

Surfactant treatment before first breath for respiratory distress syndrome in preterm lambs: comparison of a peptide-containing synthetic lung surfactant with porcine-derived surfactant

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Background: In a recent study utilizing a saline-lavaged adult rabbit model, we described a significant improvement in systemic oxygenation and pulmonary shunt after the instillation of a novel synthetic peptide-containing surfactant, Synsurf. Respiratory distress syndrome in the preterm lamb more closely resembles that of the human infant, as their blood gas, pH values, and lung mechanics deteriorate dramatically from birth despite ventilator support. Moreover, premature lambs have lungs which are mechanically unstable, with the advantage of being able to measure multiple variables over extended periods. Our objective in this study was to investigate if Synsurf leads to improved systemic oxygenation, lung mechanics, and histology in comparison to the commercially available porcine-derived lung surfactant Curosurf[®] when administered before first breath in a preterm lamb model.

Materials and methods: A Cesarean section was performed under general anesthesia on 18 time-dated pregnant Dohne Merino ewes at 129–130 days gestation. The premature lambs were delivered and ventilated with an expiratory tidal volume of 6–8 mL/kg for the first 30 minutes and thereafter at 8–10 mL/kg. In a randomized controlled trial, the two surfactants tested were Synsurf and Curosurf[®], both at a dose of 100 mg/kg phospholipids (1,2-dipalmitoyl-L- α -phosphatidylcholine; 90% in Synsurf, 40% in Curosurf[®]). A control group of animals was treated with normal saline. Measurements of physiological variables, blood gases, and lung mechanics were made before and after surfactant and saline replacement and at 15, 30, 45, 60, 90, 120, 180, 240 and 300 minutes after treatment. The study continued for 5 hours.

Results: Surfactant treatment led to a significant improvement in oxygenation within 30 minutes, with the Synsurf group and the Curosurf[®] group having significantly higher ratios between arterial partial pressure of oxygen/fraction of inspired oxygen ($\text{PaO}_2/\text{FiO}_2$; $P = 0.021$) compared to that of the control (saline-treated) animals. Dynamic compliance improved in the three groups over time, with no intergroup differences. All of the surfactant-treated animals survived, and one in the saline group died before the study ended. Histology between groups was not different, showing mild–moderate injury patterns.

Discussion: Treatment with surfactants before first breath clearly resulted in improved systemic oxygenation within 30 minutes of instillation. Both Synsurf- and Curosurf[®]-treated animals experienced similar and more sustained improvement in oxygenation and decreased calculated shunt compared to saline-treated animals.

Keywords: respiratory distress syndrome, preterm lambs, pulmonary surfactant, gas exchange, oxygenation

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Introduction

The essential role of hydrophobic surfactant proteins SP-B and SP-C in natural surfactant function has been well described.^{1–3} Increasingly, the recombinant production of these hydrophobic proteins has been the focus for the development of synthetic surfactants, especially after the protein-free synthetic surfactants were shown to perform somewhat inferiorly to mammalian-derived surfactants in the acute management of new-born infants with respiratory distress syndrome (RDS).⁴ Some *in vitro* studies have shown that mixtures of phospholipids with SP-B yield surfactants that are more efficacious than SP-C-based surfactants, with SP-B-based preparations displaying markedly lower susceptibility to fibrinogen inhibition than SP-C preparations.^{5,6} However, head-to-head trials comparing various animal-derived surfactants that provide varying amounts of SP-B or phospholipids have shown minor differences in outcomes related to the management of RDS or none at all. Post hoc analysis of data from one study showed better survival using a high initial dose of Curosurf[®] (Chiesi Farmaceutici SpA, Parma, Italy) when compared with Survanta[®] (Abbott Laboratories, Abbott Park, North Chicago, IL, USA).⁷ New surfactant preparations that include peptides or whole proteins that mimic endogenous surfactant protein have recently been developed and tested.^{8–10} A new-generation synthetic surfactant that contains a peptide mimicking the action of SP-B, Surfaxin (Discovery Laboratories, Warrington, PA, USA), performed better than a protein-free synthetic surfactant (Exosurf; GlaxoKline-Smith, Brentford, UK), and similarly to animal-derived surfactants (Survanta[®] and Curosurf[®]).¹¹ A Cochrane review identified two studies that compared protein-containing synthetic surfactants to animal-derived surfactant preparations. In a meta-analysis of these two studies, infants who received protein-containing synthetic surfactant compared to animal-derived surfactant extract did not demonstrate significant differences in primary outcomes, ie, death and chronic lung disease.¹²

In a recent study utilizing a saline-lavaged adult rabbit model, we described a significant improvement in systemic oxygenation and pulmonary shunt after the instillation of a novel synthetic peptide-containing surfactant, Synsurf.⁸ Our objective in the present study was to investigate if Synsurf leads to improved systemic oxygenation, lung mechanics, and histology in comparison to the commercially available porcine-derived lung surfactant Curosurf[®] when administered before first breath in a preterm lamb model.

Materials and methods

Surfactant preparations

Synsurf was prepared as described previously.⁸ Briefly, dipalmitoylphosphatidylcholine (DPPC), hexadecanol, and phosphatidylglycerol were mixed in a 10:1.1:1 ratio (w/w) in chloroform. The organic solvent was then removed by rotary evaporation, and the mixture was dried under a continuous stream of nitrogen at room temperature. Poly-L-lysine (~100–120 residues) was mixed with poly-L-glutamate (~80 residues) and incubated at 37°C in 0.1 M NaCl to give a complex which is 50% neutralized. The dried phospholipid film was then hydrated with the polymer mixture (3% by weight of the phospholipid concentration) and gently mixed in the presence of glass beads. The mixture was ultrasonicated on ice under a stream of nitrogen (20 watts for 7 × 13 seconds at 60-second intervals). Thereafter, 24 mg of tyloxapol was added to the preparation, and the tube was sealed under nitrogen before use. The molecular structure of the two-stranded polymer and possible interactions with DPPC is shown in Figure 1. Both Synsurf and Curosurf[®] (Chiesi Farmaceutici SpA) were used at a dose of 100 mg/kg phospholipids.

Delivery and ventilation of lambs

Animal care and experimental procedures were performed under approval from the Faculty of Medicine and Health Sciences Research Committee of Stellenbosch University. Eighteen time-dated pregnant Dohne Merino ewes at 129–130 days gestation (normal gestation 148–154 days) were anesthetized with intravenous 2.5% thiopental sodium, intubated, and mechanically ventilated with inhalation of 1%–2% halothane in oxygen. A Cesarean section was performed, and the fetal head and neck were exteriorized. To withdraw lung fluid, subcutaneous lignocaine (2%) was infiltrated into the midtracheal region with a 24-gauge needle attached to a syringe. Ten to 15 mL of fetal lung fluid was sampled to determine lung maturity (lamellar body count) and to remove excessive lung fluid. A neck incision was then made, followed by a tracheotomy. An uncuffed 4 mm or 4.5 mm endotracheal tube was placed and advanced to 4 cm. The umbilical cord was cut, the fetus was then delivered, weighed, dried, and placed in a supine position on warming pads under a radiant warmer. Within 2 minutes after umbilical cord ligation, before first breath, two of the groups received 100 mg/kg of either Synsurf or Curosurf[®]. The third group (control) received normal saline. The lambs were anesthetized and sedated with continuous infusion of midazolam (Dormicum[®] [Roche, Basel, Switzerland], 0.1 mg/kg/hour)

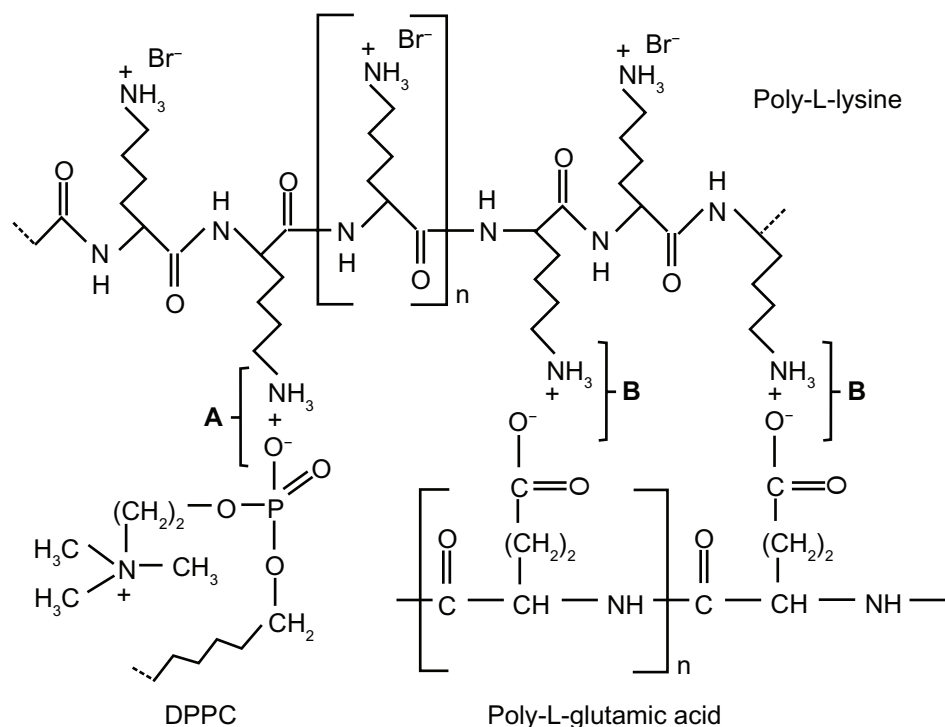


Figure 1 Molecular structures of poly-L-lysine, poly-L-glutamic acid, and dipalmitoylphosphatidylcholine (DPPC).

Notes: Possible electrostatic interactions between lysyl side chains of poly-L-lysine and DPPC (**A**), and strong electrostatic interaction between poly-L-lysine and poly-L-glutamic acid forming a two-stranded construct (**B**).

and morphine (5–10 $\mu\text{g}/\text{kg}/\text{hour}$). Supplemental intermittent ketamine dosages (10 mg/kg) were administered when spontaneous breathing was observed on the flow curves. Animals were paralyzed with intravenous pancuronium bromide (0.1 mg/kg , hourly). A rectal thermistor was placed to continuously monitor body temperature, which was maintained at 38°C–39°C. To minimize lung injury, ventilation during the first 30 minutes after birth was initiated with time-cycled, pressure-limited assist control ventilation (AVEA[®] ventilator system; CareFusion, San Diego, CA, USA). The expiratory tidal volume was set at 6–8 mL/kg and was then increased to 8–10 mL/kg . Hereafter, the ventilator settings were held constant throughout the study at an inspired O_2 fraction (FiO_2) of 1.0, a rate of 50 breaths/minute, an inspiratory time of 0.50 seconds, and a positive end-expiratory pressure of 4 $\text{cm H}_2\text{O}$. Immediately after ventilation was started, a 5 French catheter was passed via the femoral artery into the aorta for monitoring blood pressure and heart rate and for blood gas measurements. Each lamb received a continuous infusion of 5% dextrose water that was begun at the start of the study. Lambs were assigned to one of three groups (six lambs/group). Measurements of physiological variables, blood gases, and lung mechanics (tidal volume, dynamic compliance of the respiratory system [C_{dyn}]) were made

before and after surfactant and saline replacement and at 15, 30, 45, 60, 90, 120, 180, 240 and 300 minutes after treatment. After 5 hours, all live animals were killed by lethal injection of intra-arterial 15% potassium chloride. The chest wall was opened and quasistatic maximal inspiratory capacity of the intact lung at 35 $\text{cm H}_2\text{O}$ peak plateau pressure and positive end-expiratory pressure of zero was determined after exsanguination. In lambs without a pneumothorax, the lung (right lung caudal, middle, and cranial lobes) was then gravitationally filled with formaldehyde (4%) and inflated under a constant pressure of 25 $\text{cm H}_2\text{O}$. This lung was used for histology and morphometry. Total lung water was determined after the animals were killed by calculating the difference between the wet weight minus the weight of the lung exposed to 80°C until the weight was constant for 48 hours (~4–6 days). It is expressed as the lung wet weight/dry weight ratio.

Statistical analysis

STATISTICA version 10 (StatSoft, Tulsa, OK, USA) and GraphPad Prism 5 (GraphPad Software Inc, La Jolla, CA, USA) were used to determine comparability of the experimental groups before and after surfactant instillation. Changes of variables between the groups were analyzed with one-way analysis

of variance (and nonparametric). The Kruskal–Wallis test and Dunn's multiple comparison test were used as the discriminating post-test. Data are expressed as mean \pm standard deviation. Significant differences were accepted at P -values < 0.05 .

Mean linear intercept analysis

The mean linear intercept as an indicator of alveolar airspace size was calculated from counting lines of defined length as previously described by Dunnill.¹³ Briefly, the lines were randomly placed on every lung section, and the number of intersections crossing the lines were counted. The mean linear intercept is calculated from the length of the lines multiplied by the number of lines, divided by the sum of all counted intercepts.

Results

Respiratory outcomes

At randomization, arterial blood gases, ventilator indexes, and hemodynamic variables were similar for all three groups. Table 1 gives an overall summary of the pretreatment parameters of the three groups. All lambs were severely surfactant-deficient as their lamellar body counts were less than 15,000/ μ L and lecithin/sphingomyelin ratios were less than 2. The surfactant deficiency was further reflected in the overall low mean C_{dyn} (mean for $n = 18$, 0.31 ± 0.09 mL/cmH₂O · kg) and poor oxygenation status as reflected by a low arterial/alveolar ratio (mean for $n = 18$, 0.03 ± 0.01) and a high oxygenation index (mean for $n = 18$, 53.69 ± 32.1). There were no significant differences between the three groups.

Table 1 Indexes, lung mechanics, cord blood gasses, and calculated shunt

Variable	Synsurf (n = 6)	Curosurf® (n = 6)	Saline (n = 6)
L/S ratio	0.97 \pm 0.09	0.97 \pm 0.05	1.15 \pm 0.09
Lamellar body count/ μ L	5333 \pm 3933	5500 \pm 1049	5500 \pm 2881
PaO ₂ (mmHg)	27.63 \pm 9.3	19.25 \pm 9.2	27.00 \pm 9.0
PCO ₂ (mmHg)	43.80 \pm 9.2	47.35 \pm 11.5	50.11 \pm 7.2
TCO ₂ (mmol/l)	26.57 \pm 4.4	26.92 \pm 1.5	27.15 \pm 1.7
C_{dyn} (mL/cmH ₂ O · kg)	0.27 \pm 0.06	0.31 \pm 0.05	0.38 \pm 0.12
a/A ratio	0.04 \pm 0.02	0.03 \pm 0.01	0.04 \pm 0.01
pH	7.39 \pm 0.07	7.34 \pm 0.10	7.32 \pm 0.07
OI	44.06 \pm 14.8	73.89 \pm 48.2	43.11 \pm 15.2
Q ₃ /Q _t	62.02 \pm 7.0	67.62 \pm 8.7	62.62 \pm 7.7

Notes: Baseline measurements were compared using nonparametric Kruskal–Wallis test. Data expressed as a mean \pm standard deviation (t -test). Curosurf® manufactured by Chiesi Farmaceutici SpA, Parma, Italy.

Abbreviations: a/A ratio, arterial/alveolar ratio; C_{dyn} , dynamic respiratory compliance; L/S, lecithin/sphingomyelin; OI, oxygenation index; PaO₂, arterial partial pressure of O₂; PCO₂, arterial partial pressure of CO₂; Q₃/Q_t, pulmonary shunt; TCO₂, total CO₂.

Following instillation of surfactant, the Synsurf group and the Curosurf® group experienced significant improvement in oxygenation (arterial partial pressure of oxygen [PaO₂]/FiO₂ ratio, $P = 0.021$) within 30 minutes (Table 2 and Figure 2A), compared to that of the saline-treated animals. Moreover, both surfactant-treated groups showed better oxygenation over the time period of the study. For Synsurf-treated animals, oxygenation was significantly better compared to the saline-treated animals at timepoints 30, 45, 60, 90, 120 and 180 minutes. With exception of the 120-minute timepoint, the finding was similar for the Curosurf® versus saline-treated

Table 2 Physiological variables, lung mechanics, and postnatal blood gasses

Variable	Synsurf group (n = 6)	Curosurf® group (n = 6)	Saline group (n = 6)
Weight (kg)	3.4 \pm 0.4	3.2 \pm 0.5	3.4 \pm 0.7
Tracheal fluid (mL/kg)	4.43 \pm 1.6	3.34 \pm 1.6	4.47 \pm 1.8
Vte (mL/kg)			
30 min	7.77 \pm 0.4	7.61 \pm 0.6	7.95 \pm 1.0
90 min	8.88 \pm 1.0	9.48 \pm 0.6	9.28 \pm 0.8
180 min	8.93 \pm 0.9	9.77 \pm 0.5	9.13 \pm 0.5
300 min	8.45 \pm 1.1	9.43 \pm 1.3	9.46 \pm 0.7
MABP (mmHg)			
30 min	59.17 \pm 7.6	62.50 \pm 4.9	63.67 \pm 8.8
90 min	57.17 \pm 5.0	61.00 \pm 8.0	55.17 \pm 1.2
180 min	53.83 \pm 5.1	58.33 \pm 5.0	50.67 \pm 7.4
300 min	48.50 \pm 5.8	48.83 \pm 9.9	48.60 \pm 4.3
PaO ₂ /FiO ₂ (mmHg)			
30 min	311.13 \pm 102.73	346.38 \pm 126.40	160.38 \pm 89.11
90 min	245.59 \pm 136.04	265.13 \pm 87.65	117.30 \pm 93.64
180 min	205.00 \pm 155.96	189.88 \pm 113.22	59.63 \pm 43.96
300 min	288.88 \pm 137.13	133.75 \pm 110.64	190.90 \pm 149.58
PaCO ₂ (mmHg)			
30 min	68.74 \pm 24.4	70.66 \pm 17.6	80.13 \pm 27.0
90 min	45.69 \pm 7.7	53.08 \pm 18.0	52.28 \pm 17.2
180 min	45.55 \pm 8.0	51.56 \pm 19.3	47.09 \pm 19.9
300 min	52.73 \pm 24.9	63.50 \pm 16.2	54.39 \pm 23.53
TCO ₂ (mmol/L)			
30 min	24.7 \pm 3.5	26.4 \pm 3.2	25.2 \pm 3.6
90 min	24.8 \pm 2.3	25.5 \pm 2.0	22.6 \pm 2.8
180 min	25.8 \pm 1.1	24.1 \pm 1.9	22.6 \pm 3.3
300 min	26.2 \pm 2.3	26.8 \pm 3.4	24.0 \pm 2.5
C_{dyn} (mL/cmH ₂ O · kg)			
30 min	0.34 \pm 0.05	0.34 \pm 0.03	0.33 \pm 0.15
90 min	0.39 \pm 0.06	0.43 \pm 0.04	0.37 \pm 0.08
180 min	0.43 \pm 0.07	0.48 \pm 0.05	0.39 \pm 0.10
300 min	0.45 \pm 0.09	0.48 \pm 0.05	0.47 \pm 0.07

Notes: Data expressed as mean \pm standard deviation (t -test). Bolded PaO₂/FiO₂ values reflect significant higher ratios ($P = 0.021$). Curosurf® manufactured by Chiesi Farmaceutici SpA, Parma, Italy.

Abbreviations: C_{dyn} , dynamic respiratory compliance; MABP, mean arterial blood pressure; PaCO₂, arterial partial pressure of CO₂; PaO₂/FiO₂, arterial partial pressure of oxygen/fraction of inspired oxygen ratio; TCO₂, total CO₂; Vte, expiratory tidal volume.

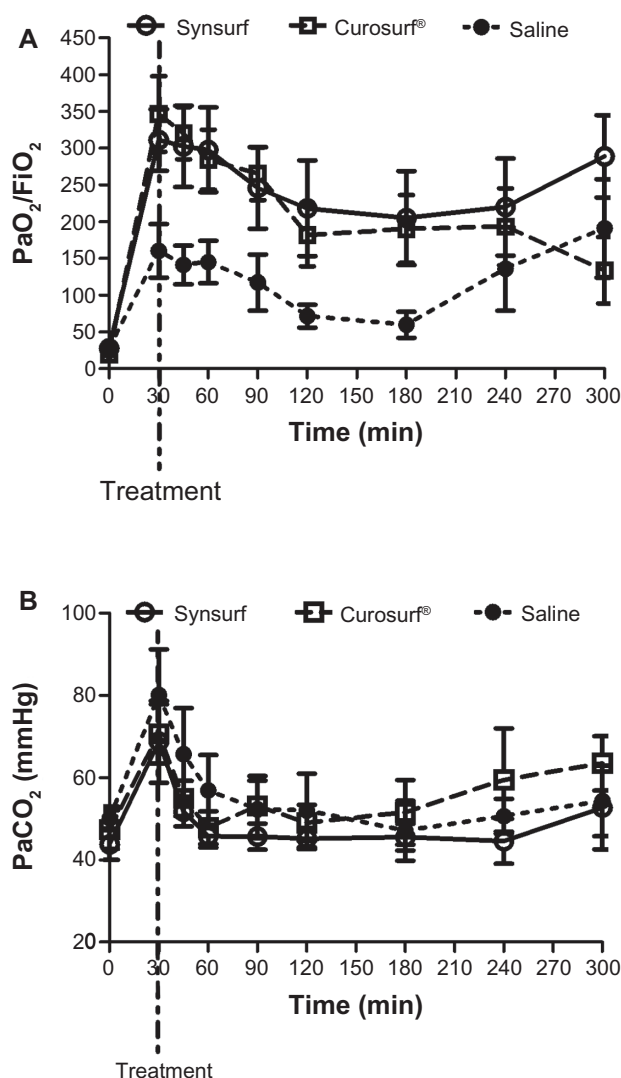


Figure 2 Time profiles for PaO₂/FiO₂ and PaCO₂.

Notes: Time profile of oxygenation as depicted by a comparison of the sequential measurements of the ratios of arterial PaO₂ to inspired oxygen fractions FiO₂ (A). Significant timepoint differences between the groups were analyzed by mixed model repeated measures analysis of variance and are described in the results. Arterial PaCO₂ before and after treatment with Synsurf or Curosurf[®] or saline (B). Values are shown as the mean ± standard deviation (*t*-test). Lambs were treated before first breath. Curosurf[®] manufactured by Chiesi Farmaceutici SpA, Parma, Italy.

Abbreviations: FiO₂, fraction of inspired oxygen; PaCO₂, arterial partial pressure of CO₂; PaO₂, arterial partial pressure of oxygen.

animals. However, at 300 minutes the Synsurf-treated animals had a significantly better oxygenation status when compared to Curosurf[®] ($P = 0.014863$, determined by mixed model repeated measures analysis of variance). A decrease in calculated pulmonary shunt (time period 0–300 minutes; Figure 3) in the Synsurf, Curosurf[®], and saline-treated group of animals was observed (intergroup differences: Synsurf 18.82 ± 16.19 versus Curosurf[®] 22.35 ± 18.42 , $P = 0.09$; and Synsurf versus saline 34.41 ± 12.52 , $P = 0.003$; Friedman analysis of variance). At 300 minutes the mean calculated value for Synsurf was 18.44%, versus 36.29% and 32.63% for the Curosurf[®] group

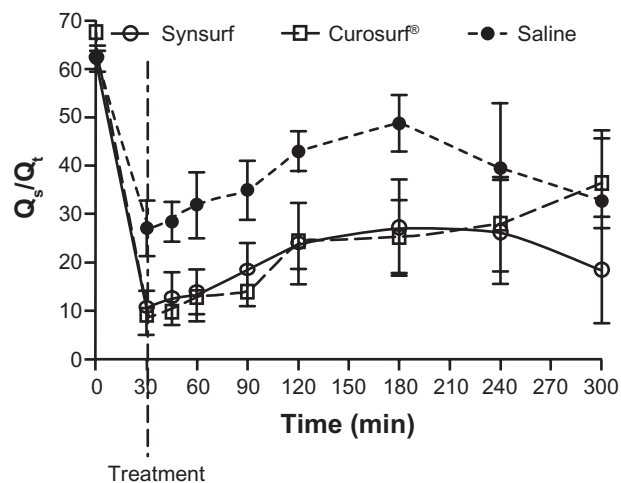


Figure 3 Time profile of a comparison of pulmonary shunt (Q_s/Q_t) between lamb groups before and after administration of Synsurf or Curosurf[®] or saline.

Notes: Values are shown as the mean ± standard deviation. Q_s/Q_t was lower for the Synsurf group at 180 minutes ($P < 0.05$ by two-way analysis of variance, versus saline) and at 300 minutes ($P = 0.0002$, *t*-test, versus saline and Curosurf[®] groups). Curosurf[®] manufactured by Chiesi Farmaceutici SpA, Parma, Italy.

and the saline-treated group, respectively. Significant lower pulmonary shunt values for Synsurf-treated animals occurred at 180 minutes ($P < 0.05$, two-way analysis of variance, Synsurf versus the saline group) and at 300 minutes ($P = 0.0002$, *t*-test, Synsurf versus the saline group and the Curosurf[®] group). The arterial partial pressure of carbon dioxide (PaCO₂) also decreased from about 70 mmHg to about 58 mmHg in the surfactant-treated animals (Figure 2B).

Pulmonary mechanics

To avoid compromising the effect of surfactant rescue treatment and to minimize lung over-distension, initial ventilation with an expiratory tidal volume of 6–8 mL/kg and thereafter 8–10 mL/kg was used. As a consequence, we found no significant differences between the Synsurf and Curosurf[®] groups for expiratory tidal volumes (Figure 4A) as well as for the arterial PaCO₂ values (Figure 2B) between the start of the study and at 300 minutes. Total C_{dyn} (Figure 4B) steadily increased in all of the study groups over time between timepoint 0 minutes (start of experiment) and 300 minutes ($P = 0.0012$, $P = 0.00026$, and $P = 0.0268$ for Synsurf, Curosurf[®], and saline, respectively). There were no significant intergroup differences at 300 minutes (Synsurf C_{dyn} 0.45 ± 0.09 , Curosurf[®] C_{dyn} 0.48 ± 0.05 , and saline C_{dyn} 0.47 ± 0.07). All of the surfactant-treated animals survived and one in the saline group died before the study ended. However, two lambs developed a pneumothorax during the study, one each in the saline group and Synsurf-treated group. Representative lungs in the three groups were used for quasistatic lung volume measurements, as determined

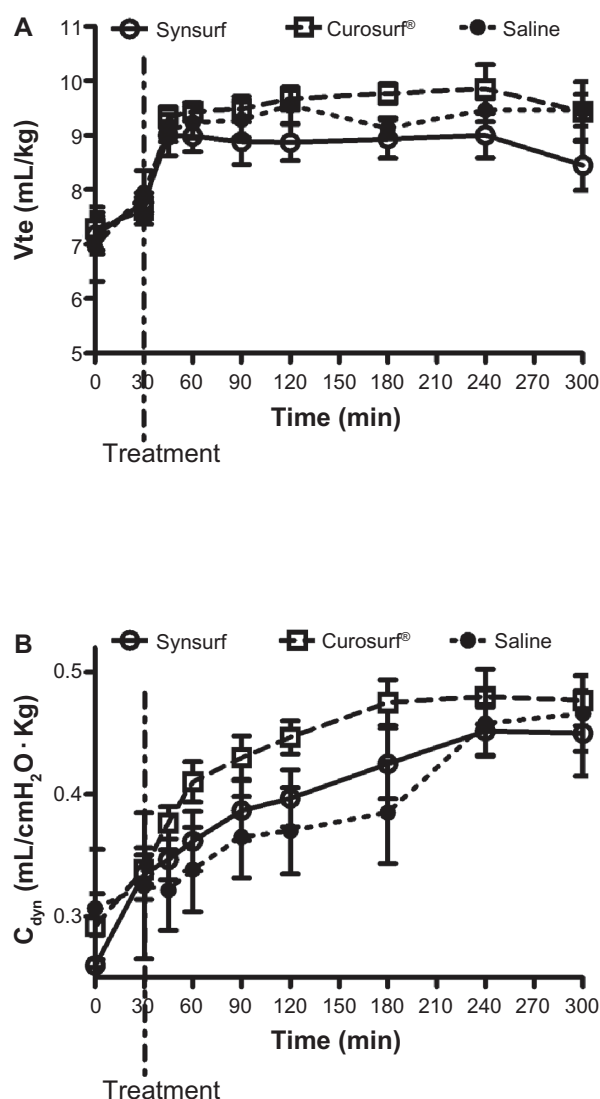


Figure 4 Time profiles of expiratory tidal volume (V_{te}) and dynamic respiratory compliance (C_{dyn}) of lambs treated with Synsurf or Curosurf[®] or saline before first breath.

Notes: Time profile of V_{te} of lambs treated with Synsurf or Curosurf[®] or saline before first breath (A). Values are shown as the mean \pm standard deviation ($P > 0.05$ by two-way analyses of variance in all three groups). Sequential measurements show that C_{dyn} increased significantly between time points 0 minutes and 300 minutes in all three groups (B) (Synsurf $P = 0.0012$; Curosurf[®] $P = 0.00026$; saline $P = 0.0268$ determined by t -test). Curosurf[®] manufactured by Chiesi Farmaceutici SpA, Parma, Italy.

in the open-chest state for maximal inspiratory capacity. Values were 36.22 ± 10.21 mL/kg for a saline-treated lamb, 33.95 ± 8.03 mL/kg for a Curosurf[®]-treated lamb, and 32.96 ± 1.98 mL/kg for a Synsurf-treated lamb. There were no statistical intergroup differences.

Indicators of inflammation

Systemic inflammation

The cord blood had the typical lymphocytosis of fetal blood for all study groups (Table 3). After 5 hours of ventilation the numbers of white blood cells changed significantly only in the

Table 3 Cord and peripheral white blood cell (WBC) and differential count

Cell type	Synsurf (n = 6)	Curosurf [®] (n = 6)	Saline (n = 6)
Cord blood			
WBC $\times 10^9/L$	1.9 ± 1.3	1.2 ± 0.7	1.9 ± 1.3
% neutrophils	20.3 ± 15.2	24.9 ± 25.2	20.3 ± 15.2
% lymphocytes	65.9 ± 12.6	65.9 ± 21.1	65.9 ± 12.6
Count at 300 min			
WBC $\times 10^9/L$	1.8 ± 1.7	2.5 ± 2.1	1.8 ± 1.7
% neutrophils	42.8 ± 38.7	$63.5 \pm 23.1^*$	42.8 ± 38.7
% lymphocytes	52.6 ± 35.5	31.8 ± 23.7	52.6 ± 35.5

Notes: Data expressed as mean \pm standard deviation, and $*P < 0.05$ by t -test. Curosurf[®] manufactured by Chiesi Farmaceutici SpA, Parma, Italy.

Curosurf[®]-treatment group. In this group, the mean peripheral white blood cell count rose from 1.2 to 2.5 ($P = 0.0236$), with significant intragroup increase in the neutrophil count. In all of the groups, the predominant shift from a lymphocytosis in cord blood to a neutrophilic predominance occurred after 5 hours of ventilation. There were, however, no significant differences between the groups.

Lung inflammation and appearance and morphology

The macroscopic appearance of lungs of the majority of lambs was categorized as mild, moderate, or severe (widespread) atelectasis as reflected by the extent of visible dark areas (Figure 5). There were no clear visible group differences with regard to the distribution of the injury. In almost all of the groups, except for one lamb in the Curosurf[®] group, the posterior regions of the lungs, especially the lower and inferior regions of the lobes, showed atelectasis (dark areas) with or without small hemorrhages. Of the Synsurf-treated lambs, two had mild (Figure 5A) and four had moderate atelectasis (Figure 5B). Of the Curosurf[®]-treated lambs, one had a normal, uniform, expanded lung appearance (Figure 5C), three had mild (Figure 5D), and two had moderate (Figure 5E) atelectasis. The lungs of only four saline-treated animals were available; one showed mild atelectasis (Figure 5F), two had moderate (Figure 5G), and one had severe (Figure 5H) atelectasis. Representing the average size of the alveoli, measurements of the mean linear intercepts (Figure 6) of the Curosurf[®] group and the Synsurf group were higher than that of the saline group. However, this was not statistically significant (intergroup differences: Synsurf 48.68 ± 5.16 versus Curosurf[®] 49.82 ± 8.78 , $P = 0.804$; Synsurf versus saline 43.10 ± 3.79 , $P = 0.115$; and Curosurf[®] versus saline, $P = 0.192$), and therefore excluded alveolar damage as a result of differences in applied volume/pressure during ventilation. Enlargement

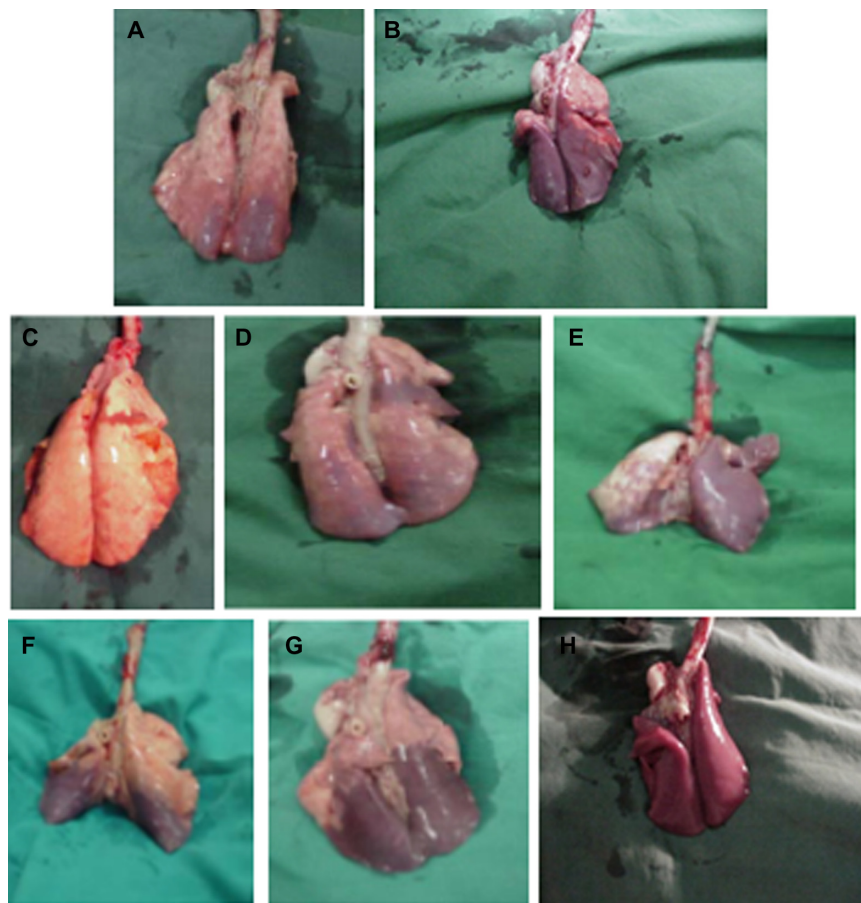


Figure 5 Representative photographs of the appearance of lungs of Synsurf- (A and B), Curosurf®- (C–E), and saline-treated lambs (F–H).

Notes: Dark color in animal lungs demonstrates injured/atelectatic areas. (A) mild in 2, (B) moderate in 4, (C) normal uniform expanded lung in 1, (D) mild in 3, (E) moderate in 2, (F) mild in 1, (G) moderate in 2, (H) severe in 1. Curosurf® manufactured by Chiesi Farmaceutici SpA, Parma, Italy.

of histomorphology microscopic images (Figure 7) in the different groups showed no accumulation of alveolar macrophages or white blood cells in the alveoli of the animals that received Curosurf® or Synsurf.

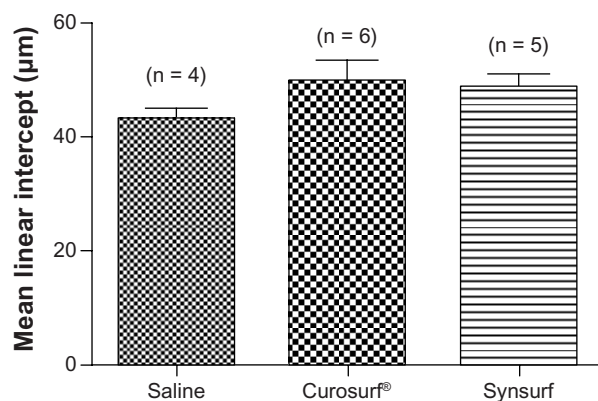


Figure 6 Mean linear intercepts of airspaces for Synsurf-, Curosurf®, and saline-treated animals.

Notes: Values are shown as mean \pm standard error of the mean. Within the groups, the mean linear intercepts values were not statistically significantly different. Curosurf® manufactured by Chiesi Farmaceutici SpA, Parma, Italy.

Calculations for postmortem lung water content, expressed as the lung wet weight/dry weight ratio (Synsurf 6.23 ± 0.65 , Curosurf® 5.67 ± 1.12 , and saline 6.49 ± 1.57), showed no significant difference between any of the groups.

Discussion

We have demonstrated that a novel synthetic peptide-containing surfactant (Synsurf), resulted in a more sustained, improved oxygenation response compared to that of Curosurf®- or saline-treated premature lambs when treated before first breath.

Commercially available surfactants for replacement therapy are still mainly mammalian-derived, and their superiority over nonprotein-containing surfactants in the treatment of neonatal RDS has repeatedly been shown.^{12,14,15} More recently, synthetic formulations containing an SP-C and/or SP-B analog have been developed and tested against mammalian-derived surfactants. Davis et al evaluated the efficacy of recombinant SP-C surfactant against natural sheep surfactant in vitro and in vivo using ventilated preterm lambs

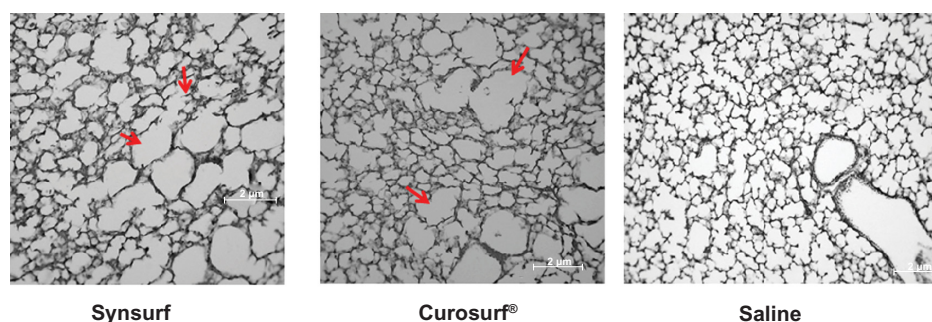


Figure 7 Lung histology findings in representative animals of each treatment group.

Notes: Staining was done with hematoxylin and eosin. Original magnification $\times 40$. Localized areas of parenchymal damage occurred in the lungs of the Curosurf®- and Synsurf-treated groups (arrows) and are surrounded by normal parenchyma. No such damage is visible in the lungs of the saline group. Scale bar: 2 μm . Curosurf® manufactured by Chiesi Farmaceutici SpA, Parma, Italy.

and rabbits.¹⁶ They found that both surfactants were sensitive to inhibition by plasma and had similar lung mechanics, lung volume measurement, and indicators of lung injury. Interestingly, a synthetic formulation (CHF5633) developed by Chiesi Farmaceutici SpA was recently compared with Survanta®, but not with Curosurf®, in extremely premature lambs (day 124).¹⁰ The authors found that the SP-B/SP-C analog-containing synthetic surfactant (200 mg/kg CHF5633) improved lung functions (tidal volumes at 2 minutes and dynamic lung compliance at 300 minutes) over that of Survanta® (100 mg/kg). Both surfactant-treated groups showed large variations in alveolar size, with patchy atelectasis in the peripheral lung and similar degrees of lung inflammation at 5 hours.

Of all the protein components in pulmonary surfactant, SP-B and SP-C have an essential function in the spreading, adsorption, and stability of surfactant lipids. Fourier transform infrared spectroscopy as well as infrared studies of porcine SP-B demonstrate that it possesses approximately 45% α -helical content, four amphipathic helices, and a β -sheet secondary structure (22%) interacting with areas of the phospholipid bilayers.^{17,18} Various research groups have chemically prepared peptides with sequences based on SP-B.^{19,20} Recently, the successful treatment of preterm infants with RDS was demonstrated with a peptide/phospholipid mixture KL₄-surfactant prepared by Cochrane et al.⁹ This peptide, that possibly mimics the pattern of hydrophobic and hydrophilic residues in SP-B, is 21 amino acids long and contains five basic lysine (hydrophilic) residues, which interact with the negative charges of phospholipid polar head groups, and 16 leucine (hydrophobic) residues that interact with the acyl side chains of the phospholipids.^{21–23} Moreover, it was designed with the intent to stabilize the phospholipid bilayer through its interaction with lipid head groups and portions of the fatty acid acyl chains. An initial study on the mechanism of interaction of KL₄ peptide with phospholipid multilayers

showed that it predominantly has α -helical content with a transmembrane orientation in lipid multilayers.²¹ However, in follow up studies it was shown that with DPPC films, the peptide adopted an antiparallel β -sheet structure.⁹

In our experiments, we reasoned that the polypeptide poly-L-lysine, which is widely used in membrane research due to its interaction with acidic lipids, was worth studying as a KL₄ alternative. To test this idea, poly-L-lysine (~100–120 residues) was mixed with poly-L-glutamic acid (~80 residues) to give a complex (~50% neutralisation) which is held together by strong electrostatic interaction (Figure 1). The rationale behind using polymers of poly-L-lysine complexed with poly-L-glutamic acid was to have a two-stranded polymer containing a percentage of basic lysyl side chains participating in the formation of protein-lipid bonds, as well as some degree of hydrophobicity in the neutralized double-stranded portion of the molecule. It should be noted that reports indicate that electrostatic binding is coupled with hydrophobic effects in protein-lipid bonding and that electrostatic interaction precedes hydrophobic interaction.^{24,25} Moreover, poly-L-lysine also becomes increasingly helical with charge cancellation above 50%.²⁶ At present, in preliminary experiments using Fourier transform infrared spectroscopy (van Zyl, unpublished data, 2012), we have found results which suggest the existence of molecular zipper-like properties of poly-L-lysine and poly-L-glutamic acid, which was first described by Dzwolac and Marszalek.²⁷ In these preliminary experiments, at neutral pD (experiments were carried out in D₂O), we also found that the two form sequenceless poly-L-lysine-poly-L-glutamic acid polymers in the random coil conformation (amide I' band) self-assemble to form an antiparallel β -sheet with a characteristic splitting of the amide I' band into a major and minor spectral component. However, a persisting amide I' band still reflected residual loose chains (random coil characteristics). At present, we suggest that these chains

of the polymer mixture will favor the exposure of the basic-charged surface groups on the lysine side chains, whereby the construct could interact flexibly with other molecules such as phosphatidyl glycerol to perform a functional role in phospholipid monolayers. Furthermore, in agreement with other reports, we also assume that part of the beneficial effect of poly-L-lysine could relate to the interaction of the charged lysine amino groups with the anionic lipid headgroups and the ability to dehydrate the lipid bilayers.^{28,29} Furthermore, from the finding that positive charges are important for maintaining the structure and function of SP-C,^{30,31} the argument can be made that the overall positive character of poly-L-lysine residues in Synsurf could contribute to the mimicking of SP-C structural and/or functional properties. This could also facilitate the action of Synsurf in our preterm lamb study, as it was shown that in the presence of phosphatidylglycerol, SP-C decreased the energy barrier limiting the adsorption of phospholipids to the air/liquid interface, possibly by introducing electrostatic interactions. As to the exact mechanism of action of the poly-L-lysine–poly-L-glutamic acid construct, further studies are in progress.

The recently developed Synsurf was also tested earlier, *in vivo* in an adult rabbit saline-lavaged model.⁸ In that study, we showed significant improvement in oxygenation and pulmonary shunt in the group of animals treated with Synsurf. The premature lamb model is an alternative test system, with a clinical and pathological course similar to that described for preterm newborns with RDS. We therefore conducted the present randomized trial to compare systemic oxygenation and lung mechanics in surfactant-deficient preterm lambs after treatment with either Synsurf (synthetic) or porcine-derived Curosurf[®] (natural) and saline as control before first breath.

We removed 10–20 mL/kg lung fluid before instilling vehicle and initiating ventilation in order to avoid ventilation on top of the fluid volume and thus injuring the lung. Furthermore, vehicle at equivalent dosages was administered as bolus before first breath in order to facilitate more uniform spreading, followed by ventilation with lower tidal volume ventilation for the first 30 minutes, again to minimize injurious ventilation.³² Despite attempts to improve distribution of surfactant in a uniform manner, inhomogeneity probably still occurred during ventilation, as between gestation days 120–130 the preterm lamb has lungs that are differentially matured, *ie*, the upper portions of the lungs are functionally more mature than the lower portions, and possibly receive better tidal volume ventilation.³³ This was demonstrated by the distribution of visible dark areas of the excised lungs

in the present study. Lambs were ventilated in the supine position, and it was evident that the dark areas which reflect atelectasis were mostly found in the posterior and posterior-inferior regions of the lungs, with no significant differences in distribution between the groups. Nevertheless, before receiving surfactant, all of the lambs had significant surfactant deficiency. Lambs that received surfactant responded with a marked increase in oxygenation and decrease in calculated shunt within 30 minutes of birth. Over the study period, replacement with Synsurf or Curosurf[®] resulted in a sustained, improved oxygenation response compared to saline-treated animals. The improved oxygenation was accompanied by a significantly improved calculated shunt over time in all three groups. However, at 300 minutes, Synsurf-treated lambs had a significantly higher PaO₂/FiO₂ ratio accompanied by a lower calculated shunt (Synsurf versus saline at 180 minutes and Synsurf versus saline- and Curosurf[®]-treated lambs at 300 minutes; Figures 2A and 3). Notably, the Synsurf-treated lambs not only had a more sustained improvement in oxygenation, they also had lower tidal volumes and associated lower PaCO₂ levels (Figures 4A and 2B), similar to what was described by Wolfson *et al* for lambs treated with lucinactant in their study.³⁴

Furthermore, although C_{dyn} increased significantly only in the surfactant-treated groups, their time profiles did not differ significantly from that of saline-treated animals. In the present study, this trend (Figure 4B versus Figure 2B) is similar to the previous study that we conducted in adult rabbits, where we observed an increase in C_{dyn} in relation to a concurrent decrease in oxygenation over time.⁸

Our findings of modestly improved C_{dyn} values over the 5-hour study period in all three study groups is of interest because the changes in C_{dyn} were not accompanied by similar improvement in oxygenation. In fact, the Curosurf[®]-treated lambs experienced the steadiest improvement in C_{dyn}, yet their timepoint-corresponding PaO₂/FiO₂ ratios were decreasing. Only in the Synsurf-treated group at timepoint 300 minutes did the improved PaO₂/FiO₂ ratio correlate significantly with their corresponding improved C_{dyn} values ($R^2 = 0.7047$; $P = 0.0366$). A novel peptide-containing synthetic surfactant, Surfaxin[®], a SP-B mimetic, was recently approved by the US Food and Drug Administration. This surfactant was evaluated against porcine- and bovine-derived surfactants in the 125–128-day preterm lamb model in regard to its potential to modulate lung inflammation following initiation of mechanical ventilation.³⁴ Surfactant treatment after commencing ventilation improved lung compliance to a similar extent compared to no surfactant treatment. In

agreement with our findings, the researchers also described “uncoupling” between sustained improved oxygenation and lung mechanics in lucinactant-treated animals. They suggested that the uncoupling is due to the presence of SP-B or SP-B-mimicking peptide, KL4, in the surfactants, which support alveolar capillary shape by reducing surface tension, thereby improving pulmonary blood flow and shunt.³⁵ Our study showed uncoupling between C_{dyn} and oxygenation and calculated intrapulmonary shunt, favoring the Synsurf-treated animals over that of Curosurf[®]- and saline-treated lambs.

Discrepancy between the time course of PaO_2 and that of C_{dyn} was also described by Häfner et al in premature lambs at 124–127 days gestation after treatment with bovine lung surfactant at 50 mg/kg body weight.³⁶ Cummings et al reported contradictory findings in preterm lambs treated with a mammalian-derived product (calf lung surfactant extract) before first breath.³⁷ They found significantly improved calculated compliance values, accompanied by improved oxygenation status in preterm lambs (127 days gestation) treated before commencing with artificial ventilation. Interestingly, in that study, the superior oxygenation status and improved calculated compliance observed between lambs treated before first breath and those treated after 5 minutes of ventilation could not be explained by differences in histopathology. The authors therefore speculated that uneven surfactant distribution in an air-filled lung as compared to a liquid-filled lung could have produced the differences.

Does exogenous surfactant instilled before initiating ventilation (prophylactic treatment) protect the immature lung against ventilation-induced lung injury? Contradictory and at times confusing evidence exists, with some research groups showing no significant injury, while others find an incomplete protective effect of natural surfactants that is linked to specific time frames after birth. For instance, Björklund et al studied immature lambs at 127 days gestation.³⁸ They found that three of four lambs treated with high dosage Curosurf[®] before delivery (235 mg/kg and 265 mg/kg) or before first breath (200 mg/kg) showed no evidence of lung injury after 4 hours of ventilation. In another study, the same research group administered Curosurf[®] (200 mg/kg) to lambs at various time intervals after birth at 127 days gestation and then subjected the lambs to various inflation treatment groups.³⁹ Although the group did not publish clear quantitative histological data, they found that although surfactant treatment before first breath did not prevent lung injury, it did protect against severe lung injury. In contrast to these findings, Ikegami et al showed that extreme styles of ventilation had

minimal effects on lung function, surfactant function, or metabolism in their study group of 131 ± 1 days preterm lambs treated with sheep surfactant before initiation of ventilation.⁴⁰ Somewhat inexplicably, in that study, lambs ventilated with low-rate (15 breaths/min) and high tidal volume ventilation (15 mL/kg) had higher PaO_2/FiO_2 ratios for lower mean airway pressures at 24 hours of age compared to a high frequency ventilation and high rate positive pressure ventilation group. The same research group studied 126- or 127-day gestation lambs and assessed tidal volume effects on surfactant treatment responses with the initiation of ventilation.⁴¹ In one of their study arms, lambs were randomized to surfactant (Survanta[®] 100 mg/kg) treatment before initiation of ventilation and then ventilated with 6 mL/kg, 12 mL/kg, or 20 mL/kg tidal volumes for 30 minutes and thereafter with 10 mL/kg tidal volumes to 6 hours of age. In contrast to their previous study findings, lambs initially ventilated with 6 mL/kg had lower dry-to-wet ratios and significantly less recovery of protein in alveolar washes compared to the 12 and 20 mL/kg tidal volume groups. The study showed that the combination of surfactant treatment at birth, followed by ventilation with tidal volumes <12 mL/kg decreases the leak of albumin into the preterm lung and is therefore less injurious.

Lung inflammation and systemic activation of inflammation follows tidal volume ventilation-induced stretch of immature lungs within 5–15 minutes after birth.^{41–43} However, when low tidal volumes (5–9 mL/kg) are targeted and maintained throughout a 5-hour protocol, minimal lung inflammation, expressed as bronchoalveolar lavage fluid inflammatory cells, or expression of proinflammatory cytokines are found.¹⁰

Jaarsma et al demonstrated a drop in polymorphonuclear leukocytes (PMNs) within 5–10 minutes of birth in ventilated 132-day premature lambs as a marker of inflammation.⁴³ Carlton et al described that circulating PMNs transiently disappeared from blood within 2 hours after birth due to sequestration in the lungs of preterm lambs (127 \pm 1 days gestation) with RDS.⁴⁴ They also found that the circulating PMN cell numbers returned to prenatal values by 6–8 hours after birth.⁴⁴ The peptide-containing synthetic surfactant lucinactant, has recently been shown to be associated with less systemic (plasma IL [interleukin]-6 and IL-8) and lung inflammation (proinflammatory markers), better lung expansion, and lower ventilator requirements than no surfactant treatment or treatment with porcine- and bovine-derived surfactants.³⁴ We found that a systemic inflammatory response

as reflected by changes in PMNs was only detectable in the Curosurf® group, but although the total white blood cell count and neutrophil count in this group increased significantly between birth and 5 hours of life, it was no different from the values found in the other two groups at 5 hours. In both the saline- and Synsurf-treated groups, no significant changes in neutrophil counts or total white blood cell count occurred. Our findings are therefore in keeping with those described by Carlton et al, showing comparative PMNs and total white blood cell count numbers between cord blood (prenatal) and 5 hours of life.⁴⁴ Furthermore, we found no relationship between changes in neutrophil counts and PaO₂/FiO₂ or PaCO₂ respectively, in order to demonstrate an influence of systemic inflammation in the lung on gas exchange and a relationship with respiratory compromise. Moreover, there was no accumulation of alveolar macrophages or white blood cells in the alveoli of the animals that received Curosurf® or Synsurf (histology of saline-instilled lungs were similar to that of surfactant-treated lungs). Taken together with the findings of a relative lack of lung inflammation, we speculate that surfactant administration before first breath did not exert a clear anti-inflammatory protective effect, but possibly the initiation of ventilation utilizing a low tidal volume may have played a role.

Why did the Synsurf-treated animals have a more sustained oxygenation response and significantly better PaO₂/FiO₂ ratio at 300 minutes? Explanations may include earlier studies that have shown that clinical responses after surfactant treatment in experimental animals is unpredictable when based on in vitro (bench) properties, and surfactants behave differently with respect to loss from the lungs and sensitivity to soluble airway proteins.^{45,46} In other words, the surfactant with the best in vitro characteristics may not necessarily perform better than others in the preterm lamb model.

Conclusion

To conclude, treatment with surfactants before first breath clearly resulted in improved systemic oxygenation, with the response of Synsurf-treated animals being more sustained at 300 minutes. The functional importance of the synthetic peptide complex in Synsurf was therefore apparent from the results obtained with a control group and the group of lambs treated with an equivalent dosage of Curosurf®.

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Disclosure

The authors report no conflicts of interest in this work. The surfactant has been patented by InnovUS (Stellenbosch University).

References

1. Pérez-Gil J. Structure of pulmonary surfactant membranes and films: the role of proteins and lipid-protein interactions. *Biochim Biophys Acta*. 2008;1778(7–8):1676–1695.
2. Whitsett JA, Weaver TE. Hydrophobic surfactant proteins in lung function and disease. *N Engl J Med*. 2002;347(26):2141–2148.
3. Hawgood S. Surfactant protein B: structure and function. *Biol Neonate*. 2004;85(4):285–289.
4. Moya FR, Gadzinowski J, Bancalari E, et al; International Surfaxin Collaborative Study Group. A multicenter, randomized, masked, comparison trial of lucinactant, colfosceril palmitate, and beractant for the prevention of respiratory distress syndrome among very preterm infants. *Pediatrics*. 2005;115(4):1018–1029.
5. Wang Z, Gurel O, Baatz JE, Notter RH. Differential activity and lack of synergy of lung surfactant proteins SP-B and SP-C in interactions with phospholipids. *J Lipid Res*. 1996;37(8):1749–1760.
6. Seeger W, Günther A, Thede C. Differential sensitivity to fibrinogen inhibition of SP-C- vs SP-B-based surfactants. *Am J Physiol*. 1992; 262(3 Pt 1):L286–L291.
7. Ramanathan R, Rasmussen MR, Gerstmann DR, Finer N, Sekar K; North American Study Group. A randomized, multicenter masked comparison trial of poractant alfa (Curosurf) versus beractant (Survanta) in the treatment of respiratory distress syndrome in preterm infants. *Am J Perinatol*. 2004;21(3):109–119.
8. van Zyl JM, Smith J, Hawtrey A. The effect of a peptide-containing synthetic lung surfactant on gas exchange and lung mechanics in a rabbit model of surfactant depletion. *Drug Des Devel Ther*. 2013;7: 139–148.
9. Cochrane CG, Revak SD, Merritt TA, et al. The efficacy and safety of KL4-surfactant in preterm infants with respiratory distress syndrome. *Am J Respir Crit Care Med*. 1996;153(1):404–410.
10. Sato A, Ikegami M. SP-B and SP-C containing new synthetic surfactant for treatment of extremely immature lamb lung. *PLoS ONE*. 2012;7(7):e39392.
11. Moya F, Maturana A. Animal-derived surfactants versus past and current synthetic surfactants: current status. *Clin Perinatol*. 2007;34(1): 145–177, viii.
12. Pfister RH, Soll RF, Wiswell T. Protein containing synthetic surfactant versus animal derived surfactant extract for the prevention and treatment of respiratory distress syndrome. *Cochrane Database Syst Rev*. 2007;(4):CD006069.
13. Dunnill MS. Quantitative methods in the study of pulmonary pathology. *Thorax*. 1962;17(4):320–328.
14. Halliday HL. Surfactants: past, present and future. *J Perinatol*. 2008; 28(Suppl 1):S47–S56.
15. Suresh GK, Soll RF. Lung surfactants for neonatal respiratory distress syndrome: animal-derived or synthetic agents? *Pediatric Drugs*. 2002;4(8):485–492.
16. Davis AJ, Jobe AH, Häfner D, Ikegami M. Lung function in premature lambs and rabbits treated with a recombinant SP-C surfactant. *Am J Respir Crit Care Med*. 1998;157(2):553–559.
17. Vandenbussche G, Clercx A, Clercx M, et al. Secondary structure and orientation of the surfactant protein SP-B in a lipid environment. A Fourier transform infrared spectroscopy study. *Biochemistry*. 1992;31(38):9169–9176.

18. Andersson M, Curstedt T, Jörnvall H, Johansson J. An amphipathic helical motif common to tumourolytic polypeptide NK-lysin and pulmonary surfactant polypeptide SP-B. *FEBS Lett.* 1995;362(3):328–332.
19. Waring A, Taesch W, Bruni R, et al. Synthetic amphipathic sequences of surfactant protein-B mimic several physicochemical and in vivo properties of native pulmonary surfactant proteins. *Pept Res.* 1989;2(5):308–313.
20. Bruni R, Taesch HW, Waring AJ. Surfactant protein B: lipid interactions of synthetic peptides representing the amino-terminal amphipathic domain. *Proc Natl Acad Sci U S A.* 1991;88(16):7451–7455.
21. Gustafsson M, Vandenbussche G, Curstedt T, Ruyschaert JM, Johansson J. The 21-residue surfactant peptide (LysLeu4)4 Lys(KL4) is a transmembrane alpha-helix with a mixed nonpolar/polar surface. *FEBS Lett.* 1996;384(2):185–188.
22. Cochrane CG, Revak SD. Pulmonary surfactant protein B (SP-B): structure-function relationships. *Science.* 1991;254(5031):566–568.
23. Cochrane CG, Revak SD. Protein-phospholipid interactions in pulmonary surfactant. The Parker B. Francis Lectureship. *Chest.* 1994;105(Suppl 3):57S–62S.
24. Cai P, Flach CR, Mendelsohn R. An infrared reflection-absorption spectroscopy study of the secondary structure in (KL4)4 K, a therapeutic agent for respiratory distress syndrome, in aqueous monolayers with phospholipids. *Biochemistry.* 2003;42(31):9446–9452.
25. Hammes GG, Schullery SE. Structure of macromolecular aggregates. II. Construction of model membranes from phospholipids and polypeptides. *Biochemistry.* 1970;9(13):2555–2563.
26. Kimelberg HK, Papahadjopoulos D. Interactions of basic proteins with phospholipid membranes. Binding and changes in the sodium permeability of phosphatidylserine vesicles. *J Biol Chem.* 1971;246(4):1142–1148.
27. Dzwolak W, Marszalek PE. Zipper-like properties of [poly(L-lysine) + poly(L-glutamic acid)] β -pleated molecular self-assembly. *Chem Commun (Camb).* 2005;28(44):5557–5559.
28. Ciferri A, Puett D, Rajagh L, Hermans J. Potentiometric titrations and the helix-coil transition of poly(L-glutamic acid) and poly-L-lysine in aqueous salt solutions. *Biopolymers.* 1968;6(8):119–136.
29. Carrier D, Dufourcq J, Faucon J-F, Pérolet M. A fluorescence investigation of the effects of polylysine on dipalmitoylphosphatidylglycerol bilayers. *Biochim Biophys Acta.* 1985;820(1):131–139.
30. Carrier D, Pérolet M. Investigation of polylysine-dipalmitoylphosphatidylglycerol interactions in model membranes. *Biochemistry.* 1986;25(14):4167–4174.
31. Creuwels LA, Boer EH, Demel RA, van Golde LM, Haagsman HP. Neutralization of the positive charges of surfactant protein C. Effects on structure and function. *J Biol Chem.* 1995;270(27):16225–16229.
32. Ueda T, Ikegami M, Rider ED, Jobe AH. Distribution of surfactant and ventilation in surfactant-treated preterm lambs. *J Appl Physiol.* 1994;76(1):45–55.
33. Brumley GW, Chernick V, Hodson WA, Normand C, Fenner A, Avery ME. Correlations of mechanical stability, morphology, pulmonary surfactant, and phospholipid content in the developing lamb lung. *J Clin Invest.* 1967;46(5):863–873.
34. Wolfson MR, Wu J, Hubert TL, Gregory TJ, Mazela J, Shaffer TH. Lucinactant attenuates pulmonary inflammatory response, preserves lung structure, and improves physiologic outcomes in a preterm lamb model of RDS. *Pediatr Res.* 2012;72(4):375–383.
35. Ikegami M, Weaver TE, Grant SN, Whitsett JA. Pulmonary surfactant surface tension influences alveolar capillary shape and oxygenation. *Am J Respir Cell Mol Biol.* 2009;41(4):433–439.
36. Häfner D, Hanauer G, Kilian U, Beume R, Dillmann G. Randomized placebo-controlled study on the characteristics and duration of action of surfactant treatment in premature lambs. *Pulm Pharmacol.* 1993;6(4):255–262.
37. Cummings JJ, Holm BA, Nickerson PA, Ferguson WH, Egan EA. Pre- versus post-ventilatory surfactant treatment in surfactant-deficient preterm lambs. *Reprod Fertil Dev.* 1995;7(5):1333–1338.
38. Björklund LJ, Ingimarsson J, Curstedt T, Larsson A, Robertson B, Werner O. Lung recruitment at birth does not improve lung function in immature lambs receiving surfactant. *Acta Anaesthesiol Scand.* 2001;45(8):986–993.
39. Ingimarsson J, Björklund LJ, Curstedt T, et al. Incomplete protection by prophylactic surfactant against the adverse effects of large lung inflations at birth in immature lambs. *Intensive Care Med.* 2004;30(7):1446–1453.
40. Ikegami M, Wada K, Emerson GA, Rebello CM, Hernandez RE, Jobe AH. Effects of ventilation style on surfactant metabolism and treatment response in preterm lambs. *Am J Respir Crit Care Med.* 1998;157(2):638–644.
41. Wada K, Jobe AH, Ikegami M. Tidal volume effects on surfactant treatment responses with the initiation of ventilation in preterm lambs. *J Appl Physiol.* 1997;83(4):1054–1061.
42. Jaarsma AS, Braaksma MA, Geven WB, Van Oeveren W, Oetomo SB. Early activation of inflammation and clotting in the preterm lamb with neonatal RDS: comparison of conventional ventilation and high frequency oscillatory ventilation. *Pediatr Res.* 2001;50(5):650–657.
43. Jaarsma AS, Braaksma MA, Geven WB, van Oeveren W, Bambang Oetomo S. Activation of the inflammatory reaction within minutes after birth in ventilated preterm lambs with neonatal respiratory distress syndrome. *Biol Neonate.* 2004;86(1):1–5.
44. Carlton DP, Albertine KH, Cho SC, Lont M, Bland RD. Role of neutrophils in lung vascular injury and edema after premature birth in lambs. *J Appl Physiol.* 1997;83(4):1307–1317.
45. Bernhard W, Mottaghian J, Gebert A, Rau GA, von Der HARDT H, Poets CF. Commercial versus native surfactants. Surface activity, molecular components, and the effect of calcium. *Am J Respir Crit Care Med.* 2000;162(4 Pt 1):1524–1533.
46. Ikegami M, Agata Y, Elkady T, Hallman M, Berry D, Jobe A. Comparison of four surfactants: in vitro surface properties and responses of preterm lambs to treatment at birth. *Pediatrics.* 1987;79(1):38–46.

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