Community-Acquired Pneumonia in the Conjugate Vaccine Era

Derek J. Williams^{1,2} and Samir S. Shah^{3,4,5}

¹Division of Hospital Medicine, The Monroe Carell Jr Children's Hospital at Vanderbilt, and ²Department of Pediatrics, Vanderbilt University School of Medicine, Nashville, Tennessee; Divisions of ³Infectious Diseases and ⁴Hospital Medicine, Cincinnati Children's Hospital Medical Center, and ⁵Department of Pediatrics, University of Cincinnati College of Medicine, Ohio

Corresponding Author: Derek J. Williams, MD, MPH, 1161 21st Ave. South, CCC 5311 Medical Center North, Nashville, TN 37232. E-mail: derek.williams@vanderbilt.edu.

Received September 26, 2012; accepted October 5, 2012.

Community-acquired pneumonia (CAP) remains one of the most common serious infections encountered among children worldwide. In this review, we highlight important literature and recent scientific discoveries that have contributed to our current understanding of pediatric CAP. We review the current epidemiology of childhood CAP in the developed world, appraise the state of diagnostic testing for etiology and prognosis, and discuss disease management and areas for future research in the context of recent national guidelines.

Key words. Pneumonia; Review; Epidemiology; Diagnostics; Therapeutics

Defining the Disease

Risk Factors

Young age is perhaps the single most important risk factor for the development of pneumonia, and age is also an important predictor of etiology as well as outcomes. Most affected children in epidemiologic studies of pneumonia are less than 5 years of age. Disease incidence among this age group is higher than at any other age, including adults over 65 years [1]. Among older children, disease burden is substantially decreased and continues to decline through young adulthood. Other risk factors include poor nutritional status, low housing quality (eg, overcrowding, environmental exposures), secondhand smoke exposure, comorbidities such as asthma, and preceding upper respiratory tract infection [2–7].

Signs and Symptoms

Clinical signs and symptoms associated with pneumonia include fever, cough, tachypnea, grunting, retractions, hypoxia, abdominal pain, decreased breath sounds, and crackles or rales [8–16]. In the absence of at least 1 of these signs or symptoms, the likelihood of pneumonia is exceptionally low. Several definitions have been proposed to identify community-acquired pneumonia (CAP) [17]. The World Health Organization (WHO) criteria are widely used in the developing world and are intended to identify young children with high suspicion of clinical

pneumonia in resource-limited settings in which radiography and laboratory testing are not available [18]. According to these criteria, children less than 5 years of age with cough or difficulty breathing and with documented tachypnea at rest are classified as having pneumonia. Severe pneumonia is defined as the presence of chest indrawing, and very severe pneumonia is defined as chest indrawing and either an inability to drink, severe malnutrition, stridor at rest, convulsions, or change in mental status. Applying the WHO case definition to a cohort of children less than 5 years of age presenting to a US emergency department with suspicion of pneumonia, Wingerter et al [19] reported a sensitivity and specificity of 34% and 74%, respectively, for the WHO case definition when using radiographically confirmed pneumonia as the reference standard. Reducing the age-based respiratory rate by 10 breaths per minute to define tachypnea for the WHO case definition improved sensitivity (54%-67%) but reduced specificity (41%-62%), whereas addition of fever did not substantially alter performance characteristics. A similar study conducted in Brazil reported sensitivity and specificity of 84% and 19%, respectively, for radiographically confirmed pneumonia; addition of fever to the WHO case definition improved specificity (46%) without impacting sensitivity (81%) [20]. Thus, although the WHO case definition may be

Journal of the Pediatric Infectious Diseases Society, Vol. 1, No. 4, pp. 314–28, 2012. DOI:10.1093/jpids/pis101 © The Author 2012. Published by Oxford University Press on behalf of the Pediatric Infectious Diseases Society. All rights reserved. For Permissions, please e-mail: journals.permissions@oup.com. appropriate in resource-limited settings, its application in the developed world is more problematic. The case definition is highly dependent on documented tachypnea, the most sensitive clinical sign for diagnosing pneumonia (40%–78% for radiographic-confirmed pneumonia) [21– 24]. However, specificity of tachypnea alone is relatively poor, meaning that most children with tachypnea will not have pneumonia. The recent studies by Wingerter et al [19] and Cardoso et al [20] suggest that the addition of fever to the WHO case definition would not impact sensitivity and may improve specificity.

Radiologic Imaging

Chest radiography is routinely used to confirm the diagnosis of CAP in the developed world. Radiographically confirmed pneumonia is a more specific definition that does not appreciably impact sensitivity, although occasionally radiographic findings lag behind clinical features. Chest radiography may also provide etiologic clues (Table 1; Figures 1-3). Nevertheless, without a pathogen isolated from the lower respiratory tract, the "reference standard" definition of fever, respiratory signs and symptoms, and acute infiltrates on chest radiography is not absolute, because other illnesses may cause similar presentations (eg, acute asthma exacerbated by viral upper respiratory illness) and radiographic interpretation is subject to human variation. Radiography should not supplant clinical judgment, especially in the outpatient setting where it may serve only to delay therapy, increase costs, and expose children to unnecessary ionizing radiation. Rather, chest radiography should be reserved for those children in whom the diagnosis is in question, or for those more critically ill children who require hospitalization. In the latter, chest radiography may reveal complications requiring intervention or prolonged antibiotic therapy (eg, effusion, abscess) or findings suggestive of certain etiologies (eg, pneumatoceles in staphylococcal pneumonia). Chest radiography is also essential in epidemiologic and clinical studies to ensure accurate disease identification, and the WHO has developed criteria to standardize chest radiograph interpretation for the diagnosis of pneumonia in children [25]. These standardized definitions have also been applied in the clinical setting. The finding of alveolar infiltrate had substantial interrater agreement ($\kappa = 0.58-0.73$ across studies) [26-28]. In contrast, the finding of interstitial infiltrate had poor interobserver reliability. These studies highlight the importance of clarifying the presence or absence of alveolar infiltrates in particular when interpreting chest radiographs.

Chest ultrasound has been proposed as an alternative to chest radiography in diagnosing pneumonia. Several studies have demonstrated ultrasound to be equivalent to traditional chest radiography for detecting signs of lung consolidation and superior in the detection of pleural effusions [29–31]. In addition, chest ultrasound may facilitate detection of retrocardiac and lung base opacities, areas difficult to discern with radiography, and does not expose children to ionizing radiation. Chest computed tomography provides an exceptionally detailed view of the lung parenchyma, bronchial tree, pleural spaces, and adjacent structures, but at the expense of high levels of ionizing radiation. As a result, computed tomography is not routinely indicated although it may offer valuable additional information in selected cases, such as children with recurrent or nonresponding pneumonia.

Pneumonia Disease Burden in the Developed World Epidemiology

In the United States alone, there are nearly 2 million outpatient visits and more than 150 000 hospitalizations annually for childhood CAP, and pneumonia remains the leading indication for pediatric hospitalization outside of the newborn period [32-34]. Before the introduction of the heptavalent pneumococcal conjugate vaccine (PCV7), all-cause pneumonia disease rates of 15-49 per 1000 children less than 5 years of age and 11-16 per 1000 children 5 years of age and older were reported in Europe and North America [1, 35, 36]. Hospitalization rates of 3-7 per 1000 children less than 5 years of age and 0.6 per 1000 children 5 years of age and older were also reported [35, 37]. After introduction of PCV7 in the United States, hospitalization rates for CAP and pneumonia-associated complications among young children decreased by 39% and 36%, respectively [33, 38]. However, hospitalization rates for older children remained stable [33].

Currently, 2 large-scale, prospective, epidemiologic studies of pediatric CAP are being conducted in the United States and abroad. The Centers for Disease Control and Prevention-sponsored Etiology of Pneumonia in the Community Study will enroll an estimated 2500 children of all ages hospitalized with CAP in 3 US cities [39]. Likewise, the Pneumonia Etiology Research for Child Health study seeks to enroll 6000 children less than 5 years of age from 7 countries in Africa and Asia [40]. Both studies include comprehensive etiologic and epidemiologic assessments and both use age-matched controls. These studies represent 2 of the largest prospective studies of pediatric CAP ever assembled and will add much to our understanding of disease epidemiology.

Microorganism	Epidemiologic, Clinical, and Radiographic Findings	Diagnostic Methods
Viruses Respiratory syncytial virus Rhinovirus Parainfluenza virus I, II, III Human metapneumovirus Influenza virus A and B Adenovirus Coronavirus Human bocavirus	Viruses represent the most commonly detected pathogens among children less than 5 years of age with CAP, and typically follow predictable seasonal patterns. Up to one third may present with bacterial coinfection. Slow onset, with presence of both upper and lower respiratory tract signs and symptoms, including wheezing, is common. Radiography typically reveals bilateral interstitial infiltrates, although alveolar infiltrates may be seen occasionally.	PCR of sample from nasopharynx/oropharynx (or lower respiratory tract). Single and multiplex platforms are available. Rapid antigen testing is available for influenza and respiratory syncytial virus, although test sensitivity may be low, particularly for influenza. Other diagnostic methods include viral culture of respiratory secretions, blood immunoassay, or paired serology
Bacteria Streptococcus pneumoniae Staphylococcus aureus Streptococcus pyogenes Haemophilus influenzae Moraxella catarrhalis	<i>Streptococcus pneumoniae</i> remains the most common cause of bacterial pneumonia in children. Classic bacterial pneumonia presents with rapid onset of fever, ill appearance, cough, and lower respiratory tract signs and symptoms. Alveolar and lobar infiltrates, with or without effusion, are the most common findings on chest radiography. Abscess, necrotizing pneumonia, and pneumatoceles may occur with severe infections, especially those caused by <i>S aureus</i> , <i>S pyogenes</i> , and occasionally <i>S pneumoniae</i> .	Culture of blood, sputum, lung aspirate, pleural fluid. Rapid urine antigen testing available for <i>S pneumoniae is</i> not recommended for children due to poor specificity, although it may be applied to samples from blood, lower respiratory tract, or pleural fluid. PCR of samples from blood, lower respiratory tract, pleural fluid. PCR is not recommended for upper respiratory tract samples. Paired serology is helpful in epidemiologic studies.
Atypical Bacteria Mycoplasma pneumoniae Chlamydophila pneumoniae	<i>Mycoplasma pneumoniae</i> is considered one of the most common pathogens among school age children and adolescents, and it may be underappreciated in young children. <i>Chlamydophila pneumoniae</i> is considered less common. Classic atypical presentation is gradual onset of low grade fever, rhinorrhea, and cough. Wheezing is also common. Bilateral interstitial infiltrates are the most common radiographic finding, although lobar, alveolar, or nodular patterns are also seen.	<i>Mycoplasma pneumoniae</i> —Culture-based are techniques impractical. A cold agglutination test not recommended. A number of rapid serologic tests are available, but test characteristics vary widely. Sensitivity of PCR from respiratory samples varies, and testing is not widely available. <i>Chlamydophila pneumoniae</i> —No FDA-approved diagnostic testing approved for clinical use. Paired serologic assays (micro-immunofluorescence) preferred but are impractical in clinical setting. PCR testing not widely available.

Table 1. Common Microorganisms Causing Pediatric Community-Acquired Pneumonia

Abbreviations: CAP, community-acquired pneumoia; FDA, US Food and Drug Administration; PCR, polymerase chain reaction.

Pneumonia-Associated Complications

Pneumonia-associated complications can be classified as local, systemic, or metastatic. Parapneumonic effusion is the most common local complication, occurring in up to 25% of children hospitalized with pneumonia; its presence usually signifies bacterial pneumonia, particularly disease caused by S pneumoniae, S aureus, or S pyogenes, although other pathogens, including M pneumoniae, may occasionally be complicated by small effusions [41]. Most effusions are small and do not require drainage. However, approximately 5% of hospitalized children with pneumonia develop larger effusions or empyema that can lead to respiratory compromise [33]. Lung abscess and bronchopleural fistulae are uncommon. Systemic complications, including sepsis and respiratory failure, occur in 5%-30% of children hospitalized with pneumonia, which occurs much more frequently among young children [33, 41]. After the introduction of PCV7, systemic complications have declined, although these declines have been offset by substantial increases in local complications, especially

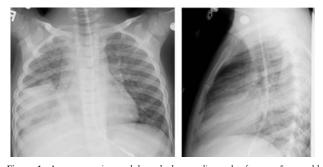


Figure 1. Anteroposterior and lateral chest radiographs from a 5-year-old child with pneumococcal pneumonia. Dense consolidation is apparent in the right middle and lower lobes. Blood culture was positive for *Streptococcus pneumoniae*.

parapneumonic effusion and empyema [33, 42–46]. Secondary sites of infection (eg, meningitis) and other metastatic complications (eg, pneumococcal endocarditis or hemolytic uremic syndrome) are rare [47–49]. Mortality in the developed world is also rare. In the United States, less than 1% of children with CAP severe enough to require hospitalization die, and this result is a nearly 100% reduction in pneumonia-associated mortality over the past 50 years [38, 50].

Pathogens

Viruses

Respiratory viruses unquestionably play a major role in pediatric pneumonia, either alone or in combination with bacteria. Respiratory viruses are ubiquitous pathogens of the upper respiratory tract and cause pneumonia by progressive invasion of the respiratory epithelium, leading to diffuse inflammation that eventually overwhelms normal host defenses to infect the lower airways. Histologically, viral CAP is characterized by diffuse interstitial and

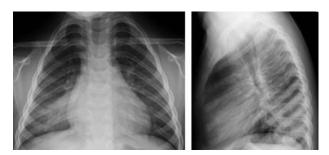


Figure 3. Anteroposterior and lateral chest radiographs from a 3-year-old child with *Mycoplasma pneumoniae* pneumonia. Interstitial infiltrates are apparent bilaterally. Focal consolidation is also appreciated in the right lower lobe. Nasopharyngeal polymerase chain reaction was positive for *M pneumoniae*.

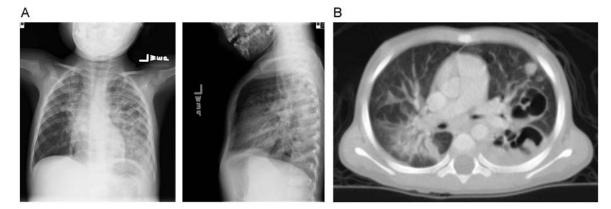


Figure 2. Anteroposterior and lateral chest radiographs (*A*) and chest computed tomography (*B*) from a 5-year-old child with necrotizing community-acquired pneumonia. Nasopharyngeal polymerase chain reaction was positive for parainfluenza virus III. Blood culture was positive for methicillin-resistant *Staphylococcus aureus*. Plain films reveal bilateral, multifocal infiltrates and a small left pleural effusion. A cystic focus is apparent in the left upper lobe. Computed tomography confirms dense consolidation in the left lower lobe, right upper, and right lower lobes, as well as numerous satellite nodules in the left lower and upper lobes. Evidence of bronchiectasis is also noted. Multifocal cystic lesions are apparent bilaterally, left greater than right.

parenchymal lymphocytic proliferation [51, 52]. Respiratory syncytial virus (RSV), parainfluenza viruses (PIV), influenza, adenovirus, and others have long been implicated as important pathogens [41, 53-58] (Table 1). Pathogens recently implicated as a cause of pneumonia include human metapneumovirus (hMPV) [59], human bocavirus (hBoV) [60], and novel rhinovirus and coronavirus species [61]. Studies using polymerase chain reaction (PCR)-based detection methods have documented viruses in 60%-80% of children with CAP [62-67], confirming the importance of RSV as the major cause of viral pneumonia (19%-42% of pneumonia cases) and also highlighting the role of rhinovirus (14%–26%), hBoV (2%–18%), hMPV (2%–12%), PIV (2%–11%), influenza (3%-10%), and adenovirus (2%-18%). Viral pathogens represent the most common cause of CAP in children less than 5 years of age, and especially those less than 2 years of age. Tsolia et al [68] also documented a high prevalence of viral pneumonia among older children who were hospitalized with CAP, although in contrast to young children, RSV was rarely detected (3%), whereas rhinovirus accounted for nearly half of the viruses detected.

A consequence of highly sensitive diagnostics is the potential identification of colonizers and asymptomatic shedding from nonsterile sites rather than true causative pathogens. Epidemiologic studies that have included asymptomatic controls document a high prevalence of viral pathogens among controls (10%-47%), although viral detections are generally lower than that detected among cases (16%-70%) [66, 67, 69, 70]. These studies indicate that RSV is the only viral pathogen that is consistently associated with disease. Results for other pathogens are variable. Rhinovirus and hBoV are often implicated in infections with 2 or more pathogens [62, 63], both are frequent causes of upper respiratory tract infections, and both exhibit prolonged shedding after acute infection [71, 72]. Nonetheless, rhinovirus has been recovered from the lower airways of experimentally infected adult volunteers [73-75], and serologic evidence of acute infection with hBoV has been reported in children with CAP [76, 77]. Thus, although it appears that these pathogens contribute to lower respiratory tract infections, their exact roles remain unknown.

Streptococcus pneumoniae and the Impact of Pneumococcal Conjugate Vaccine

Streptococcus pneumoniae is the major causative pathogen of bacterial pneumonia among children of any age outside of the newborn period, and it is implicated in as many as half of all pneumonia cases [41, 53–58] (Table 1; Figure 1). Pneumococci are frequent colonizers of the upper respiratory tract, and they invade the lung

by aspiration or inhalation, often during periods of impaired pulmonary host defenses such as a viral upper respiratory tract infection. Proliferation in the lower airways results in alveolar inflammation, which is characterized by neutrophil activation and exudative fluid accumulation that spreads rapidly to adjacent alveoli and typically results in lobar pneumonia [52, 78, 79]. PCV7 has greatly reduced the incidence of invasive pneumococcal disease (IPD) and all-cause pneumonia among children. By 2007, rates of laboratory-confirmed pneumococcal pneumonia among US children less than 5 years of age had declined by 50% (from 16.3 to 8 per 100 000) compared with preconjugate vaccine years 1998-99 [80]. Postlicensure observational studies demonstrate similar reductions of 20%-52% for all-cause pneumonia hospitalization rates in the United States and England; admission rates for pneumococcal pneumonia have declined by 57%-73% [33, 38, 81].

Concomitant with declines in IPD after PCV7 licensure, the proportion of isolates considered to be penicillin-nonsusceptible have decreased. This decrease is not surprising because nearly 80% of penicillinnonsusceptible isolates were caused by PCV7 serotypes in the preconjugate vaccine era. In 1998, 24% of all IPD isolates were considered to be penicillin-nonsusceptible (32% among children less than 5 years of age), and 14% of isolates were considered to be multidrug-resistant [82]. By 2004, penicillin-nonsusceptible isolates had declined by 57% (81% among children less than 2 years of age) and multidrug-resistant isolates had declined by 59% [83]. In 2008, the minimum inhibitory concentration breakpoints for reporting susceptibility of nonmeningitis pneumococcal infections to intravenous penicillin were revised, citing clinical studies that documented clinical cure in patients with pneumococcal pneumonia treated with intravenous penicillin despite documented reduced susceptibility in vitro [84]. The effect was an immediate further reduction in the rate of pneumococcal isolates classified as penicillin-nonsusceptible for infections outside the central nervous system [85].

After widespread use of PCV7, nonvaccine serotypes emerged rapidly [86–88]. Recent data from the Active Bacterial Core surveillance (ABCs) program demonstrated that over 80% of IPD cases prelicensure were caused by PCV7 serotypes. By 2007, only 2% of cases were caused by PCV7 serotypes. Of the nonvaccine serotypes, 19A was responsible for nearly one-half of laboratoryconfirmed IPD cases reported to ABCs in 2007 compared with 3% prelicensure [80]. Serotype 19A is also associated with the rapid emergence of penicillin- and multidrug-resist strains and is implicated frequently in severe disease [86–91]. Nevertheless, the increase in nonvaccine serotypes is modest compared with the reduction in PCV7 serotypes, such that actual disease incidence remains significantly reduced. Serotype 19A along with several other important serotypes are also included in the 13-valent PCV (PCV13) licensed in 2010 in the United States and should lead to further declines in the incidence of IPD [92]. The proportion of penicillinnonsusceptible isolates will likely continue to decline because nearly all nonsusceptible isolates in 2008 were caused by the additional serotypes contained in the PCV13 [93].

Staphylococcus aureus, Streptococcus pyogenes, Haemophilus influenzae, and Other Pathogens

Following *S pneumoniae*, *S aureus* and *S pyogenes* are the next most frequent causes of bacterial pneumonia (Table 1). Although historically occurring much less frequently than *S pneumoniae*, both *S aureus* and *S pyogenes* have been associated with significant morbidity. Both are frequently associated with preceding viral infection (eg, influenza, measles, varicella), and both often rapidly progress causing bacteremia, septic shock, local tissue necrosis, pleural effusion, and lung abscess [52] (Figure 2).

The emergence of community-associated methicillinresistant S aureus (CA-MRSA) in the 1990s heralded a new epidemic of staphylococcal disease that has had a substantial influence on the epidemiology and management of pediatric CAP. Between 2002 and 2007, discharges for MRSA pneumonia more than doubled at US children's hospitals [94]. True to the typical phenotype of invasive staphylococcal infections, MRSA pneumonia is often severe with high rates of both local and systemic complications [95]. The increase in CA-MRSA also parallels the rapid increase in parapneumonic empyema [42, 96-98], and it is often the most commonly isolated pathogen in MRSA-prevalent regions [99]. As a result, empiric therapy for severe or complicated pneumonia almost always requires consideration of S aureus, including MRSA in prevalent regions. Despite the increased incidence of staphylococcal disease, molecular diagnostics suggest that pneumococcus remains an important cause of complicated pneumonia, especially in cases of culturenegative empyema [100].

Haemophilus influenzae type B (HiB) disease, once a cause of significant morbidity and mortality among children, is now rare as a result of the widespread use of conjugate HiB vaccines. Non-type B *H influenzae* remain frequent colonizers and pathogens of the upper respiratory tract, but they only occasionally cause lower tract disease. Likewise, *Moraxella catarrhalis* is occasionally implicated in pediatric CAP. Both pathogens are implicated in less

than 10% of pediatric pneumonia cases among recent epidemiologic studies [53, 57, 58]. Outside of the neonatal period, other causes of CAP are quite rare, occurring occasionally in specialized populations (eg, immunocompromised) or specific regions where pathogens may be endemic (eg, histoplasma) (Table 2).

Atypical Bacterial Pathogens

Mycoplasma pneumoniae is a frequent cause of bacterial pneumonia; Chlamydophila pneumoniae is much less common (Table 1). Mycoplasma pneumoniae and C pneumoniae are implicated in 2%-30% and 1%-14% of pneumonia cases, respectively, typically being much more frequently isolated among older children and those in ambulatory settings [41, 53-58, 101]. However, several studies have reported high rates of atypical bacterial infections among preschool age children with rates similar to that of older children [41, 102, 103]. Mycoplasma pneumoniae infections occur sporadically throughout the year, although epidemic outbreaks occur regularly every few years, and it is during these epidemics when young children may be disproportionately affected [103]. Thus, although atypical pathogens are more frequently encountered among school age children, they remain important considerations among younger children. Distinguishing between atypical pathogens and other bacterial causes is also difficult (Figure 3). Although the classic presentation of CAP caused by atypical pathogens is distinct from that of other bacterial pathogens, atypical disease may present very similarly to classic bacterial pneumonia, and M pneumoniae infection is occasionally severe with complications including parapneumonic effusion, lung abscess, and neurologic sequelae [102, 104-107].

Viral and Bacterial Coinfections

Respiratory viruses frequently precede the development of bacterial pneumonia [5, 6, 108]. Viral-bacterial coinfections are well described, particularly with influenza and pneumococcal or staphylococcal pneumonia. During the 1918 influenza pandemic, more than 500 000 US deaths were attributed to influenza, and it is believed that the majority of deaths were a result of bacterial coinfection [109]. Epidemiologic studies using sensitive molecular diagnostics also reveal a high prevalence of coinfections (20%-35%) [41, 53, 57, 58, 62, 63, 65, 68]. In vitro and animal studies suggest that the virulence of both bacterial and viral pathogens is enhanced in coinfections, exerting a synergistic effect on the host organism [110]. Several clinical studies document worse outcomes among children with coinfections compared with those without identified viral coinfection [109, 111-114]. As diagnostic methods improve, our

Microorganism	Comment			
Viruses				
Varicella zoster virus	Potential complication after primary varicella infection. Often severe and associated with secondary bacterial infection.			
Measles virus	Rubeola. Pneumonia is a frequent complication.			
Hantavirus	Hantavirus pulmonary syndrome. Rodent exposure.			
Bacteria				
Bordetella pertussis	Pneumonia uncommon manifestation. Bacterial coinfection may be severe, especially in infants.			
Group B Streptococci	Neonatal pneumonia and sepsis.			
Listeria monocytogenes	Neonatal pneumonia and sepsis.			
Gram-negative enterics	Neonatal pneumonia and sepsis. Potential pathogens in aspiration pneumonia.			
Chlamydia trachomatis	Cause of afebrile pneumonia in young infants less than 3 months of age.			
Anaerobes (oral flora)	Potential pathogens in aspiration pneumonia.			
Legionella pneumophila	Legionnaires' disease. Rare in children but associated with community outbreaks. Exposure to contaminated artificial freshwater systems.			
Coxiella burnetii	Q fever. Exposure to wild and domesticated herbivores or unpasteurized dairy (eg, cattle, sheep, goats). Also potential bioterrorism agent.			
Chlamydia psittaci	Psittacosis. Bird (eg, pet birds, pigeons) exposure.			
Francisella tularensis	Tularemia. Rabbit exposure.			
Yersinia pestis	Pneumonic plague. Rodent flea exposure.			
Bacillus anthracis	Anthrax. "Woolsorters' disease." Wild and domesticated herbivore (eg, cattle, sheep, goats) exposure. Also potential bioterrorism agent.			
Leptospira interrogans Mycobacterium tuberculosis	Leptospirosis. Exposure to urine of wild and domestic animals carrying the bacterium. Rare in the US children. Usually associated with high-risk exposures.			
Brucella abortus	Brucellosis. Exposure to wild and domesticated animals or unpasteurized dairy (eg, cattle, sheep, pigs, goats, deer,			
	dogs).			
Fungi				
Histoplasma capsulatum	Histoplasmosis. Exposure to bird or bat droppings (eg, poultry/bird roosts, caves). Endemic to eastern and centra United States.			
Blastomyces dermatitidis	Blastomycosis. Environmental exposure to fungal spores (wooded areas). Endemic to Southeastern and Midwestern United States.			
Cryptococcus neoformans	Cryptococcosis. Exposure to soil contaminated with bird droppings. Significant pathogen nearly exclusively among immunocompromised.			
Coccidioides immitis	Coccidiomycosis. "Valley fever." Environmental exposure to fungal spores (dry, dusty environments). Endemic to Southwestern United States			

Table 2. Rare Microorganisms Causing Pediatric Community-Acquired Pneumonia or Occurring in Specialized Populations

understanding of the pathogenesis of viral-bacterial coinfections and their impact on outcomes will improve.

Culture-Based Methods

Diagnostic Testing for Etiology and Prognosis

Determining microbiologic etiology is essential for tailoring antimicrobial coverage, predicting outcomes, and understanding the changing epidemiology of CAP (Table 1). Yet, accurate determination of etiology for most cases of pediatric pneumonia remains elusive. Reasons for this include difficulty in obtaining culture material from the primary site of infection, use of antimicrobials before obtaining samples for analysis, lack of sensitive and rapid bacterial diagnostic tests, and difficulties distinguishing pathogenic from colonizing bacteria. Even when a viral pathogen is implicated, distinguishing viral CAP from viral-bacterial coinfections is problematic. Moreover, most children recover quickly with commonly used empiric antibiotics, and some argue that routine use of comprehensive diagnostics would increase costs without improving care or changing management decisions. Thus, the perceived need to comprehensively assess microbiologic etiology is low.

Culture-based methods, although considered as the gold standard for pathogen identification from normally sterile sites, are insensitive for bacterial pneumonia. Less than 10% of children with blood cultures obtained yield a causative pathogen [45, 115-120]. Moreover, nonpathogenic contaminants are recovered frequently. Pleural fluid cultures are more commonly positive (10%– 25% of cases) despite nearly always being collected after initiation of antimicrobial therapy, presumably due to higher concentrations of organisms as well as prolonged time to sterilization compared with blood [45, 115, 121-123]. Culture and microbiologic examination of sputum is rarely performed in children due to the inability of young children to expectorate and concerns for poor specimen quality. However, among older children, obtaining high-quality sputum could aid in the recovery of bacterial pathogens. Induction of sputum production with hypertonic saline may facilitate obtaining sputa from young children, but whether this method produces useful proportions of high-quality samples remains to be determined [67, 124]. Culture of material obtained directly from the lower respiratory tract (eg, bronchoalveolar lavage, transthoracic aspiration), although likely sensitive and less subject to upper airway contamination, is impractical in most cases.

Serologic Studies

Serologic studies are available for a number of pathogens, but widespread adoption of serologic methods is hampered by the need for both acute and convalescent samples, lack of test results in a clinically relevant time frame, and lack of specific serologic targets for certain pathogens. Nonetheless, high-quality paired sera often contribute important information for epidemiologic purposes and should be included in research studies of pneumonia etiology.

Antigen Detection

Antigen detection is also useful in selected settings. Rapid antigen tests for the detection of RSV and influenza obtained from upper respiratory tract samples are highly specific, although test sensitivity varies and may be particularly poor for influenza [125, 126]. In contrast, urinary antigen testing for *S pneumoniae* (Binax NOW) has high sensitivity but poor specificity in children, resulting in false-positive results attributable to pneumococcal colonization of the upper airway [127, 128]. Application of the Binax NOW platform is also adaptable to other sample types, and it has proven both highly sensitive and specific in pleural fluid samples [129].

Molecular Diagnostics

Rapid molecular diagnostics, particularly PCR, also have potential for improving microbiologic yield over traditional methods. PCR testing is now available for nearly all clinically important respiratory pathogens, in both single and multiplex formats. PCR is also used to quantify pathogen load and determine antimicrobial resistance patterns. The nasopharynx is the preferred source for identifying pathogens not likely to colonize the upper airways (ie, viruses, M pneumoniae, C pneumoniae). Testing for bacterial pathogens from these samples is problematic because distinguishing causative agent from colonizer is often impossible. Quantitative real-time PCR may prove to be useful in this regard, as recently demonstrated for S pneumoniae [130]. Pathogen load may also predict illness severity [131, 132]. Thus, further refinement of quantitative methods could be beneficial in assessing etiology and predicting outcomes.

Pneumococcal PCR from blood samples has received much attention in recent years. Although issues regarding specificity of the early PCR targets have largely been overcome, clinical sensitivity of these assays is 40%–100% when using culture-confirmed pneumococcal bacteremia as the reference standard; expanding the reference standard to include sputum culture and urine antigen testing among adults, clinical sensitivity decreases to 26%–32% [133–138]. Conversely, most PCR-positive blood samples in pediatric studies are among children with negative blood cultures; whether these all represent causative pathogens versus carriage remains to be determined [135, 137–139].

Molecular testing of pleural fluid is attractive because sensitivity is likely improved over blood PCR and less subject to issues of specificity. Both broad range and species-specific bacterial PCR have been applied to pleural fluid samples. Using PCR to target the 16S ribosomal RNA gene common to a wide range of bacteria, Saglani et al [140] and Le Monnier et al [129] documented significant increases in pathogen recovery with this technique (69%-100%) compared with bacterial culture alone (19%–58%). Gollomp et al [141] documented only modest improvements of 16S PCR over bacterial culture. Using species-specific PCR targets, Blaschke et al [100] increased etiologic yield to 84% compared with 35% for culture-based methods alone among 63 children with pneumonia complicated by empyema. Recovery of S pneumoniae doubled from 35% to 71% using PCR, confirming the importance of this pathogen in severe pneumonia. Serotype 19A, responsible for 23% of S pneumoniae isolates recovered by culture, represented only 2% of culture-negative isolates recovered by PCR. Likewise, only 2% of culture-negative empyema cases were attributed to S aureus, whereas this pathogen was responsible for 18% of culture-positive cases. Although the impact of pneumococcal serotype 19A and S aureus should not be minimized, this study demonstrates the potential bias associated with reliance on culture-based methods and the need for improved diagnostics.

Biomarkers

Laboratory markers (eg, white blood cell count, C-reactive protein) are also often used to assist in predicting etiology, although these tests generally lack sufficient sensitivity or specificity to guide decision-making and are rarely helpful in confirming the diagnosis [142–149]. These tests may be helpful in selected cases when measured serially to track disease progression and response to therapy.

Procalcitonin is another potential biomarker for bacterial pneumonia. Procalcitonin is elevated in serious bacterial infections, and it has been used to predict neonatal sepsis and bacterial meningitis [150–153]. In most reported studies, procalcitonin is significantly higher among children with bacterial (ie, pneumococcal) pneumonia compared with pneumonia caused by viral or atypical pathogens [144, 145, 154–157]. Cutoff values ranging from 0.5 ng/mL to 2 ng/mL have been proposed to distinguish bacterial pneumonia from other pathogens, although these values lack sufficient sensitivity and specificity for routine clinical use. Lower cutoff values demonstrate improved sensitivity [157], and they may prove useful in reducing antimicrobial use as demonstrated in a randomized controlled trial [158].

Other Diagnostic Methods

Other diagnostic methods on the horizon include genomic and proteomic analyses. Host gene expression analyses using blood microarray distinguish between individuals with acute viral or bacterial infections and healthy controls and may also predict illness severity [159–162]. Urinary metabolic profiles and breath analysis may also one day prove useful for diagnostic purposes [163, 164], although all of these techniques require additional investigation.

Disease Management and National Guidelines

Despite a better understanding of CAP epidemiology and its evolution in the conjugate vaccine era, optimal disease management remains a challenge. Although our appreciation for viral CAP has grown, it remains difficult to distinguish bacterial from viral etiologies, and even when bacterial disease seems obvious, insensitive diagnostics make precise pathogen detection challenging. Furthermore, the heterogeneous nature of the disease leads to a wide spectrum of illness severity, rendering outcome prediction difficult. The net effect is clinical decision-making that often lacks evidence, leading to wide variation in disease management, resource utilization, and outcomes [165]. In the United States, national guidelines for the management of CAP in children were published jointly by the Pediatric Infectious Diseases Society and the Infectious Diseases Society of America [166]. The guidelines are comprehensive in scope and are an excellent resource for both clinicians and investigators, which hopefully will assist in standardizing management strategies. However, the presence of evidence-based national guidelines will not lead to improved care and outcomes unless widespread local adoption and implementation efforts are used [167, 168].

Several recommendations in the guidelines center on antimicrobial management, emphasizing the use of narrow spectrum antibiotics (ie, penicillin and aminopenicillins) for the majority of cases of bacterial pneumonia (Table 3). Strong consideration for withholding antibiotics among children less than 5 years of age with mild CAP is also recommended. Controversy over these points can be expected because this approach represents a major departure from typical practice in the United States Nonetheless, these recommendations are well supported by evidence, including recognition of the importance of viral CAP among young children; the dramatic reductions in IPD incidence and concomitant declines in penicillin resistance after introduction of PCV7 —a trend likely to continue with PCV13; and the proven effectiveness of appropriately dosed narrow spectrum therapy against nonmeningitic pneumococcal infections in the absence of high-level penicillin resistance.

In contrast, several of the guidelines' recommendations were supported by poor-quality evidence. The guideline committee highlighted this challenge by identifying areas for future research. Blood cultures are recommended for all children hospitalized with CAP even though the sensitivity of this test is poor; most children with suspected bacterial pneumonia have negative blood cultures. However, very little data are available to guide clinicians in selecting populations in whom blood cultures may be most useful. Macrolide therapy is recommended for CAP caused by atypical pathogens, although without microbiologic confirmation, distinguishing atypical from viral and typical bacterial pathogens remains challenging. Evidence demonstrating substantial effectiveness of macrolides for primary treatment of atypical CAP is also lacking. Recommendations for severe CAP are also less clear, including the use of adjunctive immunomodulatory therapies for very severe CAP [169] and optimal management approaches for CAP complicated by pleural effusion or empyema. Simply defining CAP disease severity using objective criteria would be useful. A number of clinical severity scores are available for adult CAP (eg, CURB-65 [170] Pneumonia Severity Index [171]), which predict outcomes. These scores are used for risk stratification in research studies, and help to inform site of care decisions and tailor antimicrobial therapies in clinical care [172, 173]. The majority of these adult severity scores are designed to predict disease mortality and include predictive factors that are not relevant to children, thus limiting their utility in pediatric populations. Unfortunately, no pediatric CAP severity scores have been validated in the developed world, which is a major impediment to advancing our understanding of disease severity and improving outcomes for pediatric CAP.

Conclusions

Over the last 2 decades, our understanding of the epidemiology of pediatric CAP and the varied nature of the disease has grown substantially. The introduction of conjugate vaccines targeting HiB and *S pneumoniae* has dramatically reduced the incidence of invasive disease caused by these pathogens, although pneumococcus remains the most common bacterial cause of pediatric

Population		Bacterial Pneumonia	Atypical Pneumonia	Influenza Pneumonia
Outpatient Neonates–3 months Preschool (<5 years)	Preferred alternative(s)	Outpatient therapy not recommended Amoxicillin Amoxicillin/clavulanate; Levofloxacin for children	Azithromycin Clarithromycin or erythromycin	Oseltamivir
5–17 years	Preferred alternative(s)	with serious penicillin allergy Amoxicillin Amoxicillin/clavulanate; Levofloxacin for children with serious penicillin allergy	Azithromycin Clarithromycin or erythromycin; Doxycycline if >7 years of age	Oseltamivir Zanamivir if ≥ 7 years of age
Inpatient		1 07	,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,	
Neonates	Preferred alternative(s)	Ampicillin + gentamicin Ampicillin + cefotaxime	N/A	No FDA-approved antiviral drugs for children <1 year of age. Oseltamivir dosing recommendations are available at www.cdc.gov
1–3 months	Preferred alternative(s)	Cefotaxime Azithromycin if suspect; <i>Chlamydia trachomatis</i> or <i>Bordetella pertussis</i>	N/A	No FDA-approved antiviral drugs for children <1 year of age. Oseltamivir dosing recommendations are available at www.cdc.gov
3 months to 17 years, fully immunized, local epidemiology indicates low prevalence of penicillin-nonsusceptible <i>Streptococcus pneumoniae</i>	Preferred alternative(s)	Ampicillin or Penicillin G Ceftriaxone or cefotaxime Antistaphylococcal coverage for suspected <i>Staphylococcus aureus</i> , including clindamycin or vancomycin in MRSA prevalent regions; Levofloxacin for children with serious penicillin allergy	Azithromycin Clarithromycin or erythromycin; Doxycycline if >7 years of age; Levofloxacin for those who have reached skeletal maturity	Oseltamivir Zanamivir if ≥7 years of age
3 months to 17 years, not fully immunized, or local epidemiology indicates moderate to high prevalence of penicillin-nonsusceptible <i>S pneumoniae</i>	Preferred alternative(s)	Ceftriaxone or cefotaxime Levofloxacin Antistaphylococcal coverage for suspected <i>S aureus</i> , including clindamycin or vancomycin in MRSA prevalent regions	Azithromycin Clarithromycin or erythromycin; Doxycycline if >7 years of age; Levofloxacin for those who have reached skeletal maturity	Oseltamivir Zanamivir if ≥ 7 years of age

Table 3. Empiric Antimicrobial Strategies for Pediatric Community-Acquired Pneumonia^a

Abbreviations: FDA, US Food and Drug Administration; MRSA, methicillin-resistant Staphylococcus aureus.

^aAdapted from "Table 7: Empiric Therapy for Pediatric Community-Acquired Pneumonia (CAP)." John S. Bradley, Carrie L. Byington, Samir S. Shah, et al. The Management of Community-Acquired Pneumonia in Infants and Children Older Than 3 Months of Age: Clinical Practice Guidelines by the Pediatric Infectious Diseases Society and the Infectious Diseases Society of America. Clin Infect Dis 2011; 53:e34. CAP. In addition, the emergence of CA-MRSA has led to an epidemic of serious infections. Finally, improvements in diagnostic methods have confirmed the importance of respiratory viruses, both singularly and in mixed infections, and improved our understanding of the role of *M pneumoniae* and other atypical pathogens as important causes of pediatric CAP.

Despite these many advances, uncertainty abounds. Diagnostics with the ability to rapidly and comprehensively assess etiology and predict outcomes at the point of care are urgently needed. Highly sensitive and specific diagnostics are essential to improving disease management and informing our knowledge of disease pathogenesis and outcomes for viruses, bacteria, and mixed infections. Treatment studies of both old and new antimicrobials and adjunctive therapies are imperative for facilitating appropriate antimicrobial selection, combating antimicrobial resistance, and optimizing outcomes. Finally, epidemiologic studies using state-of-the-art diagnostics and populationbased surveillance must continue to monitor disease trends and inform public policy.

Acknowledgments

Potential conflict of interest. All authors: No reported conflicts.

All authors have submitted the ICMJE Form for Disclosure of Potential Conflicts of Interest. Conflicts that the editors consider relevant to the content of the manuscript have been disclosed.

References

- Jokinen C, Heiskanen L, Juvonen H, et al. Incidence of community-acquired pneumonia in the population of four municipalities in eastern Finland. Am J Epidemiol 1993. 137: 977–88.
- Grant CC, Emery D, Milne T, et al. Risk factors for community-acquired pneumonia in pre-school-aged children. J Paediatr Child Health 2012. 48:402–12.
- Heiskanen-Kosma T, Korppi M, Jokinen C, Heinonen K. Risk factors for community-acquired pneumonia in children: a population-based case-control study. Scand J Infect Dis 1997. 29:281–5.
- Teepe J, Grigoryan L, Verheij TJ. Determinants of community-acquired pneumonia in children and young adults in primary care. Eur Respir J 2010. 35:1113–7.
- Talbot TR, Poehling KA, Hartert TV, et al. Seasonality of invasive pneumococcal disease: temporal relation to documented influenza and respiratory syncytial viral circulation. Am J Med 2005. 118:285–91.
- Ampofo K, Bender J, Sheng X, et al. Seasonal invasive pneumococcal disease in children: role of preceding respiratory viral infection. Pediatrics 2008. 122:229–37.
- Camargo CA Jr, Ganmaa D, Frazier AL, et al. Randomized trial of vitamin D supplementation and risk of acute respiratory tract infection in Mongolia. Pediatrics 2012. 130: e561–7.
- Redd SC, Vreuls R, Metsing M, et al. Clinical signs of pneumonia in children attending a hospital outpatient department in Lesotho. Bull World Health Organ 1994. 72:113–8.

- Lynch T, Platt R, Gouin S, et al. Can we predict which children with clinically suspected pneumonia will have the presence of focal infiltrates on chest radiographs? Pediatrics 2004. 113(3 Pt 1):e186–9.
- Mathews B, Shah S, Cleveland RH, et al. Clinical predictors of pneumonia among children with wheezing. Pediatrics 2009. 124:E29–6.
- Neuman MI, Monuteaux MC, Scully KJ, Bachur RG. Prediction of pneumonia in a pediatric emergency department. Pediatrics 2011. 128:246–3.
- Mahabee-Gittens EM, Grupp-Phelan J, Brody AS, et al. Identifying children with pneumonia in the emergency department. Clin Pediatr (Phila) 2005. 44:427–35.
- Murphy CG, van de Pol AC, Harper MB, Bachur RG. Clinical predictors of occult pneumonia in the febrile child. Acad Emerg Med 2007. 14:243–9.
- Campbell H, Byass P, Forgie IM, Lloyd-Evans N. Clinical signs of pneumonia in children. Lancet 1989. 1:899–900.
- Lozano JM, Steinhoff M, Ruiz JG, et al. Clinical predictors of acute radiological pneumonia and hypoxaemia at high altitude. Arch Dis Child 1994. 71:323–7.
- Bilkis MD, Gorgal N, Carbone M, et al. Validation and development of a clinical prediction rule in clinically suspected community-acquired pneumonia. Pediatr Emerg Care 2010. 26:399–405.
- 17. Lynch T, Bialy L, Kellner JD, et al. A systematic review on the diagnosis of pediatric bacterial pneumonia: when gold is bronze. PLoS One 2010. 5:pe11989.
- World Health Organization. Department of Child and Adolescent Health and Development. Pocket Book of Hospital Care for Children: Guidelines for the Management of Common Illnesses with Limited Resources. Geneva: World Health Organization, 2005.
- Wingerter SL, Bachur RG, Monuteaux MC, Neuman MI. Application of the world health organization criteria to predict radiographic pneumonia in a US-based pediatric emergency department. Pediatr Infect Dis J 2012. 31:561–4.
- Cardoso MR, Nascimento-Carvalho CM, Ferrero F, et al. Adding fever to WHO criteria for diagnosing pneumonia enhances the ability to identify pneumonia cases among wheezing children. Arch Dis Child 2011. 96:58–61.
- Palafox M, Guiscafre H, Reyes H, et al. Diagnostic value of tachypnoea in pneumonia defined radiologically. Arch Dis Child 2000. 82:41–5.
- Shah S, Bachur R, Kim D, Neuman MI. Lack of predictive value of tachypnea in the diagnosis of pneumonia in children. Pediatr Infect Dis J 2010. 29:406–9.
- Taylor JA, Del Beccaro M, Done S, Winters W. Establishing clinically relevant standards for tachypnea in febrile children younger than 2 years. Arch Pediatr Adolesc Med 1995. 149: 283–7.
- 24. Oostenbrink R, Thompson M, Lakhanpaul M, et al. Children with fever and cough at emergency care: diagnostic accuracy of a clinical model to identify children at low risk of pneumonia [published online ahead of print August 3, 2012]. Eur J Emerg Med 2012; doi:10.1097/MEJ.0b013e32835771fd.
- Cherian T, Mulholland EK, Carlin JB, et al. Standardized interpretation of paediatric chest radiographs for the diagnosis of pneumonia in epidemiological studies. Bull World Health Organ 2005. 83:353–9.
- Neuman MI, Lee EY, Bixby S, et al. Variability in the interpretation of chest radiographs for the diagnosis of pneumonia in children. J Hosp Med 2012. 7:294–8.
- Ben Shimol S, Dagan R, Givon-Lavi N, et al. Evaluation of the World Health Organization criteria for chest radiographs for pneumonia diagnosis in children. Eur J Pediatr 2012. 171:369–4.

- Xavier-Souza G, Vilas-Boas AL, Fontoura MS, et al. The inter-observer variation of chest radiograph reading in acute lower respiratory tract infection among children. [Published ahead of print] Pediatr Pulmonol 2012. doi:10.1002/ ppul.22644.
- Caiulo VA, Gargani L, Caiulo S, et al. Lung ultrasound characteristics of community-acquired pneumonia in hospitalized children. [Published ahead of print] Pediatr Pulmonol 2012. doi:10.1002/ppul.22585.
- Iuri D, De Candia A, Bazzocchi M. Evaluation of the lung in children with suspected pneumonia: usefulness of ultrasonography. Radiol Med 2009. 114:321–30.
- Copetti R, Cattarossi L. Ultrasound diagnosis of pneumonia in children. Radiol Med 2008. 113:190–8.
- 32. AHRQ. National Estimates on Use of Hospitals by Children from the HCUP Kids' Inpatient Database (KID). 2009; Available at: http://hcupnet.ahrq.gov/HCUPnet.jsp?Id= F3D2E7DF566C8BCC&Form=SelPAT&JS=Y&Action=%3E% 3ENext%3E%3E&_InPatChar=Yes&_InHospChar=Yes&_ PatChar=. Accessed August 20, 2012.
- Lee GE, Lorch SA, Sheffler-Collins S, et al. National hospitalization trends for pediatric pneumonia and associated complications. Pediatrics 2010. 126:204–13.
- Kronman MP, Hersh AL, Feng R, et al. Ambulatory visit rates and antibiotic prescribing for children with pneumonia, 1994– 2007. Pediatrics 2011. 127:411–8.
- Nelson JC, Jackson M, Yu O, et al. Impact of the introduction of pneumococcal conjugate vaccine on rates of community acquired pneumonia in children and adults. Vaccine 2008. 26: 4947–54.
- Weigl JA, Bader HM, Everding A, Schmitt HJ. Population-based burden of pneumonia before school entry in Schleswig-Holstein, Germany. Eur J Pediatr 2003. 162:309–16.
- Senstad AC, Suren P, Brauteset L, et al. Community-acquired pneumonia (CAP) in children in Oslo, Norway. Acta Paediatr 2009. 98:332–6.
- Grijalva CG, Nuorti JP, Arbogast PG, et al. Decline in pneumonia admissions after routine childhood immunisation with pneumococcal conjugate vaccine in the USA: a time-series analysis. Lancet 2007. 369:1179–86.
- Jain S. Findings from the Etiology of Pneumonia in the Community (EPIC) Study. Pediatric Academic Societies; April 30 2012, Boston, MA.
- Levine OS, OBrien KL, Deloria-Knoll M, et al. The Pneumonia Etiology Research for Child Health Project: a 21st century childhood pneumonia etiology study. Clin Infect Dis 2012. 54(Suppl 2):S93–101.
- Michelow IC, Olsen K, Lozano J, et al. Epidemiology and clinical characteristics of community-acquired pneumonia in hospitalized children. Pediatrics 2004. 113:701–7.
- Grijalva CG, Nuorti JP, Zhu Y, Griffin MR. Increasing incidence of empyema complicating childhood communityacquired pneumonia in the United States. Clin Infect Dis 2010. 50:805–13.
- 43. Hendrickson DJ, Blumberg DA, Joad JP, et al. Five-fold increase in pediatric parapneumonic empyema since introduction of pneumococcal conjugate vaccine. Pediatr Infect Dis J 2008. 27:1030–2.
- Li ST, Tancredi DJ. Empyema hospitalizations increased in US children despite pneumococcal conjugate vaccine. Pediatrics 2010. 125:26–33.
- 45. Byington CL, Spencer LY, Johnson TA, et al. An epidemiological investigation of a sustained high rate of pediatric parapneumonic empyema: risk factors and microbiological associations. Clin Infect Dis 2002. 34:434–40.

- Byington CL, Korgenski K, Daly J, et al. Impact of the pneumococcal conjugate vaccine on pneumococcal parapneumonic empyema. Pediatr Infect Dis J 2006. 25:250–4.
- 47. Banerjee R, Hersh AL, Newland J, et al. Streptococcus pneumoniae-associated hemolytic uremic syndrome among children in North America. Pediatr Infect Dis J 2011. 30: 736–9.
- 48. Huang YH, Lin TY, Wong KS, et al. Hemolytic uremic syndrome associated with pneumococcal pneumonia in Taiwan. Eur J Pediatr 2006. 165:332–5.
- 49. Waters AM, Kerecuk L, Luk D, et al. Hemolytic uremic syndrome associated with invasive pneumococcal disease: the United kingdom experience. J Pediatr 2007. 151:140–4.
- Dowell SF, Kupronis BA, Zell ER, Shay DK. Mortality from pneumonia in children in the United States, 1939 through 1996. N Engl J Med 2000. 342:1399–407.
- Long SS, Pickering LK, Prober CG. Principles and Practice of Pediatric Infectious Diseases. 2nd ed. New York: Churchill Livingstone; 2003.
- 52. Feigin RD, Cherry JD. Textbook of Pediatric Infectious Diseases. 3rd ed. Philadelphia: Saunders; **1992**.
- Juven T, Mertsola J, Waris M, et al. Etiology of community-acquired pneumonia in 254 hospitalized children. Pediatr Infect Dis J 2000. 19:293–8.
- Drummond P, Clark J, Wheeler J, et al. Community acquired pneumonia-a prospective UK study. Arch Dis Child 2000. 83: 408–12.
- 55. Wubbel L, Muniz L, Ahmed A, et al. Etiology and treatment of community-acquired pneumonia in ambulatory children. Pediatr Infect Dis J 1999. 18:98–104.
- 56. Claesson BA, Trollfors B, Brolin I, et al. Etiology of community-acquired pneumonia in children based on antibody responses to bacterial and viral antigens. Pediatr Infect Dis J 1989. 8:856–62.
- 57. Heiskanen-Kosma T, Korppi M, Jokinen C, et al. Etiology of childhood pneumonia: serologic results of a prospective, population-based study. Pediatr Infect Dis J 1998. 17: 986–91.
- Don M, Fasoli L, Paldanius M, et al. Aetiology of community-acquired pneumonia: serological results of a paediatric survey. Scand J Infect Dis 2005. 37:806–12.
- 59. van den Hoogen BG, de Jong JC, Groen J, et al. A newly discovered human pneumovirus isolated from young children with respiratory tract disease. Nat Med **2001**. 7:719–24.
- Allander T, Tammi MT, Eriksson M, et al. Cloning of a human parvovirus by molecular screening of respiratory tract samples. Proc Natl Acad Sci USA 2005. 102:12891–6.
- Greenberg SB. Update on rhinovirus and coronavirus infections. Semin Respir Crit Care Med 2011. 32:433–46.
- 62. Garcia-Garcia ML, Calvo C, Pozo F, et al. Spectrum of respiratory viruses in children with community acquired pneumonia. Pediatr Infect Dis J **2012**. 31:808–13.
- Esposito S, Daleno C, Prunotto G, et al. Impact of viral infections in children with community-acquired pneumonia: results of a study of 17 respiratory viruses. [Published ahead of print] Influenza Other Respi Viruses 2012. doi:10.1111/j. 1750-2659.2012.00340.x.
- 64. Wolf DG, Greenberg D, Shemer-Avni Y, et al. Association of human metapneumovirus with radiologically diagnosed community-acquired alveolar pneumonia in young children. J Pediatr **2010**. 156:115–20.
- 65. Cilla G, Onate E, Perez-Yarza EG, et al. Viruses in community-acquired pneumonia in children aged less than 3 years old: high rate of viral coinfection. J Med Virol 2008. 80:1843–9.

- Hammitt LL, Kazungu S, Morpeth SC, et al. A preliminary study of pneumonia etiology among hospitalized children in Kenya. Clin Infect Dis 2012. 54(Suppl 2):S190–9.
- 67. Mermond S, Zurawski V, D'Ortenzio E, et al. Lower respiratory infections among hospitalized children in New Caledonia: a pilot study for the Pneumonia Etiology Research for Child Health project. Clin Infect Dis 2012. 54(Suppl 2):S180–9.
- Tsolia MN, Psarras S, Bossios A, et al. Etiology of community-acquired pneumonia in hospitalized school-age children: evidence for high prevalence of viral infections. Clin Infect Dis 2004. 39:681–6.
- 69. Fry AM, Lu X, Olsen SJ, et al. Human rhinovirus infections in rural Thailand: epidemiological evidence for rhinovirus as both pathogen and bystander. PLoS One **2011**. 6:e17780.
- Berkley JA, Munywoki P, Ngama M, et al. Viral etiology of severe pneumonia among Kenyan infants and children. JAMA 2010. 303:2051–7.
- Brieu N, Guyon G, Rodiere M, et al. Human bocavirus infection in children with respiratory tract disease. Pediatr Infect Dis J 2008. 27:969–73.
- Martin ET, Fairchok MP, Kuypers J, et al. Frequent and prolonged shedding of bocavirus in young children attending daycare. J Infect Dis 2010. 201:1625–32.
- Gern JE, Calhoun W, Swenson C, et al. Rhinovirus infection preferentially increases lower airway responsiveness in allergic subjects. Am J Respir Crit Care Med 1997. 155:1872–6.
- Simons E, Schroth MK, Gern JE. Analysis of tracheal secretions for rhinovirus during natural colds. Pediatr Allergy Immunol 2005. 16:276–8.
- 75. Papadopoulos NG, Bates PJ, Bardin PG, et al. Rhinoviruses infect the lower airways. J Infect Dis 2000. 181:1875–84.
- Don M, Soderlund-Venermo M, Valent F, et al. Serologically verified human bocavirus pneumonia in children. Pediatr Pulmonol 2010. 45:120–6.
- 77. Nascimento-Carvalho CM, Cardoso MR, Meriluoto M, et al. Human bocavirus infection diagnosed serologically among children admitted to hospital with community-acquired pneumonia in a tropical region. J Med Virol 2012. 84:253–8.
- Shah SS. Pediatric Practice. Infectious Disease. New York: McGraw-Hill Medical; 2009.
- 79. Long SS, Pickering LK, Prober CG. Principles and Practice of Pediatric Infectious Diseases. 2003; 2nd:xxix, 1645 p. Available at: http://home.mdconsult.com/start_session?autologin=true& user=VANDYGENERIC&password=generic&clogin-chang=false& targeturl=/public/book/view%3Ftitle%3Dlong.
- Pilishvili T, Lexau C, Farley MM, et al. Sustained reductions in invasive pneumococcal disease in the era of conjugate vaccine. J Infect Dis 2010. 201:32–41.
- Zhou F, Kyaw MH, Shefer A, et al. Health care utilization for pneumonia in young children after routine pneumococcal conjugate vaccine use in the United States. Arch Pediatr Adolesc Med 2007. 161:1162–8.
- Whitney CG, Farley MM, Hadler J, et al. Increasing prevalence of multidrug-resistant Streptococcus pneumoniae in the United States. N Engl J Med 2000. 343:1917–24.
- Kyaw MH, Lynfield R, Schaffner W, et al. Effect of introduction of the pneumococcal conjugate vaccine on drug-resistant *Streptococcus pneumoniae*. N Engl J Med 2006. 354: 1455–63.
- 84. CLSI. Performance Standards for Antimicrobial Susceptibility Testing; Eighteenth Informational Supplement. 2008.
- CDC. Effects of new penicillin susceptibility breakpoints for Streptococcus pneumoniae—United States, 2006–2007. MMWR 2008. 57:1353–5.
- 86. Pelton SI, Huot H, Finkelstein JA, et al. Emergence of 19A as virulent and multidrug resistant Pneumococcus in

Massachusetts following universal immunization of infants with pneumococcal conjugate vaccine. Pediatr Infect Dis J 2007. 26:468–72.

- Hicks LA, Harrison LH, Flannery B, et al. Incidence of pneumococcal disease due to non-pneumococcal conjugate vaccine (PCV7) serotypes in the United States during the era of widespread PCV7 vaccination, 1998–2004. J Infect Dis 2007. 196: 1346–54.
- Hsu KK, Shea KM, Stevenson AE, Pelton SI. Changing serotypes causing childhood invasive pneumococcal disease: Massachusetts, 2001–2007. Pediatr Infect Dis J 2010. 29: 289–93.
- Moore MR, Gertz RE Jr, Woodbury RL, et al. Population snapshot of emergent Streptococcus pneumoniae serotype 19A in the United States, 2005. J Infect Dis 2008. 197: 1016–27.
- 90. Techasaensiri C, Messina AF, Katz K, et al. Epidemiology and evolution of invasive pneumococcal disease caused by multidrug resistant serotypes of 19A in the 8 years after implementation of pneumococcal conjugate vaccine immunization in Dallas, Texas. Pediatr Infect Dis J 2010. 29:294–300.
- 91. Messina AF, Katz-Gaynor K, Barton T, et al. Impact of the pneumococcal conjugate vaccine on serotype distribution and antimicrobial resistance of invasive *Streptococcus pneumoniae* isolates in Dallas, TX, children from 1999 through 2005. Pediatr Infect Dis J 2007. 26:461–7.
- CDC. Licensure of a 13-valent pneumococcal conjugate vaccine (PCV13) and recommendations for use among children—Advisory Committee on Immunization Practices (ACIP), 2010. MMWR 2010. 59:258–61.
- Hampton LM, Farley MM, Schaffner W, et al. Prevention of antibiotic-nonsusceptible Streptococcus pneumoniae with conjugate vaccines. J Infect Dis 2012. 205:401–11.
- Gerber JS, Coffin SE, Smathers SA, Zaoutis TE. Trends in the incidence of methicillin-resistant *Staphylococcus aureus* infection in children's hospitals in the United States. Clin Infect Dis 2009. 49:65–71.
- 95. Carrillo-Marquez MA, Hulten KG, Hammerman W, et al. *Staphylococcus aureus* pneumonia in children in the era of community-acquired methicillin-resistance at Texas Children's Hospital. Pediatr Infect Dis J 2011. 30:545–50.
- Buckingham SC, King MD, Miller ML. Incidence and etiologies of complicated parapneumonic effusions in children, 1996 to 2001. Pediatr Infect Dis J 2003. 22:499–504.
- Alfaro C, Fergie J, Purcell K. Emergence of communityacquired methicillin-resistant Staphylococcus aureus in complicated parapneumonic effusions. Pediatr Infect Dis J 2005. 24: 274–6.
- Grijalva CG, Zhu Y, Nuorti JP, Griffin MR. Emergence of parapneumonic empyema in the USA. Thorax 2011. 66: 663–8.
- Gonzalez BE, Hulten KG, Dishop MK, et al. Pulmonary manifestations in children with invasive community-acquired *Staphylococcus aureus* infection. Clin Infect Dis 2005. 41: 583–90.
- Blaschke AJ, Heyrend C, Byington CL, et al. Molecular analysis improves pathogen identification and epidemiologic study of pediatric parapneumonic empyema. Pediatr Infect Dis J 2011. 30:289–94.
- 101. Korppi M, Heiskanen-Kosma T, Kleemola M. Incidence of community-acquired pneumonia in children caused by *Mycoplasma pneumoniae*: serological results of a prospective, population-based study in primary health care. Respirology 2004. 9:109–14.
- 102. Principi N, Esposito S, Blasi F, Allegra L. Role of *Mycoplasma* pneumoniae and *Chlamydia pneumoniae* in children with

community-acquired lower respiratory tract infections. Clin Infect Dis 2001. 32:1281–9.

- 103. Eun BW, Kim NH, Choi EH, Lee HJ. *Mycoplasma pneumoniae* in Korean children: the epidemiology of pneumonia over an 18-year period. J Infect 2008. 56:326–31.
- 104. Vervloet LA, Vervloet VE, Tironi Junior M, Ribeiro JD. Mycoplasma pneumoniae-related community-acquired pneumonia and parapneumonic pleural effusion in children and adolescents. J Bras Pneumol 2012. 38:226–36.
- 105. Leonardi S, del Giudice MM, Spicuzza L, et al. Lung abscess in a child with *Mycoplasma pneumoniae* infection. Eur J Pediatr **2010**. 169:1413–5.
- 106. Walter ND, Grant GB, Bandy U, et al. Community outbreak of *Mycoplasma pneumoniae* infection: school-based cluster of neurologic disease associated with household transmission of respiratory illness. J Infect Dis **2008**. 198:1365–74.
- 107. Esposito S, Blasi F, Bellini F, et al. Mycoplasma pneumoniae and Chlamydia pneumoniae infections in children with pneumonia. Mowgli Study Group. Eur Respir J 2001. 17:241–5.
- Peltola VT, McCullers JA. Respiratory viruses predisposing to bacterial infections: role of neuraminidase. Pediatr Infect Dis J 2004. 23(1 Suppl):S87–97.
- Morens DM, Taubenberger JK, Fauci AS. Predominant role of bacterial pneumonia as a cause of death in pandemic influenza: implications for pandemic influenza preparedness. J Infect Dis 2008. 198:962–70.
- 110. McCullers JA. Insights into the interaction between influenza virus and pneumococcus. Clin Microbiol Rev 2006. 19: 571–82.
- 111. Reed C, Kallen AJ, Patton M, et al. Infection with community-onset *Staphylococcus aureus* and influenza virus in hospitalized children. Pediatr Infect Dis J **2009**. 28:572–6.
- 112. Schrag SJ, Shay DK, Gershman K, et al. Multistate surveillance for laboratory-confirmed, influenza-associated hospitalizations in children: 2003–2004. Pediatr Infect Dis J **2006**. 25: 395–400.
- 113. Williams DJ, Hall M, Brogan TV, et al. Influenza coinfection and outcomes in children with complicated pneumonia. Arch Pediatr Adolesc Med 2011. 165:506–12.
- 114. Louie JK, Acosta M, Winter K, et al. Factors associated with death or hospitalization due to pandemic 2009 influenza A (H1N1) infection in California. JAMA 2009. 302:1896–902.
- 115. Shah SS, Dugan MH, Bell LM, et al. Blood cultures in the emergency department evaluation of childhood pneumonia. Pediatr Infect Dis J 2011. 30:475–9.
- 116. Shah SS, Alpern ER, Zwerling L, et al. Risk of bacteremia in young children with pneumonia treated as outpatients. Arch Pediatr Adolesc Med 2003. 157:389–92.
- 117. Grant CC, Harnden A, Mant D, et al. Why do children hospitalised with pneumonia not receive antibiotics in primary care? Arch Dis Child **2012**. 97:21–7.
- Bonadio WA. Bacteremia in febrile children with lobar pneumonia and leukocytosis. Pediatr Emerg Care 1988. 4:241–2.
- 119. Hickey RW, Bowman MJ, Smith GA. Utility of blood cultures in pediatric patients found to have pneumonia in the emergency department. Ann Emerg Med **1996**. 27:721–5.
- 120. Tsarouhas N, Shaw KN, Hodinka RL, Bell LM. Effectiveness of intramuscular penicillin versus oral amoxicillin in the early treatment of outpatient pediatric pneumonia. Pediatr Emerg Care 1998. 14:338–41.
- 121. Hoff SJ, Neblett WW, Edwards KM, et al. Parapneumonic empyema in children: decortication hastens recovery in patients with severe pleural infections. Pediatr Infect Dis J 1991. 10:194–9.
- 122. Freij BJ, Kusmiesz H, Nelson JD, McCracken GH Jr. Parapneumonic effusions and empyema in hospitalized

children: a retrospective review of 227 cases. Pediatr Infect Dis **1984**. 3:578–91.

- 123. Picard E, Joseph L, Goldberg S, et al. Predictive factors of morbidity in childhood parapneumonic effusion-associated pneumonia: a retrospective study. Pediatr Infect Dis J 2010. 29:840–3.
- 124. Lahti E, Peltola V, Waris M, et al. Induced sputum in the diagnosis of childhood community-acquired pneumonia. Thorax 2009. 64:252–7.
- 125. Uyeki TM, Prasad R, Vukotich C, et al. Low sensitivity of rapid diagnostic test for influenza. Clin Infect Dis 2009. 48: e89–92.
- 126. Self WH, McNaughton CD, Grijalva CG, et al. Diagnostic performance of the BinaxNow Influenza A&B rapid antigen test in ED patients [published online ahead of print July 13, 2012]. Amer J Emerg Med 2012. doi:10.1016/j.ajem.2012. 04.018.
- 127. Murdoch DR, Laing RT, Mills GD, et al. Evaluation of a rapid immunochromatographic test for detection of Streptococcus pneumoniae antigen in urine samples from adults with community-acquired pneumonia. J Clin Microbiol 2001. 39:3495–8.
- 128. Dowell SF, Garman RL, Liu G, et al. Evaluation of Binax NOW, an assay for the detection of pneumococcal antigen in urine samples, performed among pediatric patients. Clin Infect Dis 2001. 32:824–5.
- 129. Le Monnier A, Carbonnelle E, Zahar JR, et al. Microbiological diagnosis of empyema in children: comparative evaluations by culture, polymerase chain reaction, and pneumococcal antigen detection in pleural fluids. Clin Infect Dis **2006**. 42:1135–40.
- 130. Albrich WC, Madhi SA, Adrian PV, et al. Use of a rapid test of pneumococcal colonization density to diagnose pneumococcal pneumonia. Clin Infect Dis **2012**. 54:601–9.
- Werno AM, Anderson TP, Murdoch DR. Association between pneumococcal load and disease severity in adults with pneumonia. J Med Microbiol 2012. 61(Pt 8):1129–35.
- 132. Houben ML, Coenjaerts FE, Rossen JW, et al. Disease severity and viral load are correlated in infants with primary respiratory syncytial virus infection in the community. J Med Virol 2010. 82:1266–71.
- 133. Carvalho Mda G, Tondella ML, McCaustland K, et al. Evaluation and improvement of real-time PCR assays targeting lytA, ply, and psaA genes for detection of pneumococcal DNA. J Clin Microbiol 2007. 45:2460–6.
- 134. Abdeldaim G, Herrmann B, Molling P, et al. Usefulness of real-time PCR for lytA, ply, and Spn9802 on plasma samples for the diagnosis of pneumococcal pneumonia. Clin Microbiol Infect **2010**. 16:1135–41.
- 135. Dagan R, Shriker O, Hazan I, et al. Prospective study to determine clinical relevance of detection of pneumococcal DNA in sera of children by PCR. J Clin Microbiol **1998**. 36:669–73.
- 136. Avni T, Mansur N, Leibovici L, Paul M. PCR using blood for diagnosis of invasive pneumococcal disease: systematic review and meta-analysis. J Clin Microbiol 2010. 48:489–96.
- 137. Resti M, Moriondo M, Cortimiglia M, et al. Communityacquired bacteremic pneumococcal pneumonia in children: diagnosis and serotyping by real-time polymerase chain reaction using blood samples. Clin Infect Dis 2010. 51:1042–9.
- Esposito S, Marchese A, Tozzi AE, et al. Bacteremic pneumococcal community-acquired pneumonia in children less than 5 years of age in Italy. Pediatr Infect Dis J 2012. 31:705–10.
- 139. Azzari C, Cortimiglia M, Moriondo M, et al. Pneumococcal DNA is not detectable in the blood of healthy carrier children by real-time PCR targeting the lytA gene. J Med Microbiol 2011. 60(Pt 6):710–4.

- 140. Saglani S, Harris KA, Wallis C, Hartley JC. Empyema: the use of broad range 16S rDNA PCR for pathogen detection. Arch Dis Child 2005. 90:70–3.
- 141. Gollomp K, Rankin SC, White C, et al. Broad-range bacterial polymerase chain reaction in the microbiologic diagnosis of complicated pneumonia. [Published ahead of print] J Hosp Med 2011. doi:10.1002/jhm.911.
- Furer V, Raveh D, Picard E, et al. Absence of leukocytosis in bacteraemic pneumococcal pneumonia. Prim Care Respir J 2011. 20:276–81.
- 143. Esposito S, Bosis S, Cavagna R, et al. Characteristics of *Streptococcus pneumoniae* and atypical bacterial infections in children 2–5 years of age with community-acquired pneumonia. Clin Infect Dis 2002. 35:1345–52.
- 144. Toikka P, Irjala K, Juven T, et al. Serum procalcitonin, C-reactive protein and interleukin-6 for distinguishing bacterial and viral pneumonia in children. Pediatr Infect Dis J 2000. 19:598–602.
- 145. Cevey-Macherel M, Galetto-Lacour A, Gervaix A, et al. Etiology of community-acquired pneumonia in hospitalized children based on WHO clinical guidelines. Eur J Pediatr 2009. 168:1429–36.
- Juven T, Mertsola J, Toikka P, et al. Clinical profile of serologically diagnosed pneumococcal pneumonia. Pediatr Infect Dis J 2001. 20:1028–33.
- 147. Flood RG, Badik J, Aronoff SC. The utility of serum C-reactive protein in differentiating bacterial from nonbacterial pneumonia in children: a meta-analysis of 1230 children. Pediatr Infect Dis J 2008. 27:95–9.
- 148. Virkki R, Juven T, Rikalainen H, et al. Differentiation of bacterial and viral pneumonia in children. Thorax 2002. 57:438–41.
- 149. Heiskanen-Kosma T, Korppi M. Serum C-reactive protein cannot differentiate bacterial and viral aetiology of community-acquired pneumonia in children in primary healthcare settings. Scand J Infect Dis 2000. 32:399–402.
- 150. Gendrel D, Bohuon C. Procalcitonin as a marker of bacterial infection. Pediatr Infect Dis J 2000. 19:679–87; quiz 688.
- 151. Gendrel D, Raymond J, Assicot M, et al. Measurement of procalcitonin levels in children with bacterial or viral meningitis. Clin Infect Dis **1997**. 24:1240–2.
- Gendrel D, Assicot M, Raymond J, et al. Procalcitonin as a marker for the early diagnosis of neonatal infection. J Pediatr 1996. 128:570–3.
- 153. Assicot M, Gendrel D, Carsin H, et al. High serum procalcitonin concentrations in patients with sepsis and infection. Lancet 1993. 341:515–8.
- 154. Korppi M, Remes S. Serum procalcitonin in pneumococcal pneumonia in children. Eur Respir J 2001. 17:623–7.
- 155. Don M, Valent F, Korppi M, et al. Efficacy of serum procalcitonin in evaluating severity of community-acquired pneumonia in childhood. Scand J Infect Dis 2007. 39:129–37.
- 156. Korppi M, Don M, Valent F, Canciani M. The value of clinical features in differentiating between viral, pneumococcal and atypical bacterial pneumonia in children. Acta Paediatr 2008. 97:943–7.
- 157. Nascimento-Carvalho CM, Cardoso MR, Barral A, et al. Procalcitonin is useful in identifying bacteraemia among children with pneumonia. Scand J Infect Dis 2010. 42:644–9.

- 158. Esposito S, Tagliabue C, Picciolli I, et al. Procalcitonin measurements for guiding antibiotic treatment in pediatric pneumonia. Respir Med 2011. 105:1939–45.
- 159. Zaas AK, Chen M, Varkey J, et al. Gene expression signatures diagnose influenza and other symptomatic respiratory viral infections in humans. Cell Host Microbe 2009. 6: 207–17.
- Ramilo O, Allman W, Chung W, et al. Gene expression patterns in blood leukocytes discriminate patients with acute infections. Blood 2007. 109:2066–77.
- 161. Banchereau R, Jordan-Villegas A, Ardura M, et al. Host immune transcriptional profiles reflect the variability in clinical disease manifestations in patients with *Staphylococcus aureus* infections. PLoS One **2012**. 7:e34390.
- 162. Berry MP, Graham CM, McNab FW, et al. An interferoninducible neutrophil-driven blood transcriptional signature in human tuberculosis. Nature 2010. 466:973–7.
- 163. Slupsky CM, Rankin KN, Fu H, et al. Pneumococcal pneumonia: potential for diagnosis through a urinary metabolic profile. J Proteome Res 2009. 8:5550–8.
- Corradi M, Mutti A. Exhaled breath analysis: from occupational to respiratory medicine. Acta Biomed 2005. 76(Suppl 2):20–9.
- 165. Brogan TV, Hall M, Williams DJ, et al. Variability in processes of care and outcomes among children hospitalized with community-acquired pneumonia. Pediatr Infect Dis J 2012. 31:1036–41.
- 166. Bradley JS, Byington CL, Shah SS, et al. The management of community-acquired pneumonia in infants and children older than 3 months of age: clinical practice guidelines by the Pediatric Infectious Diseases Society and the Infectious Diseases Society of America. Clin Infect Dis 2011. 53:e25–76.
- 167. Newman RE, Hedican EB, Herigon JC, et al. Impact of a guideline on management of children hospitalized with community-acquired pneumonia. Pediatrics **2012**. 129:e597–604.
- 168. Paul R, Neuman MI, Monuteaux MC, Melendez E. Adherence to PALS sepsis guidelines and hospital length of stay. Pediatrics 2012. 130:e273–80.
- Wunderink RG, Mandell L. Adjunctive therapy in community-acquired pneumonia. Semin Respir Crit Care Med 2012. 33:311–8.
- 170. Lim WS, van der Eerden MM, Laing R, et al. Defining community acquired pneumonia severity on presentation to hospital: an international derivation and validation study. Thorax 2003. 58:377–82.
- 171. Fine MJ, Auble TE, Yealy DM, et al. A prediction rule to identify low-risk patients with community-acquired pneumonia. N Engl J Med **1997**. 336:243–50.
- 172. Chalmers JD, Singanayagam A, Akram AR, et al. Safety and efficacy of CURB65-guided antibiotic therapy in communityacquired pneumonia. J Antimicrob Chemother 2011. 66: 416–23.
- 173. Renaud B, Coma E, Labarere J, et al. Routine use of the Pneumonia Severity Index for guiding the site-of-treatment decision of patients with pneumonia in the emergency department: a multicenter, prospective, observational, controlled cohort study. Clin Infect Dis 2007. 44:41–9.