



POSTER PRESENTATION

Open Access

Optimization and validation of the HIV-1 neutralizing antibody assay in A3R5 cells

M Sarzotti-Kelsoe^{1*}, X Daniell¹, CA Todd¹, M Bilaska¹, C LaBranche¹, LG Perez¹, C Ochsenbauer², J Kappes², W Rountree¹, DA Ozaki¹, JH Kim³, R McLinden³, T Denny¹, DC Montefiori¹

From AIDS Vaccine 2012
Boston, MA, USA. 9-12 September 2012

Background

A3R5 is a highly sensitive cell line for the detection of neutralizing antibodies (Nabs) against tier 2 strains of HIV-1. This cell line is particularly useful for the detection of weak Nab responses in preclinical and clinical trials of candidate HIV-1 vaccines. All methods used for endpoint analyses in clinical trials should be validated and demonstrably fit for purpose, in compliance with ICH Q2 (R1) guidelines. Here we describe the optimization/qualification and validation of the HIV-1 Nab Assay in A3R5 cells.

Methods

A3R5 is a human lymphoblastoid cell line naturally expressing CD4 and CXCR4 and engineered to express CCR5. Nab assays in A3R5 cells are performed with Env. IMC.LucR viruses containing a reporter gene in the viral genome, whose expression is induced by viral Tat protein soon after infection. Luciferase activity is quantified by luminescence and is directly proportional to the number of infectious virus particles present in the viral inoculum. The assay is performed in 96-well culture plates for high throughput capacity.

Results

We determined the stability of the cell line over time in culture for receptor and coreceptor expression, susceptibility to infection, and sensitivity to neutralization. The assay was optimized for cell density, input virus dose, length of incubation time and use of DEAE-dextran. We also determined the stability of a set of reference reagents, for validation experiments and for future competency and proficiency testing that express a broad spectrum of neutralization phenotypes. A prospective validation plan with

pre-set pass/fail criteria was composed and implemented that addressed key assay parameters, including accuracy, precision, limit of detection and quantitation, specificity, linearity and range, robustness and specificity. Results of validation experiments were statistically analyzed and used to generate a final validation document.

Conclusion

This validated assay will be used to identify correlates of protection in HIV vaccine trials conducted globally.

Author details

¹Duke University Medical Center, Durham, NC, USA. ²University of Alabama, Birmingham, AL, USA. ³U.S. Military HIV Research Program, WRAIR, Silver Spring, MD, USA.

Published: 13 September 2012

doi:10.1186/1742-4690-9-S2-P69

Cite this article as: Sarzotti-Kelsoe *et al.*: Optimization and validation of the HIV-1 neutralizing antibody assay in A3R5 cells. *Retrovirology* 2012 9(Suppl 2):P69.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit



¹Duke University Medical Center, Durham, NC, USA
Full list of author information is available at the end of the article