Comparative analysis using K-mer and K-flank patterns provides evidence for CpG island sequence evolution in mammalian genomes

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ABSTRACT

CpG islands are GC-rich regions often located in the 5' end of genes and normally protected from cytosine methylation in mammals. The important role of CpG islands in gene transcription strongly suggests evolutionary conservation in the mammalian genome. However, as CpG dinucleotides are over-represented in CpG islands, comparative CpG island analysis using conventional sequence analysis techniques remains a major challenge in the epigenetics field. In this study, we conducted a comparative analysis of all CpG island sequences in 10 mammalian genomes. As sequence similarity methods and character composition techniques such as information theory are particularly difficult to conduct, we used exact patterns in CpG island sequences and single character discrepancies to identify differences in CpG island sequences. First, by calculating genome distance based on rank correlation tests, we show that k-mer and k-flank patterns around CpG sites can be used to correctly reconstruct the phylogeny of 10 mammalian genomes. Further, we used various machine learning algorithms to demonstrate that CpG islands sequences can be characterized using k-mers. In addition, by testing a human model on the nine different mammalian genomes, we provide the first evidence that k-mer signatures are consistent with evolutionary history.

INTRODUCTION

The dinucleotide sequence CpG (cytosine followed by a guanine, coupled by a phosphodiester bond) is a target for DNA methylation. The cytosine residue in CpG sites is frequently modified to form 5-methylcytosine, and 70-80% of CpG dinucleotides in the mammalian genomes are methylated (1). DNA methylation is essential for proper mammalian development and plays crucial roles in imprinting, maintaining genomic stability and many other biological processes (2). In addition, aberrant DNA methylation changes have been detected in several diseases (3-5). In the human genome, spontaneous deamination of methylated cytosine in the context of CpG dinucleotides results in the creation of thymine $(C \rightarrow T)$ and under-representation of CG dinucleotides over evolutionary time (known as CG suppression) (6, 7). In fact, the frequency of CpG sites in vertebrate genomes is only about a fifth of the expected frequency, given the GC content of the genome. Although CpG sites are under-represented in genomes overall, clusters of CpGs known as CpG islands are observed, and these are normally protected from methylation (8). The vast majority of genes are associated with a CpG islands, and $\sim 40\%$ of gene promoters contain a CpG island (9), including the 5' ends of housekeeping genes and many tissue-specific genes in vertebrates (10). Recently, human promoters were classified as high CpG content (about 70%) versus 30% low CpG promoters (CpG content characteristic of the overall genome) (11). Comparative studies (12) on CpG island promoter organization, in terms of protein-DNA interactions and patterns of expression, recently reported a strong link between CpG islands and

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evolution and that accumulation of CpG islands at transcription start sites (TSS) is a vertebrate-specific genomic feature. Those authors (12) suggested that CpG islands at TSS are a consequence of warm-blooded vertebrate evolution, presumably for efficient regulation of transcription in large genomes. On the other hand, CpG islands could have played a direct role in evolution of warm-blooded vertebrates, perhaps contributing to the gain of placenta, a hallmark of eutherian mammals (13). In support of the latter, a relationship between evolution of CpG island promoter function and gene expression in mammalian heart was recently reported (14).

To date, objective definitions of CpG islands are limited. Gardiner-Garden and Frommer (15) described CpG islands as a region with at least 200 bp, a GC content >50% and an observed/expected CpG ratio >60%. Takai and Jones (16) revised the definition of CpG islands as DNA regions with at least 500 bp, a GC content >55% and an observed/expected CpG ratio >65% were more likely to be true CpG islands associated with the 5' end regions of genes. They also enhanced the ability to detect CpG islands by excluding other GC-rich genomic sequences such as Alu repeats (17, 18). Despite significant efforts to define CpG islands, it remains a challenge to perform computational CpG island analysis using conventional sequence analysis methods.

To overcome this barrier, we propose novel oligomercounting approaches for the comparative analysis of all CpG island sequences in 10 mammalian genomes. These two new approaches use exact sequence patterns in CpG island sequences called k-mer and k-flank. After counting the k-mers and k-flanks, pattern counting was used to reconstruct 10 mammalian phylogenies and for machine learning analysis. We demonstrate that k-mers are characteristic of CpG island sequences and also show that k-mer data are consistent with evolutionary history of 10 mammalian genomes. To our knowledge, this extensive study represents the first comparative analysis of CpG island sequences in mammalian genomes.

MATERIALS AND METHODS

Raw data

From the UCSC Genome Browser, CpG island sequences in 10 mammalian were downloaded and used for the analysis. Table 1 shows 10 mammalian reference genomes and their versions from which the CpG island sequences are taken.

BLAST and information theoretic approaches are not effective for CpG island sequence analysis

Despite previous studies on CpG island sequences, it is unclear why CpG island regions are found more frequently in mammalian genomes compared with other genomes. In addition, while studies exist on CpG island sequences and evolution (19, 20) in primates, comparative mammalian studies are lacking, perhaps owing to the difficulty of performing computational analysis of CpG island sequences containing over-represented CpG dinucleotides. In this regard, conventional sequence analysis techniques are not effective for the comparative analysis of highly similar CpG island sequences (shown in the next section).

We used BLAST for CpG island sequence analysis. Owing to over-represented CpG dinucleotides, CpG island sequences are very similar to each other, and sequence similarity-based methods like BLAST are not effective for performing comparative CpG island sequences analysis. We took another traditional approach and computed relative entropy between 10 mammalian species. As shown in Table 2, computing the relative entropy was ineffective in showing significant differences among CpG island sequences. Shown below is the computational procedure we performed for the relative entropy between species.

- (i) Let P and Q be probability distributions of CpG island for chimp and human
- (ii) Get probabilities of A, G, T, C for chimp and human in each CpG island sequence. Let them be p(A), p(G), p(T), p(C) and q(A), q(G), q(T), q(C)
- (iii) Relative entropy is calculated by
- (iv) $\sum_{x \in \{A,T,G,C\}} p(x) \ln(p(x)/q(x))$
- (v) Repeat (i) to (iii) for all species pairwise

The k-mer and k-flank approaches

As shown in the previous section, over-represented CpG dinucleotides make it difficult to perform analysis of CpG islands. To overcome this hurdle, we used two new oligomer-counting approaches, exact sequence patterns called k-mer and k-flank, for the comparative analysis of all CpG island sequences in 10 mammalian genomes. The first model, k-mer, gives a general descriptor of the oligomer landscape in the entire CpG island sequence. Given a sequence S, k-mers are sub-sequences of S of length k, also known as oligomers for small k. For each CpG island sequence, sliding windows of length k are moved across the CpG island sequence from the 5' end to 3' end, and each k-mer occurrence is counted. Determining the number of occurrences of specific k-mers in a sequence is called k-mer counting or oligomer counting, and can provide descriptive information about the DNA sequence (6, 21). To better characterize and describe CpG island sequences, we used k-mer counting techniques and frequency measurements to perform a comparative analysis of the CpG islands. The second oligomer-counting approach, called k-flanks, records the DNA sequence of k length directly upstream and downstream of each CpG site in a CpG island sequence. This oligomer model is stricter and specifically describes the DNA bases directly adjacent to CpG sites. Figure 1 illustrates the definition of k-mer and k-flank. In this study, we counted 3-10 k-mers and k-flanks. Once k-mers and k-flanks were counted, we used pattern counting for the reconstruction of 10 mammalian phylogenies and also for machine learning analysis to show that (i) k-mers are characteristic of CpG island sequences and (ii) k-mer data are consistent with evolutionary history of 10 mammalian genomes. As far as we know,

this study is the first extensive comparative analysis of CpG island sequences.

Collecting k-mer/k-flank frequencies and common k-mers/k-flanks

- (i) Count k-mer and k-flank frequencies in CpG island for each species
- (ii) Collect k-mers and k-flanks common in all 10 mammalian genomes and their frequencies

Once the common k-mers/k-flanks are collected, they are sorted according to their frequencies. Based on the ranks of common k-mers/k-flanks, their ranks are marked. Since these k-mers/k-flanks are in common across all 10 mammalian genomes, the order of k-mers/ k-flanks in each species becomes a permutation of ranks of common k-mers/k-flanks. This k-mer/k-flank ranking method based on the common k-mer/k-flank ranks

Table 1. Ten mammalian CpG islands sequence

Species	Data version
Chimp	CGSC 2.1.3/panTro3
Cow	Bos taurus UMD 3.1/bosTAu6
Dog	Broad/canFam2
Human	CRCh37/hg19
Marmoset	WUGSC 3.2/calJac3
Mouse	NCBI37/mm9
Opossum	Broad/monDom5
Pig	SGSC Sscrofa9.2/susScr2
Rat	Baylor 3.4/rn4
Rhesus	MGSC Merged 1.0/rheMac2

comes from an assumption that all these mammalian species are closely related from the evolutionary perspective. Figure 2 illustrates the experimental protocol.

Correlation between species

k-mer/k-flank selection for correlation

Based on the common k-mers/k-flanks, k-mers/k-flanks in the each species were selected for correlation analysis between species. That is, for the correlation coefficient computation, each species has the same set of k-mers/ k-flanks but a different rank order 2.

Kendall tau rank correlation coefficient with merge sort

Kendalls tau is a method to measure rank correlation, first discussed by G.T. Fechner in 1900 and rediscovered by M.G. Kendall in 1938 (23, 24). It is a statistic used to measure the association between two measured quantities, effectively measuring rank correlation.

RESULTS AND DISCUSSION

Reconstruction of the phylogenetic tree based on the distance matrix

Distance matrix can be directly obtained from the correlation coefficient value computed by Kendalls tau method. If the correlation coefficient value is τ , the distance matrix will be $(1 - \tau)$.

Neighbour-joining algorithm

We applied a neighbour-joining algorithm (22) to reconstruct phylogenetic trees using matrix of pairwise

Table 2. Relative entropy between species

Species	Chimp	Cow	Dog	Human	Marmoset	Mouse	Opossum	Pig	Rat	Rhesus
Chimp	0.00E + 00	6.52E-05	1.24E-05	2.61E-05	3.38E-04	2.61E-05	8.42E-04	7.18E-04	4.15E-04	2.20E-06
Cow	6.51E-05	0.00E + 00	1.34E-04	1.71E-04	1.07E-04	1.31E-05	4.39E-04	3.54E-04	1.56E-04	4.59E-05
Dog	1.24E-05	1.34E-04	0.00E + 00	4.47E-06	4.80E-04	7.28E-05	1.06E-03	9.19E-04	5.69E-04	2.41E-05
Human	2.62E-05	1.72E-04	4.48E-06	0.00E + 00	5.48E-04	1.00E-04	1.15E-03	1.00E-03	6.43E-04	4.12E-05
Marmoset	3.37E-04	1.07E-04	4.77E-04	5.44E-04	0.00E + 00	1.80E-04	1.12E-04	7.23E-05	6.83E-06	2.88E-04
Mouse	2.61E-05	1.31E-05	7.26E-05	1.00E-04	1.81E-04	0.00E + 00	5.79E-04	4.75E-04	2.35E-04	1.33E-05
Opossum	8.35E-04	4.36E-04	1.04E-03	1.14E-03	1.12E-04	5.75E-04	0.00E + 00	8.32E-06	8.11E-05	7.58E-04
Pig	7.12E-04	3.52E-04	9.11E-04	9.97E-04	7.21E-05	4.72E-04	8.34E-06	0.00E + 00	4.62E-05	6.40E-04
Rat	4.13E-04	1.56E-04	5.64E-04	6.38E-04	6.83E-06	2.34E-04	8.12E-05	4.63E-05	0.00E + 00	3.57E-04
Rhesus	2.20E-06	4.60E-05	2.41E-05	4.11E-05	2.90E-04	1.33E-05	7.64E-04	6.45E-04	3.59E-04	0.00E + 00

No significant difference enough to represent CpG island sequence features.



Figure 1. Definition of K-mer and K-flank.

evolutionary between distance. Our objective here was to define distance between genomes by using the rank-sum tests of conserved k-mers.

Visualize phylogenetic tree

Using the Newick format data, we used the on-line phylogeny drawing tool, called PHY-FI (25). A phylogenetic tree using top ranked 64 3-mers is shown in Figure 3, which is consistent with the evolutionary history of the 10 mammalian genomes. Additional results using different k-mers and k-flanks are available at http://biohealth.snu. ac.kr/wiki/index.php/PhylogeneticTree



Figure 2. Get common k-mer(flank) exist in all the species. Each alphabet stands for the k-mer(flank) sequences. Based on the k-mer(flank) rank order in common, k-mer(flank) order in each species are set. Rank difference between common and other species are computed.

Machine learning analysis of CpG island sequence using k-mer as features

In the previous section, the k-mer/k-flank rank method was used to reconstruct a phylogenetic tree based on their sequence pattern. To investigate further relationships among CpG island sequence patterns and evolutionary relationship between species, we performed machine learning analysis on the CpG island sequences using the same k-mer frequency approach.

Machine learning algorithms

We performed the machine learning analysis using representative algorithms including Random forest (RF), naïve bayes (NB), support vector machine (SVM) and radial basis function network (RBF) implemented in Weka package (26). We did not use the artificial neural network (ANN) algorithm, as it is exceptionally time consuming. In the machine learning analysis, statistically significant k-mers were used as features, and CpG island sequences from each species were used as class.

Positive and negative data set

Based on the k-th order, the frequency of each k-mers in the CpG islands was counted and used as the positive data set to represent CpG island sequence. We used Markov model (MM) in seq++ package (27) to generate the negative data set. From CpG island sequences, the Markov model parameters were estimated and the negative sequences were generated using the Markov model. We generated random sequences of the same length as original CpG island sequences to make the positive and negative data set was one to one. Using the positive and negative data sets, we performed a 10-fold cross validation to evaluate machine learning models to characterize CpG island sequences.



Figure 3. Phylogenetic tree using top ranked 64 3-mers.

Feature selections

To select statistically significant k-mers, t-tests were used and a *P*-value of 0.05 was used as a cut-off value. For the machine learning models, only statistically significant k-mers were used.

MM order and k-mer selection

Selecting the appropriate k-mer length and the order of MM for generating random sequence are critically important. Thus, we investigated the number of statistically significant k-mers between original CpG island sequences and random sequences from varying degrees of MM. Table 3 shows the relationship between the orders of MM and k-mer lengths.

Results of machine learning on k-mer as a human CpG island sequence feature

To investigate predictive power of the machine learning models, we tested all possible combinations of the order of MM and k-mer length (Figures 4–11). Overall, SVM and RF performed better than NB and RBF, achieving prediction accuracies between 0.8 and 0.9. This result shows that CpG island sequences in humans contain distinctive k-mer patterns and are not random sequences.

Analysis of CpG island sequences in nine mammalian genomes

We next extended the k-mer pattern analysis method used for the human to the other mammalian species (Table 2). We fixed the machine learning algorithm as SVM, because it showed the best performance in our previous analysis. We also fixed parameters as 4-mer and the 2nd order for



Figure 4. Machine learning performance with 4-mer as features using 2nd order MM random set.

Table 3. Relation between MM order and k-mer

MM-order	2-mer	3-mer	4-mer	5-mer	6-mer	7-mer
lst	0	50	202	830	2992	9570
2nd	0	0	193	699	2444	7950
3rd	0	0	0	456	1826	6120
4th	0	0	0	7	947	4707
5th	0	0	0	0	38	3520
6th	0	0	0	0	0	230

Number of significant k-mer filtered by *t*-test with P < 0.05.

Table 4. Number of existing k-mer pattern in each species

	Chimp	Cow	Dog	Human	Marmoset	Mouse	Opossum	Pig	Rat	Rhesus
Total (255)	215	204	179	222	211	215	200	182	216	204

Among total possible combination of 4-mer pattern (255), each species contains different number of k-mer pattern (P < 0.05).

Table 5. Number of common k-mers between species

Species	Chimp	Cow	Dog	Human	Marmoset	Mouse	Opossum	Pig	Rat	Rhesus
Chimp	215/100%	197/88.7%	169/75.1%	213/95.0%	196/85.2%	200/86.9%	182/78.1%	173/77.2%	196/83.4%	202/93.0%
Cow	197/88.7%	204/100%	166/76.4%	200/88.4%	184/79.6%	190/82.9%	181/81.1%	174/82.0%	188/81.0%	192/88.8%
Dog	169/75.1%	166/76.4%	179/100%	171/74.3%	159/68.8%	165/72.0%	153/67.6%	164/83.2%	162/69.5%	162/73.3%
Human	213/95.0%	200/88.4%	171/74.3%	222/100%	197/83.4%	204/87.5%	186/78.8%	175/76.4%	201/84.8%	203/91.0%
Marmoset	196/85.2%	184/79.6%	159/68.8%	197/83.4%	211/100%	190/80.5%	178/76.3%	165/72.3%	191/80.9%	190/84.4%
Mouse	200/86.9%	190/82.9%	165/72.0%	204/87.5%	190/80.5%	215/100%	175/72.9%	168/73.3%	201/87.3%	191/83.7%
Opossum	182/78.1%	181/81.1%	153/67.6%	186/78.8%	178/76.3%	175/72.9%	200/100%	158/70.5%	178/74.7%	176/77.1%
Pig	173/77.2%	174/82.0%	164/83.2%	175/76.4%	165/72.3%	168/73.3%	158/70.5%	182/100%	168/73.0%	170/78.7%
Rat	196/83.4%	188/81.0%	162/69.5%	201/84.8%	191/80.9%	201/87.3%	178/74.7%	168/73.0%	216/100%	189/81.8%
Rhesus	202/93.0%	192/88.8%	162/73.3%	203/91.0%	190/84.4%	191/83.7%	176/77.1%	170/78.7%	189/81.8%	204/100%

(Number of common k-mer)/(Percentage of common k-mer). Evolutionarily closer genome pairs retain higher percentage of common k-mer pattern.



Figure 5. Machine learning performance with 5-mer as features using 2nd order MM random set.



Figure 6. Machine learning performance with 6-mer as features using 2nd order MM random set.



Figure 7. Machine learning performance with 5-mer as features using 3rd order MM random set.

the MM negative data set, as 4-mer is the smallest k-mer to get meaningful result and at least 2nd order for the MM is required to simulate dinucleotide characteristics of CpG sites in CpG islands sequence. Figure 12 shows the trained machines performance for other species. The overall performance was determined to be between 75–80%, demonstrating that the CpG island sequences contain certain unique pattern in each of the species.



Figure 8. Machine learning performance with 6-mer as features using 3rd order MM random set.



Figure 9. Machine learning performance with 6-mer as features using 4th order MM random set.



Figure 10. Accuracy change over different k-mer length with 2nd order MM as negative set.

Analysis of CpG island sequences using common k-mer patterns between species

Previous machine learning analysis showed that CpG island sequences of all species contained distinct and non-random k-mer pattern. To further investigate the k-mers, we analyzed the similarity k-mer patterns between species. The number of k-mer patterns in each



Figure 11. Accuracy change over different k-mer length with different MM order using SVM.



Figure 12. Machine learning k-mer analysis on each species.

species is conserved (Table 4); in addition, k-mer patterns were shared between each species, ranging from 67 to 100% (Table 5).

Comparison between species I: applying a human model to other species

To compare CpG island sequences among species, we used a human model of 4-mer and the 2nd order MM background model to classify CpG island sequences in nine mammalian genomes. In this case, the human CpG island sequences were used as the training data, and CpG island sequences in other species served as the test data. The experimental procedure is summarized in Figure 13. We predicted the predictive power of the



Figure 14. Result of applying a human model to different species.



Figure 13. Applying human CpG island model to different species.



Figure 15. Machine learning analysis human as positive data set and others as negative data set.



Figure 16. Result of machine learning analysis human as positive data set and other species as negative data set. The result is consistent with evolutionary history since when two species are close, e.g., human versus chimp, the prediction accuracy is low while two species are distant, e.g., human versus opossum, the prediction accuracy is high.

human model to decrease as the evolutionary distance increased between species. As shown in Figure 14, the results are consistent with evolutionary history.

Comparison between species II: models between human and other species

To further compare CpG island sequences among species, we used the human CpG island sequences as the positive data set and other species sequences as the negative data set and performed 10-fold cross validation experiments. Figure 15 illustrates the experimental scheme. The result in Figure 16 is consistent with evolutionary history: prediction accuracy was low for close species (e.g., human versus chimp), and high prediction accuracy was observed for distant species, e.g., human versus opossum.

CONCLUSION

CpG island sequences play critical roles in development and disease biology. Despite the number of important analytical studies on CpG island sequence characteristics, no comparative analysis exists on CpG island sequences among different species. One possible reason is conventional sequence analysis techniques are currently ineffective for analyzing highly biased character composition of CpG island sequences. In this article, we proposed new approaches using exact patterns of CpG island sequence called k-mer and k-flank. By using genome distance based on rank correlation tests, we show that k-mer and k-flank patterns nearby CpG sites can correctly reconstruct the phylogeny of 10 mammalian genomes. We further report that k-mers, by using various machine learning algorithms, can be used to characterize CpG islands sequences. Conserved k-mers mean conservation of short sequence in CpG island sequences. Thus, our findings of conserved k-mers in CpG island sequences extend our current knowledge of CpG islands as CpG over-represented sequences to partially conserved sequences. In addition, human model testing on nine additional mammalian genomes confirms that k-mers indeed are signatures consistent with their evolutionary history. We conclude for the first time that CpG islands sequences of 10 mammalian genomes contain evolutionary evidence for non-random pattern characteristics.

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