

Factors Associated with Serum Levels of Carcinoembryonic Antigen in Healthy Non-smokers

Original
Article

Jung-Im No, Jung-Yoon Yang, Han Jae Hyun, Choi Si Yeon, Hee-Jeong Choi*

Department of Family Medicine, Eulji University Hospital, Daejeon, Korea

Background: Carcinoembryonic antigen (CEA), a serological marker of malignant tumors, demonstrates a modest increase under nonmalignant conditions such as aging and smoking. Also, recent study results suggested that serum CEA levels may be related to insulin resistance or low-grade inflammation. The aim of this study was to investigate the factors associated with serum CEA levels in healthy non-smokers.

Methods: Data was gathered from 21,501 adults aged 20 and over. We excluded 19,081 subjects who had omissions in anthropometric measurements or laboratory tests, or who had previous or current smoking history.

Results: The mean CEA level was 1.18 ± 0.85 ng/dL in males and 0.93 ± 0.73 ng/dL in females. After adjustment for age, CEA level was positively correlated with fasting glucose, glycosylated hemoglobin (HbA1C), high density lipoprotein (HDL) cholesterol, estimated glomerular filtration rate in males. In females, CEA level was positively correlated with fasting glucose, HbA1C, HDL cholesterol, aspartate aminotransferase, and high-sensitivity C-reactive protein. In both gender groups, HbA1C had a strong influence on CEA levels when all other variables were included in the regression model ($P < 0.05$).

Conclusion: Within normal range, serum CEA levels were significantly associated with HbA1C levels but not with homeostasis model assessment of insulin resistance in the non-smoking population. These findings suggest that serum CEA levels are influenced by the glucose level itself instead of insulin resistance.

Keywords: Carcinoembryonic Antigen; Tumor Marker; Insulin Resistance; Cardiovascular Diseases

INTRODUCTION

Carcinoembryonic antigen (CEA), an oncofetal glycoprotein, is a widely used tumor marker due to its high expression in

adenocarcinomas, particularly colorectal cancer. However, it is also expressed in normal mucosal cells¹⁾ and CEA elevation is associated with several non-neoplastic conditions, including chronic renal failure, hypothyroidism, chronic obstructive pulmonary disease, obesity, aging, and cigarette smoking.²⁻⁶⁾

Recently, several studies have reported that the serum level of CEA was independently associated with metabolic syndrome, atherosclerosis, and insulin resistance.⁷⁻¹⁰⁾ Although the function of CEA has not been fully clarified, previous studies have reported that CEA stimulates monocytes and macrophages to trigger the production of pro-inflammatory cytokines and endothelial adhesion molecules.^{11,12)} Also, these pro-inflammatory features of CEA suggest that CEA could be related with insulin resistance or metabolic syndrome because insulin resistance and low-

Received: August 24, 2012, Accepted: November 1, 2013

*Corresponding Author: Hee-Jeong Choi

Tel: +82-42-611-3230, Fax: +82-42-611-3776

E-mail: ohinia@daum.net

Korean Journal of Family Medicine

Copyright © 2013 The Korean Academy of Family Medicine

© This is an open-access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0>) which permits unrestricted noncommercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

grade inflammation have been considered important pathogenic components of metabolic syndrome.¹³⁾

Because serum CEA levels are often slightly elevated in healthy cigarette smokers, only some information is available on the possible association between serum CEA levels and insulin resistance or low-grade inflammation in non-smokers including males. The aim of this study was to investigate whether the insulin resistance or low-grade inflammation was associated with increased serum levels of CEA in healthy non-smoking males and females.

METHODS

1. Study Subjects

Data was gathered from 21,501 adults aged 20 and over. All subjects had visited the health promotion center of Eulji University Hospital for a periodic health examination between January 2010 and July 2011. Among these subjects, we excluded 19,081 subjects who met one of the following conditions: omissions in anthropometric measurements or laboratory tests ($n = 16,534$); previous or current history of cancers ($n = 56$); previous or current smoking history ($n = 2,319$); pregnancy ($n = 2$); abnormal liver or renal function ($n = 26$); high-sensitivity C-reactive protein (hsCRP) > 1 mg/dL ($n = 41$); or a history of taking medications for diabetes ($n = 103$). The final analytic sample was confined to 2,420 subjects. This study was approved by the institutional review board of Eulji University Hospital.

2. Data Collection and Anthropometric Measurements

All subjects completed self-reported questionnaires regarding lifestyle-related information, including their smoking, alcohol drinking, and exercise habits. Detailed medical histories were collected from the medical interviews. A smoker was defined as a current smoker and an ex-smoker who had stopped smoking within the last 6 months. Alcohol consumption was defined as more than two bottles (one bottle in females) of soju (a type of distilled spirit containing 56.8 g of pure alcohol) consumed per week. Regular exercise was defined as 30 minutes or more at a time, three times per week regularly. Height and weight were measured in subjects wearing a light robe and no shoes.

Waist circumference was measured and body mass index was calculated as body weight (kg) divided by height squared (m^2). Blood pressure and heart rate were measured with an automatic instrument (BP-8800C; Colin Electronics, Komaki, Japan) in a sitting position on at least two different occasions after 10 minutes of rest.

Blood samples were collected after an overnight fast (> 12 hours) and analyzed within 3 hours after the sampling. Complete blood cell counts were quantified by an automated blood cell counter (ADVIA 2120i analyzer; SIEMENS Diagnostics, Tarrytown, NY, USA). Blood chemistry was measured by an enzymatic technique using an ADVIA 2400 analyzer. The method used to measure the low density lipoprotein cholesterol was the homogeneous assay from International Reagents Corporation. High-sensitivity C-reactive protein (hsCRP) was measured via a turbidmetric immunoassay using a Hitachi 7170S device (Hitachi Hi-Tech, Tokyo, Japan). Estimated glomerular filtration rate (eGFR) was calculated by simplified modification of diet in the renal disease equation. CEA and plasma insulin levels were measured by chemiluminescence immunoassays using an ADVIA Centaur XP device (Siemens Healthcare Diagnostics, Los Angeles, CA, USA). Glycosylated hemoglobin (HbA1C) was assayed by ion-exchange high-performance liquid chromatography using an HLC-723 G7 machine (Tosoh Corporation, Tokyo, Japan), which was calibrated to the Diabetes Control and Complication Trial standard. The homeostasis model assessment of insulin resistance (HOMA-IR) was calculated as $[\text{fasting insulin (mIU/mL)} \cdot \{\text{fasting glucose (mg/dL)}\} / 18.1] / 22.5$.

3. Definition of Variables

Metabolic syndrome was defined based on the modified National Cholesterol Education Program's Adult Treatment Panel III definition¹⁴⁾ and according to the definition of the Korean Society for the Study of Obesity.¹⁵⁾ Subjects were classified as having metabolic syndrome if any three of the following were present: abdominal obesity (waist circumference ≥ 90 cm in male, ≥ 85 cm in female); high blood pressure (systolic blood pressure ≥ 130 mm Hg or diastolic blood pressure ≥ 85 mm Hg); high fasting glucose (≥ 100 mg/dL); high triglyceride (≥ 150 mg/dL); or low high density lipoprotein (HDL) cholesterol (< 40 mg/dL in males and < 50 mg/dL in females).

4. Statistical Analyses

All statistical analyses were performed using SPSS ver. 14.0 (SPSS Inc., Chicago, IL, USA). Skewed data were transformed to the natural logarithm scale prior to statistical analysis. To compare the general characteristics of the study subjects according to gender, independent t-test and chi-square tests were used for continuous and categorical variables, respectively. Correlation coefficients between CEA levels and other variables

were calculated by partial correlation analysis after adjusting for age. Multivariate linear regression analysis was performed to identify the major determinants of CEA levels. In this model, the independent variables were chosen based on a partial correlation analysis. The mean values of CEA according to HbA1C levels were compared by analysis of variance. A probability value of $P < 0.05$ was considered significant.

Table 1. General characteristics of study subjects

Variable	Male (n = 543)	Female (n = 1,877)	P-value*
Age (y)	46.8 ± 11.0	47.0 ± 11.5	0.760
Body mass index (kg/m ²)	24.2 ± 3.1	22.9 ± 3.1	<0.001
Waist (cm)	81.1 ± 7.4	72.9 ± 7.2	<0.001
Drinking	278 (51.2)	397 (21.2)	<0.001
Regular exercise	320 (58.9)	1,100 (58.6)	0.351
Metabolic syndrome	61 (11.4)	211 (11.2)	0.912
Systolic blood pressure (mm Hg)	129.2 ± 15.2	121.2 ± 16.6	<0.001
Diastolic blood pressure (mm Hg)	76.0 ± 10.1	69.3 ± 10.2	<0.001
Heart rate (beat/min)	64.0 ± 9.8	64.5 ± 9.3	0.261
Fasting glucose (mg/dL)	91.0 ± 20.9	86.1 ± 14.7	<0.001
Glycosylated hemoglobin (%)	5.58 ± 0.91	5.53 ± 0.55	0.074
Insulin (mU/L)	6.9 ± 4.2	6.7 ± 4.0	0.360
HOMA-IR	1.57 ± 1.08	1.46 ± 1.00	0.022
Total cholesterol (mg/dL)	190.4 ± 32.7	187.6 ± 33.8	0.088
Triglyceride (mg/dL)	129.2 ± 76.3	105.2 ± 62.5	<0.001
HDL cholesterol (mg/dL)	49.1 ± 10.5	56.0 ± 13.4	<0.001
LDL cholesterol (mg/dL)	120.4 ± 30.5	112.7 ± 31.3	<0.001
Aspartate aminotransferase (U/L)	25.3 ± 8.2	21.9 ± 7.1	<0.001
Alanine aminotransferase (U/L)	28.5 ± 15.0	18.9 ± 10.3	<0.001
γ-glutamyltransferase (U/L)	39.4 ± 36.8	19.8 ± 22.2	<0.001
Creatinine (mg/dL)	0.93 ± 0.12	0.67 ± 0.10	<0.001
Estimated GFR (mL/min)	95.8 ± 15.2	104.2 ± 19.7	<0.001
White blood cell (×10 ³ /μL)	5.94 ± 1.40	5.62 ± 1.50	<0.001
Erythrocyte sediment rate (mm/h)	6.5 ± 6.5	12.6 ± 9.9	<0.001
High-sensitivity C-reactive protein (mg/dL)	0.13 ± 0.14	0.11 ± 0.12	<0.001
Carcinoembryonic antigen (ng/dL)	1.18 ± 0.85	0.93 ± 0.73	<0.001

Values are presented as mean ± SD or number (%).

HOMA-IR: homeostasis model assessment of insulin resistance, HDL: high density lipoprotein, LDL: low density lipoprotein, GFR: estimated glomerular filtration rate.

*By independent t-test for continuous variables and by chi-square test for categorical variables.

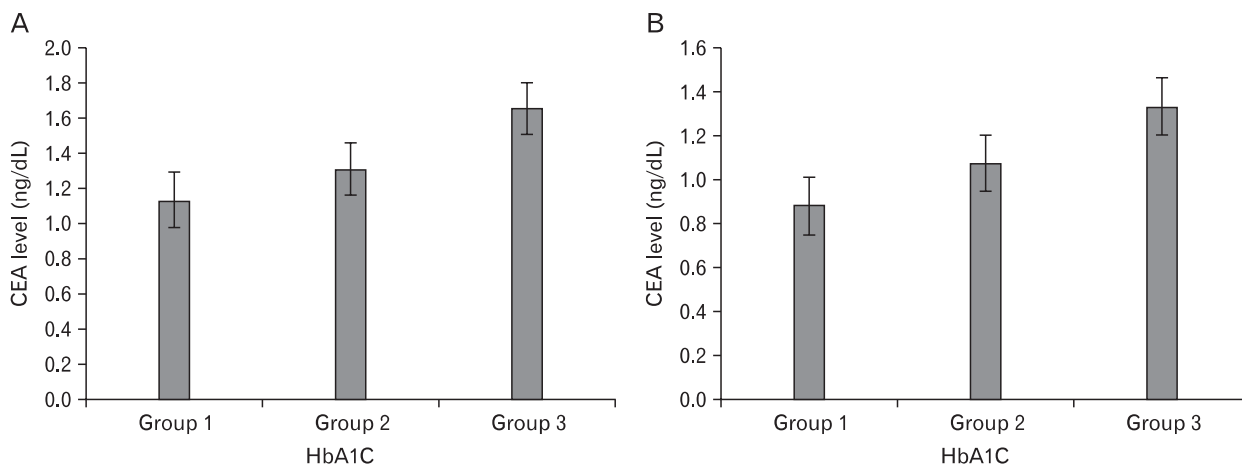


Figure 1. Mean values of carcinoembryonic antigen (CEA) according to glycosylated hemoglobin (HbA1C) groups in males (A) and females (B). Values are presented as the mean ± SE. P-value < 0.005 by analysis of variance in both (A) and (B). The groups were defined as follows: group 1, HbA1C < 5.7%; group 2, 5.7% ≤ HbA1C < 6.5%; group 3, HbA1C ≥ 6.5%.

Table 2. Partial correlation coefficients between CEA levels and other variables

Variable	Male		Female	
	r	P-value*	r	P-value*
Body mass index	-0.026	0.555	0.000	0.987
Waist	-0.022	0.610	0.012	0.594
Systolic blood pressure	0.055	0.204	0.020	0.386
Diastolic blood pressure	0.009	0.843	0.029	0.213
Heart rate	0.044	0.305	-0.022	0.338
Fasting glucose	0.125	0.004	0.052	0.026
Glycosylated hemoglobin	0.110	0.010	0.066	0.004
HOMA-IR [†]	0.010	0.814	-0.018	0.447
Total cholesterol	0.026	0.544	0.018	0.447
Triglyceride [†]	0.019	0.566	-0.011	0.629
HDL cholesterol	0.097	0.024	0.077	0.001
LDL cholesterol	0.042	0.329	0.010	0.665
Aspartate aminotransferase	0.067	0.119	0.057	0.013
Alanine aminotransferase	0.058	0.183	0.037	0.109
γ-glutamyltransferase	0.029	0.501	0.031	0.187
Creatinine	-0.123	0.004	-0.028	0.224
Estimated GFR	0.116	0.007	0.034	0.145
White cell count	-0.010	0.824	0.011	0.645
Erythrocyte sediment rate	-0.004	0.932	-0.031	0.176
High-sensitivity C-reactive protein [†]	0.048	0.264	0.065	0.005

CEA: carcinoembryonic antigen, HOMA-IR: homeostasis model assessment of insulin resistance, HDL: high density lipoprotein, LDL: low density lipoprotein, GFR: estimated glomerular filtration rate.

*By partial correlation after adjustment for age. [†]Values and CEA levels were analyzed after log-transformation.

RESULTS

The mean ages of the 543 males (22%) and 1,877 females (78%) were 46.8 ± 11.0 and 47.0 ± 11.5 years, respectively. Thirty-two percent of female subjects were menopausal. There was no difference in the prevalence of metabolic syndrome according to gender. The mean CEA level was 1.18 ± 0.85 ng/dL in males and 0.93 ± 0.73 ng/dL in females ($P < 0.001$). The mean CEA levels of females with and without metabolic syndrome were 0.91 ± 0.73 ng/dL and 1.07 ± 0.70 ng/dL, respectively ($P < 0.05$). This difference was significant only in females (data not shown). Fasting glucose levels and hsCRP levels were significantly higher in females. On the other hand, there were no differences in HbA1C. General characteristics of the study subjects are presented in Table 1. Figure 1 shows the mean levels of CEA according to HbA1C in each gender. The mean levels of CEA levels were significantly higher in the highest HbA1C group ($P < 0.005$) (Figure 1).

After adjustment for age, CEA level was positively correlated with fasting glucose, HbA1C, HDL cholesterol, eGFR in males. In females, CEA level was positively correlated with fasting glucose, HbA1C, HDL cholesterol, aspartate aminotransferase,

and hsCRP (Table 2).

Table 3 shows that HbA1C had the strongest influence on CEA levels when age, waist and all other variables were included in the regression models in each gender. Also, in males, eGFR and age were associated with CEA levels ($P < 0.05$). On the other hand, age, HDL cholesterol, hsCRP, and exercise were associated with CEA levels in females ($P < 0.05$). Waist, blood pressure, triglyceride levels, and HOMA-IR were not associated with CEA levels in either gender. All together, these variables explain up to 3.3% and 8.3% of the variation in the CEA levels in males and females, respectively (Table 3).

DISCUSSION

Although of little use in detecting early colorectal cancer, CEA is a useful indicator for surveillance of patients with diagnosed colorectal cancer. Slightly elevated CEA levels are associated with several conditions,²⁻⁶⁾ but very little information exists about the significance of mildly elevated CEA levels over an extended period in individuals who do not have cancer. Recently, some epidemiological studies have demonstrated a

Table 3. Multivariate regression analyses with CEA level as a dependent variable

Variable	R ²	B	SE	Beta	P-value*
Male					
Constant	0.033	-0.870	0.194		<0.001
HbA1C	0.033	0.057	0.020	0.123	0.005
Estimated GFR	0.033	0.003	0.001	0.114	0.011
Age	0.033	0.004	0.002	0.091	0.043
Female					
Constant	0.083	-0.952	0.137		<0.001
HbA1C	0.083	0.042	0.021	0.49	0.042
Age	0.083	0.010	0.001	0.249	0.001
HDL cholesterol	0.083	0.003	0.001	0.087	<0.001
hsCRP [†]	0.083	0.092	0.033	0.066	0.006
Exercise	0.083	-0.043	0.021	-0.046	0.039

CEA: carcinoembryonic antigen, HbA1C: glycosylated hemoglobin, GFR: estimated glomerular filtration rate, HDL: high density lipoprotein, hsCRP: high-sensitivity C-reactive protein, LDL: low density lipoprotein, HOMA-IR: homeostasis model assessment of insulin resistance.

*By multiple regression analysis including age, waist, drinking, exercise, diastolic blood pressure, HDL cholesterol, LDL cholesterol, triglyceride, HbA1C, HOMA-IR, aspartate aminotransferase, estimated GFR, and hsCRP. [†]Values and CEA levels were analyzed after log-transformation.

possible association between increased CEA levels and metabolic syndrome or atherosclerosis.⁷⁻¹¹ Lee et al.⁷ reported that CEA level was independently associated with metabolic syndrome after adjustment for all covariates in Korean female non-smokers. For the same reason, Kim et al.⁸ suggested that metabolic syndrome may lead to a misunderstanding of CEA levels. Ishizaka et al.⁹ reported that CEA was associated with carotid atherosclerosis in Japanese males without serious health problems. The increase in serum CEA was found to be associated with an increased prevalence of carotid plaque independent of blood pressure, fasting glucose, serum lipids, and inflammatory markers. Bae et al.¹⁰ reported that CEA level was associated with arterial stiffness in healthy Korean males and females after adjustment for all covariates. These data indicate the plausibility that serum CEA may be associated with increased insulin resistance and low-grade inflammation.

Previous studies have suggested that CEA may stimulate macrophages or monocytes via binding to the CEA receptor which results in the production of pro-inflammatory cytokines such as tumor necrosis factor- α , interleukin-1b and -6, reactive oxygen species, eicosanoids, nitric oxide, and carbon monoxide, and up-regulation of endothelial adhesion molecules, which may play a role in metabolic disturbances and cardiovascular disease in addition to cancer.^{11,12} Low-grade inflammation is now recognized as the central player in the pathogenesis of atherosclerosis, insulin resistance, and glucose intolerance.^{16,17} Also serum CEA levels were increased in patients with chronic inflammatory disorders such as inflammatory bowel disease, collagen disease, and chronic viral hepatitis.^{2,18}

In this study, we investigated the factors associated with CEA levels and whether insulin resistance or low-grade inflammation was associated with high CEA levels in healthy non-smokers including non-smoking males. Although HbA1C and age were shared determinants of CEA levels in both genders, the other factors associated with CEA levels were different in each gender. In male subjects, eGFR was associated with CEA levels, but CEA levels were associated with HDL cholesterol, hsCRP, and exercise status in females. Contrary to expectations, the HOMA-IR was not associated with CEA levels in each gender. This finding suggests that the factor affecting CEA levels is glucose levels rather than insulin resistance. On the other hand, hsCRP is associated with CEA levels in females, but not in males. Shio et al.¹⁹ reported

that the serum CEA level was significantly associated with fasting glucose levels in diabetic patients. Another study had revealed that serum levels of CEA and carbohydrate antigen 19-9 (CA19-9), a tumor marker of pancreatic cancer, could be influenced by high glucose levels and HbA1C in diabetic patients.²⁰ These results may be due to similar components such as glucose, glycoprotein, and lipid of CEA, CA19-9, and HbA1C.²¹

Several variables including HbA1C were associated with CEA levels, and these variables explain up to 3.3% and 8.3% of the variation in the CEA levels in male and female gender, respectively. This finding suggests that many other factors seem to have a stronger influence on CEA levels.

This study has a number of strengths. First, this study subjects include a large number of non-smoking males, though a fewer number than males. Second, there are several factors that influence serum CEA levels and its metabolism. For this reason, we excluded all subjects that had any smoking history including ex-smokers and abnormal liver or kidney function test results, in order to eliminate the influence of these conditions. Third, we also excluded all subjects that had any diabetic medications which may influence HOMA-IR in the identification of the association between insulin resistance and serum CEA levels. For the same reason, we exclude all subjects whose levels of hsCRP > 1 mg/dL, as this may indicate active inflammatory or infectious conditions.

At the same time, this study has some limitations. First, because the study was cross-sectional, we cannot verify the causality between the each variable and the elevation of serum CEA. Second, we cannot rule out selection bias in our subjects, who had visited a health promotion center. Therefore, these results cannot be generalized to other population groups. Third, serum CEA is known to be elevated in other nonmalignant conditions, such as hypothyroidism and end-stage lung diseases. We did not include these disorders as confounding variables, because their prevalence is considered to be very low among study populations.

In conclusion, within the normal range, serum CEA levels were significantly associated with HbA1C levels but not with HOMA-IR in the non-smoking population. These findings suggest that serum CEA levels are influenced by the glucose level itself rather than insulin resistance.

CONFLICT OF INTEREST

No potential conflict of interest relevant to this article was reported.

REFERENCES

1. Yalta K, Yilmaz A, Turgut OO, Erselcan T, Yilmaz MB, Karadas F, et al. Evaluation of tumor markers CA-125 and CEA in acute myocardial infarction. *Adv Ther* 2006;23:1052-9.
2. Ruibal Morell A. CEA serum levels in non-neoplastic disease. *Int J Biol Markers* 1992;7:160-6.
3. Amino N, Kuro R, Yabu Y, Takai SI, Kawashima M, Morimoto S, et al. Elevated levels of circulating carcinoembryonic antigen in hypothyroidism. *J Clin Endocrinol Metab* 1981;52:457-62.
4. Bulut I, Arbak P, Coskun A, Balbay O, Annakkaya AN, Yavuz O, et al. Comparison of serum CA 19.9, CA 125 and CEA levels with severity of chronic obstructive pulmonary disease. *Med Princ Pract* 2009;18:289-93.
5. Park JS, Choi GS, Jang YS, Jun SH, Kang H. Influence of obesity on the serum carcinoembryonic antigen value in patients with colorectal cancer. *Cancer Epidemiol Biomarkers Prev* 2010;19:2461-8.
6. Herbeth B, Bagrel A. A study of factors influencing plasma CEA levels in an unselected population. *Oncodev Biol Med* 1980;1:191-8.
7. Lee JW, Park KD, Im JA, Hwang HJ, Kim SH. Serum carcinoembryonic antigen is associated with metabolic syndrome in female Korean non-smokers. *Clin Chim Acta* 2011;412:527-30.
8. Kim KN, Joo NS, Je SY, Kim KM, Kim BT, Park SB, et al. Carcinoembryonic antigen level can be overestimated in metabolic syndrome. *J Korean Med Sci* 2011;26:759-64.
9. Ishizaka N, Ishizaka Y, Toda E, Koike K, Yamakado M, Nagai R. Are serum carcinoembryonic antigen levels associated with carotid atherosclerosis in Japanese men? *Arterioscler Thromb Vasc Biol* 2008;28:160-5.
10. Bae U, Shim JY, Lee HR, Shin JY. Serum carcinoembryonic antigen level is associated with arterial stiffness in healthy Korean adult. *Clin Chim Acta* 2013;415:286-9.
11. Ganguly A, Yeltsin E, Robbins J. Identification of a carcinoembryonic antigen binding protein on monocytes. *Biochem Biophys Res Commun* 2003;311:319-23.
12. Aarons CB, Bajenova O, Andrews C, Heydrick S, Bushell KN, Reed KL, et al. Carcinoembryonic antigen-stimulated THP-1 macrophages activate endothelial cells and increase cell-cell adhesion of colorectal cancer cells. *Clin Exp Metastasis* 2007;24:201-9.
13. Kirk EP, Klein S. Pathogenesis and pathophysiology of the cardiometabolic syndrome. *J Clin Hypertens (Greenwich)* 2009;11:761-5.
14. Grundy SM, Brewer HB Jr, Cleeman JI, Smith SC Jr, Lenfant C; American Heart Association, et al. Definition of metabolic syndrome: report of the National Heart, Lung, and Blood Institute/American Heart Association conference on scientific issues related to definition. *Circulation* 2004;109:433-8.
15. Lee SY, Park HS, Kim DJ, Han JH, Kim SM, Cho GJ, et al. Appropriate waist circumference cutoff points for central obesity in Korean adults. *Diabetes Res Clin Pract* 2007;75:72-80.
16. Tjong AY, Brieger D. Inflammation and coronary artery disease. *Am Heart J* 2005;150:11-8.
17. Shoelson SE, Lee J, Goldfine AB. Inflammation and insulin resistance. *J Clin Invest* 2006;116:1793-801.
18. Booth SN, King JP, Leonard JC, Dykes PW. Serum carcinoembryonic antigen in clinical disorders. *Gut* 1973;14: 794-9.
19. Shio H, Iizuka K, Majima Y. Serum CEA level in diabetic patients. *Intern Med (Japan)* 1982;49:377-80.
20. Kim KC, Kim HB, Suh JK, Lee SS, Lew YS, Lee SJ. The relationship between blood glucose and tumor marker (CEA, CA19-9) in patients with NIDDM. *J Korean Diabetes Assoc* 1991;15:85-9.
21. Bernheim F, Bernheim ML, Wilbur KM. The reaction between thiobarbituric acid and the oxidation products of certain lipides. *J Biol Chem* 1948;174:257-64.