

# Supporting Information

## Identification of inosine monophosphate dehydrogenase as a potential target for anti-mpox virus agents

Takayuki Hishiki<sup>1†</sup>, Takeshi Morita<sup>1†</sup>, Daisuke Akazawa<sup>1¶</sup>, Hirofumi Ohashi<sup>1¶</sup>, Eun-Sil Park<sup>2</sup>, Michiyo Kataoka<sup>3</sup>, Junki Mifune<sup>1</sup>, Kaho Shionoya<sup>4,5</sup>, Kana Tsuchimoto<sup>1</sup>, Shinjiro Ojima<sup>1</sup>, Aa Haeruman Azam<sup>1</sup>, Shogo Nakajima<sup>4</sup>, Madoka Kawahara<sup>1,6</sup>, Tomoki Yoshikawa<sup>6</sup>, Masayuki Shimojima<sup>6</sup>, Kotaro Kiga<sup>1</sup>, Ken Maeda<sup>2</sup>, Tadaki Suzuki<sup>3</sup>, Hideki Ebihara<sup>6</sup>, Yoshimasa Takahashi<sup>1</sup>, Koichi Watashi<sup>1,4,5,7#</sup>

<sup>1</sup>Research Center for Drug and Vaccine Development, National Institute of Infectious Diseases, Tokyo 162-8640, Japan, <sup>2</sup>Department of Veterinary Science, National Institute of Infectious Diseases, Tokyo 162-8640, Japan, <sup>3</sup>Department of Pathology, National Institute of Infectious Diseases, Tokyo 162-8640, Japan, <sup>4</sup>Department of Virology II, National Institute of Infectious Diseases, Tokyo 162-8640, Japan, <sup>5</sup>Department of Applied Biological Science, Tokyo University of Science, Noda 278-8510, Japan, <sup>6</sup>Department of Virology I, National Institute of Infectious Diseases, Tokyo 162-8640, Japan, <sup>7</sup>MIRAI, Japan Science and Technology Agency (JST), Saitama 332-0012, Japan

<sup>†¶</sup>These authors contributed equally to this work.

<sup>#</sup>Corresponding author: Koichi Watashi, Ph.D.

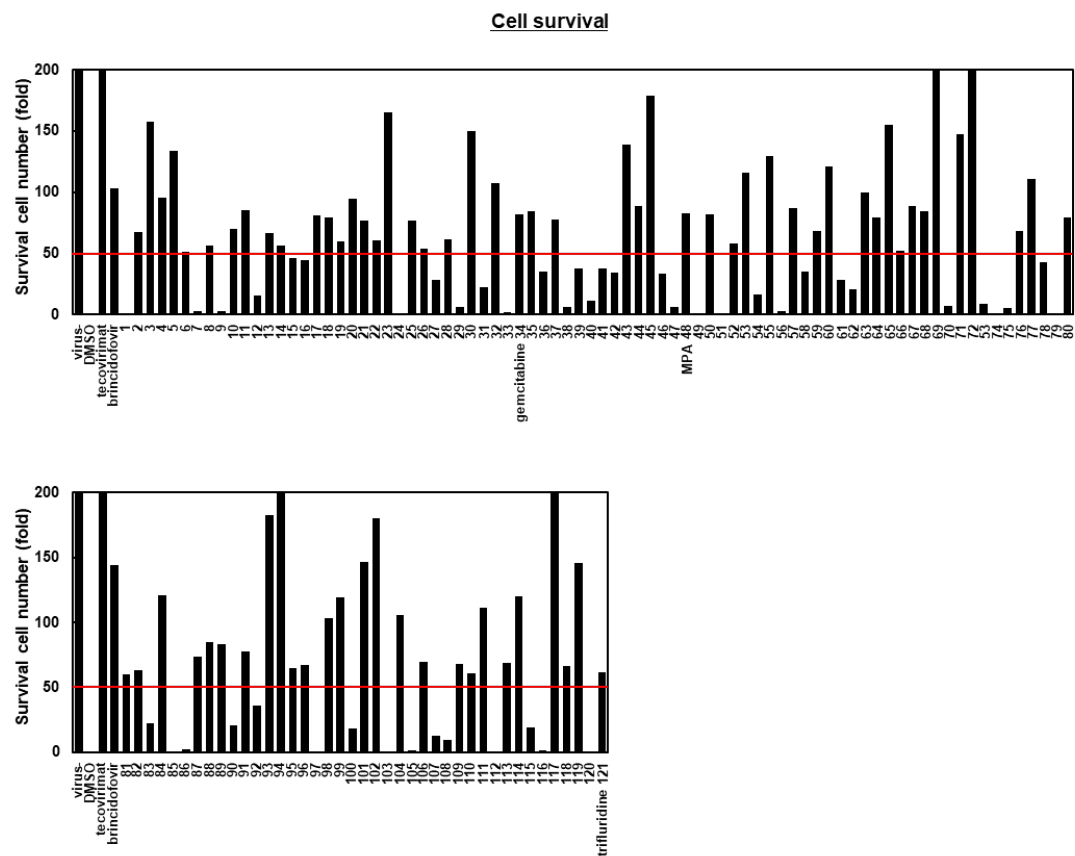
Research Center for Drug and Vaccine Development, National Institute of Infectious Diseases, 1-23-1 Toyama, Shinjuku-ku, Tokyo 162-8640, Japan  
Phone: +81-3-5285-1111; E-mail: kwatashi@niid.go.jp

### Table of contents

**Fig. S1, S2, S3, S4, and S5**

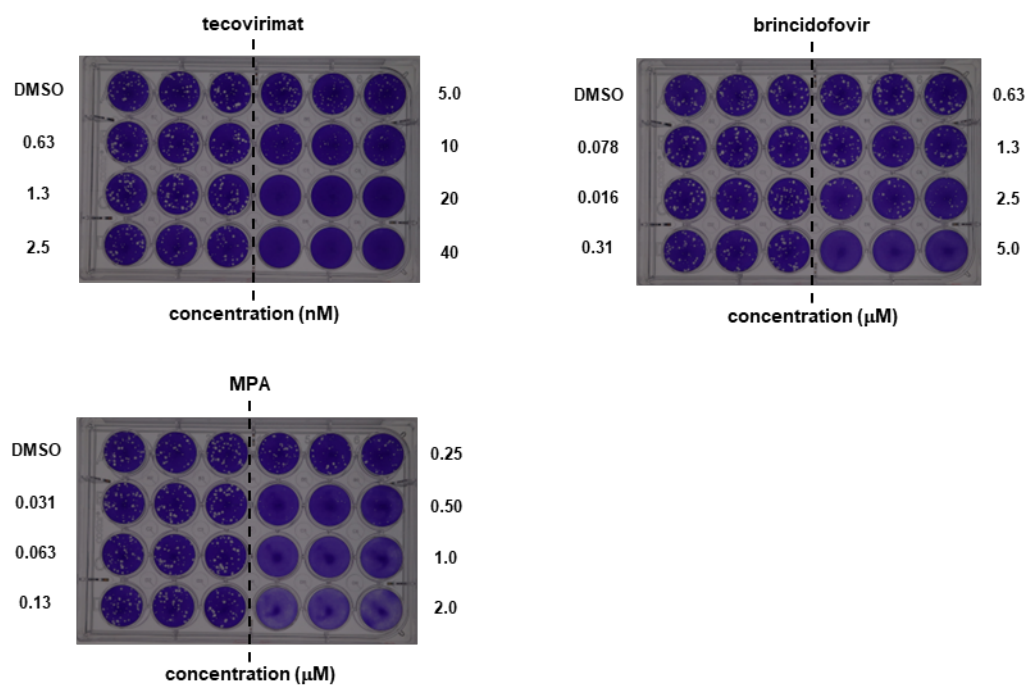
**Table S1 and S2**

Fig. S1



**Fig. S1 Primary screening of the compound library in MPXV-infected VeroE6 cells.** VeroE6 cells were infected with or without MPXV at an MOI of 0.1 and treated with compounds at 10  $\mu$ M (2 compounds treated at 2  $\mu$ M are shown in Table S1) or 0.1% DMSO. Tecovirimat and brincidofovir were used as positive controls. After 72 h of infection, the number of surviving cells was quantified by DAPI staining using a high-content imaging analyzer. The red line shows a 50-fold higher cell survival rate relative to that of the DMSO control in MPXV-infected cells. The numbers (1 to 121) in x-axis corresponds to the compounds listed in Table S1.

**Fig. S2**

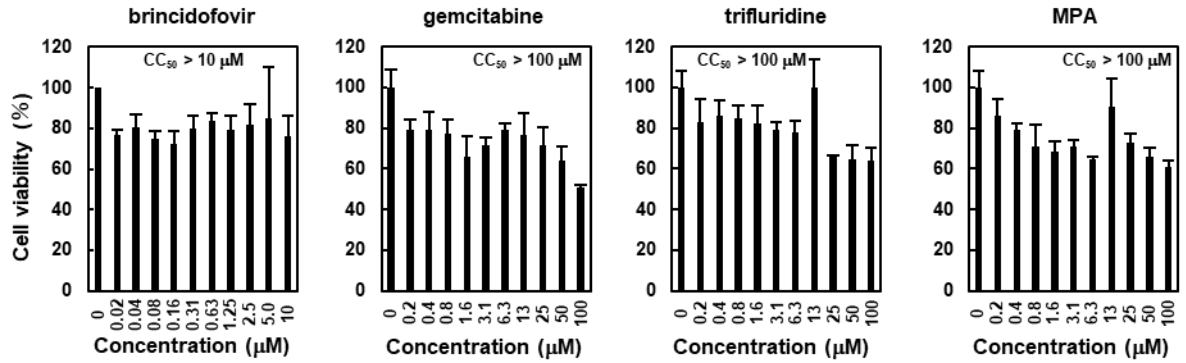


**Fig. S2 Evaluation of antiviral activity by plaque reduction assay**

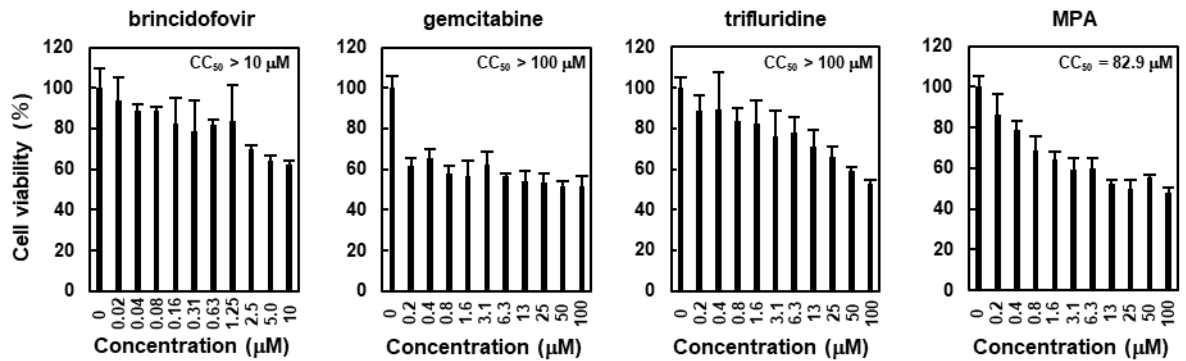
VeroE6 cells were seeded at  $1 \times 10^5$  cells/well in a 24-well plate. Cells were infected with MPXV together with the indicated concentrations of compounds. After 72 h of infection, cells were fixed and stained with crystal violet to count the plaque number.

Fig. S3

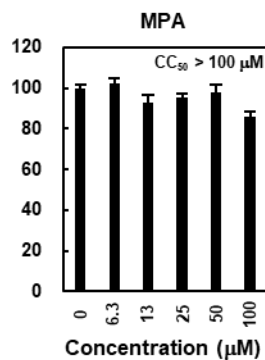
**(A) Low cell number condition (VeroE6)**



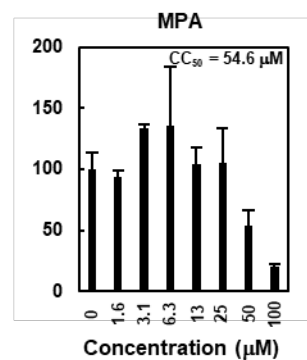
**(B) Low cell number condition (Huh7)**



**(C) Primary human cells (PHH)**



**(D) Primary human cells (PBMC)**

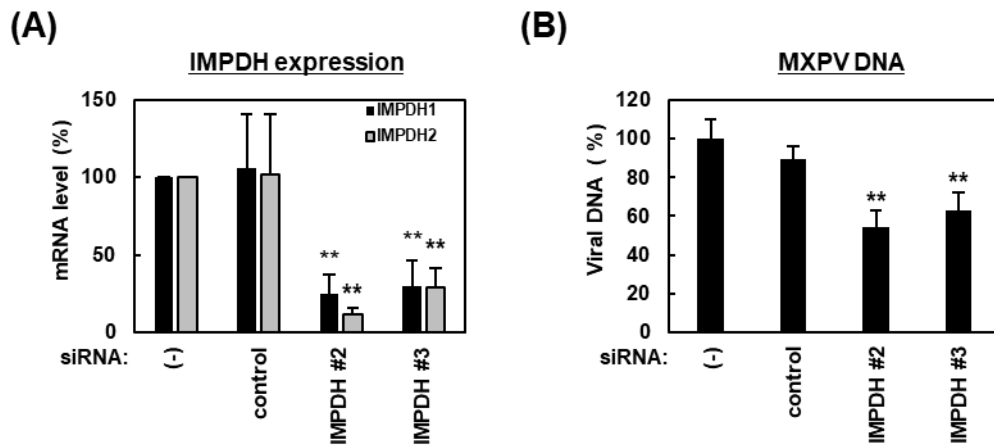


**Fig. S3 Cytotoxicity assay in various cells**

(A) VeroE6 and (B) Huh7 cells were seeded at  $5 \times 10^3$  cells/well in a 96-well plate as low cell number condition (normally,  $2 \times 10^4$  cells/well). (C) Primary human hepatocytes (PHH) were seeded at  $7 \times 10^4$  cells/well and (D) primary human peripheral blood mononuclear cells (PBMC) were seeded at  $1 \times 10^5$  cells/well in a 96-well plate,

respectively. These cells were incubated with the indicated concentrations of compound for 72 h were subjected to the detection of cell viability. The y-axis shows values relative to that of the DMSO-treated cells as a control.

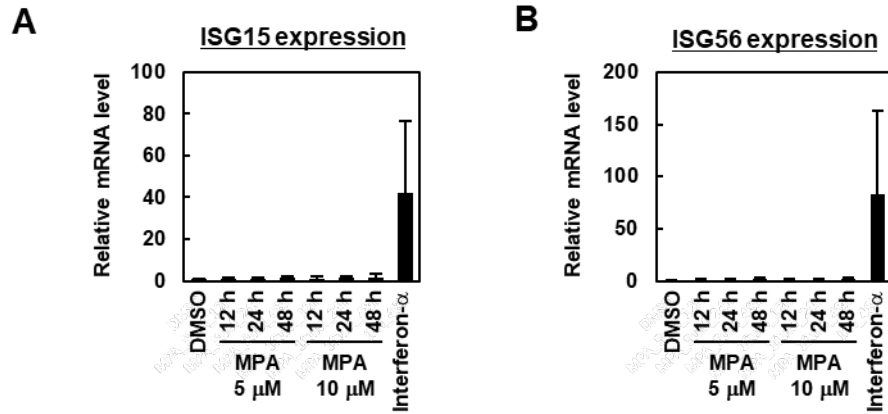
**Fig. S4**



**Fig. S4 Suppression of IMPDH expression reduces viral DNA**

(A) Huh7 cells were transfected with or without [(-)] siRNA targeting IMPDH (IMPDH) or randomized control siRNA (control). At 48 h post-transfection, intracellular RNA for IMPDH1, IMPDH2, and actin were detected by real-time RT-PCR. The y-axis shows the value relative to that for the untransfected cells. (B) Intracellular MPXV DNA levels at 72 h post-transfection with siRNA were quantified by real-time PCR and are shown as the percentage relative to that of the untransfected cells. Statistical significance is shown.

**Fig. S5**



**Fig. S5 Expression of interferon-stimulated genes (ISGs) upon MPA treatment.**

Huh7 cells were incubated with MPA (5 or 10  $\mu$ M), interferon- $\alpha$  (1000 U/ml), or DMSO (0.1%). After incubation with MPA for 12, 24, and 48 h or with interferon- $\alpha$  or DMSO for 48 h, intracellular RNA was extracted and mRNA expression levels of ISG15 (A) and ISG56 (B) were measured by real-time RT-PCR. The Y-axis shows the value relative to that of DMSO-treated cells as a control.

**Table S1 List of drugs in the library**

1	ABT-737
2	Linifanib
3	Dovitinib
4	Dasatinib
5	Gefitinib
6	Luminespib
7	MLN8054
8	Cabozantinib
9	Mocetinostat
10	BMS-754807
11	Tanespimycin
12	Delanzomib
13	Ganetespib
14	Onalespib
15	ABT-751
16	BIIB021
17	WZ8040
18	ENMD-2076
19	Cladribine
20	Methotrexate
21	Clofarabine
22	YM201636
23	OSI-930
24	Etoposide
25	KU-0063794
26	Vincristine sulfate
27	BX-912
28	Floxuridine
29	Genistein
30	SP600125
31	HMN-214
32	Fludarabine
33	Selisistat
34	Gemcitabine



35	Adefovir Dipivoxil
36	Azacitidine
37	Cyclocytidine HCl
38	Atorvastatin Calcium
39	Gandotinib
40	Ixazomib
41	Ixazomib Citrate
42	Avasimibe
43	OSI-420
44	UK 383367
45	Apigenin
46	Phloretin
47	Tolbutamide
48	Mycophenolic acid
49	MG-132
50	OSI-027
51	URB597
52	PF-04929113
53	WYE-125132
54	ICG-001
55	Ibrutinib
56	KW-2478
57	Mardepodect
58	KX2-391
59	AMG-900
60	MK-2461
61	Nocodazole
62	RITA
63	Vistusertib
64	Lonafarnib
65	AZD4547
66	TAE226
67	TPCA-1
68	StemRegenin 1
69	Golvatinib

70	ML130
71	WHI-P154
72	CCG 50014
73	Niclosamide
74	Anagrelide HCl
75	Fexofenadine HCl
76	Cabozantinib malate
77	Nifuroxazide
78	PD168393
79	Oprozomib
80	PP1
81	XL888
82	SC144
83	KPT-185
84	SKI II
85	Skepinone-L
86	KPT-276
87	CNX-774
88	NMS-E973
89	Rociletinib
90	TG003
91	PTC-209
92	Sorafenib
93	CGP 57380
94	AR-A014418
95	VER-49009
96	Triapine
97	Afatinib Dimaleate
98	Tenovin-1
99	Tyrphostin AG 1296
100	Butein
101	Ivacaftor
102	Vidarabine
103	Teniposide
104	Cyclosporin A

105	Kaempferol
106	PAC-1
107	Azaguanine-8
108	Bergapten
109	NSC 319726
110	PD153035
111	Miconazole Nitrate
112	Navitoclax
113	Cytarabine
114	Erlotinib HCl (2 $\mu$ M)
115	Torin 1 (2 $\mu$ M)
116	Drospirenone
117	Idoxuridine
118	Ciclopirox
119	Econazole nitrate
120	Norethindrone acetate
121	Trifluridine

Compounds were treated at 10  $\mu$ M, with the exception of 114 and 115, which were treated at 2  $\mu$ M.

**Table S2 Comparison of antiviral activity determined by plaque assay and that by viral DNA quantification**

Compound	IC <sub>50</sub> of plaque	IC <sub>50</sub> of viral DNA	Significance
tecovirimat (nM)	8.97 $\pm$ 1.56	4.60 $\pm$ 1.41	N.S.
brincidofovir ( $\mu$ M)	2.58 $\pm$ 0.13	2.10 $\pm$ 0.69	N.S.
MPA ( $\mu$ M)	0.34 $\pm$ 0.09	0.26 $\pm$ 0.02	N.S.

**Table S2**

In comparison of IC<sub>50</sub> values between viral DNA levels and plaque reduction, the statistical analysis was performed using GraphPad Prism 9 software, and significance was determined using Mann-Whitney U test. N.S.; not significant.