6363(02)00809-X
- Baatscheer, A., Schumacher, C. A., Belterman, C. N. W., Coronel, R., and Fiolet, J. W. T. (2003c). S R calcium handling and calcium after-transients in a rabbit model of heart failure. *Heart Fail.* 58, 99–108. doi: 10.1016/S0008-6363(02)00854-4
- Baatscheer, A., Schumacher, C. A., Van Borren, M. M. G., Belterman, C. N. W., Coronel, R., Ophof, T., et al. (2005). Chronic inhibition of Na^+/H^+ -exchanger attenuates cardiac hypertrophy and prevents cellular remodeling in heart failure. *Cardiovasc. Res.* 65, 83–92. doi: 10.1016/j.cardiores.2004.09.024
- Baatscheer, A., Schumacher, C. A., Wüst, R. C. I., Fiolet, J. W. T., Stienen, G. J. M., Coronel, R., et al. (2017). Empagliflozin decreases myocardial cytoplasmic Na^+ through inhibition of the cardiac Na^+/H^+ exchanger in rats and rabbits. *Diabetologia* 60, 568–573. doi: 10.1007/s00125-016-4134-x
- Baatscheer, A., and Van Borren, M. M. (2008). Sodium ion transporters as new therapeutic targets in heart failure. *Cardiovasc. Hematol. Agents Med. Chem.* 6, 229–236. doi: 10.2174/187152508785909546
- Banerjee, S. K., McGaffin, K. R., Pastor-Soler, N. M., and Ahmad, F. (2009). SGLT1 is a novel cardiac glucose transporter that is perturbed in disease states. *Cardiovasc. Res.* 84, 111–118. doi: 10.1093/cvr/cvp190
- Bay, J., Kohlhaas, M., and Maack, C. (2013). Intracellular Na^+ and cardiac metabolism. *J. Mol. Cell. Cardiol.* 61, 20–27. doi: 10.1016/j.yjmcc.2013.05.010
- Beauloye, C., Bertrand, L., Horman, S., and Hue, L. (2011). AMPK activation, a preventive therapeutic target in the transition from cardiac injury to heart failure. *Cardiovasc. Res.* 90, 224–233. doi: 10.1093/cvr/cvr034
- Beer, M., Seyfarth, T., Sandstede, J., Landschütz, W., Lipke, C., Köstler, H., et al. (2002). Absolute concentrations of high-energy phosphate metabolites in normal, hypertrophied, and failing human myocardium measured noninvasively with 31P-SLOOP magnetic resonance spectroscopy. *J. Am. Coll. Cardiol.* 40, 1267–1274. doi: 10.1016/S0735-1097(02)02160-5
- Bers, D. M. (2014). Cardiac sarcoplasmic reticulum calcium leak: basis and roles in cardiac dysfunction. *Annu. Rev. Physiol.* 76, 107–127. doi: 10.1146/annurev-physiol-020911-153308
- Bertero, E., and Maack, C. (2018). Calcium signaling and reactive oxygen species in Mitochondria. *Circ. Res.* 122, 1460–1478. doi: 10.1161/CIRCRESAHA.118.310082
- Boudina, S., Sena, S., Theobald, H., Sheng, X., Wright, J. J., Hu, X. X., et al. (2007). Mitochondrial energetics in the heart in obesity-related. *Diabetes* 56, 2457–2466. doi: 10.2337/db07-0481.Additional
- Bugger, H., and Abel, E. D. (2014). Molecular mechanisms of diabetic cardiomyopathy. *Diabetologia* 57, 660–671. doi: 10.1007/s00125-014-3171-6
- Byrne, N. J., Parajuli, N., Levasseur, J. L., Boisvenue, J., Beker, D. L., Masson, G., et al. (2017). Empagliflozin prevents worsening of cardiac function in an experimental model of pressure overload-induced heart failure. *JACC Basic Transl. Sci.* 2, 347–354. doi: 10.1016/j.jacbs.2017.07.003
- Cassidy, S., Hallsworth, K., Thoma, C., MacGowan, G. A., Hollingsworth, K. G., Day, C. P., et al. (2015). Cardiac structure and function are altered in type 2 diabetes and Non-alcoholic fatty liver disease and associate with glycemic control. *Cardiovasc. Diabetol.* 14:23. doi: 10.1186/s12933-015-0187-2
- Cavender, M. A., Steg, P. G., Smith, S. C., Eagle, K., Ohman, E. M., Goto, S., et al. (2015). Impact of diabetes mellitus on hospitalization for heart failure, cardiovascular events, and death: outcomes at 4 years from the reduction of atherothrombosis for continued health (REACH) registry. *Circulation* 132, 923–931. doi: 10.1161/CIRCULATIONAHA.114.014796
- Chen, S., Khan, Z. A., Karmazyn, M., and Chakrabarti, S. (2007). Role of endothelin-1, sodium hydrogen exchanger-1 and mitogen activated protein kinase (MAPK) activation in glucose-induced cardiomyocyte hypertrophy. *Diabetes Metab. Res. Rev.* 23, 356–367. doi: 10.1002/dmrr.689
- Cordero, M. D., Williams, M. R., and Ryffel, B. (2018). AMP-activated protein kinase regulation of the NLRP3 inflammasome during aging. *Trends Endocrinol. Metab.* 29, 8–17. doi: 10.1016/j.tem.2017.10.009
- Coronel, R., Wilders, R., Verkerk, A. O., Wiegierinck, R. F., Benoist, D., and Bernus, O. (2013). Electrophysiological changes in heart failure and their implications for arrhythmogenesis. *Biochim. Biophys. Acta* 1832, 2432–2441. doi: 10.1016/j.bbdis.2013.04.002
- Cubbon, R. M., Adams, B., Rajwani, A., Mercer, B. N., Patel, P. A., Gherardi, G., et al. (2013). Diabetes mellitus is associated with adverse prognosis in chronic heart failure of ischaemic and non-ischaemic aetiology. *Diabetes Vasc. Dis. Res.* 10, 330–336. doi: 10.1177/1479164112471064
- Cui, G. M., Zhao, Y. X., Zhang, N. N., Liu, Z. S., Sun, W. C., and Peng, Q. S. (2013). Amiloride attenuates lipopolysaccharide-accelerated atherosclerosis via inhibition of NHE1-dependent endothelial cell apoptosis. *Acta Pharmacol. Sin.* 34, 231–238. doi: 10.1038/aps.2012.155
- Darmellah, A., Baetz, D., Prunier, F., Tamareille, S., Rücker-Martin, C., and Feuvray, D. (2007). Enhanced activity of the myocardial Na^+/H^+ exchanger contributes to left ventricular hypertrophy in the Goto-Kakizaki rat model of type 2 diabetes: critical role of Akt. *Diabetologia* 50, 1335–1344. doi: 10.1007/s00125-007-0628-x
- Despa, S. (2018). Myocyte $[\text{Na}^+]$ dysregulation in heart failure and diabetic cardiomyopathy. *Front. Physiol.* 9:1303. doi: 10.3389/fphys.2018.01303
- Despa, S., Islam, M. A., Weber, C. R., Pogwizd, S. M., and Bers, D. M. (2002). Intracellular Na^+ concentration is elevated in heart failure but Na/K pump function is unchanged. *Circulation* 105, 2543–2548. doi: 10.1161/01.CIR.0000016701.85760.97
- Despa, S., Lingrel, J. B., and Bers, D. M. (2012). Na^+/K^+ -ATPase 2-isoform preferentially modulates Ca_2^+ transients and sarcoplasmic reticulum Ca_2^+ release in cardiac myocytes. *Cardiovasc. Res.* 95, 480–486. doi: 10.1093/cvr/cvs213
- Dey, S., DeMazumder, D., Sidor, A., Foster, D. B., and O'Rourke, B. (2018). Mitochondrial ROS drive sudden cardiac death and chronic proteome remodeling in heart failure: novelty and significance. *Circ. Res.* 123, 356–371. doi: 10.1161/CIRCRESAHA.118.312708
- Di Franco, A., Cantini, G., Tani, A., Coppini, R., Zecchi-Orlandini, S., Raimondi, L., et al. (2017). Sodium-dependent glucose transporters (SGLT) in human ischemic heart: a new potential pharmacological target. *Int. J. Cardiol.* 243, 86–90. doi: 10.1016/j.ijcard.2017.05.032
- Dmitrieva, N. I., and Burg, M. B. (2015). Elevated sodium and dehydration stimulate inflammatory signaling in endothelial cells and promote atherosclerosis. *PLoS One* 10:e0128870. doi: 10.1371/journal.pone.0128870
- El Messaoudi, S., Nederlof, R., Zuurbier, C. J., van Swieten, H. A., Pickkers, P., Noyez, L., et al. (2015). Effect of metformin pretreatment on myocardial injury during coronary artery bypass surgery in patients without diabetes (MetCAB): a double-blind, randomised controlled trial. *Lancet Diabetes Endocrinol.* 3, 615–623. doi: 10.1016/S2213-8587(15)00121-7
- El-daly, M., Krishna, V., Venu, P., Mihara, K., Kang, S., Fedak, P. W. M., et al. (2018). Hyperglycaemic impairment of PAR2-mediated vasodilation: prevention by inhibition of aortic endothelial sodium-glucose-co-Transporter-2 and minimizing oxidative stress. *Vascul. Pharmacol.* 109, 56–71. doi: 10.1016/j.vph.2018.06.006

- Engelhardt, S. (2002). Inhibition of $\text{Na}^+\text{-H}^+$ exchange prevents hypertrophy, fibrosis, and heart failure in beta1-adrenergic receptor transgenic mice. *Circ. Res.* 90, 814–819. doi: 10.1161/01.RES.0000014966.97486.C0
- Erickson, J. R., Pereira, L., Wang, L., Han, G., Ferguson, A., Dao, K., et al. (2013). Diabetic hyperglycemia activates CaMKII and arrhythmias by O linked glycosylation. *Nature* 502, 372–376. doi: 10.1038/nature12537
- Feldstein, C. A. (2002). Salt intake and hypertension therapy. *J. Hum. Hypertens.* 16, 48–51. doi: 10.1038/sj/jhh/1001342
- Ferrannini, G., Hach, T., Crowe, S., Sanghvi, A., Hall, K. D., and Ferrannini, E. (2015). Energy balance after sodium–glucose cotransporter 2 inhibition. *Diabetes Care* 38, 1730–1735. doi: 10.2337/dc15-0355
- Foretz, M., Guigas, B., Berbrand, L., Pollak, M., and Viollet, B. (2014). Metformin: from mechanisms of action to therapies. *Cell Metab.* 20, 953–966. doi: 10.1016/j.cmet.2014.09.018
- Frati, G., Schirone, L., Chimenti, I., Yee, D., Biondi-Zoccai, G., Volpe, M., et al. (2017). An overview of the inflammatory signalling mechanisms in the myocardium underlying the development of diabetic cardiomyopathy. *Cardiovasc. Res.* 113, 378–388. doi: 10.1093/cvr/cvx011
- Garcia-Dorado, D., Ruiz-Meana, M., Inseste, J., Rodriguez-Sinovas, A., and Piper, H. M. (2012). Calcium-mediated cell death during myocardial reperfusion. *Cardiovasc. Res.* 94, 168–180. doi: 10.1093/cvr/cvs116
- Gaspari, T., Spizzo, I., Liu, H., Hu, Y., Simpson, R. W., Widdop, R. E., et al. (2017). Dapagliflozin attenuates human vascular endothelial cell activation and induces vasorelaxation: a potential mechanism for inhibition of atherosclerosis. *Diabetes Vasc. Dis. Res.* 15, 64–73. doi: 10.1177/1479164117733626
- Gélinas, R., Mailleux, F., Dontaine, J., Bultot, L., Demeulder, B., Ginion, A., et al. (2018). AMPK activation counteracts cardiac hypertrophy by reducing O-GlcNAcylation. *Nat. Commun.* 9:374. doi: 10.1038/s41467-017-02795-4
- Glancy, B., and Balaban, R. S. (2012). Role of mitochondrial Ca_2^+ in the regulation of cellular energetics. *Biochemistry* 51, 2959–2973. doi: 10.1021/bi2018909
- Hamouda, N. N., Sydorenko, V., Qureshi, M. A., Alkaabi, J. M., Oz, M., and Howarth, F. C. (2014). Dapagliflozin reduces the amplitude of shortening and Ca_2^+ transient in ventricular myocytes from streptozotocin-induced diabetic rats. *Mol. Cell. Biochem.* 400, 57–68. doi: 10.1007/s11010-014-2262-5
- Han, Y., Cho, Y.-E., Ayon, R., Guo, R., Youssef, K. D., Pan, M., et al. (2015). SGLT inhibitors attenuate NO-dependent vascular relaxation in the pulmonary artery but not in the coronary artery. *Am. J. Physiol. Lung Cell. Mol. Physiol.* 309, L1027–L1036. doi: 10.1152/ajplung.00167.2015
- Hawley, S. A., Ford, R. J., Smith, B. K., Gowans, G. J., Mancini, S. J., Pitt, R. D., et al. (2016). The Na^+ /glucose cotransporter inhibitor canagliflozin activates AMPK by inhibiting mitochondrial function and increasing cellular AMP levels. *Diabetes Metab. Res. Rev.* 65, 2784–2794. doi: 10.2337/db16-0058
- He, C., Li, H., Viollet, B., Zou, M. H., and Xie, Z. (2015). AMPK suppresses vascular inflammation in vivo by inhibiting signal transducer and activator of transcription-1. *Diabetes Metab. Res. Rev.* 64, 4285–4297. doi: 10.2337/db15-0107
- He, F. J., and MacGregor, G. A. (2018). Role of salt intake in prevention of cardiovascular disease: controversies and challenges. *Nat. Rev. Cardiol.* 15, 371–377. doi: 10.1038/s41569-018-0004-1
- Ingwall, J. S. (2009). Energy metabolism in heart failure and remodelling. *Cardiovasc. Res.* 81, 412–419. doi: 10.1093/cvr/cvn301
- Janse, M. J., Vermeulen, J. T., Opthof, T., Coronel, R., Wilms-Schopman, F. J., Rademaker, H. M. E., et al. (2001). Arrhythmogenesis in heart failure. *J. Cardiovasc. Electrophysiol.* 12, 496–499. doi: 10.1046/j.1540-8167.2001.00496.x
- Jespersen, N. R., Lassen, T. R., Hjortbak, M. V., Stottrup, N. B., and Botker, H. E. (2017). Sodium glucose transporter 2 (SGLT2) inhibition does not protect the myocardium from acute ischemic reperfusion injury but modulates post-ischemic mitochondrial function. *Cardiovasc. Pharmacol. Open Access.* 6, 2–4. doi: 10.4172/2329-6607.1000210
- Jia, G., Hill, M. A., and Sowers, J. R. (2018). Diabetic cardiomyopathy: an update of mechanisms contributing to this clinical entity. *Circ. Res.* 122, 624–638. doi: 10.1161/CIRCRESAHA.117.311586
- Joshi, M., Kotha, S. R., Malireddy, S., Selvaraju, V., Satoskar, A. R., Palesty, A., et al. (2014). Conundrum of pathogenesis of diabetic cardiomyopathy: role of vascular endothelial dysfunction, reactive oxygen species, and mitochondria. *Mol. Cell. Biochem.* 386, 233–249. doi: 10.1007/s11010-013-1861-x
- Kannel, W. B., Hjortland, M., and Castelli, W. P. (1974). Role of diabetes in congestive heart failure: the Framingham study. *Am. J. Cardiol.* 34, 29–34. doi: 10.1016/0002-9149(74)90089-7
- Karmazyn, M. (2013). NHE-1: still a viable therapeutic target. *J. Mol. Cell. Cardiol.* 61, 77–82. doi: 10.1016/j.yjmcc.2013.02.006
- Kashiwagi, Y., Nagoshi, T., Yoshino, T., Tanaka, T. D., Ito, K., Harada, T., et al. (2015). Expression of SGLT1 in human hearts and impairment of cardiac glucose uptake by phlorizin during ischemia-reperfusion injury in mice. *PLoS One* 10:e0130605. doi: 10.1371/journal.pone.0130605
- Kasichayanula, S., Liu, X., Zhang, W., Pfister, M., Reece, S. B., Aubry, A. F., et al. (2011). Effect of a high-fat meal on the pharmacokinetics of dapagliflozin, a selective SGLT2 inhibitor, in healthy subjects. *Diabetes Obes. Metab.* 13, 770–773. doi: 10.1111/j.1463-1326.2011.01397.x
- Kusumoto, K., Haist, J. V., and Karmazyn, M. (2001). Na^+ /H⁺ exchange inhibition reduces hypertrophy and heart failure after myocardial infarction in rats. *Am. J. Physiol. Heart. Circ. Physiol.* 280, H738–H745. doi: 10.1152/ajpheart.2001.280.2.H738
- Lambert, R., Srodulskic, S., Peng, X., Margulies, K. B., Despa, F., and Despa, S. (2015). Intracellular Na^+ concentration ($[\text{Na}^+]_i$) is elevated in diabetic hearts due to enhanced Na^+ -glucose cotransport. *J. Am. Heart Assoc.* 4, 1–11. doi: 10.1161/JAHA.115.002183
- Lanaspa, M. A., Kuwabara, M., Andres-Hernando, A., Li, N., Cicerchi, C., Jensen, T., et al. (2018). High salt intake causes leptin resistance and obesity in mice by stimulating endogenous fructose production and metabolism. *Proc. Natl. Acad. Sci. U.S.A.* 115:201713837. doi: 10.1073/pnas.1713837115
- Lastra, G., Dhuper, S., Johnson, M. S., and Sowers, J. R. (2010). Salt, aldosterone, and insulin resistance: impact on the cardiovascular system. *Nat. Rev. Cardiol.* 7, 577–584. doi: 10.1038/nrcardio.2010.123
- Lee, T. M., Chang, N. C., and Lin, S. Z. (2017). Dapagliflozin, a selective SGLT2 inhibitor, attenuated cardiac fibrosis by regulating the macrophage polarization via STAT3 signaling in infarcted rat hearts. *Free Radic. Biol. Med.* 104, 298–310. doi: 10.1016/j.freeradbiomed.2017.01.035
- Levent, E., Rodgers, C. T., Clarke, W. T., Mahmod, M., Ariga, R., Francis, J. M., et al. (2016). Cardiac energetics, oxygenation, and perfusion during increased workload in patients with type 2 diabetes mellitus. *Eur. Heart J.* 37, 3461–3469. doi: 10.1093/eurheartj/ehv442
- Li, C.-Y., Wang, L.-X., Dong, S.-S., Hong, Y., Zhou, X.-H., Zheng, W.-W., et al. (2018). Phlorizin exerts direct protective effects on palmitic acid (PA)-induced endothelial dysfunction by activating the PI3K/AKT/eNOS signaling pathway and increasing the levels of nitric oxide (NO). *Med. Sci. Monit. Basic Res.* 24, 1–9. doi: 10.12659/MSMBR.907775
- Li, J., Wang, Y., Wang, Y., Wen, X., Ma, X. N., Chen, W., et al. (2015). Pharmacological activation of AMPK prevents Drp1-mediated mitochondrial fission and alleviates endoplasmic reticulum stress-associated endothelial dysfunction. *J. Mol. Cell. Cardiol.* 86, 62–74. doi: 10.1016/j.yjmcc.2015.07.010
- Liu, T., and Rourke, B. O. (2008). Enhancing mitochondrial Ca_2^+ uptake in myocytes from failing hearts restores energy supply and demand matching. *Circ. Res.* 103, 279–288. doi: 10.1161/CIRCRESAHA.108.175919
- Luo, M., Guan, X., Luczak, E. D., Lang, D., Kutschke, W., Gao, Z., et al. (2013). Diabetes increases mortality after myocardial infarction by oxidizing CaMKII. *J. Clin. Invest.* 123, 1262–1274. doi: 10.1172/JCI65268
- Mancini, S. J., Boyd, D., Katwan, O. J., Strembitska, A., Almabrouk, T. A., Kennedy, S., et al. (2018). Canagliflozin inhibits interleukin-1 β -stimulated cytokine and chemokine secretion in vascular endothelial cells by AMP-activated protein kinase-dependent and -independent mechanisms. *Sci. Rep.* 8:5276. doi: 10.1038/s41598-018-23420-4
- Mentzer, R. M., Bartels, C., Bolli, R., Boyce, S., Buckberg, G. D., Chaitman, B., et al. (2008). Sodium-hydrogen exchange inhibition by cariporide to reduce the risk of ischemic cardiac events in patients undergoing coronary artery bypass grafting: results of the EXPEDITION study. *Ann. Thorac. Surg.* 85, 1261–1270. doi: 10.1016/j.athoracsur.2007.10.054
- Miki, T., Yuda, S., Kouzu, H., and Miura, T. (2013). Diabetic cardiomyopathy: pathophysiology and clinical features. *Heart Fail. Rev.* 18, 149–166. doi: 10.1007/s10741-012-9313-3
- Munasinghe, P. E., Riu, F., Dixit, P., Edamatsu, M., Saxena, P., Hamer, N. S. J., et al. (2016). Type-2 diabetes increases autophagy in the human heart through promotion of Beclin-1 mediated pathway. *Int. J. Cardiol.* 202, 13–20. doi: 10.1016/j.ijcard.2015.08.111

- Muñoz-Planillo, R., Kuffa, P., Martínez-Colón, G., Smith, B., Rajendiran, T., and Núñez, G. (2013). K⁺ efflux is the common trigger of NLRP3 inflammasome activation by bacterial toxins and particulate matter. *Immunity* 38, 1142–1153. doi: 10.1016/j.immuni.2013.05.016
- Münzel, T., Gori, T., Keaney, J. F., Maack, C., and Daiber, A. (2015). Pathophysiological role of oxidative stress in systolic and diastolic heart failure and its therapeutic implications. *Eur. Heart J.* 36, 2555–2564. doi: 10.1093/eurheartj/ehv305
- Murphy, E., and Eisner, D. A. (2009). Regulation of intracellular and mitochondrial sodium in health and disease. *Circ. Res.* 104, 292–303. doi: 10.1161/CIRCRESAHA.108.189050
- Mustroph, J., Wagemann, O., Lucht, C. M., Trum, M., Hammer, K. P., Martin, C., et al. (2018). Empagliflozin reduces Ca/calmodulin-dependent kinase II activity in isolated ventricular cardiomyocytes. *ESC Hear. Fail.* 5, 642–648. doi: 10.1002/ehf2.12336
- Neal, B., Perkovic, V., Mahaffey, K. W., de Zeeuw, D., Fulcher, G., Erondu, N., et al. (2017). Canagliflozin and cardiovascular and renal events in type 2 diabetes. *N. Engl. J. Med.* 377, 644–657. doi: 10.1056/NEJMoa1611925
- Neef, S., Dybkova, N., Sossalla, S., Ort, K. R., Fluschnik, N., Neumann, K., et al. (2010). CaMKII-dependent diastolic SR Ca₂⁺ leak and elevated diastolic Ca₂⁺ levels in right atrial myocardium of patients with atrial fibrillation. *Circ. Res.* 106, 1134–1144. doi: 10.1161/CIRCRESAHA.109.203836
- Neubauer, S. (2007). The failing heart — an engine out of fuel. *N. Engl. J. Med.* 356, 1140–1151. doi: 10.1056/NEJMra063052
- Oelze, M., Kröller-Schön, S., Welschof, P., Jansen, T., Hausding, M., Mikhed, Y., et al. (2014). The sodium-glucose co-transporter 2 inhibitor empagliflozin improves diabetes-induced vascular dysfunction in the streptozotocin diabetes rat model by interfering with oxidative stress and glucotoxicity. *PLoS One* 9:e112394. doi: 10.1371/journal.pone.0112394
- Packer, M. (2017). Activation and inhibition of sodium-hydrogen exchanger is a mechanism that links the pathophysiology and treatment of diabetes mellitus with that of heart failure. *Circulation* 136, 1548–1559. doi: 10.1161/CIRCULATIONAHA.117.030418
- Paulus, W. J., and Tschöpe, C. (2013). A novel paradigm for heart failure with preserved ejection fraction: comorbidities drive myocardial dysfunction and remodeling through coronary microvascular endothelial inflammation. *J. Am. Coll. Cardiol.* 62, 263–271. doi: 10.1016/j.jacc.2013.02.092
- Pogwizd, S. M., Sipido, K. R., Verdonck, F., and Bers, D. M. (2003). Intracellular Na in animal models of hypertrophy and heart failure: contractile function and arrhythmogenesis. *Cardiovasc. Res.* 57, 887–896. doi: 10.1016/S0008-6363(02)00735-6
- Riehle, C., and Abel, E. (2016). Insulin signaling and heart failure. *Circ. Res.* 118, 1151–1169. doi: 10.1161/CIRCRESAHA.116.306206
- Sasahara, T., Yayama, K., Matsuzaki, T., Tsutsui, M., and Okamoto, H. (2013). Na⁺/H⁺ exchanger inhibitor induces vasorelaxation through nitric oxide production in endothelial cells via intracellular acidification-associated Ca₂⁺ mobilization. *Vascul. Pharmacol.* 58, 319–325. doi: 10.1016/j.vph.2012.11.004
- Scheen, A. J. (2014). Evaluating SGLT2 inhibitors for type 2 diabetes: pharmacokinetic and toxicological considerations. *Expert Opin. Drug Metab. Toxicol.* 10, 647–663. doi: 10.1517/17425255.2014.873788
- Sena, C. M., Pereira, A. M., and Seica, R. (2013). Endothelial dysfunction - A major mediator of diabetic vascular disease. *Biochim. Biophys. Acta* 1832, 2216–2231. doi: 10.1016/j.bbdis.2013.08.006
- Shah, M. S., and Brownlee, M. (2017). Molecular and cellular mechanisms of cardiovascular disorders in diabetes. *Circ. Res.* 118, 1808–1829. doi: 10.1161/CIRCRESAHA.116.306923.Molecular
- Sharma, A., Tate, M., Mathew, G., Vince, J. E., Ritchie, R. H., and De Haan, J. B. (2018). Oxidative stress and NLRP3-inflammasome activity as significant drivers of diabetic cardiovascular complications: therapeutic implications. *Front. Physiol.* 9:114. doi: 10.3389/fphys.2018.00114
- Steven, S., Oelze, M., Hanf, A., Kröller-Schön, S., Kashani, F., Roohani, S., et al. (2017). The SGLT2 inhibitor empagliflozin improves the primary diabetic complications in ZDF rats. *Redox Biol.* 13, 370–385. doi: 10.1016/j.redox.2017.06.009
- Suzuki, M., Honda, K., Fukazawa, M., Ozawa, K., Hagita, H., Kawai, T., et al. (2012). Tofogliflozin, a potent and highly specific sodium/glucose cotransporter 2 inhibitor, improves glycaemic control in diabetic rats and mice. *J. Pharmacol. Exp. Ther.* 341, 692–701. doi: 10.1124/jpet.112.191593
- Tanajak, P., Sa-nguanmoo, P., Sivasinprasasn, S., Thummasorn, S., Siri-Angkul, N., Chattipakorn, S. C., et al. (2018). Cardioprotection of dapagliflozin and vildagliptin in rats with cardiac ischemia-reperfusion injury. *J. Endocrinol.* 236, 69–84. doi: 10.1530/JOE-17-0457
- ten Hove, M., van Emous, J. G., and van Echteld, C. J. A. (2003). Na⁺ overload during ischemia and reperfusion in rat hearts: comparison of the Na⁺/H⁺ exchange blockers EIPA, cariporide and eniporide. *Mol. Cell. Biochem.* 250, 47–54. doi: 10.1023/A:1024985931797
- Thérout, P., Chaitman, B. R., Danchin, N., Erhardt, L., Meinertz, T., Schroeder, J. S., et al. (2000). Inhibition of the sodium-hydrogen exchanger with cariporide to prevent myocardial infarction in high-risk ischemic situations. main results of the GUARDIAN trial. *Circulation* 102, 3032–3038. doi: 10.1161/01.CIR.102.25.3032
- Tian, R., Musi, N., D'Agostino, J., Hirshman, M. F., and Goodyear, L. J. (2001). Increased adenosine monophosphate-activated protein kinase activity in rat hearts with pressure-overload hypertrophy. *Circulation* 104, 1664–1669. doi: 10.1161/hc4001.097183
- Uthman, L., Baartscheer, A., Bleijlevens, B., Schumacher, C. A., Fiolet, J. W. T., Koeman, A., et al. (2018a). Class effects of SGLT2 inhibitors in mouse cardiomyocytes and hearts: inhibition of Na⁺/H⁺ exchanger, lowering of cytosolic Na⁺ and vasodilation. *Diabetologia* 61, 722–726. doi: 10.1007/s00125-017-4509-7
- Uthman, L., Homayr, A., Hollmann, M. W., Zuurbier, C. J., and Weber, N. C. (2018b). Administration of SGLT2 inhibitor empagliflozin against TNF- α induced endothelial dysfunction in human venous and arterial endothelial cells. *FASEB J.* 32:569.4.
- Uthman, L., Nederlof, R., Eerbeek, O., Baartscheer, A., Buchholtz, N., Coronel, R., et al. (2018c). Empagliflozin effects on ischemic contracture and I/R injury in isolated mouse hearts perfused with or without insulin. *FASEB J.* 32:lb292.
- Van Heerebeek, L., Hamdani, N., Handoko, M. L., Falcao-Pires, I., Musters, R. J., Kupreishvili, K., et al. (2008). Diastolic stiffness of the failing diabetic heart: importance of fibrosis, advanced glycation end products, and myocyte resting tension. *Circulation* 117, 43–51. doi: 10.1161/CIRCULATIONAHA.107.728550
- Van Steenberghe, A., Balteau, M., Ginion, A., Ferté, L., Battault, S., Ravenstein, C. D., et al. (2017). Sodium-myoinositol cotransporter-1, SMT1, mediates the production of reactive oxygen species induced by hyperglycemia in the heart. *Sci. Rep.* 7:41166. doi: 10.1038/srep41166
- Varma, U., Koutsifeli, P., Benson, V. L., Mellor, K. M., and Delbridge, L. M. D. (2018). Molecular mechanisms of cardiac pathology in diabetes – Experimental insights. *Biochim. Biophys. Acta* 1864, 1949–1959. doi: 10.1016/j.bbdis.2017.10.035
- Vornov, J. J., Thomas, A. G., and Jo, D. (1996). Protective effects of extracellular acidosis and blockade of sodium/hydrogen ion exchange during recovery from metabolic inhibition in neuronal tissue culture. *J. Neurochem.* 67, 2379–2389. doi: 10.1046/j.1471-4159.1996.67062379.x
- Vrhovac, I., Eror, D. B., Klessen, D., Burger, C., Breljak, D., Kraus, O., et al. (2015). Localizations of Na⁺-D-glucose cotransporters SGLT1 and SGLT2 in human kidney and of SGLT1 in human small intestine, liver, lung, and heart. *Pflügers Arch. Eur. J. Physiol.* 467, 1881–1898. doi: 10.1007/s00424-014-1619-7
- Wan, Z., Wen, W., Ren, K., Zhou, D., Liu, J., Wu, Y., et al. (2018). Involvement of NLRP3 inflammasome in the impacts of sodium and potassium on insulin resistance in normotensive Asians. *Br. J. Nutr.* 119, 228–237. doi: 10.1017/S0007114517002926
- Wang, S., Peng, Q., Zhang, J., and Liu, L. (2008). Na⁺/H⁺ exchanger is required for hyperglycaemia-induced endothelial dysfunction via calcium-dependent calpain. *Cardiovasc. Res.* 80, 255–262. doi: 10.1093/cvr/cvn179
- WHO (2012). *Guideline: Sodium Intake for Adults and Children*. Geneva: World Health Organization.
- Ye, Y., Bajaj, M., Yang, H. C., Perez-Polo, J. R., and Birnbaum, Y. (2017). SGLT-2 inhibition with dapagliflozin reduces the activation of the Nlrp3/ASC inflammasome and attenuates the development of diabetic cardiomyopathy in mice with type 2 diabetes. further augmentation of the effects with saxagliptin, a DPP4 inhibitor. *Cardiovasc. Drugs Ther.* 31, 119–132. doi: 10.1007/s10557-017-6725-2
- Yokoyama, H., Gunasegaram, S., Harding, S. E., and Avkiran, M. (2000). Sarcolemmal Na⁺/H⁺ exchanger activity and expression in human ventricular

- myocardium. *J. Am. Coll. Cardiol.* 36, 534–540. doi: 10.1016/S0735-1097(00)00730-0
- Zeymer, U., Suryapranata, H., Monassier, J. P., Opolski, G., Davies, J., Rasmanis, G., et al. (2001). The Na⁺/H⁺ exchange inhibitor eniporide as an adjunct to early reperfusion therapy for acute myocardial infarction. This manuscript is dedicated to the memory of Karl-Ludwig Neuhaus (1944–2000). *J. Am. Coll. Cardiol.* 38, E1644–E1650. doi: 10.1016/S0735-1097(01)01608-4
- Zhou, H., Wang, S., Zhu, P., Hu, S., Chen, Y., and Ren, J. (2018). Empagliflozin rescues diabetic myocardial microvascular injury via AMPK-mediated inhibition of mitochondrial fission. *Redox Biol.* 15, 335–346. doi: 10.1016/j.redox.2017.12.019
- Zhou, L., Cryan, E. V., D'Andrea, M. R., Belkowsky, S., Conway, B. R., and Demarest, K. T. (2003). Human cardiomyocytes express high level of Na⁺/glucose cotransporter 1 (SGLT1). *J. Cell. Biochem.* 90, 339–346. doi: 10.1002/jcb.10631
- Zinman, B., Wanner, C., Lachin, J. M., Fitchett, D., Bluhmki, E., Hantel, S., et al. (2015). Empagliflozin, cardiovascular outcomes, and mortality in type 2 diabetes. *N. Engl. J. Med.* 373, 2117–2128. doi: 10.1056/NEJMoa1504720

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Uthman, Baartscheer, Schumacher, Fiolet, Kuschma, Hollmann, Coronel, Weber and Zuurbier. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.