

## Invited Mini Review

## Investigating the role of Sirtuins in cell reprogramming

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Cell reprogramming has been considered a powerful technique in the regenerative medicine field. In addition to diverse its strengths, cell reprogramming technology also has several drawbacks generated during the process of reprogramming. Telomere shortening caused by the cell reprogramming process impedes the efficiency of cell reprogramming. Transcription factors used for reprogramming alter genomic contents and result in genetic mutations. Additionally, defective mitochondria functioning such as excessive mitochondrial fission leads to the limitation of pluripotency and ultimately reduces the efficiency of reprogramming. These problems including genomic instability and impaired mitochondrial dynamics should be resolved to apply cell reprogramming in clinical research and to address efficiency and safety concerns. Sirtuin (NAD<sup>+</sup>-dependent histone deacetylase) has been known to control the chromatin state of the telomere and influence mitochondria function in cells. Recently, several studies reported that Sirtuins could control for genomic instability in cell reprogramming. Here, we review recent findings regarding the role of Sirtuins in cell reprogramming. And we propose that the manipulation of Sirtuins may improve defects that result from the steps of cell reprogramming. [BMB Reports 2018; 51(10): 500-507]

## INTRODUCTION

Cell reprogramming techniques have emerged with novel techniques to treat a variety of human diseases in the regenerative medicine field (1). In the reprogramming process, 'immortality' is regarded as a key to develop rejuvenation strategies (2). Takahashi et al. stated that cell reprogramming using four transcription factors such as Oct4, Sox2, Klf4, and c-Myc could convert terminally differentiated cells into

induced pluripotent stem cells (iPSCs) (1). The pluripotency of iPSCs has opened up numerous possibilities for regenerative medicine to treat many diseases (3). Despite the powerful ability of iPSCs to treat numerous diseases, major concerns in recent iPSCs research include enhancing reprogramming efficiency and genomic stability. Genomic instability in iPSCs is generated in several steps of the cell reprogramming process (4). Cellular reprogramming goes through an intricate process that is similar to biological pathways of tumorigenesis (5). The essential factors for cell reprogramming are associated with tumorigenesis. For example, c-Myc and Klf4 play central roles in tumorigenesis, and Oct4 acts as an important initiator for germ cell tumors (5). In addition, to inducing changes in the original cell identity, cell reprogramming needs reactivation of the telomerase to continue to survive (6). Maintenance of telomere as an enzyme for telomere elongation is important for genomic stability during reprogramming (7). Telomerase is reactivated during reprogramming and the length and epigenetic state of the telomere contributes to rejuvenation in iPSCs. Shortening of the telomeres influences the reprogramming efficiency and the quality of the iPSCs (8). The strategy to solve the genome instability in cell reprogramming research for application in disease modeling and clinical cell therapy (9). During cell reprogramming, cells experience a metabolic shift into the glycolytic state (10). Oxidative stress and DNA damage from the cell reprogramming process results in a metabolic imbalance (11). Because of these metabolic shifts, mitochondrial activity is hampered and cannot react when energy is demanded due to cellular respiration. The reduction of mitochondrial activity during cell reprogramming is a matter that should be resolved for increasing iPSCs efficiency. Sirtuins known as histone deacetylases are relevant to the control of longevity, energy metabolism, and cell development in mammals (12). It was reported that sirtuins can affect the fate of stem cells through deacetylation of histone and non-histone proteins involved in gene expression (13). Recent studies demonstrated that the deficiency of Sirtuins influences reprogramming efficiency (14) and contributes to genomic instability, which as we noted, is an important issue in the cell reprogramming process (15). Here, we review evidence on the significant role of Sirtuins in the cell reprogramming process.

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## GENOMIC INSTABILITY IN CELL REPROGRAMMING

Genomic instability occurs during the cell reprogramming process (16). A number of studies report that after reprogramming iPSCs exhibit the genomic abnormalities such as chromosomal aberrations (17). Because of the transcription factors used in cell reprogramming cells have an increased risk of both tumor formation and genetic mutation (18). Telomerase is significantly upregulated during cell programming (8). Pluripotent cells show high activity of telomerase responsible for synthesizing telomeres in the reprogramming process (19). The iPSCs generation process showed that telomerase reverse transcriptase was upregulated in cells during cellular reprogramming (1). Telomerase activity and telomere length affect the state of pluripotency (20). In cell reprogramming, reactivation of telomerase has been shown to promote efficiency of iPSC reprogramming by maintaining telomere length and self-renewal potential for a relatively long time (21). Upon reprogramming, telomere lengthening is affected by a decrease of DNA methylation (22) and a reduction of methylation in histone H3 at lysine 9 (H3K9) m3 and histone H4 at lysine 20 (H4K20) m3 (8). Some studies investigated the differences in the telomere dynamics during reprogramming (21). Telomere shortening is a crucial issue in reprogramming process in that it hampers sufficient iPSCs generation. During the cell reprogramming process the proliferation rate increases causing replication stress and genomic structural variation (23). Additionally, recent studies show that pluripotent stem cells have an abnormal cell-cycle regulation such as a shorter G1 phase. The ataxia telangiectasia mutated Rad3 (ATR)-mediated checkpoint pathway is an essential replication stress response that generates genomic instability during reprogramming (24). Other studies report that Checkpoint kinase 1 (CHK1) overexpression could enhance both the reprogramming efficiency and the iPSCs quality (25). Abnormal cell cycle regulation is a distinct feature and the control over it is considered important to current reprogramming research. Accordingly, to realize the application of iPSCs in clinical research, we need a comprehensive understanding of genetic instability and should find an appropriate solution for it in cell reprogramming.

## MITOCHONDRIAL DYNAMICS IN CELL REPROGRAMMING

Mitochondria is a multifunctional organelle and plays a crucial role in many cellular mechanisms such as energy production, apoptosis, reactive oxygen species (ROS) production, senescence, and metabolism (26). Mitochondrial homeostasis has been shown to be essential for maintenance of a pluripotent state. Ji et al. report that a decrease of ROS production in the mitochondria could improve iPSCs quality (27). Also excessive mitochondrial fission and knockdown of the mitochondrial DNA polymerase could trigger a lack of pluripotency (28). Tricyclic antidepressant (TCA)-derived

cytosolic acetyl-CoA is essential for maintaining histone acetylation and an open chromatin state during cell reprogramming (29). Reprogramming somatic cells into iPSCs triggers impairment of the mitochondrial network during the reprogramming process (30). Besides, during cell reprogramming, cells show particular characteristics including immature and globular mitochondria (31), and poorly developed cristae (32). Reduced expression 1 (REX1) known as a pluripotency marker regulates cell fate through its effect on mitochondrial dynamics (33). The knockdown of Dynamin-related GTPases-1 (DRP1) triggers the elongation of the mitochondrial network (34) and regulates membrane dynamics in a variety of cellular mechanisms and in mitochondrial fusion (35). One study demonstrated that the DRP1-GTPase inhibitor impedes cell reprogramming of human fibroblasts to iPSCs. The mechanistic target of rapamycin (mTOR) promotes cellular homeostasis and multiple signaling events that affect reprogramming (1). Inhibition of mTOR leads to an immediate decrease in mitochondrial respiration (36) and subsequently influences the generation of iPSCs (37). Taken together, cell reprogramming influences abnormal mitochondria function and homeostasis, and mitochondrial dynamics should be a focus for future cell reprogramming research.

## SIRTUINS IN GENOMIC INSTABILITY DERIVED FROM CELL REPROGRAMMING

Sirtuins as an NAD<sup>+</sup>-dependent histone deacetylase have been involved in the improvement of longevity and metabolism in mammals (38). Given that histone acetylation is associated with gene activation (39), Sirtuins act as an epigenetic regulator of gene expression by histone deacetylation (40). Sirtuins have been shown to be essential for the silent chromatin state of the ribosomal RNA genes and telomeres. In mammals, Sirt6 has been reported to maintain telomeric chromatin and to enhance replicative capacity (41). According to cell reprogramming research the activation of Sirtuins considerably enhances the efficiency of cell reprogramming (42). Several studies demonstrate that the inhibition of histone deacetylases leads to increases of histone acetylation levels, chromatin opening, and ultimately could enhance efficiency of cell reprogramming (43). Sirtuins could possibly control the chromatin state by modulating the activation of enzymes such as H4K16Ac (44) and H3K4me3 that can upregulate cell reprogramming. Sirt1 is intimately linked to the maintenance of human embryonic stem cells pluripotency by inactivating p53 (45). Besides stem cells derived from Sirt6, knockout mice cells exhibit expression of Oct4, Sox2 and Nanog and present Sirt6's function in balance between pluripotency and differentiation (46). Sirt1 could lead to the deacetylation of Sox2 (14) and Sirt1's overexpression induces the demethylation of the Oct4 promoter (47) and also affects reprogramming efficiency. Myc stability, important in cell reprogramming, could also be regulated by Sirt2 (48). Sirt1 deacetylates c-Myc

by interacting physically with the C-terminus of c-Myc (49). Sirt1 induces p53 translocation into the mitochondria (50) and modulates Nanog expression (51) and is an important reprogramming factor. Judging by the metabolic state of the cell, Sirt1 can affect the epigenome change and the activity of chromatin-modifying enzymes (52). Sirt1 histone deacetylase regulates the epigenetical change and gene expressions in cells by translating a metabolic shift in the reprogramming process (53). A recent study showed that Sirt6 inhibits the transcription of Hypoxia-inducible factors (HIF1)-alpha and Myc (54). Sirt6 is essential for the maintenance of the telomere position in cells (55) and the deficiency of it leads to DNA damage and genomic instability (15). In addition, Sirt6 protects cells against stress by repairing DNA damage and preserving telomere integrity and controlling metabolic homeostasis (56). Sirt6 can deacetylate lysine 9 on histone H3 (H3K9Ac) (41) and lysine 56 on histone H3 (H3K56Ac) (57). And Sirt6 can recruit the chromatin remodeler Sucrose Nonfermenting Protein 2 Homolog (SNF2H) (58). As we have seen, Sirtuins influence cell reprogramming efficiency by regulating the activities of histone deacetylases, by controlling the chromatin state of telomere, and by being involved in metabolic shifts during cell reprogramming (Fig. 1).

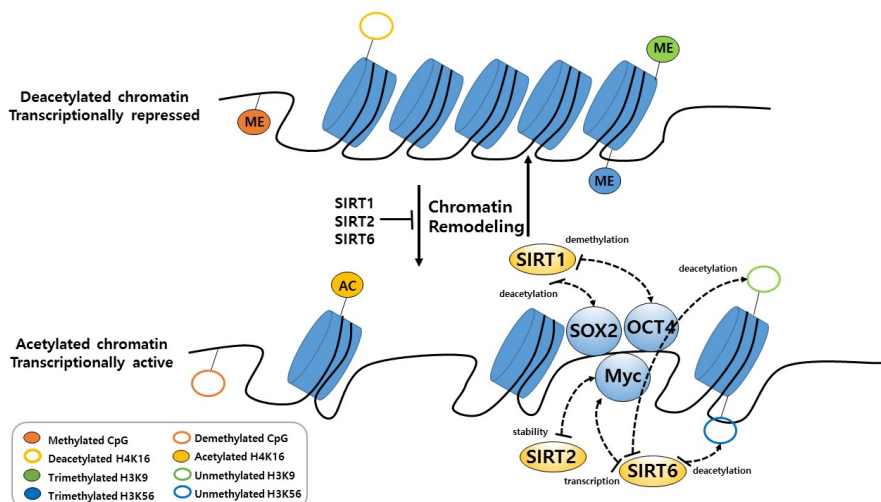
## ACTIVATORS AND INHIBITORS OF SIRTUINS

Several compounds are known to be activators of Sirtuin. Resveratrol (3,5,4'-trihydroxy-trans-stilbene), SRT1720, Oxazolo [4,5-b] pyridines derivative, imidazole [1,2-b] thiazole derivative, and 1,4-dihydropyridine (DHP) derivatives are typical compounds that are known activators of Sirtuins (59). The exact mechanisms of Sirt1 activation by these activators is still unclear, but many of them seem to activate Sirt1 through allosteric activation, particularly, the resveratrol mediate

activation of Protein Kinase AMP-Activated Catalytic Subunit Alpha 2 (AMPK), which is an initial sensor that increases NAD<sup>+</sup> levels leading to activation of Sirt1 (60).

The metabolic effects of Resveratrol, the most common Sirtuin activator, relate to the cAMP level elevation in muscles (61). Also the general health in mice fed with a high caloric diet improved and they showed a marked reduction in signs of aging (62). These results open the possibility of clinical use of commercial micronized Respiratory Syncytial Virus (RSV) formulation, SRT501, for lowering blood glucose and improving insulin sensitivity in patients with type 2 diabetes (63). Moreover, SRT1720 has been shown to induce cell death in multiple myeloma cells (64) and significantly decrease tumor growth in a preclinical evaluation for cancer treatment (65). Also, as a new activators of Sirt1 unrelated to Resveratrol, a series of oxazolo pyridines was identified for potential therapeutic targets to treat different diseases (66). For example, compound 29 showed antidiabetic activity in types 2 diabetes (67) and SRT2104 was tested in a clinical trial of patients with metabolic inflammatory (68) and cardiovascular diseases (69).

In contrast, Splitomicin, HR73, Sirtinol, AGK2, Cambinol, Salermide, Tenovin, and Suramin are inhibitors of Sirtuin (70). The reaction mechanism of Sirtuins is the cleavage of nicotinamide (NIC) from NAD<sup>+</sup> whereas ADP-ribose binds its acetyl-peptide with the formation of an o-alkylamidate intermediate. Sirtuin inhibitors hamper cleavage of NIC from NAD<sup>+</sup>. Suramin is an especially potent inhibitor of Sirt1, Sirt2 and Sirt5 (71). It inhibits NAD<sup>+</sup>-dependent deacetylase activity with an IC<sub>50</sub> value of 22uM leading to mitochondrial dynamics disruptions (72). Several studies revealed that pharmacological inhibition of Sirtuin1 by Sirtinol inhibits prostate cancer cell proliferation in which Sirtuin1 is highly enriched (73). Moreover, Salermide, a sirtinol derivative, induces cell death via inhibiting MAP kinases erk1/2, p38 and JNK



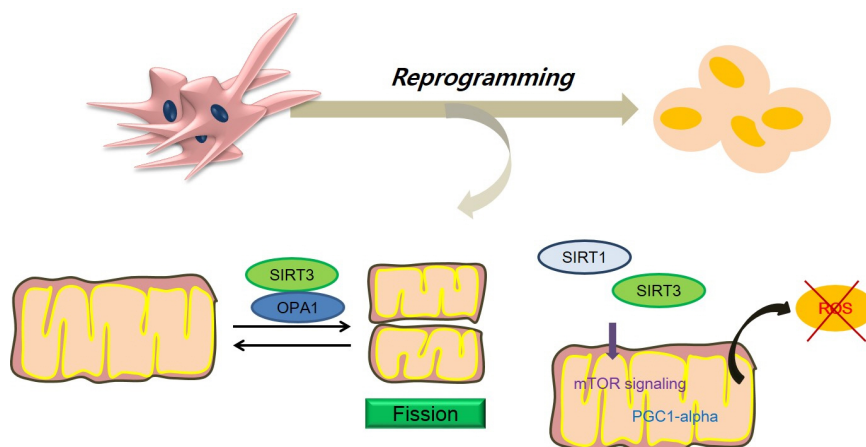
**Fig. 1.** The function of sirtuins on genome stability. Sirt 1, 2, and 6 control the chromatin state by regulating the activation of enzymes during chromatin remodeling. Sirt1 removes acetylates in Sox2 and Myc removes methylates in Oct4. Also, Sirt2 modulates the stability of the Myc protein. SIRT6 can deacetylate H3K9Ac and H3K56Ac and is involved in the transcription of c- Myc. AC: Acetylation, ME: Methylation, SIRT: Sirtuin.

paring Sirtuin1 and Sirtuin2 in various human cancer cell lines derived from leukemia, lymphoma, colon, and breast primary malignancies (74). 6-Chloro-2,3,4,9-tetrahydro-1 H-Carbazole-1-carboxamide (EX527) is also known as a Sirtuin1 inhibitor and EX-527/SEN0014196 reduced neuronal death caused by mutant Huntington proteins in cell-based assays in preclinical studies of Huntington's disease (75). More importantly, activation and inhibition of Sirtuin by small molecules is a complicated process and the effects of activation and inhibition of Sirtuin occasionally depend on the physiological state of the specific cells for its activity. For instance, increased activity of SIRT1 after treatment with resveratrol in the immediate immune response reduced the NFκB activation in the NFκB-dependent inflammatory genes in microglia and neuronal loss (76). This suggests that Sirt1 is working as anti-inflammatory mediator, whereas decreasing Sirt1 activity by sirtinol potentiates inflammatory responses, presumably occur via Sirt1-mediated deacetylation of p65 (77). Moreover, unlike Sirt1 effects on the inflammation, Sirt6 activity is positive for a given pro-inflammatory gene expression up-regulating Tumor necrosis factor alpha (TNFα) and Interferon Production Regulator (IFN $\gamma$ ) synthesis on both innate and adaptive immune cells (78). Such complexity of sirtuin activity in the various physiological states of cells lead to the difficulties of Sirtuin activators or inhibitors in determining the desired outcome of cell reprogramming efforts.

### SIRTUINS IN MITOCHONDRIA DYNAMICS DURING CELL REPROGRAMMING

Mitochondrial dynamics are controlled by many cellular proteins such as fission proteins DRP1 (79), fusion proteins Mitofusins 1 and 2 (Mfn1/2) (80), and optic atrophy 1 (OPA1) proteins (81). The mitochondrial network was reported to have changed during the cell-cycle progression and mitosis processes (82). Mitochondrial distribution during mitosis acts in a critical role during asymmetric cell division in stem cells.

Mitochondrial fusion, fission, and biogenesis are linked with mitochondrial dynamics as well (83). One study showed that mitochondrial fission and fusion contributes to the maintenance of pluripotency (84). Loss of mitochondrial fusion proteins such as Mfn1/2 (85) leads to a metabolic transition by activating HIF-1 $\alpha$  signaling in iPSC reprogramming (86). Sirt1 can exert nuclear localization signals and nuclear export signals and can come and go between the cytoplasm and the nucleus (87). Sirt3 regulates the activity of both mitochondrial enzymes (88) and mitochondrial biogenesis through activation of the Peroxisome proliferator-activated receptor gamma coactivator 1-alpha (PGC1-alpha) (89). OPA1 is a GTPase anchored to the mitochondria's inner membrane and is linked to the maintenance of mitochondria crista structure and protection of cells against stimuli (90). Sirt3 has been known to bind directly to OPA1 and subsequently modulates mitochondrial dynamics (91). Sirt1 can enhance mitochondrial function by involving PI3K/Beclin 1 and mTOR signaling (92). Additionally, Sirt1 can destruct damaged mitochondria through a mitophagy process (93). Mitophagy is dependent on the activities of specific factors such as PTEN-induced putative kinase 1 (PINK1) and E3 ubiquitin ligase Parkin (94). According to genetic research, Sirtuins affect mitophagy by inhibiting mitochondrial defects in PINK1-null mutants (95). In addition, Sirt1 suppresses the activity of the HIF1-alpha (96) that inhibits mitochondrial function. Sirt3 is known as a powerful regulator of the ROS detoxification via deacetylation of Mangan-Superoxide Dismutase (MnSOD) in mitochondria (97). Sirt3 eliminates excessive ROS production through activation of a Forkhead box O3 FOXO3-alpha (98) and then regulates mitochondrial dynamics (99). Proceeding from what has been said above, Sirtuins may affect cell reprogramming efficiency through the regulation of mitochondrial dynamics including the regulation of fission proteins, the regulation of mitophagy, the modulation of mTOR signaling, and the control of ROS production (Fig. 2).



**Fig. 2.** The relationship between sirtuins and mitochondrial dynamics caused by cell reprogramming. Cell reprogramming leads to mitochondrial dynamics such as changes in fission and fusion. The mitochondrial dynamics are linked with the maintenance of the pluripotent state. Sirtuins regulates mitochondria fission by binding with fission proteins such as OPA1 proteins. Also, sirtuins promote mTOR signaling, the activity of PGC1-alpha, and ultimately eliminate ROS production during cell reprogramming. mTOR: The mechanistic target of rapamycin, OPA1: Optic atrophy 1, PGC1-alpha: Peroxisome proliferator-activated receptor gamma coactivator 1-alpha, ROS: reactive oxygen species.

## PERSPECTIVES AND CONCLUSIONS

In conclusion, cell reprogramming has limitations including genomic instability and impaired mitochondrial dynamics. Until now, the appropriate solution to overcome these limitations was not fully investigated. Sirtuins contribute to genomic stability and mitochondrial dynamics through several signaling reactions and the activation of enzymes. After examining the roles of Sirtuins, we propose further research should look at the multiple other functions of Sirtuins in cell reprogramming. We suggest investigating more advanced manipulation of Sirtuins in cell reprogramming and ultimately expect to promote more efficient and safe cell reprogramming processes and technology.

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## CONFLICTS OF INTEREST

The authors have no conflicting interests.

## REFERENCES

1. Takahashi K and Yamanaka S (2006) Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. *Cell* 126, 663-676
2. Rando TA and Chang HY (2012) Aging, rejuvenation, and epigenetic reprogramming: resetting the aging clock. *Cell* 148, 46-57
3. Yu J, Vodyanik MA, Smuga-Otto K et al (2007) Induced pluripotent stem cell lines derived from human somatic cells. *Science* 318, 1917-1920
4. Blasco MA, Serrano M and Fernandez-Capetillo O (2011) Genomic instability in iPS: time for a break. *EMBO J* 30, 991-993
5. Suva ML, Riggi N and Bernstein BE (2013) Epigenetic reprogramming in cancer. *Science* 339, 1567-1570
6. Ruiz S, Panopoulos AD, Herreras A et al (2011) A high proliferation rate is required for cell reprogramming and maintenance of human embryonic stem cell identity. *Curr Biol* 21, 45-52
7. Lopez-Otin C, Blasco MA, Partridge L, Serrano M and Kroemer G (2013) The hallmarks of aging. *Cell* 153, 1194-1217
8. Marion RM, Strati K, Li H et al (2009) Telomeres acquire embryonic stem cell characteristics in induced pluripotent stem cells. *Cell Stem Cell* 4, 141-154
9. Weissbein U, Benvenisty N and Ben-David U (2014) Quality control: Genome maintenance in pluripotent stem cells. *J Cell Biol* 204, 153-163
10. Folmes CD, Nelson TJ, Martinez-Fernandez A et al (2011) Somatic oxidative bioenergetics transitions into pluripotency-dependent glycolysis to facilitate nuclear reprogramming. *Cell Metab* 14, 264-271
11. Banito A, Rashid ST, Acosta JC et al (2009) Senescence impairs successful reprogramming to pluripotent stem cells. *Genes Dev* 23, 2134-2139
12. Kimura H, Hayashi-Takanaka Y, Stasevich TJ and Sato Y (2015) Visualizing posttranslational and epigenetic modifications of endogenous proteins in vivo. *Histochem Cell Biol* 144, 101-109
13. Xiao Y and Chen J (2013) Proteomics approaches in the identification of molecular signatures of mesenchymal stem cells. *Adv Biochem Eng Biotechnol* 129, 153-176
14. Mu WL, Wang YJ, Xu P et al (2015) Sox2 Deacetylation by Sirt1 Is Involved in Mouse Somatic Reprogramming. *Stem Cells* 33, 2135-2147
15. Mostoslavsky R, Chua KF, Lombard DB et al (2006) Genomic instability and aging-like phenotype in the absence of mammalian SIRT6. *Cell* 124, 315-329
16. Laurent LC, Ulitsky I, Slavin I et al (2011) Dynamic changes in the copy number of pluripotency and cell proliferation genes in human ESCs and iPSCs during reprogramming and time in culture. *Cell Stem Cell* 8, 106-118
17. Taapken SM, Nisler BS, Newton MA et al (2011) Karotypic abnormalities in human induced pluripotent stem cells and embryonic stem cells. *Nat Biotechnol* 29, 313-314
18. Araten DJ, Golde DW, Zhang RH et al (2005) A quantitative measurement of the human somatic mutation rate. *Cancer Res* 65, 8111-8117
19. Morin GB (1989) The human telomere terminal transferase enzyme is a ribonucleoprotein that synthesizes TTAGGG repeats. *Cell* 59, 521-529
20. Vaziri H, Chapman KB, Guigova A et al (2010) Spontaneous reversal of the developmental aging of normal human cells following transcriptional reprogramming. *Regen Med* 5, 345-363
21. Marion RM and Blasco MA (2010) Telomere rejuvenation during nuclear reprogramming. *Curr Opin Genet Dev* 20, 190-196
22. Yehezkel S, Rebibo-Sabbah A, Segev Y et al (2011) Reprogramming of telomeric regions during the generation of human induced pluripotent stem cells and subsequent differentiation into fibroblast-like derivatives. *Epigenetics* 6, 63-75
23. Pasi CE, Dereli-Oz A, Negrini S et al (2011) Genomic instability in induced stem cells. *Cell Death Differ* 18, 745-753
24. Lopez-Contreras AJ, Gutierrez-Martinez P, Specks J, Rodrigo-Perez S and Fernandez-Capetillo O (2012) An extra allele of Chk1 limits oncogene-induced replicative stress and promotes transformation. *J Exp Med* 209, 455-461
25. Ruiz S, Lopez-Contreras AJ, Gabut M et al (2015) Limiting replication stress during somatic cell reprogramming reduces genomic instability in induced pluripotent stem cells. *Nat Commun* 6, 8036
26. Mishra P and Chan DC (2014) Mitochondrial dynamics and inheritance during cell division, development and disease. *Nat Rev Mol Cell Biol* 15, 634-646
27. Ji J, Sharma V, Qi S et al (2014) Antioxidant supplementation reduces genomic aberrations in human induced

- pluripotent stem cells. *Stem Cell Reports* 2, 44-51
28. Facucho-Oliveira JM, Alderson J, Spikings EC, Egginton S and St John JC (2007) Mitochondrial DNA replication during differentiation of murine embryonic stem cells. *J Cell Sci* 120, 4025-4034
  29. Moussaieff A, Rouleau M, Kitsberg D et al (2015) Glycolysis-mediated changes in acetyl-CoA and histone acetylation control the early differentiation of embryonic stem cells. *Cell Metab* 21, 392-402
  30. Prigione A, Fauler B, Lurz R, Lehrach H and Adjaye J (2010) The senescence-related mitochondrial/oxidative stress pathway is repressed in human induced pluripotent stem cells. *Stem Cells* 28, 721-733
  31. Mandal S, Lindgren AG, Srivastava AS, Clark AT and Banerjee U (2011) Mitochondrial function controls proliferation and early differentiation potential of embryonic stem cells. *Stem Cells* 29, 486-495
  32. Sathananthan H, Pera M and Trounson A (2002) The fine structure of human embryonic stem cells. *Reprod Biomed Online* 4, 56-61
  33. Han H, Irimia M, Ross PJ et al (2013) MBNL proteins repress ES-cell-specific alternative splicing and reprogramming. *Nature* 498, 241-245
  34. Wang L, Ye X, Zhao Q et al (2014) Drp1 is dispensable for mitochondria biogenesis in induction to pluripotency but required for differentiation of embryonic stem cells. *Stem Cells Dev* 23, 2422-2434
  35. Otera H and Mihara K (2011) Molecular mechanisms and physiologic functions of mitochondrial dynamics. *J Biochem* 149, 241-251
  36. Ramanathan A and Schreiber SL (2009) Direct control of mitochondrial function by mTOR. *Proc Natl Acad Sci U S A* 106, 22229-22232
  37. Wang S, Xia P, Ye B, Huang G, Liu J and Fan Z (2013) Transient activation of autophagy via Sox2-mediated suppression of mTOR is an important early step in reprogramming to pluripotency. *Cell Stem Cell* 13, 617-625
  38. Greer EL, Dowlatshahi D, Banko MR et al (2007) An AMPK-FOXO pathway mediates longevity induced by a novel method of dietary restriction in *C. elegans*. *Curr Biol* 17, 1646-1656
  39. Wang Z, Zang C, Rosenfeld JA et al (2008) Combinatorial patterns of histone acetylations and methylations in the human genome. *Nat Genet* 40, 897-903
  40. Lee S, Park JR, Seo MS et al (2009) Histone deacetylase inhibitors decrease proliferation potential and multilineage differentiation capability of human mesenchymal stem cells. *Cell Prolif* 42, 711-720
  41. Michishita E, McCord RA, Berber E et al (2008) SIRT6 is a histone H3 lysine 9 deacetylase that modulates telomeric chromatin. *Nature* 452, 492-496
  42. Chen T, Shen L, Yu J et al (2011) Rapamycin and other longevity-promoting compounds enhance the generation of mouse induced pluripotent stem cells. *Aging Cell* 10, 908-911
  43. Huangfu D, Maehr R, Guo W et al (2008) Induction of pluripotent stem cells by defined factors is greatly improved by small-molecule compounds. *Nat Biotechnol* 26, 795-797
  44. Vaquero A, Scher M, Lee D, Erdjument-Bromage H, Tempst P and Reinberg D (2004) Human SirT1 interacts with histone H1 and promotes formation of facultative heterochromatin. *Mol Cell* 16, 93-105
  45. Zhang ZN, Chung SK, Xu Z and Xu Y (2014) Oct4 maintains the pluripotency of human embryonic stem cells by inactivating p53 through Sirt1-mediated deacetylation. *Stem Cells* 32, 157-165
  46. Etchegaray JP, Chavez L, Huang Y et al (2015) The histone deacetylase SIRT6 controls embryonic stem cell fate via TET-mediated production of 5-hydroxymethylcytosine. *Nat Cell Biol* 17, 545-557
  47. Peng CH, Cherng JY, Chiou GY et al (2011) Delivery of Oct4 and SirT1 with cationic polyurethanes-short branch PEI to aged retinal pigment epithelium. *Biomaterials* 32, 9077-9088
  48. Liu PY, Xu N, Malyukova A et al (2013) The histone deacetylase SIRT2 stabilizes Myc oncoproteins. *Cell Death Differ* 20, 503-514
  49. Mao B, Zhao G, Lv X et al (2011) Sirt1 deacetylates c-Myc and promotes c-Myc/Max association. *Int J Biochem Cell Biol* 43, 1573-1581
  50. Vaziri H, Dessain SK, Ng Eaton E et al (2001) hSIR2(SIRT1) functions as an NAD-dependent p53 deacetylase. *Cell* 107, 149-159
  51. Han MK, Song EK, Guo Y, Ou X, Mantel C and Broxmeyer HE (2008) SIRT1 regulates apoptosis and Nanog expression in mouse embryonic stem cells by controlling p53 subcellular localization. *Cell Stem Cell* 2, 241-251
  52. Katada S, Imhof A and Sassone-Corsi P (2012) Connecting threads: epigenetics and metabolism. *Cell* 148, 24-28
  53. Ryall JG, Dell'Orso S, Derfoul A et al (2015) The NAD(+) dependent SIRT1 deacetylase translates a metabolic switch into regulatory epigenetics in skeletal muscle stem cells. *Cell Stem Cell* 16, 171-183
  54. Zwaans BM and Lombard DB (2014) Interplay between sirtuins, MYC and hypoxia-inducible factor in cancer-associated metabolic reprogramming. *Dis Model Mech* 7, 1023-1032
  55. Tennen RI, Bua DJ, Wright WE and Chua KF (2011) SIRT6 is required for maintenance of telomere position effect in human cells. *Nat Commun* 2, 433
  56. Kugel S and Mostoslavsky R (2014) Chromatin and beyond: the multitasking roles for SIRT6. *Trends Biochem Sci* 39, 72-81
  57. Michishita E, McCord RA, Boxer LD et al (2009) Cell cycle-dependent deacetylation of telomeric histone H3 lysine K56 by human SIRT6. *Cell Cycle* 8, 2664-2666
  58. Toiber D, Erdel F, Bouazoune K et al (2013) SIRT6 recruits SNF2H to DNA break sites, preventing genomic instability through chromatin remodeling. *Mol Cell* 51, 454-468
  59. Galleano I, Schiedel M, Jung M, Madsen AS and Olsen CA (2016) A Continuous, Fluorogenic Sirtuin 2 Deacetylase Assay: Substrate Screening and Inhibitor Evaluation. *J Med Chem* 59, 1021-1031
  60. Baur JA (2010) Biochemical effects of SIRT1 activators. *Biochim Biophys Acta* 1804, 1626-1634
  61. Gu XS, Wang ZB, Ye Z et al (2014) Resveratrol, an activator of SIRT1, upregulates AMPK and improves

- cardiac function in heart failure. *Genet Mol Res* 13, 323-335
62. Chaudhary N and Pfluger PT (2009) Metabolic benefits from Sirt1 and Sirt1 activators. *Curr Opin Clin Nutr Metab Care* 12, 431-437
  63. Bruckbauer A, Zemel MB, Thorpe T et al (2012) Synergistic effects of leucine and resveratrol on insulin sensitivity and fat metabolism in adipocytes and mice. *Nutr Metab (Lond)* 9, 77
  64. Chauhan D, Bandi M, Singh AV et al (2011) Preclinical evaluation of a novel SIRT1 modulator SRT1720 in multiple myeloma cells. *Br J Haematol* 155, 588-598
  65. Lahusen TJ and Deng CX (2015) SRT1720 induces lysosomal-dependent cell death of breast cancer cells. *Mol Cancer Ther* 14, 183-192
  66. Maya JD, Morello A, Repetto Y et al (2001) Trypanosoma cruzi: inhibition of parasite growth and respiration by oxazolo(thiazolo)pyridine derivatives and its relationship to redox potential and lipophilicity. *Exp Parasitol* 99, 1-6
  67. Vu CB, Bemis JE, Disch JS et al (2009) Discovery of imidazo[1,2-b]thiazole derivatives as novel SIRT1 activators. *J Med Chem* 52, 1275-1283
  68. Bonkowski MS and Sinclair DA (2016) Slowing ageing by design: the rise of NAD<sup>+</sup> and sirtuin-activating compounds. *Nat Rev Mol Cell Biol* 17, 679-690
  69. Camins A, Sureda FX, Junyent F et al (2010) Sirtuin activators: designing molecules to extend life span. *Biochim Biophys Acta* 1799, 740-749
  70. Botta G, De Santis LP and Saladino R (2012) Current advances in the synthesis and antitumoral activity of SIRT1-2 inhibitors by modulation of p53 and pro-apoptotic proteins. *Curr Med Chem* 19, 5871-5884
  71. Villalba JM and Alcain FJ (2012) Sirtuin activators and inhibitors. *Biofactors* 38, 349-359
  72. Verdin E, Hirschey MD, Finley LW and Haigis MC (2010) Sirtuin regulation of mitochondria: energy production, apoptosis, and signaling. *Trends Biochem Sci* 35, 669-675
  73. Wang TT, Schoene NW, Kim EK and Kim YS (2013) Pleiotropic effects of the sirtuin inhibitor sirtinol involves concentration-dependent modulation of multiple nuclear receptor-mediated pathways in androgen-responsive prostate cancer cell LNCaP. *Mol Carcinog* 52, 676-685
  74. Peck B, Chen CY, Ho KK et al (2010) SIRT inhibitors induce cell death and p53 acetylation through targeting both SIRT1 and SIRT2. *Mol Cancer Ther* 9, 844-855
  75. Naia L and Rego AC (2015) Sirtuins: double players in Huntington's disease. *Biochim Biophys Acta* 1852, 2183-2194
  76. Chen J, Zhou Y, Mueller-Steiner S et al (2005) SIRT1 protects against microglia-dependent amyloid-beta toxicity through inhibiting NF-kappaB signaling. *J Biol Chem* 280, 40364-40374
  77. Risitano R, Curro M, Cirmi S et al (2014) Flavonoid fraction of Bergamot juice reduces LPS-induced inflammatory response through SIRT1-mediated NF-kappaB inhibition in THP-1 monocytes. *PLoS One* 9, e107431
  78. Polyakova O, Borman S, Grimley R, Vamathevan J, Hayes B and Solari R (2012) Identification of novel interacting partners of Sirtuin6. *PLoS One* 7, e51555
  79. Mopert K, Hajek P, Frank S, Chen C, Kaufmann J and Santel A (2009) Loss of Drp1 function alters OPA1 processing and changes mitochondrial membrane organization. *Exp Cell Res* 315, 2165-2180
  80. Chen H, Detmer SA, Ewald AJ, Griffin EE, Fraser SE and Chan DC (2003) Mitofusins Mfn1 and Mfn2 coordinately regulate mitochondrial fusion and are essential for embryonic development. *J Cell Biol* 160, 189-200
  81. Chen H, Chomyn A and Chan DC (2005) Disruption of fusion results in mitochondrial heterogeneity and dysfunction. *J Biol Chem* 280, 26185-26192
  82. Christiansen EG (1949) Orientation of the mitochondria during mitosis. *Nature* 163, 361
  83. Cho DH, Nakamura T and Lipton SA (2010) Mitochondrial dynamics in cell death and neurodegeneration. *Cell Mol Life Sci* 67, 3435-3447
  84. Wilkerson DC and Sankar U (2011) Mitochondria: a sulfhydryl oxidase and fission GTPase connect mitochondrial dynamics with pluripotency in embryonic stem cells. *Int J Biochem Cell Biol* 43, 1252-1256
  85. Cipolat S, Martins de Brito O, Dal Zilio B and Scorrano L (2004) OPA1 requires mitofusin 1 to promote mitochondrial fusion. *Proc Natl Acad Sci U S A* 101, 15927-15932
  86. Son MJ, Kwon Y, Son MY et al (2015) Mitofusins deficiency elicits mitochondrial metabolic reprogramming to pluripotency. *Cell Death Differ* 22, 1957-1969
  87. Tanno M, Sakamoto J, Miura T, Shimamoto K and Horio Y (2007) Nucleocytoplasmic shuttling of the NAD<sup>+</sup>-dependent histone deacetylase SIRT1. *J Biol Chem* 282, 6823-6832
  88. Bell EL and Guarente L (2011) The SirT3 divining rod points to oxidative stress. *Mol Cell* 42, 561-568
  89. Giralt A, Hondares E, Villena JA et al (2011) Peroxisome proliferator-activated receptor-gamma coactivator-1alpha controls transcription of the Sirt3 gene, an essential component of the thermogenic brown adipocyte phenotype. *J Biol Chem* 286, 16958-16966
  90. Olichon A, Baricault L, Gas N et al (2003) Loss of OPA1 perturbs the mitochondrial inner membrane structure and integrity, leading to cytochrome c release and apoptosis. *J Biol Chem* 278, 7743-7746
  91. Park SH, Ozden O, Jiang H et al (2011) Sirt3, mitochondrial ROS, ageing, and carcinogenesis. *Int J Mol Sci* 12, 6226-6239
  92. Ou X, Lee MR, Huang X, Messina-Graham S and Broxmeyer HE (2014) SIRT1 positively regulates autophagy and mitochondria function in embryonic stem cells under oxidative stress. *Stem Cells* 32, 1183-1194
  93. Yoshii SR and Mizushima N (2015) Autophagy machinery in the context of mammalian mitophagy. *Biochim Biophys Acta* 1853, 2797-2801
  94. Eiyama A and Okamoto K (2015) PINK1/Parkin-mediated mitophagy in mammalian cells. *Curr Opin Cell Biol* 33, 95-101
  95. Koh H, Kim H, Kim MJ, Park J, Lee HJ and Chung J (2012) Silent information regulator 2 (Sir2) and Forkhead box O (FOXO) complement mitochondrial dysfunction and dopaminergic neuron loss in Drosophila PTEN-induced kinase 1 (PINK1) null mutant. *J Biol Chem* 287, 12750-12758



96. Lim JH, Lee YM, Chun YS, Chen J, Kim JE and Park JW (2010) Sirtuin 1 modulates cellular responses to hypoxia by deacetylating hypoxia-inducible factor 1alpha. *Mol Cell* 38, 864-878
97. Qiu X, Brown K, Hirschey MD, Verdin E and Chen D (2010) Calorie restriction reduces oxidative stress by SIRT3-mediated SOD2 activation. *Cell Metab* 12, 662-667
98. Sundaresan NR, Gupta M, Kim G, Rajamohan SB, Isbatan A and Gupta MP (2009) Sirt3 blocks the cardiac hypertrophic response by augmenting Foxo3a-dependent antioxidant defense mechanisms in mice. *J Clin Invest* 119, 2758-2771
99. Morigi M, Perico L, Rota C et al (2015) Sirtuin 3-dependent mitochondrial dynamic improvements protect against acute kidney injury. *J Clin Invest* 125, 715-726