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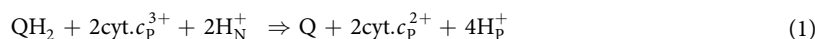
Rapid Electron Transfer within the III-IV Supercomplex in *Corynebacterium glutamicum*

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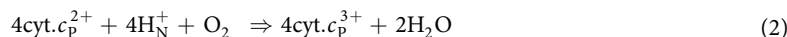
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Complex III in *C. glutamicum* has an unusual di-heme cyt. c_1 and it co-purifies with complex IV in a supercomplex. Here, we investigated the kinetics of electron transfer within this supercomplex and in the cyt. aa_3 alone (cyt. bc_1 was removed genetically). In the reaction of the reduced cyt. aa_3 with O_2 , we identified the same sequence of events as with other A-type oxidases. However, even though this reaction is associated with proton uptake, no pH dependence was observed in the kinetics. For the cyt. bc_1 -cyt. aa_3 supercomplex, we observed that electrons from the c -hemes were transferred to Cu_A with time constants 0.1–1 ms. The b -hemes were oxidized with a time constant of 6.5 ms, indicating that this electron transfer is rate-limiting for the overall quinol oxidation/ O_2 reduction activity ($\sim 210 e^-/s$). Furthermore, electron transfer from externally added cyt. c to cyt. aa_3 was significantly faster upon removal of cyt. bc_1 from the supercomplex, suggesting that one of the c -hemes occupies a position near Cu_A . In conclusion, isolation of the III-IV-supercomplex allowed us to investigate the kinetics of electron transfer from the b -hemes, via the di-heme cyt. c_1 and heme a to the heme a_3 - Cu_B catalytic site of cyt. aa_3 .

The respiratory chain in aerobic organisms is composed of a number of membrane-bound protein complexes through which electrons, originating from the oxidation of organic compounds, are transferred to finally reach O_2 . The free energy released in this process is employed to establish a proton electrochemical gradient across the membrane, which is used to synthesize ATP by the F_1F_0 ATP synthase or for secondary transmembrane transport. In mitochondria, Complex III (the cytochrome (cyt.) bc_1 -complex) of the respiratory chain links the two-electron oxidation of quinol (QH_2) to the one-electron reduction of water-soluble cyt. c in the respiratory chain:



where the subscripts N and P refer to the more negative and positive sides of the membrane, respectively. Reduced cyt. c delivers electrons to Complex IV (cytochrome c oxidase, Cyt cO), which catalyzes the reduction of dioxygen to water:



Part of the respiratory-chain enzymes in mitochondria are organized in so-called supercomplexes^{1–11}. There are also reports of supercomplexes in bacteria, for example in *Paracoccus denitrificans*. In this bacterium, depending on the detergent used, supercomplexes composed of respiratory-enzyme complexes III-IV or I-III-IV at variable stoichiometries were identified^{11,12–14}. Furthermore, in several bacterial systems electrons could be transferred directly between complexes III and IV via a membrane-anchored cyt. c ^{15–17}.

Corynebacterium glutamicum is a rod-shaped, Gram positive soil bacterium, which harbors two different terminal oxidases; an aa_3 -type Cyt cO and a bd -type menaquinol oxidase^{18–20}. The cyt. aa_3 in *C. glutamicum* is a four-subunit protein complex, comprising subunits CtaD, C, E, and F. Mass spectrometric analyses of the purified

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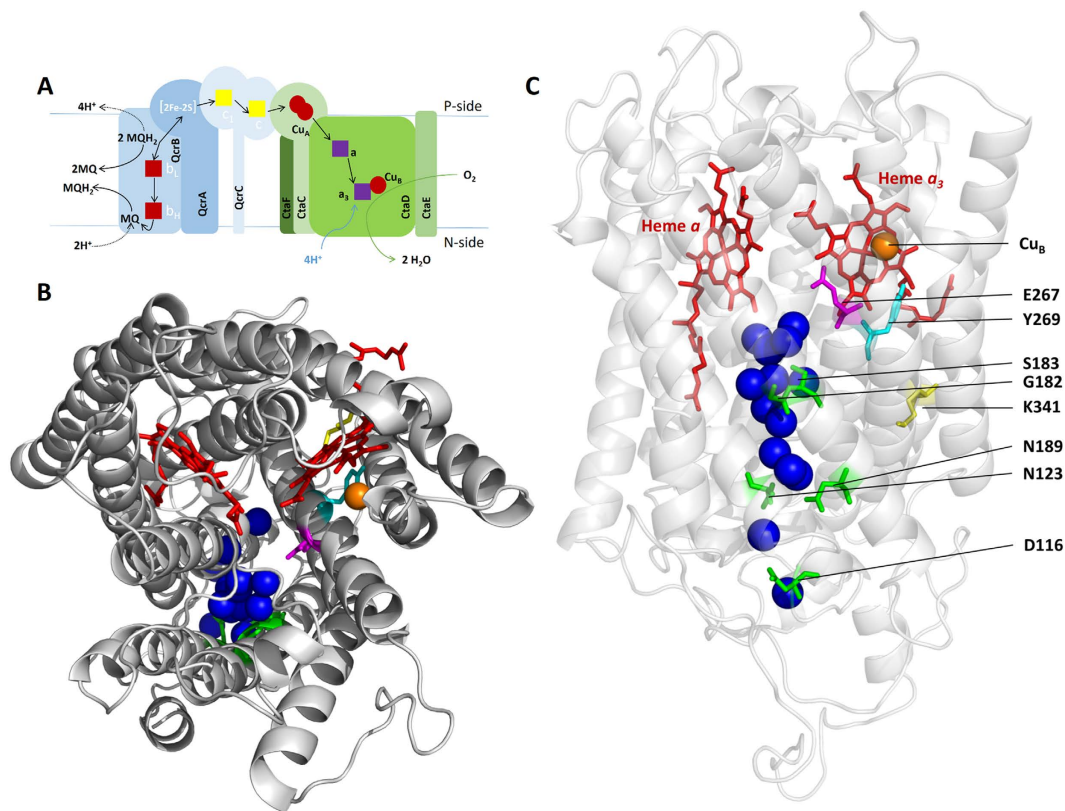
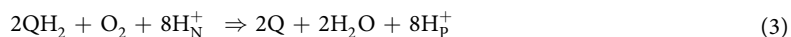


Figure 1. Schematic model of the *C. glutamicum* cyt. *bc*₁-CytcO supercomplex and a homology model of subunit I in CytcO. (A) Cyt. *bc*₁ (in blue) and CytcO (in green), represented as a supercomplex. The electron-transfer pathway is indicated by black bold arrows. Reduction of O₂ at the heme a₃-Cu_B catalytic site is accompanied by proton uptake from solution. The Figure is adapted from¹⁹. **(B,C)** A structural model of subunit I of CytcO from *C. glutamicum* **(B,C)**, top and side view, respectively), based on the structure of aa₃ (PDB ID 1M56) from *R. sphaeroides*³⁹. Water molecules found in the structure of the *R. sphaeroides* CytcO are shown in the model to indicate the location of the D proton pathway (some residues defining this pathway are shown in green, Glu267 in magenta). Tyr269 is presumably part of the catalytic site. The putative K-pathway residue Lys341 (equivalent to Lys362 in *R. sphaeroides* CytcO) is shown as a yellow stick.

CytcO revealed that instead of heme *a*, the *C. glutamicum* contains heme a_s in the active site. Furthermore, the *C. glutamicum* CytcO harbors an extra charged amino-acid cluster near the cyt. *c*-binding domain of subunit II (CtaC), which was suggested to interact with the second cyt. *c* of the cyt. *bc*₁ complex¹⁸.

Complex III in *C. glutamicum* is a three-subunit protein, containing cyt. *c*₁ (QcrC), the Rieske iron-sulfur protein (QcrA), and cyt. *b* (QcrB) (Fig. 1A)^{21,22}. The QcrC subunit contains two CXXCH heme-binding motifs, suggesting that this protein complex contains two *c*-type hemes^{19,22}, hence referred to as the di-heme *c*₁ cyt. *bc*₁ complex. Furthermore, *C. glutamicum* contains no other *c*-type hemes, which suggested that the second heme *c* in cyt. *bc*₁ shuttles electrons between complexes III and IV that form a tight supercomplex²¹. Such a supercomplex was isolated, its shape was determined using electron microscopy²³ and it was shown to exhibit quinol-oxidase activity²⁰:



The involvement of the second cyt. *c* of cyt. *bc*₁ in electron transfer between cyt. *c*₁ and Cu_A, the electron acceptor of CytcO (see below), is also supported by mutagenesis data²⁰. Quinol-oxidase activity was also found for the cyt. *bc*₁-aa₃ supercomplex from *Mycobacterium smegmatis*²⁴, a bacterium also devoid of soluble cyt. *c*.

The mechanism of the cyt. *bc*₁ complex involves a Q-cycle in which the net reaction results in oxidation of menaquinol and reduction of cyt. *c* (see Fig. 1A), linked to proton uptake at the N side and release to the P side of the membrane. The process is initiated by binding of a menaquinol at the quinone-binding site, Q_P, located near the low-potential heme b_L (see Fig. 1A)²⁵. In the next step, one electron is transferred from the menaquinol, via the Rieske protein [2Fe-2S] cluster, to cyt. *c*₁ while one electron is transferred via heme b_L and heme b_H to a menaquinone bound at a second quinone-binding site, Q_N. This bifurcated electron transfer yields reduced cyt. *c*₁, a semireduced menaquinone at the Q_N-site and release of two protons to the P-side of the membrane. After binding a second menaquinol at the Q_P-site the same process is repeated. The doubly reduced menaquinone at the Q_N site picks up two protons from the N-side of the membrane to form a menaquinol that is released into the membrane.

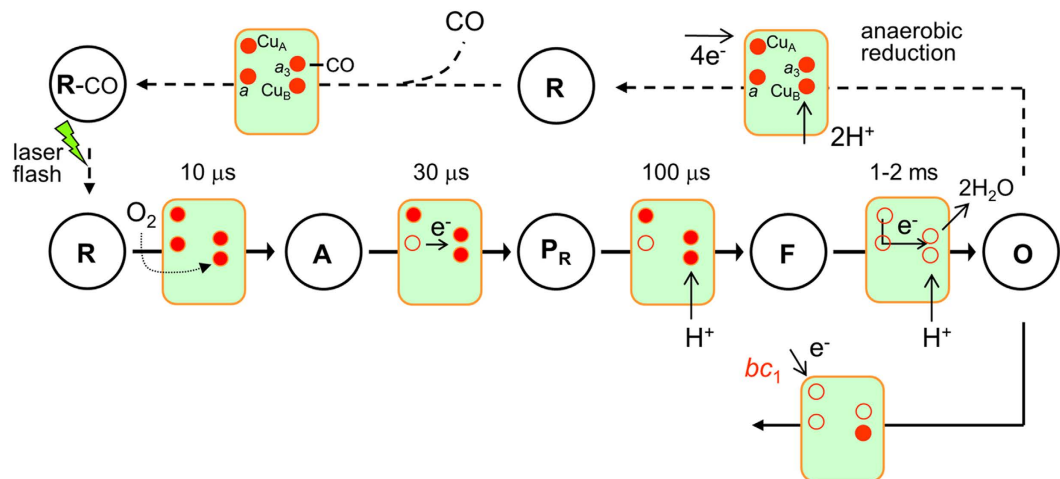


Figure 2. Schematic illustration of the reactions in Cyt cO studied in this work. The four red circles represent the redox-active metal sites as indicated (filled - reduced). During preparation of the sample, the oxidized Cyt cO (O) is reduced by 4 electrons under an atmosphere of pure N_2 (anaerobic reduction) to yield the fully reduced Cyt cO (state R). The sample is then incubated under an atmosphere of CO, which binds to heme a_3 at the catalytic site forming R-CO (see dashed line). The reaction studied in this work (indicated by a solid line) is started by photo-dissociation of the CO ligand (laser flash), yielding the reduced Cyt cO (R), which allows O_2 to bind to heme a_3 forming state A with a time constant of $\sim 10 \mu s$ at 1 mM O_2 . An electron is then transferred from heme a to the catalytic site forming the “peroxy” state called P_R with a time constant of $\sim 30 \mu s$. Next, a proton is taken up from solution forming the ferryl state F with a time constant of $\sim 100 \mu s$. In most oxidases studied to date, at the same time the electron at Cu_A equilibrates with heme a . This electron transfer is not indicated in the figure because in the *C. glutamicum* Cyt cO the equilibrium is shifted towards reduced Cu_A . In the final step of the reaction the electron from Cu_A (or the Cu_A -heme a equilibrium) is transferred to the catalytic site forming the oxidized Cyt cO (state O) in 1–2 ms. In cyt. bc_1 -Cyt cO supercomplex, as soon as Cu_A is partly oxidized ($\tau \cong 100 \mu s$), electrons are transferred from the cyt. bc_1 complex to the Cyt cO. Further electron transfer from the c and b hemes occurs over a slower time scale. For simplicity, all these electron transfers are indicated schematically in the lower part of this scheme (the time constants are given in Fig. 8). The pumped protons are not shown.

Cytochrome c oxidase, which belongs to a large family of enzymes called the heme-copper oxidases, catalyzes oxidation of cyt. c and reduction of O_2 to H_2O . Here, we refer to the *C. glutamicum* cytochrome aa_3 as a Cyt cO even though in the cyt. bc_1 -cyt. aa_3 supercomplex it receives electrons from quinol, via the cyt. bc_1 complex and not from (a water-soluble) cyt. c . The heme-copper oxidases are classified according to sequence, phylogenetic, and structural analyses into three main classes, A, B and C^{26,27}. Subunit I, the core subunit shared by all of the three oxidase types, contains a low-spin heme group and the catalytic site, which is composed of a copper ion, Cu_B , and a high-spin heme (for review on structure and function of the Cyt cOs, see^{28–37}). The fourth redox-active site, Cu_A , is found in subunit II. The most studied Cyt cOs are those from bovine heart mitochondria and the bacterial aa_3 Cyt cOs from *Paracoccus denitrificans* and *Rhodobacter sphaeroides*, which all belong to the A-class. In these Cyt cOs, electrons delivered by cyt. c are transferred consecutively to the Cu_A site, heme a and finally to the binuclear center composed of heme a_3 and Cu_B . In these bacterial Cyt cOs protons are transferred to the catalytic site through two pathways denoted by the letters D and K after conserved residues Asp132 and Lys362, respectively (numbering refers to the *R. sphaeroides* aa_3 -type Cyt cO).

Internal electron and proton-transfer reactions in Cyt cOs from several organisms have been studied in the past (see e.g.^{30,31,34,37}). An experimental technique that yields particularly detailed information about the sequence and rates of these reactions is the so-called flow-flash technique, in which the oxidative part of a reaction cycle (single turnover) of the enzyme is monitored. In this approach, the oxidase is first fully reduced by four electrons (see dashed line in Fig. 2) and incubated under an atmosphere of carbon monoxide, which binds to heme a_3 at the catalytic site. The Cyt cO-CO complex is then rapidly mixed with O_2 -containing solution followed in time by light-induced dissociation of the blocking CO ligand, which allows O_2 from the surrounding medium to bind. Initially, the reduced Cyt cO (the state is called R) binds O_2 to heme a_3 with a time constant of $\sim 10 \mu s$ (at 1 mM O_2) yielding the ferrous heme a_3 - O_2 state (state A) (see Fig. 2). Next, an electron is transferred from heme a to the catalytic site and the O=O bond is cleaved, resulting in formation of a ferryl intermediate that is called “peroxy” (P_R) for historical reasons ($\tau \cong 30 \mu s$). A proton is then taken up to the catalytic site resulting in formation of the ferryl state, F, with a time constant of $\sim 100 \mu s$ at pH 7. At the same time, an electron is transferred from Cu_A to heme a in a small fraction of the population (not shown in Fig. 2). Finally, the last electron is transferred from the Cu_A -heme a equilibrium to the catalytic site forming the oxidized Cyt cO (state O) with a time constant of ~ 1 ms at pH 7 (the time constants are those observed with the *R. sphaeroides* Cyt cO³⁸).

In the present study we used the flow-flash technique to investigate the reaction of the purified *C. glutamicum* Cyt cO, as well as the cyt. bc_1 -Cyt cO supercomplex with O_2 . The data indicate rapid electron transfer from the cyt.

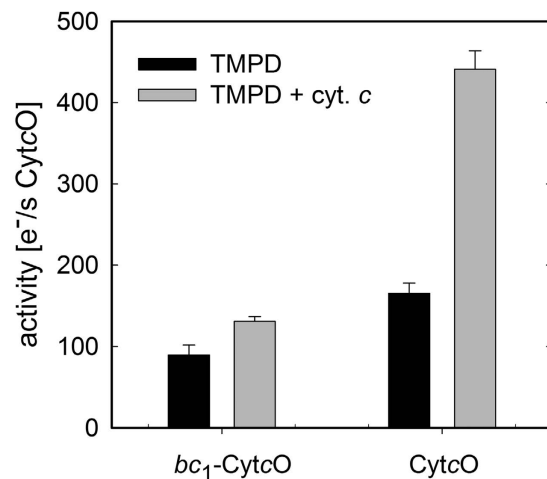


Figure 3. O₂-reduction activity upon addition of an electron donor to CytcO. The measurements were done with either CytcO or the cyt. *bc*₁-CytcO super complex using ascorbate as electron donor and, TMPD and cyt. *c* as electron mediators. The activity was determined by measuring the O₂-reduction rate ($e^-/s/CytcO$). Conditions: 100 mM Tris-HCl pH 7.5, 100 mM NaCl, 2 mM MgSO₄, 0.015% (w/v) DDM, 2 mM sodium ascorbate, 0.2 mM TMPD, with or without 24 μM cyt.*c*.

*bc*₁-complex to the CytcO, suggesting a functional supercomplex in which the additional cyt. *c* of the cyt. *bc*₁ complex acts as an electron bridge between the two respiratory-enzyme complexes. Furthermore, a comparison of the TMPD/cyt. *c*-driven O₂-reduction activities with the cyt. *bc*₁-CytcO complex and with CytcO alone, indicate that the second heme *c* of the cyt. *bc*₁ complex does bind at the surface of the CytcO, presumably near Cu_A. We also studied the electron-transfer kinetics in purified CytcO and found that reaction steps linked to proton uptake during O₂ reduction displayed pH-independent kinetics, suggesting differences in the pK_a of residues involved in proton transfer, compared to other A-type CytcOs.

Results

Sequence Alignment and Homology Modeling. To analyze the structural characteristics of CytcO from *C. glutamicum*, we performed homology modeling of the highly conserved subunit I with the three-dimensional structure of that from the *R. sphaeroides* aa₃-type CytcO³⁹ using the SwissModel program^{40–42} (Fig. 1B,C). The two protein sequences are ~40% identical and ~60% similar. On the basis of the sequence itself and this model, the *C. glutamicum* protein is identified as an A1-type CytcO. It holds the active site tyrosine (Tyr269 in *C. glutamicum*)²¹ and Glu267 in the D pathway (Glu286 in *R. sphaeroides*). Furthermore, in the *C. glutamicum* CytcO an Asp residue (Asp116) is found at the same location as Asp132, the entry point of the D pathway in the *R. sphaeroides* CytcO. Other residues that are discussed below are Asn123 and Asn189 in the *C. glutamicum* CytcO (Asn139 and Asn207, respectively, in the *R. sphaeroides* CytcO).

Purification and multiple turnover activity. We have purified the cyt. *bc*₁-aa₃ supercomplex from strain ΔC-D_{st} and the aa₃ oxidase from the cyt. *bc*₁ deficient strain ΔQ-D_{st}. The quality of the preparations was assessed using SDS gel electrophoresis (Figure S1) and dithionite-reduced minus ferricyanide-oxidized difference spectra of the two samples (Figure S2). As expected, both spectra showed the heme *a* signature (605 nm), and the supercomplex preparation displayed additional peaks for *b*-heme (560 nm) and *c*-heme (550 nm) (Figure S2). The approximately similar height of the three alpha peaks indicates the presence of a 1:1 complex of cyt. *bc*₁ (2 heme *b* and 2 heme *c*) and cytochrome aa₃ (two heme *a*). The reduced CO-bound minus reduced difference spectrum (Figure S3) displayed the characteristic features of CO binding to heme a₃.

Next, the quinol-oxidase activity of the purified supercomplex was measured by following O₂ consumption after addition of a pre-reduced menaquinone. This process involves quinol oxidation by the cyt. *bc*₁ complex followed by electron transfer to CytcO, where oxygen is reduced to water. The measured quinol-oxidase activity for the purified supercomplex was 210 ± 20 e⁻/s (SD, *n* = 4 measurements) (normalized to the total CytcO), i.e. in the same range as the published value²⁰. The activity dropped rapidly upon flash freezing and thawing the preparation. Therefore, the sample was kept at 4 °C, where no activity loss was observed in the time frame between purification and functional studies (typically ~1 day, but the preparation was stable up to 7 days, Figure S4).

CytcO activity of the purified supercomplexes and pure CytcO (without cyt. *bc*₁) was also measured using ascorbate as electron source and either TMPD, or TMPD and water-soluble cyt. *c* as electron mediators (Fig. 3). For the cyt. *bc*₁-CytcO, the TMPD activity was 90 ± 10 e⁻/s and it increased to 130 ± 10 e⁻/s (SD, *n* = 3) upon addition of free cyt. *c*, i.e. both rates were lower than the coupled quinol activity. For pure CytcO, the activity increased from 160 ± 10 e⁻/s to 440 ± 20 e⁻/s (SD, *n* = 3) upon addition of cyt. *c*, thus displaying a much greater stimulation by the soluble electron carrier.

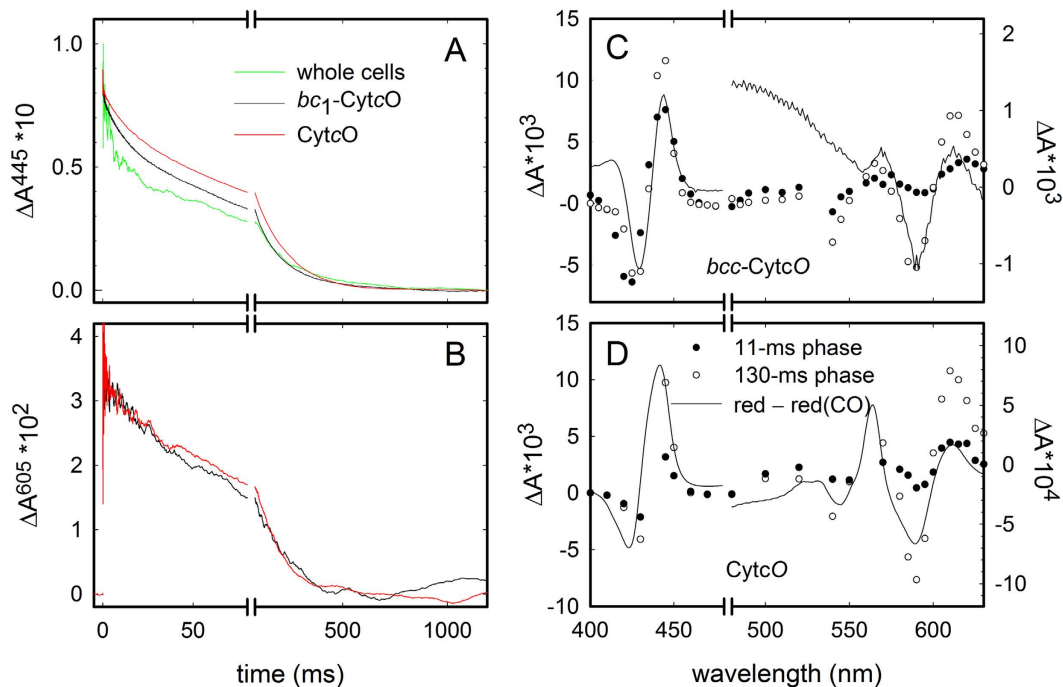


Figure 4. CO-recombination kinetics and kinetic difference spectra. Absorbance changes at 445 nm (A) and 605 nm (B) associated with light-induced CO dissociation and recombination with detergent-purified CytcO, cyt. *bc*₁-CytcO and with the ΔC -D_{St} whole cells. All traces were scaled to yield the same absorbance changes at $t = 0$. A laser artifact at $t = 0$ has been removed for clarity. Kinetic difference spectra, i.e. the amplitude of the absorbance changes of the two kinetic components (slow and fast components, empty and filled circles, respectively), for CytcO (C) and the cyt. *bc*₁-CytcO complex (D). Note the different ordinate scales before and after the break on the abscissa, respectively. The black line represents the reduced minus CO-bound difference spectrum. Experimental conditions: 100 mM Tris-HCl at pH 7.5, 1 μ M PMS, 5 mM sodium ascorbate. For the cyt. *bc*₁-CytcO supercomplex, 1 mM sodium dithionite was added to reduce the *b*-hemes.

Kinetics of CO rebinding after flash photolysis. CO binds with high affinity to heme *a*₃ in the binuclear site in the reduced CytcO. Upon illumination with a short laser flash, CO dissociates instantly, but in the absence of oxygen rebinds to the binuclear site (CO recombination). The CO-recombination kinetics was biphasic where the relative contribution of the two components varied slightly between the purified CytcO, the cyt. *bc*₁-CytcO complex and the intact membrane (Fig. 4A,B). This observation indicates the presence of two CytcO populations, also in the native membrane (see Discussion). The time constants for the fast and the slow phases were about the same for all samples (pure CytcO, the purified cyt. *bc*₁-CytcO supercomplex and whole cells), i.e., 11 ± 3 ms and 130 ± 30 ms (SD, $n = 15$) (at 1 mM CO), respectively. The kinetic difference spectra of the fast and the slow components were similar (Fig. 4C,D).

The apparent binding affinity for CO to the binuclear center of the oxidase was determined by measuring the observed CO-recombination rates for the slower kinetic phase at different CO-concentrations, for both the oxidase alone and the purified supercomplex (Figure S5). The second-order rate constants, determined from a linear fit to the data in Figure S5, were $7.6 \pm 0.2 \times 10^3 \text{ M}^{-1}\text{s}^{-1}$ and $9.4 \pm 0.2 \times 10^3 \text{ M}^{-1}\text{s}^{-1}$ for the CytcO and cyt. *bc*₁-CytcO complex, respectively. The rate of the faster component was CO-concentration independent.

Single-turnover measurements. Figure 5 shows absorbance changes after flash-induced dissociation of the CO ligand from the reduced cyt. *bc*₁-CytcO supercomplex in the presence of O₂. At 605 nm (Fig. 5A), three kinetic phases were resolved. The initial decrease in absorbance, with a time constant of $\sim 25 \mu\text{s}$ [The standard deviation of the time constants was typically 10% of the measured values ($n = 7-17$), except for the $P \rightarrow F$ reaction for which the standard deviation was 20% of the measured values (for both the pure CytcO and the supercomplex)], is attributed to oxidation of heme *a*, i.e. electron transfer from heme *a* to heme *a*₃, which yields the P_R state at the catalytic site (formation of state **A** was not resolved, see below). This component is also seen at 445 nm (initial decrease, Fig. 5B). At 605 nm, the decrease is followed by a small increase in absorbance in the time range 0.05–0.2 ms that is attributed to re-reduction of heme *a* with a time constant of 120 μs concomitant with the $P_R \rightarrow F$ reaction at the catalytic site. The slowest absorbance decrease ($\tau \cong 1.7$ ms) at 605 nm is associated with oxidation of the CytcO ($F \rightarrow O$), also seen at 445 nm (Fig. 5B). At 830 nm, oxidation of Cu_A is observed as an increase in absorbance. The small initial increase in absorbance is associated with oxidation of Cu_A during the $P_R \rightarrow F$ transition, i.e. $\tau \cong 120 \mu\text{s}$ while the major oxidation component displayed a time constant of ~ 1.7 ms (Fig. 5D).

The absorbance changes at 550 nm (Fig. 5E) are mainly attributed to *c*-heme absorption, where a decrease in absorbance is associated with oxidation of the hemes. Two kinetic components with time constants of $\sim 120 \mu\text{s}$ and ~ 1.7 ms, respectively, were observed, i.e. concomitant with electron transfer from Cu_A to heme *a* during the

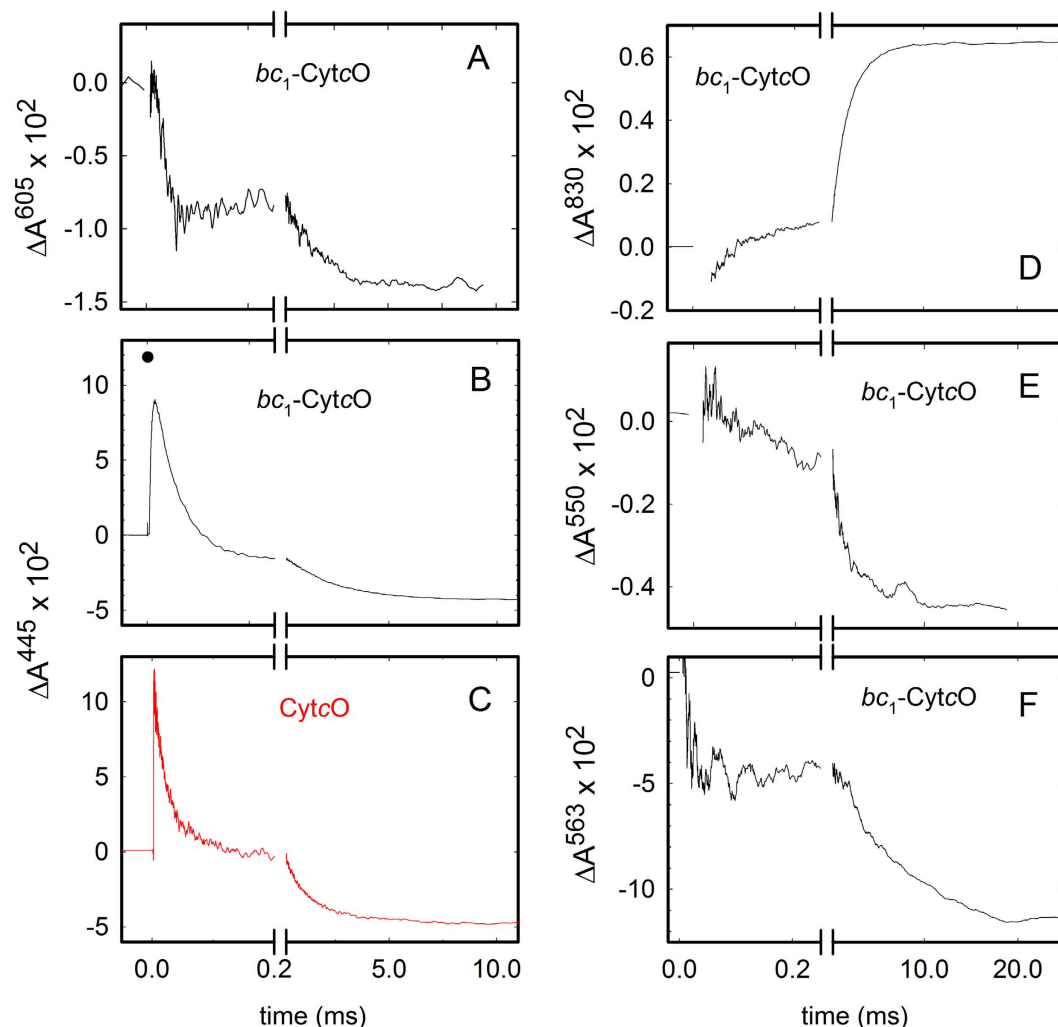


Figure 5. Absorbance changes associated with the reaction of the reduced cyt. bc_1 -Cyt cO complex and Cyt cO with O_2 . Absorbance changes were recorded over time at 605 nm (A), 445 nm (B) both associated with oxidation of a -hemes, and at 445 nm with the pure Cyt cO (C) (this trace was multiplied by a factor of two to yield approximately the same absorbance changes as those seen in (B)). The absorbance changes at 830 nm (D) are associated with oxidation of Cu_A , at 550 nm (E) oxidation of cyt. c , and at 563 nm (F) oxidation of the b -hemes. The CO ligand was dissociated at $t=0$. Experimental conditions: 100 mM Tris-HCl at pH 7.5, 4 mM ascorbate, 1 μ M PMS, $\sim 900 \mu$ M O_2 . In addition, dithionite was added to fully reduce the cyt. bc_1 complex. The Cyt cO concentration was $\sim 1 \mu$ M. A laser artifact at $t=0$ has been removed for clarity. The filled circle in (B) represents the extrapolated initial absorbance after the laser flash.

$P_R \rightarrow F$ reaction and during the $F \rightarrow O$ reaction. Finally, at 563 nm (Fig. 5F) after the unresolved initial drop in absorbance (presumably associated with a small absorbance contribution from CO dissociation), a further decrease in absorbance associated with oxidation of heme b ($\tau \cong 6.5$ ms) was observed, i.e. oxidation of heme b occurred after oxidation of the Cyt cO and the c heme (compare panels D-F in Fig. 5, after the break on the abscissa).

The concentration of reacting Cyt cO (i.e. Cyt cO from which the CO ligand is dissociated) was estimated from the change in absorbance at $t=0$ at 445 nm (Fig. 5B), which yields $\sim 0.15 \mu$ M Cyt cO (using an absorption coefficient (ϵ) of $82 \text{ mM}^{-1} \text{ cm}^{-1}$ ⁴³). The absorbance at $t=0^+$, i.e. just after CO dissociation corresponds to that of reduced Cyt cO . Consequently, the decrease in absorbance from this point until $t \cong 10$ ms, when the reaction is essentially over, corresponds to the amount oxidized Cyt cO ($\sim 0.10 \mu$ M, using $\epsilon = 164 \text{ mM}^{-1} \text{ cm}^{-1}$ ⁴⁴). Consequently, $\sim 0.05 \mu$ M ($\sim 35\%$) of the reacting Cyt cO becomes re-reduced by the cyt. bc_1 complex. From the absorbance changes at 550 nm and 563 nm we estimate that $\sim 0.025 \mu$ M ($\epsilon = 19.1 \text{ mM}^{-1} \text{ cm}^{-1}$ ²⁰) heme b and $\sim 0.028 \mu$ M ($\epsilon = 22 \text{ mM}^{-1} \text{ cm}^{-1}$ ²⁰) heme c , respectively, become oxidized, which together account for $\sim 0.05 \mu$ M Cyt cO that is re-reduced during the experiment. It should be noted that these estimations are only approximate because we were not able to accurately resolve O_2 binding to the reduced heme a_3 in all samples. In part, this problem is attributed to the requirement to add dithionite in order to fully reduce the cyt. bc_1 -Cyt cO supercomplex. Because dithionite reduces O_2 directly during mixing, the O_2 concentration was lowered before initiation

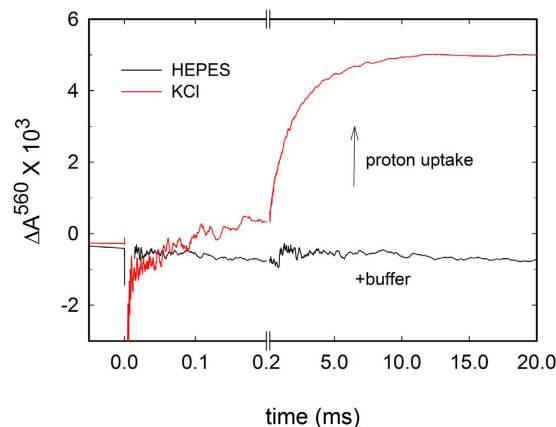


Figure 6. Absorbance changes of the pH dye phenol red, associated with proton uptake during O_2 reduction. The reaction of the reduced cyt. bc_1 -Cyt c O complex with O_2 was initiated by a laser flash at $t = 0$ (see Fig. 5). Absorbance changes were monitored over time at 560 nm in the absence and presence of buffer, respectively. Experimental conditions: 0.05% DDM, 100 μ M EDTA, 40 μ M Phenol red at pH 7.8 2 mM ascorbate, 0.2 μ M Hexamminerutheniumchloride, \sim 900 μ M O_2 and either 100 mM KCl or 100 mM HEPES. The Cyt c O concentration was \sim 5 μ M. A laser artifact at $t = 0$ has been removed for clarity.

of the reaction of Cyt c O with O_2 thereby slowing the $R \rightarrow A$ reaction. Consequently, it was difficult to resolve the associated absorbance changes from those associated with the next, $A \rightarrow P_R$ transition.

The single-turnover reaction was also studied with purified Cyt c O (Fig. 5C). Here, heme a was oxidized with a time constant of \sim 21 μ s ($R \rightarrow P_R$), followed in time by re-reduction from Cu_A with a time constant of \sim 90 μ s ($P_R \rightarrow F$) and oxidation with a time constant of \sim 1.3 ms ($F \rightarrow O$). These time constants were almost the same as those observed with the cyt. bc_1 -Cyt c O supercomplex. No changes in absorbance were observed at 550 nm nor 563 nm for the purified oxidase (data not shown). The end absorbance level at 445 nm was slightly lower with pure Cyt c O than with Cyt c O that is part of the supercomplex (c.f. Fig. 5B,C), which means that the former was more oxidized than the latter. This observation presumably reflects the re-reduction of Cyt c O by the cyt. bc_1 complex in the latter. However, as mentioned above, it was difficult to quantify the relative absorbance differences in the two samples because we were unable to confidently scale the two different traces to each other due to unresolved absorbance changes associated with O_2 binding.

Binding of water-soluble cyt. c to e.g. the bovine heart Cyt c O is highly dependent on the salt concentration, reflecting electrostatic interactions of the two proteins⁴⁵. In order to investigate whether or not the interactions of Cyt c O with cyt. c in the supercomplex could be disrupted, we studied the reaction with O_2 at increasing ionic strengths (addition of KCl). As seen in Figure S6, the amplitude of the absorbance changes associated with the $F \rightarrow O$ reaction decreased slightly (indicating more re-reduction of Cyt c O) rather than increasing with increased ionic strength (we expect more oxidation of Cyt c O upon cyt. c dissociation), which indicates that the cyt. c -Cyt c O interactions were not disrupted at high salt concentrations.

Net Proton Uptake from Solution. The protons required for the reduction of O_2 to water are taken up from the medium, a process that can be monitored during flow-flash experiments in the absence of buffer by use of a pH-sensitive dye. Using phenol red, we studied proton uptake during reaction of the reduced cyt. bc_1 -Cyt c O supercomplex with O_2 , following absorbance changes at 560 nm. As seen in Fig. 6, the absorbance increased over time, which indicates net proton uptake during the reaction. The process displayed two components with time constants of 130 μ s (\sim 25% of the total absorbance change) and 1.9 ms, i.e. they coincided with the $P_R \rightarrow F$ and $F \rightarrow O$ reactions, respectively. Upon addition of buffer the dye signal was quenched and only a very small decrease in absorbance associated with oxidation of b -hemes was observed (c.f. Fig. 6).

pH-dependence. Reactions steps that are associated with proton uptake from solution (e.g. the $F \rightarrow O$ step of the reaction of reduced Cyt c O with O_2) often display pH dependent rates^{30,46,47}. We therefore investigated the pH dependence of the reaction with O_2 of the reduced cyt. bc_1 -Cyt c O supercomplex and Cyt c O alone (Fig. 7A,B). At 445 nm, for the supercomplex, only a slight pH dependence was observed for the $F \rightarrow O$ reaction rate. As the total amplitude of the oxidation also changed, the results presumably reflect a pH-dependence in the extent of re-reduction by the cyt. bc_1 complex, i.e. “down-stream” steps of the reaction. Also with the pure Cyt c O, the $F \rightarrow O$ reaction rate was pH independent and all kinetic components displayed essentially the same amplitudes in the measured pH range (c.f. data at pH 7.5 and 8.5 in Fig. 7B).

Na^+ -dependence. Because we did not observe any significant pH-dependence in the reaction rates with O_2 , we also investigated the Na^+ -concentration dependence to test the possibility that the Cyt c O transports Na^+ (see refs 48 and 49). As can be seen in Figure S7, the addition of Na^+ had a slight effect on the time constant of the $A \rightarrow P_R$ reaction, i.e. electron transfer from heme a to the catalytic site (inset Figure S7), but this reaction step is not linked to pumping in other A-type oxidases. No effect on the kinetics of the $F \rightarrow O$ reaction was observed.

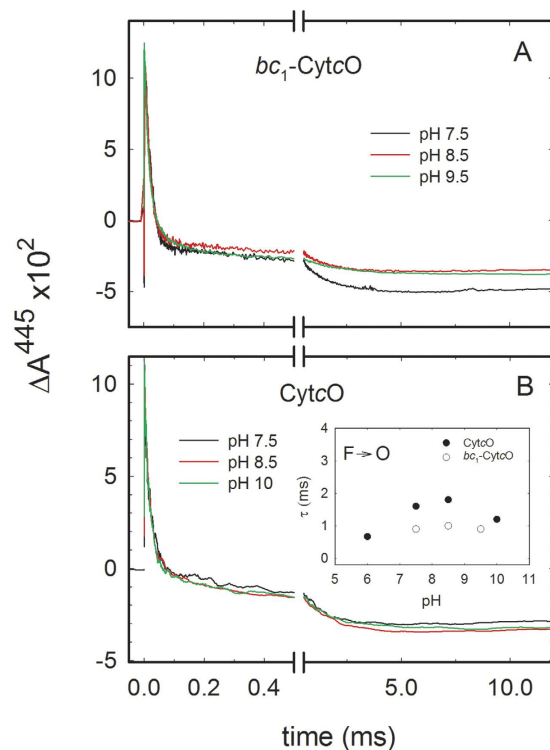


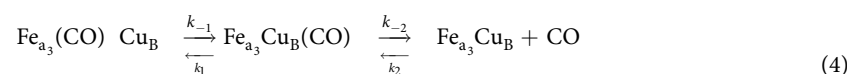
Figure 7. pH dependence of the reaction of cyt. *bc*₁-Cyt_cO (A) and Cyt_cO (B) with O₂. Kinetic data at representative pH-values are shown (color codes as indicated in the graph) and the time constants are summarized in the inset to panel B (measurements were also performed at pH 6 with the Cyt_cO). The CO ligand was dissociated at *t* = 0. Experimental conditions: ~1 μM Cyt_cO, 10–100 mM Tris-HCl (see Materials and Methods), 4 mM ascorbate, 1 μM PMS and 100 μM dithionite. The pH was set as described in the Materials and Methods section, and measured after mixing using a pH meter. A laser artifact at *t* = 0 has been removed for clarity.

Discussion

Purification and Activity. The purified cyt. *bc*₁-Cyt_cO complex displayed quinol-oxidase activity, i.e. electrons were transferred first from quinol to the cyt. *bc*₁ complex and then, via the two *c*-hemes²⁰, to the Cyt_cO, which reduces oxygen to water. The activity was ~210 e⁻/s, which is in good agreement with previously published results²⁰. The data are also consistent with those obtained for the cyt. *bc*₁-Cyt_cO supercomplex from *Mycobacterium smegmatis*²⁴, which exhibited quinol-oxidase activity even at high detergent concentrations, supporting the presence of the cyt. *bc*₁-Cyt_cO supercomplex. Both the *C. glutamicum* cyt. *bc*₁-Cyt_cO and Cyt_cO preparations exhibited TMPD-driven O₂-reduction activities. In both cases the activity increased upon addition of soluble cyt. *c*. However, the increase was larger for the pure Cyt_cO (a factor of ~2.8) than for the supercomplex (a factor of ~1.4) (but see data with the *M. smegmatis*²⁴). This observation suggests that in *C. glutamicum* cyt. *c* is more accessible for binding to the Cyt_cO upon removal of the cyt. *bc*₁ complex, which indicates that in the supercomplex one of the *c*-hemes is located near the Cyt_cO electron entry point (i.e. presumably near Cu_A). Thus, the results indicate that pure Cyt_cO (i.e. with the cyt. *bc*₁ removed) is capable of binding soluble cyt. *c* in spite of the presence of an extra loop of charged amino acids located at the cyt. *c* binding site¹⁸ (this binding can only occur in the mutant where the cyt. *bc*₁ complex is removed, i.e. not *in vivo*). Formation of a stable cyt. *bc*₁-Cyt_cO complex that is capable of transferring electrons directly from cyt. *bc*₁ to Cyt_cO is also consistent with the rapid electron transfer from the *b* and *c*-hemes to Cyt_cO (see below).

We note that the TMPD and cyt. *c*-oxidation activities measured here are higher than those presented previously^{18,20}. The discrepancy is presumably due to differences in experimental conditions. While in the earlier studies the rate was derived from changes in the concentration of reduced electron donor, here the data was obtained by measuring the O₂-reduction rate at a constant concentration of the reduced electron donor (with excess ascorbate).

Ligand Binding to Cyt_cO. Results from earlier studies with e.g. the bovine heart oxidase indicate that upon pulsed illumination the CO ligand dissociates from heme *a*₃ and binds transiently to Cu_B before it dissociates into solution⁵⁰:



Dissociation of CO from heme a_3 in the dark is very slow ($k_{-1} \cong 0.03 \text{ s}^{-1}$), but upon illumination the ligand moves to Cu_B in $\ll 10 \text{ ns}$, if the light intensity of the pulse is strong enough (such as in this study). The dissociation rate constant from Cu_B , k_{-2} , is $\sim 7 \cdot 10^5 \text{ s}^{-1}$ with the bovine heart Cyt $c\text{O}^{50}$. Recombination of CO occurs via Cu_B with a second-order process ($k_2 \cong 1 \cdot 10^8 \text{ M}^{-1} \text{ s}^{-1}$). The rate for internal CO transfer from Cu_B to heme a_3 , k_1 , is $\sim 10^3 \text{ s}^{-1}$. The observed rate of CO recombination is approximately given by the fraction of Cu_B with bound CO (middle state in scheme 1) multiplied by the rate of CO transfer from Cu_B to heme a_3 ,

$$k_{\text{obs}} \cong \frac{k_2}{k_{-2} + k_2} k_1 \quad (5)$$

With the rate constants given above, we obtain $k_{\text{obs}} \cong 0.12 \times 1000 \text{ s}^{-1} = 120 \text{ s}^{-1}$ ($\tau \cong 8 \text{ ms}$) at 1 mM CO.

The CO-recombination kinetics measured with the *C. glutamicum* Cyt $c\text{O}$ was biphasic (see Fig. 4A). The kinetic difference spectra of the two components with the purified supercomplex and the oxidase alone were similar (see Fig. 4C,D), indicating that both components are associated with CO binding to heme a_3 after light-induced dissociation. Consequently, the data indicate the presence of two populations of Cyt $c\text{O}$ with different CO-recombination rates to heme a_3 . The smaller ($\sim 25\%$) Cyt $c\text{O}$ population, displayed a CO-concentration independent rate constant of $\sim 90 \text{ s}^{-1}$ (also the relative amplitude was independent on the CO concentration). Assuming the model in Eq. 4, this observation indicates that for this population the ratio $k_2/(k_2 + k_{-2})$ is equal to ~ 1 at all CO concentrations used in this study and that $k_1 = 90 \text{ s}^{-1}$. Alternatively, after photolysis from heme a_3 and binding to Cu_B , the CO ligand did not equilibrate with solution in this population.

The slower, major CO-recombination component displayed a CO-concentration dependent rate constant of 6.7 s^{-1} at 1 mM CO. Assuming that k_1 has the same value of 90 s^{-1} for the two Cyt $c\text{O}$ populations, the ratio $k_2/(k_2 + k_{-2}) \cong 0.07$. Thus, the difference between the two Cyt $c\text{O}$ populations reflecting the two time constants could be explained by different values of $k_2/(k_2 + k_{-2})$, i.e. by differences in CO binding to Cu_B . Because CO-recombination was biphasic both in whole cells and in the detergent-purified samples, the presence of the two components is not an artifact caused by the purification of the supercomplex or the oxidase. Instead, we speculate that the two components reflect two Cyt $c\text{O}$ populations that are present in the native membrane and that could differ, for example, in the local structure of Cu_B resulting in different CO-binding affinities. Most likely these two Cyt $c\text{O}$ populations would display different reactivity towards the natural ligand and electron acceptor, O_2 . If the relative fraction of these two populations would be modulated by the cell, this mechanism could be used to regulate electron transfer through the respiratory chain.

Reaction with O_2 . The four-electron reduction of O_2 to H_2O takes place in a number of distinct kinetic steps in which the Cyt $c\text{O}$ is gradually oxidized and O_2 is reduced. We identified the kinetic components on the basis of a comparison to data obtained earlier with other well-studied oxidases³⁸. With the pure Cyt $c\text{O}$ we observed electron transfer from heme a to the catalytic site and formation of the “peroxy” state, P_R , with a time constant of $\sim 21 \mu\text{s}$. Formation of the next, ferryl intermediate (F) displayed a time constant of $90 \mu\text{s}$. Finally, the Cyt $c\text{O}$ was oxidized forming the oxidized state (O) with a time constant of 1.3 ms. All these time constants are essentially the same as those observed previously with e.g. the well-studied Cyt $c\text{O}$ from bovine heart or *R. sphaeroides*. Consequently, the differences in CO-binding kinetics between the earlier studied A-type oxidases and *C. glutamicum* Cyt $c\text{O}$ are apparently not reflected in the kinetics of O_2 binding and reduction.

For the cyt. bc_1 -Cyt $c\text{O}$ complex, all time constants for the different reaction steps were similar to those observed with the pure Cyt $c\text{O}$, however, there were also notable differences reflecting intra-complex electron transfer. During the $\text{P}_R \rightarrow \text{F}$ reaction ($\tau \cong 120 \mu\text{s}$) an electron is transferred from Cu_A to heme a , which leaves Cu_A oxidized allowing electron transfer from cyt. c of the cyt. bc_1 complex to Cu_A . This electron transfer is rate-limited by proton uptake⁵¹ and does not occur at the same rate as when photochemically injected into Cu_A ⁵². As seen at 550 nm (Fig. 5E), cyt. c was partially oxidized over a time scale of $\sim 100 \mu\text{s}$, which indicates that Cu_A was re-reduced concomitantly with the Cu_A -to-heme a electron transfer. This interpretation is also supported by the very small extent of net Cu_A oxidation (observed at 830 nm) over the 100- μs time scale (Fig. 5D, see the small increase in absorbance). In the next step of the reaction, $\text{F} \rightarrow \text{O}$ ($\tau \cong 1.7 \text{ ms}$), the fourth electron is transferred to the catalytic site, which allows further electron transfer from heme c to the Cyt $c\text{O}$, reflected in a further decrease in absorbance at 550 nm over the time scale of the $\text{F} \rightarrow \text{O}$ reaction in Cyt $c\text{O}$. The absorbance changes at 550 nm reflect oxidation of cyt. c , but we could not determine the degree of oxidation of each of the two cyt. c s separately. Most likely these two cyt. c s are oxidized simultaneously, but not necessarily to the same degree. In the summarizing Fig. 8 we indicate that both cyt. c s are oxidized over time scales of 100 μs and 1.7 ms (approximated by 2 ms in the figure).

We also observed further electron transfer from heme b (absorbance decrease at 563 nm, Fig. 5F), which reflects electron transfer from heme b to the Cyt $c\text{O}$, but this electron transfer significantly lags behind ($\tau \cong 6.5 \text{ ms}$) that of the $\text{F} \rightarrow \text{O}$ reaction ($\tau \cong 1.7 \text{ ms}$). As described above for cyt. c , also for heme b oxidation we could not discriminate between hemes b_L or b_H and conclude only that heme b of the cyt. bc_1 is oxidized over the 6.5-ms time scale. As outlined in the Results section, the amounts of oxidized heme c and heme b approximately equal the amount of Cyt $c\text{O}$ that becomes re-reduced during or after reaction with O_2 . Furthermore, the electron-transfer time constant from heme b to Cyt $c\text{O}$ ($\tau \cong 6.5 \text{ ms}$) is approximately compatible with the overall quinol-oxidation/ O_2 reduction turnover rate of the cyt. bc_1 -Cyt $c\text{O}$ supercomplex ($\sim 210 \text{ s}^{-1}$). This rapid electron transfer with a time constant of $\sim 6.5 \text{ ms}$ corresponds to a maximum electron-transfer rate over a distance of $\sim 25 \text{ \AA}$ ⁵³. Even though the distances connecting the heme c s with their partners are not known for the *C. glutamicum* cyt. bc_1 -Cyt $c\text{O}$ complex, this distance estimation is consistent with that between heme b_L and the iron-sulfur cluster in cyt. bc_1 from e.g. *S. cerevisiae*²⁵.

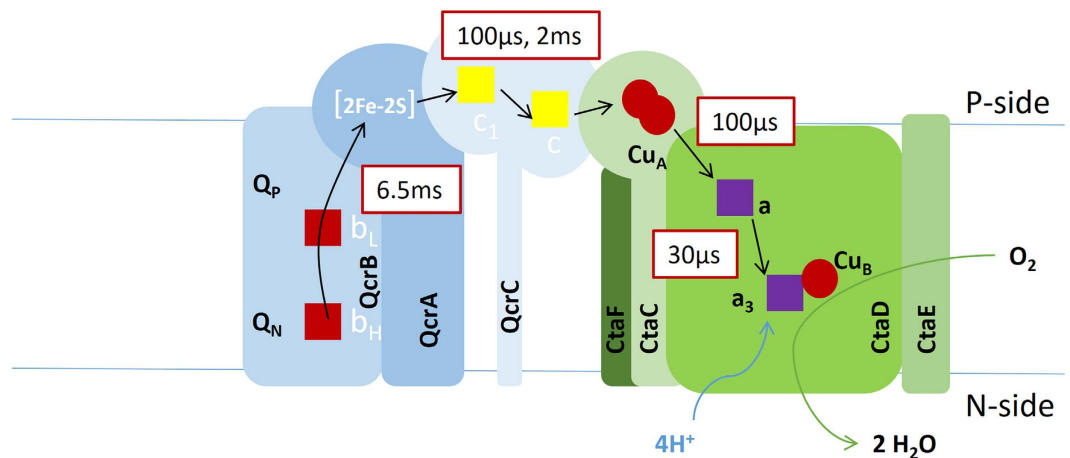


Figure 8. A schematic picture showing the approximate time constants for the electron-transfer reactions. Note that the positions of the heme groups in this picture are not compatible with the X-ray structure (e.g.³⁹); they have been adjusted for clarity. Because the data cannot discriminate between the two heme cs or the two heme bs, we have indicated the oxidation time constants for each type of heme without specifying which heme that is oxidized. Redox reactions of the quinones are not observed in these experiments.

To investigate the functional stability of the cyt. *bc*₁-CytcO supercomplex we investigated the reaction with O₂ upon increasing the ionic strength (Figure S6). Neither the amplitudes nor rates of the observed absorbance changes at 445 nm were significantly altered even at the highest KCl concentration of 1.5 M. These results suggest that the purified supercomplex is stable and remains functionally intact. Furthermore, the interactions between one of the *c* hemes of the cyt. *bc*₁ complex and CytcO seem more stable than those observed for the water-soluble cyt. *c* and CytcO from e.g. bovine heart as in the latter case the 1:1 cyt. *c*-CytcO complex dissociates at ionic strength above ~300 mM⁴⁵.

pH (in)dependence of the Reaction with O₂. As seen in Fig. 7 we did not observe any significant pH-dependence in the kinetics of the F → O reaction, neither for the pure CytcO nor for the cyt. *bc*₁-CytcO supercomplex. Slight differences were observed for the two samples (see inset to Fig. 7), which may be attributed to binding of cyt. *bc*₁ to CytcO in the supercomplex. The observation that the F → O rate is essentially pH-independent is surprising given that the reaction is associated with proton uptake (see Fig. 6) and therefore expected to display pH-dependent kinetics, as in other oxidases^{30,46,47}. One possible explanation for pH-independent kinetics of a reaction step that is linked to proton uptake is that the pK_a in this pH dependence may be outside of the accessible pH range. Alternatively, the proton uptake may not be part of the rate-limiting step, however, this explanation is less likely based on results from earlier studies with other A- and B-type oxidases where proton uptake is rate limiting^{51,54–57}.

In earlier studies with the *R. sphaeroides* CytcO it has been observed that the P_R → F rate is essentially pH-independent up to pH ~9 and then decreases with increasing pH with an apparent pK_a of 9.4⁵⁸. The F → O rate in *R. sphaeroides* CytcO displayed a more complex pH dependence and was found to titrate with two pK_as of ~9 and <6, respectively³⁰. The pK_a around 9 was attributed to residue Glu286 within the D proton pathway, which is conserved in the *C. glutamicum* CytcO (Glu267). In the *R. sphaeroides* CytcO, replacement of e.g. Asn139 or Asn207 by Asp resulted in an increase in the Glu286 pK_a; for example in the Asn139Asp variant the pK_a increased to a value above the accessible pH range^{46,59,60}. In other words, even though the reaction is associated with proton uptake in these structural variants, it did not display a pH-dependent kinetics. In the *C. glutamicum* CytcO, many but not all residues “below” the Glu267 in the D pathway are conserved. The data with the *R. sphaeroides* structural variants show that very small changes in the D pathway structure, also at a distance from the Glu, may yield pH-independent kinetics. Thus, considering the differences in the environment of Glu267 in the *C. glutamicum* CytcO it is possible that its pK_a is tuned to adopt a value that is higher than that of Glu286 in the *R. sphaeroides* CytcO. However, more experiments with single point mutations in the D pathway are necessary to confirm this speculation.

Concluding Remarks

The reaction of the purified reduced CytcO with O₂ displayed the same sequence of electron transfers as that observed previously with other bacterial and mitochondrial A-type CytcOs. However, in contrast to data obtained with the other oxidases, none of the reaction steps associated with proton uptake displayed any pH dependent rates, which is explained in terms of an elevated apparent pK_a of Glu267. The data indicate that one of the *c* hemes of cyt. *bc*₁ is bound near Cu_A, at the same site where externally added water-soluble cyt. *c* would bind. Furthermore, the data indicate that the interaction of this heme *c* with CytcO is strong in the cyt. *bc*₁-CytcO supercomplex and insensitive to changes in ionic strength. The second heme *c* of the cyt. *bc*₁-CytcO supercomplex provides a link for direct electron transfer between cyt. *bc*₁ and Cu_A, the electron acceptor of CytcO (see Fig. 8), which was also indicated from earlier mutagenesis data²⁰. Electron transfer from heme *c* to the CytcO occurred over time scales of approximately 100 μs–2 ms, while re-reduction of heme *c* by heme *b* displayed a time constant

of ~6.5 ms, which suggest that this reaction may be rate limiting for the overall quinol-oxidation O₂-reduction turnover rate. In conclusion, the isolation of a stable supercomplex from *C. glutamicum* allowed us to investigate the kinetics of electron transfer all the way from heme *b* in the cyt. *bc*₁ complex to the catalytic site of Cyt_cO, via the bridging *c* hemes.

Materials and Methods

If not stated otherwise, the chemicals were purchased from Sigma-Aldrich.

***Corynebacterium glutamicum* strains.** The *C. glutamicum* strains used for purification of Cyt_cO and the cyt. *bc*₁-Cyt_cO supercomplex were described before²⁰. The ΔC-D_{St} strain refers to the 13032Δ*ctaD* strain transformed with the pJC1-*ctaD*_{St} plasmid (Kan^R), which serves as an expression plasmid for Strep-tagged CtaD; *ctaD* is expressed from its native promoter and contains 10 additional codons at the 3'-end (AAWSHPQFEK). The ΔQ-D_{St} strain refers to the 13032Δ*qcrCAB* strain transformed with the pJC1-*ctaD*_{St} plasmid.

Culture Conditions. The cells were cultivated at 30 °C in all steps. Single *C. glutamicum* colonies were picked from BHI-Agar plates (33 g/l brain heart infusion broth, 15 g/l agar agar, 20 g/l D-(+)-glucose, 25 mg/l kanamycin) and inoculated into 10 ml BHI culture medium (33 g/l brain heart infusion broth, 20 g/l D-(+)-glucose, 25 mg/l kanamycin) at 220 rpm. After overnight growth the pre-culture was inoculated into 500 ml CGXII medium²⁰ in a 2 l Erlenmeyer flask (at 160 rpm). After the optical density at 600 nm (OD₆₀₀) reached between 25 and 30, the cells were diluted 1:20 into 2 l of CGXII medium into a 5 l baffled Erlenmeyer flask (at 130 rpm) and cultivated to an OD₆₀₀ of 15–17 before harvest.

Membrane Preparation. The cells were harvested using a Beckman centrifuge equipped with the JLA 8.1000 rotor at 7,500 rpm (~10,000 × g) for 30 min. The cells were homogenized in 4 ml cell lysis buffer (100 mM Tris-HCl pH 7.5, 5 mM MgSO₄, some crystals phenylmethanesulfonyl fluoride, some crystals DNaseI (Roche)) per 1 g of cells (wet weight), and passed through a cell disrupter 4 times at 40 kPsi (Constant Systems). Cell debris was collected by centrifugation at 27,000 × g for 20 min. (Type 45Ti rotor, Beckman) and subsequently the membranes were collected by ultracentrifugation at 150,000 × g for 90 min (Type 45Ti rotor, Beckman).

Protein Purification. The purification was done essentially as described in ref. 20 with minor modifications. Briefly, the membranes were mixed with solubilization buffer (100 mM Tris-HCl pH 7.5, 100 mM NaCl, 2 mM MgSO₄, 50 mg/l avidin (iba lifescience), 1% (w/v) DDM (GLYCON Biochemicals) at a protein concentration of 5 mg/ml, and incubated at 4 °C for 45 min under slow stirring. Unsolubilized material was removed by ultracentrifugation (180,000 × g, 20 min, 4 °C). The supernatant was collected and concentrated in an Amicon Ultra 15 ml filter spin tube with 100 kDa cutoff until the volume was around 5 ml. The concentrated supernatant was diluted tenfold in solubilization buffer without DDM (yielding a final DDM concentration of 0.1% (w/v) and concentrated again to reach a volume <15 ml. The concentrated supernatant was applied to a Gravity flow Strep-Tactin Superflow column (bed volume 5 ml, iba lifescience). Subsequently, the column was washed 3 times with 0.5 column volumes of washing buffer (100 mM Tris-HCl pH 7.5, 100 mM NaCl, 2 mM MgSO₄, 0.015% (w/v) DDM) and protein was eluted with up to 3 column volumes elution buffer (100 mM Tris-HCl pH 7.5, 100 mM NaCl, 2 mM MgSO₄, 0.015% (w/v) DDM, 2.5 mM D-desthiobiotin) and concentrated as described above. The protein samples (Cyt_cO alone green colored, *bc*₁-Cyt_cO brown colored) were stored at 4 °C.

The Cyt_cO alone, as well as the cyt. *bc*₁-Cyt_cO supercomplex, could be purified from membranes originating from the ΔQ-D_{St} and the ΔC-D_{St} *C. glutamicum* strains, respectively, both having a Strep-tag on subunit I of the Cyt_cO. The SDS-PAGE for the two purifications (supplementary Figure S1) shows that the eluate of the supercomplex purification contains the three core subunits of Cyt_cO (CtaD, C, and E), along with the three subunits of the *bc*₁-complex (QcrC, A, and B), as well as some additional subunits, which were also co-purified with the supercomplex²⁰. Accordingly, the eluate for the oxidase purification from ΔQ-D_{St} membranes contained only the three core subunits of the Cyt_cO.

Dithionite-reduced minus ferricyanide-oxidized difference spectra were recorded at room temperature using a Cary100 UV-Vis Spectrophotometer. The Cyt_cO concentration was determined from the reduced minus oxidized difference spectrum using the absorption coefficient $\Delta\epsilon^{600-630} = 3.2 \text{ mM}^{-1}\text{cm}^{-120}$.

Quinone Reduction. Instead of ubiquinone, *C. glutamicum*, as a Gram-positive bacterium, employs menaquinones as an electron carrier in the membrane⁶¹. To prepare reduced quinol, 3.7 mg 2,3-dimethyl-[1,4]naphthoquinone (Rare Chemicals GmbH) was dissolved in 1 ml N₂-saturated anhydrous cyclohexane to yield a 20 mM solution. The solution was mixed with 5 ml N₂-saturated 1 M sodium dithionite solution (in H₂O) and shaken vigorously. After phase separation, the organic phase containing the reduced 2,3-dimethyl-[1,4]naphthoquinol was removed and transferred to a 15 ml Falcon tube (all steps performed under a stream of N₂). The cyclohexane was evaporated under an N₂ stream, while the sample was kept at ~40 °C in a water bath. Subsequently, the reduced quinol was dissolved in N₂-saturated, acidified ethanol (ethanol with 10 mM HCl), aliquoted, and flash frozen in liquid nitrogen, and stored at -20 °C.

Determination of enzyme activities. Oxygen consumption in multiple turnover experiments was measured using a Clark-type oxygraph (Hansatech). The TMPD (N,N,N',N'-tetramethyl-p-phenylenediamine)-oxidase activity was measured in 100 mM Tris-HCl (pH 7.5), 100 mM NaCl, 0.015% (w/v) DDM, 0.2 mM TMPD, 2 mM sodium ascorbate. Both TMPD and ascorbate were added before the sample and background oxygen consumption was measured and the reaction was started by adding the protein sample. Cytochrome *c* driven oxidase activity was measured as above in the presence of ascorbate and TMPD, but bovine heart cyt. *c* (25 μM final concentration) was also supplied before addition of the sample. Quinol-driven oxidase activity was measured in

the same buffer (100 mM Tris-HCl (pH 7.5), 100 mM NaCl, 0.015% (w/v) DDM), but 20 μ l of the reduced quinol solution (see above) was added before the protein sample. Background oxygen consumption was measured, followed by addition of the protein sample and monitoring the enzymatic oxygen consumption. When measuring the Cyt_cO activity with ascorbate/TMPD/cyt. *c* as a substrate, this activity was insignificant. However, in the presence of quinol the background O₂-reduction rate increased significantly, due to quinol auto-oxidation, and at most it reached 70% of the rate measured after addition of the supercomplex.

Flash Photolysis and Flow-flash Experiments. The purified samples (typical protein concentrations were in the range of 2–3 μ M and 1–1.5 μ M for *bc*₁-Cyt_cO and Cyt_cO, respectively) were transferred into a Thunberg cuvette and the atmosphere was exchanged for N₂ on the vacuum line. The sample was reduced by the addition of 1 μ M phenazine methosulfate and 5 mM sodium ascorbate from the sidearm of the cuvette. In order to achieve complete reduction of all the redox-active centers in samples containing supercomplexes, 0.5–1 mM sodium dithionite (Merck Millipore) was added and the reduction state was confirmed spectrophotometrically. After complete reduction, the atmosphere was exchanged for CO on a vacuum line. To measure the pH dependence of the reaction of the reduced Cyt_cO or cyt. *bc*₁-Cyt_cO with O₂, the sample was prepared in 10 mM Tris-HCl, the atmosphere was replaced consecutively by N₂ and then CO, and the sample was reduced by 4 mM ascorbate, 1 μ M PMS and 100 μ M dithionite at pH 7.5. Samples at different, higher pH values were prepared by adding various volumes of 1 M Tris-HCl buffer at pH 10 followed by incubation for at least 1 hour. Alternatively, the sample in 10 mM Tris-HCl at pH 7.5, was reduced with 4 mM ascorbate, 1 μ M PMS and 100 μ M dithionite (under CO atmosphere) and then mixed with an O₂-saturated solution at different pH values containing 100 mM buffer. The buffering capacity of the O₂ solution was much higher (100 mM) than that of the enzyme solution (10 mM) so the final pH after mixing was determined by the former. The pH after mixing was measured using a pH meter.

The CO rebinding/recombination kinetics to the catalytic site was measured as a change in absorbance over time at several wavelengths after the photolysis by a ~10-ns laser flash ($\lambda = 532$ nm, Nd-YAG laser, Quantel; the flash-photolysis/flow-flash setup was purchased from Applied Photophysics UK). The time resolution of the set-up was ~10⁻⁷ s. The absorbance changes were then fitted to an exponential decay function using the ProK software from Applied Photophysics U.K.

In order to determine the CO concentration dependence of the CO-recombination, 1 ml of a protein sample was transferred into a Thunberg cuvette and treated as described above, except that after reduction it was left under a nitrogen atmosphere. The sample was covered with a layer of paraffin oil and CO was added in small aliquots of CO-saturated buffer (100 mM Tris-HCl (pH 7.5), 100 mM NaCl, 0.015% (w/v) DDM) with a gas-tight Hamilton syringe through the paraffin layer. The CO-recombination kinetics was studied after each CO addition using a flash-photolysis set-up (Applied Photophysics, UK).

In flow-flash experiments, the reduced and CO-blocked protein sample was mixed 1:3 with oxygen-saturated buffer (~1.2 mM O₂) in a flow-flash setup (Applied Photophysics, UK.). About 200 ms after mixing (mixing time < 10 ms) with the oxygenated buffer, CO was dissociated from the catalytic site by a short laser pulse (~10-ns laser flash ($\lambda = 532$ nm, Nd YAG-laser, Quantel). Changes in absorbance were recorded over time at different wavelengths. The data were fitted to a kinetic model using the ProK software from Applied Photophysics, UK.

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Author Contributions

P.B., C.v.B. and P.Å. planned the research; S.G. and O.F. performed the experiments; M.B. constructed the strains; P.B., S.G., C.v.B. and P.Å. wrote the manuscript; W.-C.K. and C.H. assisted in development of methods for enzyme purification.

Additional Information

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