

Supporting Information

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LncRNA *LINK-A* Remodels Tissue Inflammatory Microenvironments to Promote Obesity

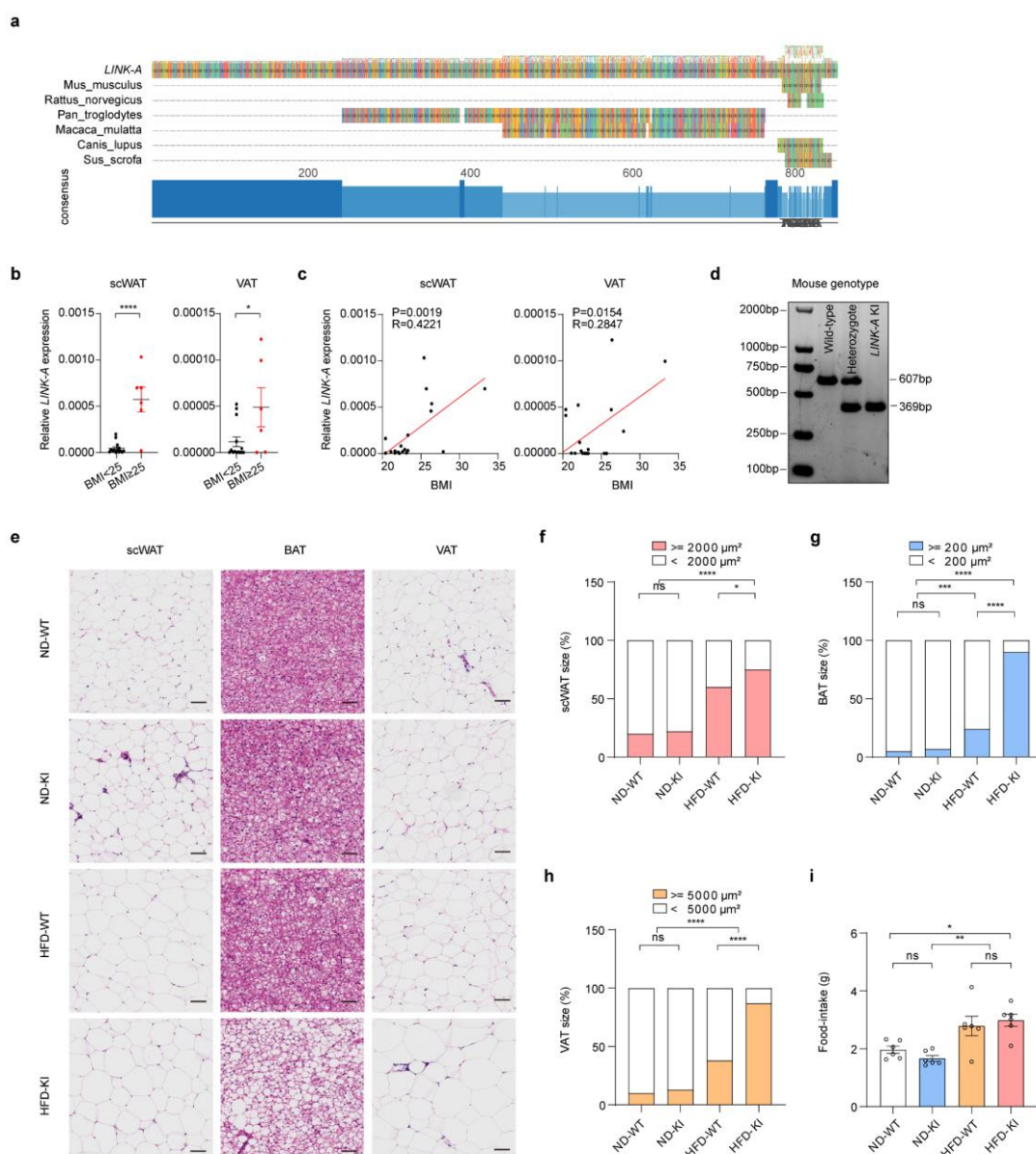
Yu Chen, Hui Chen, Ying Wang, Fangzhou Liu, Xiao Fan, Chengyu Shi, Xinwan Su, Manman Tan, Yebin Yang, Bangxing Lin, Kai Lei, Lei Qu, Jiecheng Yang, Zhipeng Zhu, Zengzhuang Yuan, Shanshan Xie, Qinming Sun, Dante Neculai, Wei Liu, Qingfeng Yan, Xiang Wang, Jianzhong Shao, Jian Liu* and Aifu Lin*

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Figures S1 to S5



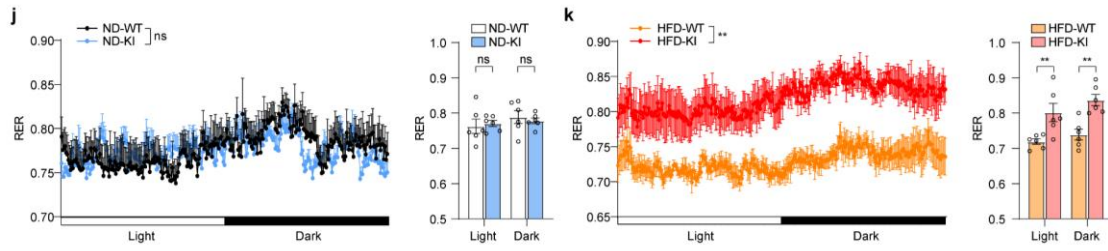


Figure S1. *LINK-A* overexpression in mice promotes HFD-induced obesity **a)** The stacked graph for *LINK-A* conserved regions of sequences in various species. To be specific, the sequence of MSA is depicted as an uninterrupted, semi-opaque line. Each line represents a sequence. A: red, C: blue, T: green, G: yellow. The Sequence logo visualization method calculates the frequency of characters in each column in the MSA, and scales and stacks the characters in each column according to the frequency. **b)** *LINK-A* expression was assessed by qRT-PCR in scWAT(left) and VAT(right) tissues of patients in the BMI < 25 and BMI $\geq$ 25 groups. Data presented as mean $\pm$ SEM, n=14, 6, Student's t-test, the data conform to normal or lognormal distribution, $*p < 0.05$, $****p < 0.0001$. **c)** Correlation analysis of *LINK-A* expression in the in scWAT(left) and VAT(right) tissues of clinical patients with patient BMI. Data presented as mean $\pm$ SEM, n=20, Pearson chi-square test, $p=0.0019$ (scWAT), $p=0.0154$ (VAT). **d)** Identification of mouse genotypes by pulsed electric field gel electrophoresis. **e–h)** Representative images of H&E-stained sections of different adipose tissues(e). The distribution of adipocyte size of scWAT(f), BAT(g), and VAT(h) of ND-WT, ND-KI, HFD-WT, and HFD-KI mice were analyzed using ImageJ. Scale bar: 50 μ m. Data presented as mean $\pm$ SEM, per group n=6, chi-square test, ns=no significance, $*p < 0.05$, $***p < 0.001$, $****p < 0.0001$. **i)** Food intake of WT or *LINK-A* KI mice fed ND or HFD. Data presented as mean $\pm$ SEM, per group n=6, one-way ANOVA, ns=no significance, $*p < 0.05$, $**p < 0.01$. **j–k)** Line graphs(left) and bar charts(right) of RER in WT and *LINK-A* KI mice fed ND(j) or HFD(k). Data presented as mean $\pm$ SEM, per group n=6, two-way ANOVA, ns=no significance, $**p < 0.01$.

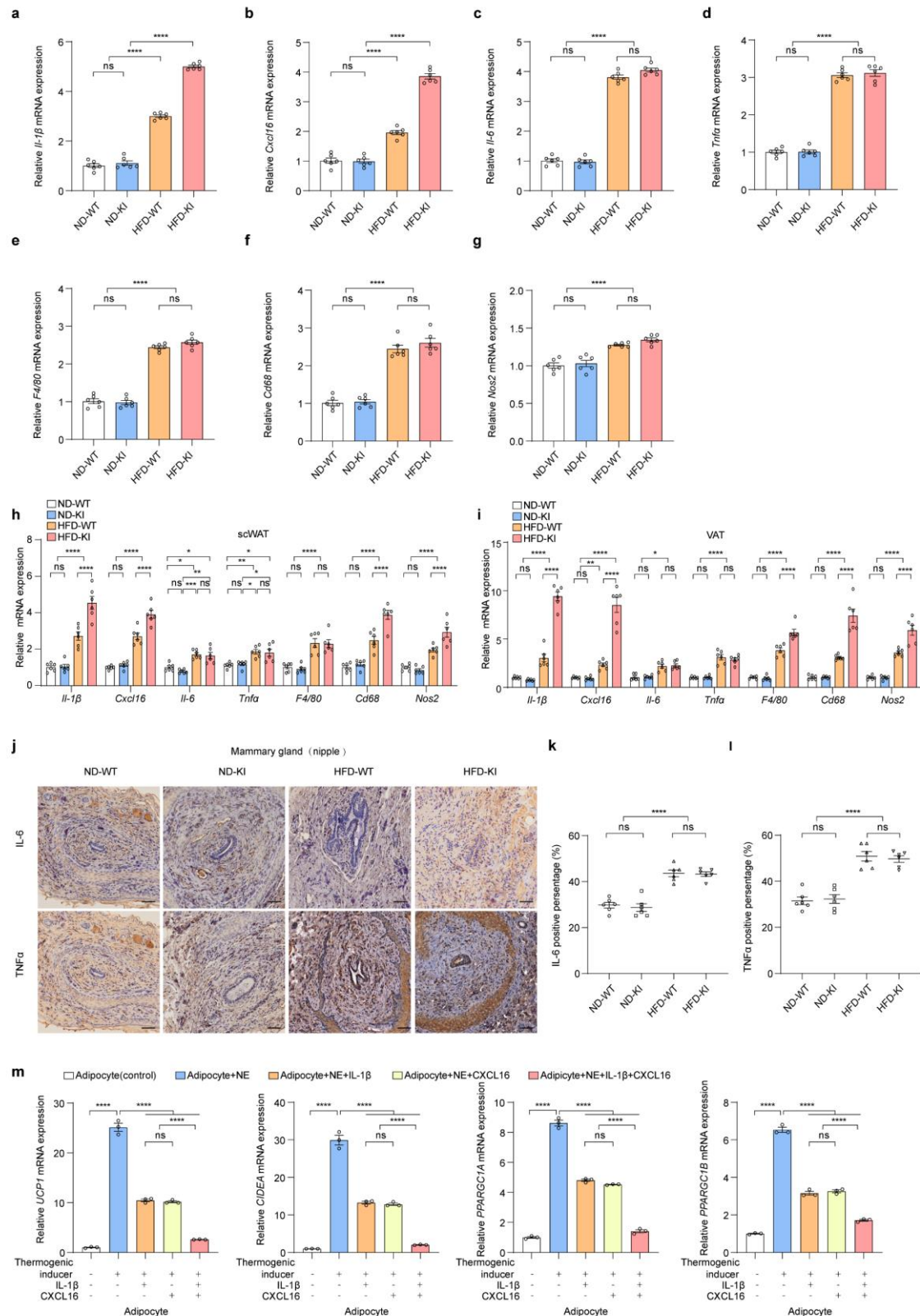


Figure S2. *LINK-A* overexpression reduces adaptive thermogenesis in HFD-fed mice by remodeling the regional inflammatory microenvironment a–g) Inflammatory factor mRNA expression levels in the mammary glands of ND-WT, ND-KI, HFD-WT, and HFD-KI mice were detected by qRT-PCR. Data presented as mean ± SEM, per group n=6, one-way ANOVA,

ns=no significance, **** $p<0.0001$. **h–i**) Inflammatory factor mRNA expression levels in the scWAT(h) and VAT(i) of ND-WT, ND-KI, HFD-WT, and HFD-KI mice were detected by qRT-PCR. Data presented as mean $\pm$ SEM, per group $n=6$, two-way ANOVA, ns=no significance, * $p<0.05$, ** $p<0.01$, *** $p<0.001$, **** $p<0.0001$. **j–l**) Representative images of IHC of mammary glands (nipple)(j). The IL-6(k) or TNF α (l) levels were analyzed using ImageJ. Scale bar: 50 μ m. Data presented as mean $\pm$ SEM, per group $n=6$, one-way ANOVA, ns=no significance, **** $p<0.0001$. **m**) The thermogenic genes mRNA levels in adipocytes treatment using IL- β or/and CXCL16 cytokines by qRT-PCR. Data presented as mean $\pm$ SEM, pooled data from three independent experiments, one-way ANOVA, ns=no significance, **** $p<0.0001$.

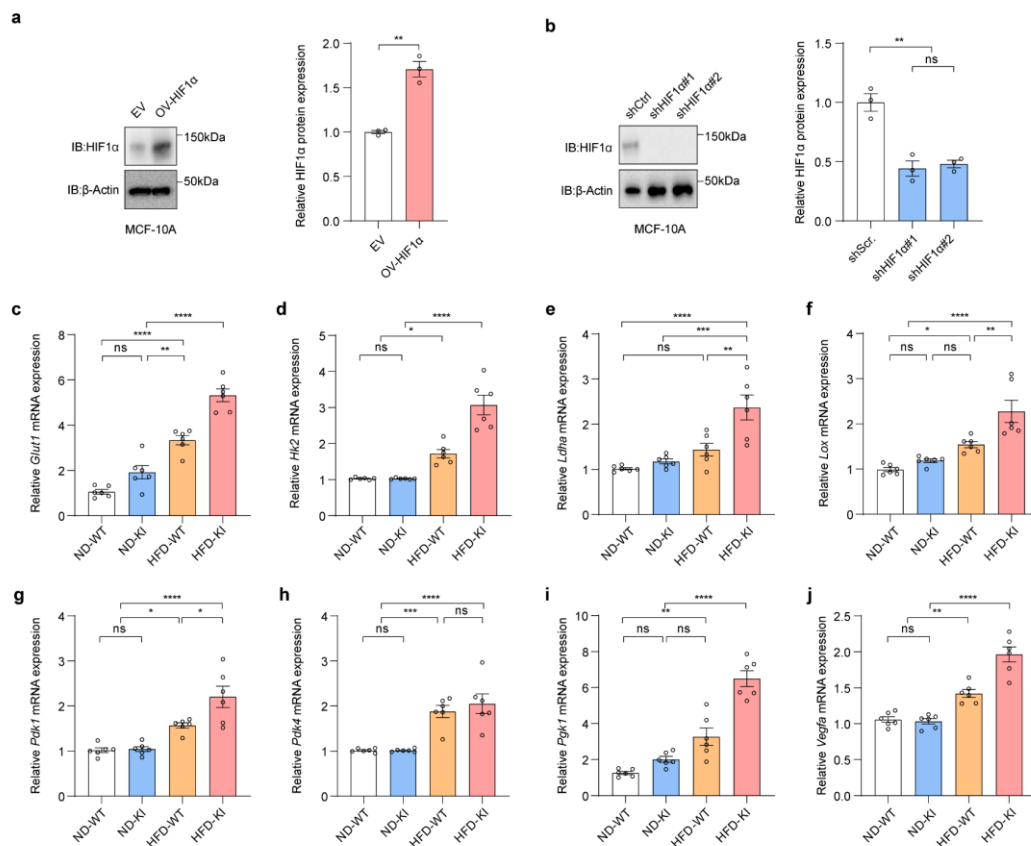


Figure S3. *LINK-A* overexpression induces inflammatory factor expression by stabilizing HIF1 α through the HFD-induced HB-EGF **a)** The HIF1 α protein levels in MCF-10A EV or HIF1 α overexpression cells were detected by immunoblot analysis, and the HIF1 α protein levels were quantified using ImageJ. Data presented as mean $\pm$ SEM, pooled data from three independent experiments, unpaired t-test, ** $p<0.01$. **b)** The HIF1 α protein levels in MCF-10A shControl (shCtrl) or shHIF1 α (shHIF1 α #1, shHIF1 α #2) cells by immunoblot analysis, and the

HIF1 α protein levels were quantified using ImageJ. Data presented as mean $\pm$ SEM, pooled data from three independent experiments, one-way ANOVA, ns=no significance, $**p<0.01$. **c–j)** The HIF1 α downstream target genes mRNA expression in the mammary glands of ND-WT, ND-KI, HFD-WT, and HFD-KI mice were detected by qRT-PCR. Data presented as mean $\pm$ SEM, per group $n=6$, one-way ANOVA, ns=no significance, $*p<0.05$, $**p<0.01$, $****p<0.0001$.

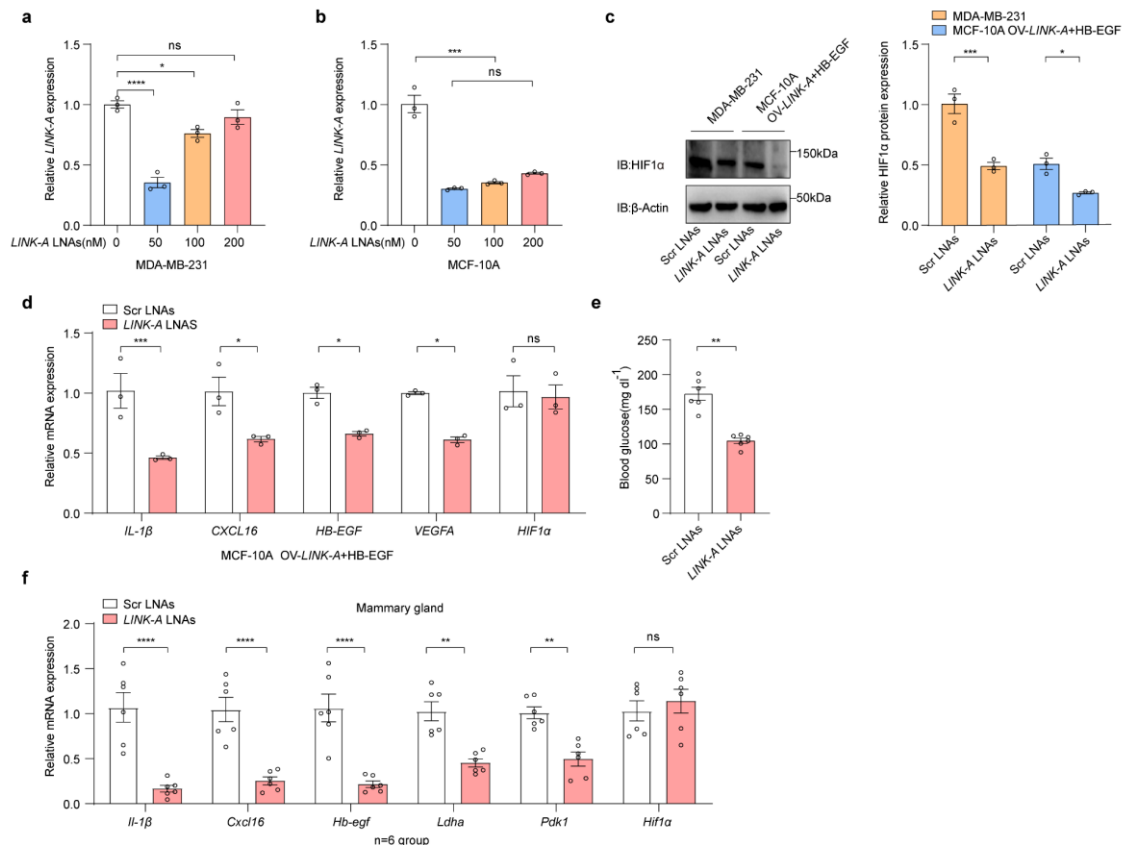


Figure S4. ASO drug inhibiting *LINK-A* attenuates obesity and metabolic disorders in mice **a-b)** The *LINK-A* levels in the MDA-MB-231(a) and MCF-10A(b) treated with different concentrations of *LINK-A* LNAs were detected by qRT-PCR. Data presented as mean $\pm$ SEM, pooled data from three independent experiments, one-way ANOVA, ns=no significance, $*p<0.05$, $***p<0.001$, $****p<0.0001$. **c)** The HIF1 α protein levels in the MDA-MB-231 and MCF-10A with or without *LINK-A* LNAs treatment were detected by immunoblot analysis, and the HIF1 α protein levels were quantified using ImageJ. Data presented as mean $\pm$ SEM, pooled data from three independent experiments, two-way ANOVA, $*p<0.05$, $***p<0.001$. **d)** The mRNA expressions of *HIF1 α* or HIF1 α downstream genes were detected by qRT-PCR. Data presented as mean $\pm$ SEM, pooled data from three independent experiments, two-way ANOVA,

ns=no significance, * $p < 0.05$, *** $p < 0.001$. **e)** The fasting glucose in HFD-KI mice with *LINK-A* LNAs or Scr LNAs treatment. Data presented as mean $\pm$ SEM, per group $n=6$, Mann-Whitney U-test, ** $p < 0.01$. **f)** The mRNA levels in the mammary gland of HFD-KI mice with *LINK-A* LNAs or Scr LNAs treatment for two weeks were detected by qRT-PCR. Data presented as mean $\pm$ SEM, per group $n=6$, two-way ANOVA, ns=no significance, ** $p < 0.01$, **** $p < 0.001$.

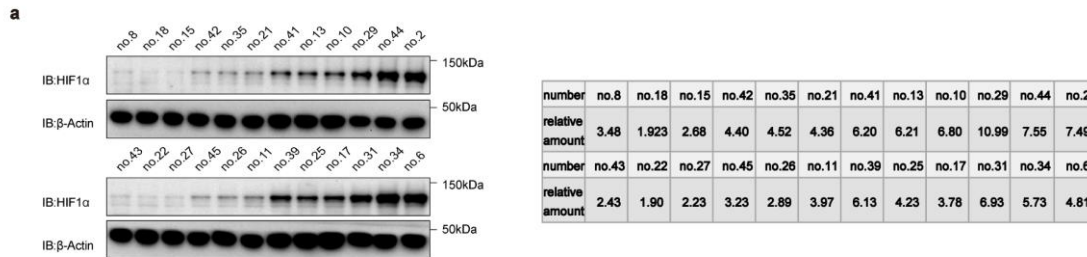


Figure S5. The expression levels of HIF1 α protein in clinical breast samples. a) The HIF1 α protein level in the breast tissue of clinical patients was detected by immunoblot analysis, and the HIF1 α protein was quantified using ImageJ. $n=24$.

Table S1 to S4

Table S1. Clinicopathological Parameters of Tissue Microarrays Used in the Study

Table S1.Clinicopathological Parameters of Tissue Microarrays Used in the Study									
Breast Tissue Microarrays Used in the Study									
number	Age	Sex	Height (m)	Weight (kg)	BMI	<i>LINK-A</i> expression	<i>IL-1β</i> mRNA expression	<i>CXCL16</i> mRNA expression	HIF1 α Protein level
1	50-60	F	1.57	55	22.31328	0.001022	0.0037399	0.0109827	N/A
2	40-50	F	1.61	66	25.46198	0.000913	0.0031317	0.0537407	7.49
3	20-30	F	1.64	61	22.67995	0.000284	0.0060806	0.0297091	N/A
4	40-50	F	1.58	49	19.62826	0.000168	0.0012892	0.0073193	N/A
5	40-50	F	1.59	50	19.7777	0.002592	0.0014706	0.0155814	N/A
6	50-60	F	1.6	55	21.48438	0.000436	0.0009997	0.0312578	4.816427
7	50-60	F	1.54	57	24.03441	0.004735	0.000963	0.220718	N/A
8	40-50	F	1.52	60	25.96953	0.000448	0.0024108	0.0109425	3.48
9	40-50	F	1.63	59	22.20633	0.000576	0.0169038	0.0176296	N/A
10	60-70	F	1.54	64	26.986	0.001717	0.0025334	0.0155565	6.8
11	40-50	F	1.52	47	20.3428	0.000457	0.0074517	0.0826515	3.97707
12	60-70	F	1.55	52	21.64412	0.002324	0.0007774	0.0181257	N/A
13	40-50	F	1.58	70	28.04038	0.005255	0.0100917	0.10785	6.213457
14	50-60	F	1.58	50	20.02884	0.00096	0.0004588	0.0074244	N/A
15	40-50	F	1.61	55	21.21832	0.000362	0.0022103	0.0862659	2.684295
16	50-60	F	1.6	55	21.48438	0.001749	0.0091718	0.1676537	N/A
17	60-70	F	1.58	52.5	21.03028	0.001136	0.0002348	0.0267928	3.783127
18	40-50	F	1.57	50	20.2848	0.001121	0.008899	0.0073349	1.923465
19	40-50	F	1.58	65	26.03749	0.000845	0.0051617	0.038061	N/A

20	30-40	F	1.55	72	29.96878	0.004556	0.0110747	0.0606632	N/A
21	60-70	F	1.58	63.5	25.43663	0.003113	0.0044138	0.07480802	4.36
22	30-40	F	1.56	49	20.13478	0.00096	0.0021717	0.0391797	1.901839
23	40-50	F	1.5	54	24	0.000664	0.0048995	0.0348943	N/A
24	40-50	F	1.67	63	22.58955	0.000457	0.0004594	0.0165293	N/A
25	40-50	F	1.59	48	18.98659	0.00078	0.0026087	0.0114104	4.234577
26	30-40	F	1.59	57	22.54658	0.003225	0.0038366	0.00423	2.891804
27	50-60	F	1.63	62	23.33547	0.002305	0.0030872	0.0187445	2.234565
28	50-60	F	1.58	54	21.63115	0.000348	0.0028461	0.0177258	N/A
29	50-60	F	1.53	69	29.47584	0.0062549	0.027552	0.154281	10.99793
30	50-60	F	1.55	45	18.73049	0.000549	0.0033284	0.0303122	N/A
31	50-60	F	1.63	76	28.60476	0.003585	0.0190846	0.0215266	6.937737
32	30-40	F	1.62	52	19.81405	0.001378	0.0081364	0.035292	N/A
33	40-50	F	1.59	58	22.94213	0.002143	0.0033893	0.0658015	N/A
34	40-50	F	1.65	65	23.87511	0.000614	0.0032358	0.0470298	5.739
35	30-40	F	1.56	56	23.01118	0.000501	0.0127047	0.0249603	4.523458
36	30-40	F	1.61	66	25.46198	0.000688	0.0083113	0.0731988	N/A
37	40-50	F	1.63	55	20.70082	0.000914	0.0081261	0.0139521	N/A
38	40-50	F	1.57	54	21.90758	0.002851	0.0006296	0.019462	N/A
39	50-60	F	1.53	57	24.34961	0.002625	0.0068028	0.0312146	6.136
40	50-60	F	1.5	47	20.88889	0.002739	0.00408	0.0339054	N/A
41	40-50	F	1.6	65	25.39063	0.000292	0.0051109	0.0163632	6.2
42	50-60	F	1.67	51	18.28678	0.001512	0.0061444	0.0054143	4.400865
43	50-60	F	1.59	58	22.94213	0.000794	0.0160813	0.0275414	2.437536
44	40-50	F	1.6	73	28.51563	0.00155	0.018981	0.0566803	7.558398
45	30-40	F	1.56	54	22.18935	0.001332	0.0052964	0.1131605	3.234554

Adipose Tissue Microarrays Used in the Study

number	Age	Sex	Height (m)	Weight (kg)	BMI	<i>LINK-A</i> expression (scWAT)	<i>LINK-A</i> expression (VAT)
46	60-70	F	1.58	55	22	0.00001394	0.00005223
47	40-50	F	1.56	56.7	23.3	0.0001983	0.000004327
48	60-70	F	1.56	64	26.3	0.0004575	0.00004717
49	30-40	F	1.63	59	22.2	0.000001446	0.00001199
50	40-50	F	1.64	75	27.9	0.000019	0.00002397
51	40-50	F	1.61	53	20.4	0.000004771	4.74566E-05
52	80-90	F	1.45	70	33.3	0.0006986	0.000099413
53	50-60	F	1.55	49	20.4	0.0001618	0.00004101
54	30-40	F	1.67	62.5	22.4	0.00008105	0.000002567
55	40-50	F	1.58	66	26.4	0.0005373	0.0001225
56	20-30	F	1.63	62	23.3	0.0000301961	0.0000000968
57	30-40	F	1.63	60	22.6	0.0000195398	0.0000001573
58	30-40	F	1.60	58.5	22.9	0.0000435141	0.0000001278
59	50-60	F	1.57	52	21.1	0.0000168279	0.0000004449
60	50-60	F	1.60	65	25.4	0.0010318467	0.0000000226
61	40-50	F	1.53	54	23.1	0.0000074659	0.0000001112
62	30-40	F	1.64	58	21.6	0.0000051467	0.0000003146
63	50-60	F	1.58	54	21.6	0.0000251157	0.0000003146
64	60-70	F	1.57	49	19.9	0.0000144816	0.0000010221
65	40-50	F	1.59	65	25.7	0.0006975615	0.0000000183

Table S2. Reagent or Resource

Table S2. Reagent or Resource		
Reagent or Resource	Source	Identifier
Antibodies		
anti-vinculin	Cell Signaling Technology	Cat#13901
anti-HIF1 α	Cell Signaling Technology	Cat#14179
anti- β -actin	Cell Signaling Technology	Cat#4970
anti-IL-1 β	ABclonal	Cat#A11369
anti-CXCL16	HUABIO	Cat#ER1906-85
anti-UCP1	HUABIO	Cat#HA500188
anti-Phospho-PKA R2 (S99)	HUABIO	Cat#ET1610-29
anti-PKA	HUABIO	Cat#ET1704-15
anti-Phospho-HSL(S853)	HUABIO	Cat#ET1611-19
anti-HSL	Santa Cruz Biotechnology	Cat#sc-74489
Reagents		
Liposomal Transfection Reagent	YEASEN	Cat#40802ES01
DMEM	GibcoBRL	Cat#11965
FBS	GibcoBRL	Cat#16170
Pen/Strep	GibcoBRL	Cat#10378
insulin	Sigma-Aldrich	Cat#I5500
IBMX	Sigma-Aldrich	Cat#I5879
T3	Sigma-Aldrich	Cat#T2877
Rndomethacin	Sigma-Aldrich	Cat#I7378
Rosiglitazone	Sigma-Aldrich	Cat#R2408
NE	Supelco	Cat#N-069
IL-1 β inhibitor	MCE	Cat#VX-765
CXCL16 inhibitor	Selleck	Cat#G1254032
Protease inhibitor	Roche	Cat#4693116001
TRIzol	Invitrogen	Cat#15596026CN
HiScript II qRT SuperMix	Vazyme	Cat#R222-01
SYBR qPCR Master Mix	Vazyme	Cat#Q711-012
Protein A/G PLUS-Agarose	Santa Cruz Biotechnology	Cat#sc-2003
Critical Commercial Assays		
Human IL-1 β ELISA kit	absin	Cat#abs159986
Human CXCL16 ELISA kit	R&D Systems	Cat#DCX160
Mouse IL-1 β ELISA kit	absin	Cat#abs520001
Mouse CXCL16 ELISA kit	R&D Systems	Cat#DY503
Dual-Luciferase Reporter Gene Assay Kit	YEASEN	Cat#11402ES60
BCA	Beyotime	Cat#P0009
SuperKine™ West Femto Maximum Sensitivity Substrate	ABBKINE	Cat#BMU102-CN
Histostain-Plus IHC Kit, Rabbit Primary	NeoBioscience	Cat#ENS004
Biological Samples		
Fresh frozen breast tissues	Huzhou First People's Hospital	2021kyII055
Fresh frozen adipose tissues	Affiliated Hangzhou First People's Hospital, Zhejiang University School of Medicine	ZN-20230918-0211-02

Bacterial and Virus Strains		
DH5a Competent Cells	Thermo Fisher Scientific	Cat#18265017
Experimental Models: Organisms/Strains		
Human <i>LINK-A</i> Knock in TG mice	Cyagen Biosciences Inc. (Guangzhou, China).	N/A
C57BL/6	Cyagen Biosciences Inc. (Guangzhou, China).	N/A
Mouse feed		
ND	Xietong Shengwu	Cat#1010063
HFD	Research Diets	Cat#D12492i
Oligonucleotides		
Oligonucleotides for qPCR, RNA interference	This paper	See Table S3
Software and Algorithms		
GraphPad Prism Software	GraphPad software	https://www.graphpad.com/
Leica Application Suite X	Leica Microsystems	https://www.leica-microsystems.com
ImageJ Fiji	National Institutes of Health	https://fiji.sc/
FastTree	Lawrence Berkeley National Lab.	http://meta.microbesonline.org/fasttree/

Table S3. Sequences of Oligonucleotides for qPCR and RNA Interference

Table S3. Sequences of Oligonucleotides for qPCR and RNA Interference		
Oligonucleotides Resource	Sequence	Application
<i>LINK-A</i> F1	AAGCACGTTTCCGACTTGAGTTG	Genotype PCR primer
<i>LINK-A</i> R1	GGGTGAGCATGTCTTTAATCTACC	Genotype PCR primer
<i>LINK-A</i> R2	CTCCTGACTACTCCCAGTCATAGC	Genotype PCR primer
Mouse Ucp1 qF	TTCAGGGAGAGAAACACCTGC	qRT-PCR primer
Mouse Ucp1 qR	CCTTCACGACCTCTGTAGGC	qRT-PCR primer
Mouse Cidea qF	ATCACAACCTGGCCTGGTTACG	qRT-PCR primer
Mouse Cidea qR	TACTACCCGGTGTCCATTCT	qRT-PCR primer
Mouse Ppargc1a qF	CCAGCCTCTTTGCCAGAT	qRT-PCR primer
Mouse Ppargc1a qR	GTCGCTACACCACTTCAATCCA	qRT-PCR primer
Mouse Ppargc1b qF	AACCCAACCACTCTCACAGG	qRT-PCR primer
Mouse Ppargc1b qR	ATGCTGTCTTGTGGGTAGG	qRT-PCR primer
Mouse Cox8b qF	GAACCATGAAGCCAACGACT	qRT-PCR primer
Mouse Cox8b qR	GCGAAGTTCACAGTGGTTCC	qRT-PCR primer
Mouse Cxcl16 qF	TCCTTTTCTTGTGGCGCTG	qRT-PCR primer
Mouse Cxcl16 qR	CAGCGACACTGCCCTGGT	qRT-PCR primer
Mouse Il1 β qF	AGCTTCCTTGTGCAAGTGTCT	qRT-PCR primer
Mouse Il1 β qR	GCAGCCCTTCATCTTTTGGG	qRT-PCR primer
Mouse Il6 qF	TCCTCTCTGCAAGAGACTTCC	qRT-PCR primer
Mouse Il6 qR	GTCACCAGCATCAGTCCCAA	qRT-PCR primer
Mouse Tnf α qF	GTTCTATGGCCAGACCCTCAC	qRT-PCR primer
Mouse Tnf α qR	GGCACCAGTAGTTGGTTGTCTTTG	qRT-PCR primer
Mouse F4/80 qF	CTTTGGCTATGGGCTTCCAGTC	qRT-PCR primer
Mouse F4/80 qR	GCAAGGAGGACAGAGTTATCGTG	qRT-PCR primer
Mouse Cd68 qF	CTTCTGCTGTGGAAATGCAA	qRT-PCR primer
Mouse Cd68 qR	AGAGGGGCTGGTAGGTTGAT	qRT-PCR primer

Mouse Nos2 qF	CCCTTCCGAAGTTTCAGCAGC	qRT-PCR primer
Mouse Nos2 qR	GGCTGTGAGAGCCTGGCTTTGG	qRT-PCR primer
Mouse Hbegf qF	CGGGGAGTGCAGATACCTG	qRT-PCR primer
Mouse Hbegf qR	TTCTCCACTGGTAGAGTCAGC	qRT-PCR primer
Mouse Ldha qF	TCTCGGATGTTGTGAAGGTG	qRT-PCR primer
Mouse Ldha qR	CTGCAGCTCCTTCTGGATTC	qRT-PCR primer
Mouse Pdk4 qF	TCTACAAACTCTGACAGGGCTTT	qRT-PCR primer
Mouse Pdk4 qR	CCGCTTAGTGAACACTCCTTC	qRT-PCR primer
Mouse Hk2 qF	CGTGTCCCTACCTTTGGGTT	qRT-PCR primer
Mouse Hk2 qR	CCAGGTCAAACCTCTCTCGC	qRT-PCR primer
Mouse Pdk1 qF	AGGATCAGAAACCGGCACAAT	qRT-PCR primer
Mouse Pdk1 qR	GTGCTGGTTGAGTAGCATTCTAA	qRT-PCR primer
Mouse Lox qF	ACTTCCAGTACGGTCTCCCG	qRT-PCR primer
Mouse Lox qR	GCAGCGCATCTCAGGTTGT	qRT-PCR primer
Mouse Hif1 α qF	ACCTTCATCGGAAACTCCAAG	qRT-PCR primer
Mouse Hif1 α qR	ACTGTTAGGCTCAGGTGAAC	qRT-PCR primer
Mouse Vegfa qF	CAAAAACGAAAGCGCAAGAAA	qRT-PCR primer
Mouse Vegfa qR	CGCTCTGAACAAGGCTCACA	qRT-PCR primer
Mouse Glut1 qF	GCCAGCCAAGCTCATCAATG	qRT-PCR primer
Mouse Glut1 qR	GAGGCAGTCGGACATGCTC	qRT-PCR primer
Mouse 36B4 qF	AACGGCAGCATTATAACCC	qRT-PCR primer
Mouse 36B4 qR	CGATCTGCAGACACACACTG	qRT-PCR primer
Mouse Gapdh qF	GTTGTCTCCTGCGACTTCA	qRT-PCR primer
Mouse Gapdh qR	GGTGGTCCAGGGTTTCTTA	qRT-PCR primer
Mouse 18S qF	ACAGGATTGACAGATTGA	qRT-PCR primer
Mouse 18S qR	TATCGGAATTAACCAGACA	qRT-PCR primer
Human UCP1 qF	GCTCCAGGTCCAAGGTGAATG	qRT-PCR primer
Human UCP1 qR	CAATGAATACTGCCACTCTCCA	qRT-PCR primer
Human CIDEA qF	CATGTATGAGATGTACTCCGTGTC	qRT-PCR primer
Human CIDEA qR	GAGTAGGACAGGAACCGCAG	qRT-PCR primer
Human PPARGC1A qF	TGTGCAACTCTCTGGAAC	qRT-PCR primer
Human PPARGC1A qR	TGAGGACTTGCTGAGTGGTG	qRT-PCR primer
Human PPARGC1B qF	GCTCTCCTCTTCTTCTCTC	qRT-PCR primer
Human PPARGC1B qR	ATAGAGCGTCTCCACCATCC	qRT-PCR primer
Human CXCL16 qF	GCCATCGGTTTCAGTTCA	qRT-PCR primer
Human CXCL16 qR	CAATCCCCGAGTAAGCAT	qRT-PCR primer
Human IL-1B qF	TTCGACACATGGGATAACGAGG	qRT-PCR primer
Human IL-1B qR	TTTTTGCTGTGAGTCCCGGAG	qRT-PCR primer
Human HB-EGF qF	GGACCGAAAGTCCGTGAC	qRT-PCR primer
Human HB-EGF qR	CCCGTGCTCCT CTTGTTT	qRT-PCR primer
Human HIF1 α qF	AAGTCTGCAACATGGAAGGTAT	qRT-PCR primer
Human HIF1 α qR	TGAGGAATGGGTTCACAAATC	qRT-PCR primer
Human VEGFA qF	ACCATGAACCTTCTGCTGTCTTG	qRT-PCR primer
Human VEGFA qR	ATGGCTTGAAGATGTACTCGATCTC	qRT-PCR primer
Human GAPDH qF	GTCTCCTCTGACTTCAACAGCG	qRT-PCR primer
Human GAPDH qR	ACCACCTGTTGCTGTAGCCAA	qRT-PCR primer
Human ACTIN qF	CACCATTGGCAATGAGCGGTTC	qRT-PCR primer
Human ACTIN qR	AGGTCTTTGCGGATGTCCACGT	qRT-PCR primer

Human 18S qF	GGACACGGACAGGATTGACA	qRT-PCR primer
Human 18S qR	GACATCTAAGGGCATCACAG	qRT-PCR primer
<i>LINK-A</i> qF	TCCTGCTTCTCTCACCCCTTC	ChIP-qRT-PCR primer
<i>LINK-A</i> qR	CTGTTTGTAGAGCCTCCACA	ChIP-qRT-PCR primer
HB-EGF qF	GAGACCCCTGGTATGGGG	ChIP-qRT-PCR primer
HB-EGF qR	GAGAGGGCGGCGTTCACAAT	ChIP-qRT-PCR primer
IL-1 β qF	TGGAGGCCAAAACAGAGGAGTC	ChIP-qRT-PCR primer
IL-1 β qR	TGGTGTCTGGTCTGCAGAT	ChIP-qRT-PCR primer
CXCL16 qF	CCATGCTCATCCGTCAACAAG	ChIP-qRT-PCR primer
CXCL16 qR	CCACATTTGTCTCTCGCTGC	ChIP-qRT-PCR primer
<i>LINK-A</i> LNA	TGGATAAATGAGCTGT	RNA interference
Ser LNA (Negative control)	AACACGTCTATACGC	RNA interference
shHIF1 α #1 F	CCGG AAGCCGCTGGAGACACAATCA CTCGAG TGATTGTGCTCCAGCGGCTT TTTTG	RNA interference
shHIF1 α #1 R	AATTCAAAAA AAGCCGCTGGAGACACAATCA CTCGAG TGATTGTGCTCCAGCGGCTT	RNA interference
shHIF1 α #2 F	CCGG AAGGAAGAACTATGAACATAA CTCGAG TTATGTTCAATGTTCTTCCTT TTTTG	RNA interference
shHIF1 α #2 R	AATTCAAAAA AAGGAAGAACTATGAACATAA CTCGAG TTATGTTCAATGTTCTTCCTT	RNA interference

Table S4. Sequences of Promoter

[illegible]

HB-EGF promotor sequence(2300bp):

ATGAAGCTGCTGCCGTGCTGGTGTGAAGCTCTTTCTGGCTGCAGTTCTCTCGGCACTGGTGACTGGCGAGAGCCTGGAGCGGCT
 TCGGAGAGGGCTAGCTGCTGGAACAGCAACCCGGACCTCCCACTGTATCCACGACAGCTGTACCCCTAGGAGGCGCGCGG
 GACCGGAAAGTCCGTGACTTGCAGAGGCGAGATCTGGACCTTTTGAGAGTCACTTTATCTCCAAAGCCAAAGCAAGCTGGCCACAC
 AAACAAGGAGGAGCACGGGAAAAGAAAGAAAGGCAAGGGGGTGTCAAGGAAGTGTTCCTCTCCACCCTCCCTGGGGAGAG
 CCTTGACCCCAAGGTGGCTTTGTTTTGGGGAAGCAGGTGGCCAGGCAGCTGCTGAGGGGAGGGCAAAGTCATGGCTTGGGCGGTGA
 CCATGCACGTAGGTCTTAGGCCGATACTCATCACCCAAAAGGATAATCTAACAGTGGCTCTTAGCCAAATAGGTGACAGGTGAGCT
 TCTGTCTGAAACCAGCCCACTATATCATAAAGCCTCAGATATGGCCTAGGGGAAGGCAAGCGATCTAGAGAAGTGAGAGAAAAGA
 GGCCTCTAAGGAAATAGAGTGGTCAAGTGCTTAGTGATTATTATTAACAGCAACAATAATAATGGGTACATTTATTGAGAGCTTATA
 GTGTGCTGGCACTATACTAATACTTTTATGTGTTCCCGCATTCACTTCTCAAAAAGCTCCCTACAAAAGTAGTCATTATTATTATTGTT
 ATTATTTTTTCCATTTGAGAGAGCTCAAAAAGCTGAGGCTCAGTGAAATTAAGTTTCTTGTCCAAGGTCAACAGTAAGTAGCAAAAGGC
 AGGATTCAAACTCAGGTCTGTAGTGATTCCAGTGCATATGAAACCTAGGCCTAGGTTACTGGAGCAAGTTAAGAGGGCAGGGCC
 AGACTGACCCCTGCAAAAGGTGGGGATAGGGAGGGACAGTGGGATAGACTGTGTATAAATAGCAGGCCTGTTTTATCCCTACACAC
 AGGCAGAAATCCGGCAAGTGCTCTATCTGGCACAATTTACAGGCTCTGAAAAGCCAACCCAGCCCTGTGTCAAACAACCTCTCTCCCA
 GCACGTCCCTGGAAACTGAAATACCTACTGAGACACTCATTTCTTTGGGTCAAGGACAGTGAAGTCCAGAACCCCTGCAAAAGCAC
 ATGGGTTGGAAGAGAAACACCTAGTGCTGTGAAACAGCAATCTAGGTAGCCAGCTGGGTCTATGGGAGGGTGGCTCATTTTTT
 ATGTTGAGGAAAAAGGTGGCAGTCAAAGGCAAGGAAAAAGATAAGGCTTGTTTCAACCTGTGTCTCTCAATCTCAGAAGCCT
 GGGAAATTACAACCCATAAGTTCAAAAATCTAATGTCCCTTGAGGGAAGATGGGAGGAAGCCACTCCAGAGACTAAGAGAGTAA
 CTTCCTTTGCTTTTGTGATTGATACTAAGTGCCTGCAACTTCAACTCTGGTTATCTCGGGTTTATGAGAAGCGCTGGCATTACAGT
 GTCTAATTTTTAAATGATAGATGAATTAATTATTATTGATACTTGCCGTGTTGGTAGGCCAAAAGGTGGGCTTCTGAAACTTAGAGG
 GTGACAGGGATGAGACCCCTGGTATGGGGCTTAGATGAAGCAAGAGGCCAGGCCCGGAACAGAACTCAGCTCAGCAGTCACTCA
 CAAGGCTGCAGAGTGACAGGCGTTTGGGGAAGGTAGGAACCGCTCTCCGCCACCTGCCGGTCACTAGCCCTACCCACAATCAGG
 CCGCGGCTGCCTCGGCCTGGTCCCAAAAATTGTGAACGCGCGCCTCTCTCCCGCCAGTCTCCGCGCGCGCGCGGGTTCGGGG
 GCTGCGCTGCCGCGCGCGGAGCGCGGCGAGCTTCCAGCACGGGAGGAGGGAGGGGCGCGCGCGGGCGGGCGGGCGGAGCTTCC
 GCGCTCTGAGCCTTATTCTCGGCGCGGAGCGCGGCGAGCGCTCATTCCGCCGAAGGAGCTACGCGGGCCAGCTGCTGGCTGG
 CCTGACCTAGGCGCGCGGGTCTGGGCGCGCGCGCGGGCTGAGTGAGCAAGACAAGACTCAAGAAGAGCGAGCTGCGCC
 TGGGTCCCGGCCAGGCTTGACGACAGAGCGGGCGGCGAGACGGTGCCCGCGGAATCTCTGAGCTCCGCGCCAGCTCTGGTG
 CCAGCGCCAGTGCCGCGGCTTCGAAAGTGACTGGTGCCTCGCCGCTCTCTCGGTGCGGGACCA

HB-EGF promotor truncated sequence:

GAGACCCTGGTATGGGGCTTAGATGAAGCAAGAGGCCAGGCCCGGAACAGAACTCAGCTCAGCAGTCACTCAAGGCTGCAG
 AGTGACAGGCGTTTGGGGAAGGTAGGAACCGCTCTCCGCCACCTGCCGGTCACTAGCCCTACCCACAATCAGGCCGCGGCTGCC
 TCGGCTGGTCCCAAAAATTGTGAACGCGCCCTCTC

HB-EGF promotor mutated truncated sequence:

GAGACCCTGGTATGGGGCTTAGATGAAGCAAGAGGCCAGGCCCGGTTATAAACTCAGCTCAGCAGTCACTTAAAGGCTGCAGA
 GTGACAGGCGTTTGGGGAAGGTAGGAACCGCTCTCCGCCACCTGCCGGTCACTAGCCCTACCTTAAATCAGGCCGCGGCTGCCTC
 GGCCTGGTCCCAAAAATTGTGAACGCGCCCTCTC

IL-1 β promotor sequence(2300bp):

ATGGCAGAAGTACCTGAGCTCGCCAGTGAAATGATGGCTTATTACAGTGGCAATGAGGATGACTTGTTCTTTGAAGCTGATGGCCCT
 AAACAGATGAAGTGCTCCTTCCAGGACCTGGACCTCTGCCCTCTGGATGGCGGCATCCAGCTACGAATCTCCGACCACCACTACAG
 CAAGGGCTTCAGGCAGGCGCGCTCAGTTGTTGGCCATGGACAAGCTGAGGAAGATGCTGGTTCCCTGCCACAGACCTCCAG
 GAGATGACCTAGCAGCTTCTTCCCTCATCTTTGAAGAAAGAAAGAAAGAAAGAAAGAAAGAAAGAAAGAAAGAAAGAAAGAAAGAA
 AAGAA
 GCTCTGAGGAAGGTGGCAGTTCTTACAACGGGAGAACAGTGGTTAATTTGCAAAGTGGATCCTGTGGAGGCAAAACAGAGGAGT
 CCCCAGGCCACCCAGACAGGGCTTTAGCTATCTGCAGGACCAGACACCAAAATTTAGGAGGGCTCAGTGTAGGAATGGATTAT
 GGCCTATCAAATTCACAGAACTAATCATGTTGAACAGCTTTTAGATTCTCTGGGAAATATAACTTACTAAAGATGGAGTTCTTGT
 GACTGACTCCTGATATCAAGATACTGGGAGCCAAATTAATAATCAGAAGGCTGCTTGGAGAGCAAGTCCATGAAATGCTCTTTTCC
 CACAGTAGAACCTATTTCCTCGTGTCTCAAATACTTGCACAGAGGCTCACTCCCTTGGATAATGCAGAGCGAGCAGATACCTGGC
 ACATACTAATTTGAATAAATGCTGTCAAATTTCCATTCCACCTTCAAGCAGCAAACTTACCACCTGAATGTACATGCCAGGCAC
 TGTGCTAGACTTGGCTCAAAAAGATTTCAGTTTCTTGGAGGAACAGGAGGAGCAAGGTTTCAACTCACTGCTATAAGAAAGTGTTA
 CAGGCTGGACACGGTGGCTACGCCTGTAATCCCAACACTTTGGGAGGCCGAGGCGGGCAGATCACAAGGTGAGGAGATCGAGAC
 CATCCTGGCTAACATGGTGAAACCTGTCTCTACTAAAAATACAAAAAATTAGCCGGGCGTGGCGGCAGGTGCTGTAGTCCCAGC
 TGCTGGGGAGGCTGAGGCAGGAGAATGGTGTGAACCCGGGAGGCGGAACCTTGCAAGGGGCGGAGATCGTGCCTGCACTCCAG
 CCTGGGCGACAGAGTGAGACTCTGTCTCAAAAAAAGAGTTATGATGCAGACCTGTCAAAAGAGGCAAAAGGAGGGTGT
 TCCTACACTCCAGGCACTGTTTCAACCTGGACTCTCACTTCTACAAATGGAGGGCTCCCTGGGCGAGTACCCTGGAGCAGGCA
 CTTTGCTGGTGTCTCGGTTAAAGAGAACTGATACTCTTGGTTGGTATTACCAAGAGATAGAGTCTCAGATGGATATTCTTACAGAA
 ACAATATTCCACTTTTCAGAGTTTACCAAAAAATCATTTTAGGCAGAGCTCATCTGGCATTGATCTGGTTTCATCCATGAGATTGGCTA
 GGGTAACAGCACCTGGTCTTGCAGGGTGTGTGAGCTTATCTCAGGGTTGCCCAACTCCGTGAGGAGCCTGAACCCCTGCATACC
 GTATGTTCTCTGCCCCAGCCAAGAAAGGTCAATTTCTCCTCAGAGGCTCCTGCAATTGACAGAGAGCTCCTGAGGCAGAGAACAG
 CACCAAGGTAGAGACCCACACCTCAATACAGACAGGAGGGCTATTGGCCCTTCAATTGTACCCATTATCCATCTGTAAGTGGGA
 AGATTCTAAACTTAAGTACAAAGAAAGTGAATGAAGAAAGTATGTGCATGTATAAATCTGTGTCTTCCACTTTGTCCACATATA
 CTAAATTTAAACATTCTTCAACGTGGGAAAATCCAGTATTTAATGTGACATCACTGCACAACGAATTGTCAGGAAAAACAATGCAT
 ATTTGCTGGTGATACATTTGCAAAATGTGTATAGTTTGTACTCTTCCCTTCCATGAACAGAGAAATATCTCAGTTTATTAGTC
 CCTCCCTAAGAGCTTCCACCAATACTTTTCCCTTCTTAACTGATTGTGAAATCAGGTATTCAACAGAGAAATTTCTCA
 GCCTCTACTTCTGCTTTTGAAGCCATAAAAAACAGCGAGGGAGAACTGGCAGAT

IL-1 β promotor truncated sequence:

TGGAGGCAAAACAGAGGAGTCCCTTAGGCCACCCAGACAGGGCTTTAGCTATCTGCAGGACCAGACACCA

IL-1 β promotor mutated truncated sequence:

TGGAGGCAATTATAAGGAGTCCCTTAGGCCACCCAGACAGGGCTTTAGCTATCTGCAGGACCAGACACCA

CXCL16 promotor sequence(2300bp):

[illegible]

CXCL16 promotor truncated sequence:

CCATGCTCATCCGTCAAAAGTAAGAGACAGGAAACGTGCAGGGGGGCAGCCTCTGGGGAAAGAGGGAAAAGTTGAGGTGGTAGG
GGGCAGCGAGAGACAAATGTGG

CXCL16 promoter mutated truncated sequence:

CCATGCTCATCCGTTTAAAGTAAGAGACAGGAATCTCTCAGGGGGGCAGCCTCTGGGGAAAGAGGGAAAAGTTGAGGTGGTAGGG
GGCAGCGAGAGACAAATGTGG