



Current Clinical Trials in Pemphigus and Pemphigoid

Kentaro Izumi^{1,2}, Katja Bieber² and Ralf J. Ludwig^{2*}

¹ Department of Dermatology, Hokkaido University Graduate School of Medicine, Sapporo, Japan, ² Lübeck Institute of Experimental Dermatology, Center for Research on Inflammation of the Skin, University of Lübeck, Lübeck, Germany

Autoimmune bullous dermatoses (AIBDs) are a group of rare chronic inflammatory skin diseases, which clinically manifest as blisters and erosions of the skin and/or mucosa. Immunologically, AIBDs are characterized and caused by autoantibodies targeting adhesion molecules in the skin and mucosa. According to the histological location of the blistering, AIBDs are classified into the following two main subtypes: pemphigus (intraepidermal blistering) and pemphigoid (subepidermal blistering). Most AIBDs were potentially life-threatening diseases before the advent of immunosuppressive drugs, especially systemic steroid therapies, which suppress pathogenic immunological activity. Although there have been recent advancements in the understanding of the pathogenesis of AIBDs, glucocorticosteroids and/or adjuvant immunosuppressive drugs are still needed to control disease activity. However, the long-term use of systemic immunosuppression is associated with major adverse events, including death. Based on the growing understanding of AIBD pathogenesis, novel treatment targets have emerged, some of which are currently being evaluated in clinical trials. Within this article, we review the current clinical trials involving pemphigus and pemphigoid and discuss the rationale that lead to these trials. Overall, we aim to foster insights into translational research in AIBDs to improve patient care.

Keywords: autoimmune blistering skin disease, pemphigus, pemphigoid, clinical trial, treatment

INTRODUCTION

Autoimmune bullous dermatoses (AIBDs) are a heterogeneous group of skin diseases that are characterized and caused by autoantibodies targeting adhesion molecules in the skin and/or mucous membranes. Depending on the targeted adhesion molecules and the location of the blistering, AIBDs are classified into the following two major types: pemphigus diseases with autoantibodies targeting desmosomal proteins (1) and pemphigoid diseases with autoantibodies targeting the structural proteins of dermal-epidermal junction (2). In pemphigus diseases, including pemphigus vulgaris (PV) and pemphigus foliaceus (PF), autoantibody binding leads to the disruption of epidermal adhesion, resulting in the clinical finding of flaccid blisters and the histological finding of intraepidermal blistering (1). In pemphigoid diseases, the linear deposition of autoantibodies along the dermal-epidermal junction causes subepidermal blistering, resulting in tense blisters. Based on the target molecules of the autoantibodies and the clinical manifestations, pemphigoid diseases are classified as bullous pemphigoid (BP), mucous membrane pemphigoid (MMP), epidermolysis bullosa acquisita (EBA), anti-laminin-γ1/p200 pemphigoid (p200), pemphigoid gestationis (PG), lichen planus pemphigoides (LPP), linear IgA bullous dermatosis (LAD), and dermatitis herpetiformis (DH), which is associated with gluten-sensitive enteropathy and characteristic granular IgA deposits in the upper dermis (3–5).

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> *Correspondence: Ralf J. Ludwig ralf.ludwig@uksh.de

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Although the advent of systemic steroid therapy significantly improved the prognosis of AIBDs, these groups of diseases are still potentially life-threatening, now mainly due to adverse events resulting from corticosteroid treatment (6). Due to the chronicity of AIBDs, the prolonged administration of systemic steroids is often needed to induce and maintain clinical remission, leading to various adverse effects such as cytopenia, diabetes mellitus, osteoporosis, hypertension, gastrointestinal ulcers, and infections due to immunosuppression (7). Furthermore, severe infection induced by an immunocompromised state is one of the most important causes of death during AIBD treatment (8, 9). Thus, the development of alternative treatment modalities that have fewer adverse events is urgently needed for the treatment of AIBD patients.

Based on the growing understanding of AIBD pathogenesis (1, 10, 11), novel therapeutic targets and/or treatment modalities have been identified (12-15). Some of those new treatments are currently being evaluated in clinical trials. To foster translational AIBD research, in this article, we summarize the current clinical trials involving pemphigus and pemphigoid diseases. For this purpose, we searched clinicaltrials.gov¹ and the EU clinical trials register² through December 2018 and selected clinical trials on pemphigus or pemphigoid disease with the status "recruiting (the study is currently recruiting participants)," "active, not recruiting (the study is ongoing, and participants are receiving an intervention or being examined, but not currently being recruited or enrolled)," and "completed (the study has ended normally, and participants are no longer being examined or treated)," and an initiation in 2013 or later (Tables 1, 2, Figure 1). To provide a more comprehensive overview, we list the trials before this time frame in Supplementary Tables 1, 2. In addition, we included one clinical trial that is registered with the Australia and New Zealand Clinical Trials Registry, which was recently presented at the 5th International Pemphigus and Pemphigoid Foundation Scientific Conference in Orlando (16).

PEMPHIGUS

Anti-CD20 in Pemphigus (Rituximab or Ofatumumab)

Rituximab is a human chimeric IgG1 monoclonal antibody targeting CD20, which is a cell surface marker expressed by B cells (17). Rituximab exerts its treatment effects via the depletion of B-cells following its binding to CD20. Regarding the treatment of pemphigus with rituximab, case reports have suggested the efficacy of rituximab as a second- or third-line therapy (18–20). There are two main rituximab regimens for pemphigus; one regimen, which is based on the lymphoma protocol, is composed of a total of 4 doses of 375 mg/m² weekly infusions (21), while the other regimen, which is based on the rheumatoid arthritis protocol, consists of a total of two doses of 1,000 mg (or 500 mg) biweekly intravenous infusions (22). Prior to evaluating the efficacy of a single cycle regimen of rituximab in refractory

pemphigus cases, a multicenter, single arm, phase 2/3 clinical trial was conducted in France (23). At 3 months after rituximab treatment, 18 (86%) of 21 cases achieved complete remission (CR). Furthermore, 18 (86%) of 21 cases maintained CR after a median follow-up of 34 months. In addition, 8 of these 18 cases did not receive systemic corticosteroid therapy. To evaluate the safety and efficacy of rituximab in a controlled clinical trial, a prospective, parallel-group, open-label, randomized, phase 3 clinical trial of rituximab as a first-line treatment for moderate to severe cases of pemphigus (NCT00784589) was initiated (Table 1). Patients with PV or PF were randomized to receive either oral prednisolone at 1.0 to 1.5 mg/kg/day that was tapered over 12-18 months or oral prednisolone at 0.5-1.0 mg/kg/day that was tapered over 3-6 months plus rituximab 1 g on days 0 and 14 and 0.5 g at months 12 and 18. The primary endpoint was the proportion of patients who achieved CR off-therapy at month 24. The results of this clinical trial were recently reported (24). Regarding the primary endpoint, 34% of patients in the prednisolone arm achieved CR at month 24. In the rituximab plus prednisolone arm, CR was reached by 89% of the patients; the difference was significant. Furthermore, fewer grade 3-4 adverse events were observed in the rituximab plus prednisone group than in the prednisone alone group (27 events in 16 out of 46 patients; mean 0.59 [SD 1.15] vs. 53 events in 29 out of 44 patients; mean 1.20 [1.25]). Overall, this trial demonstrated that the first-line treatment of pemphigus with rituximab and lower doses of prednisolone is more effective and safer compared to high-dose prednisolone treatment. Based on the results of this clinical trial, the Food and Drug Administration approved the expansion of health insurance coverage of rituximab for pemphigus vulgaris in the United States.

In addition to rituximab, ofatumumab, a fully human anti-CD20 monoclonal antibody, has been demonstrated to be safe and effective for the treatment of autoimmune disorders other than pemphigus or pemphigoid (25). Preclinical studies suggested that ofatumumab shows a high affinity for CD20 and activates complement-dependent cytotoxicity (26). A double-blind, randomized, placebocontrolled, phase 3 clinical trial evaluating the efficacy of ofatumumab in pemphigus was completed in January 2018 (**NCT01920477**). This clinical trial enrolled moderate

¹https://clinicaltrials.gov/

²https://www.clinicaltrialsregister.eu/

Abbreviations: AIBDs, autoimmune bullous dermatoses; PV, pemphigus vulgaris; PF, pemphigus foliaceus; BP, bullous pemphigoid; EBA, epidermolysis acquisita; p200, anti-laminin-y1/p200 pemphigoid; PG, pemphigoid gestationis; LPP, lichen planus pemphigoides; LAD, linear IgA bullous dermatosis; DH, dermatitis herpetiformis; RA, rheumatoid arthritis; MS, multiple sclerosis; CR, complete remission; MMF, mycophenolate mofetil; BAFF, B cell activating factor of the tumor necrosis factor family; BAFF-R, B cell activating factor of the tumor necrosis factor family receptor; BCR, B cell receptor; PDAI, Pemphigus Disease Area Index; FcRn, neonatal Fc receptor; IVIg, intravenous immunoglobulin; BTK, Bruton's tyrosine kinase; Tregs, regulatory T cells; GVHD, graft vs. host disease; Dsg, desmoglein; PRP, Platelet-rich plasma; CAR, chimeric antigen receptor; Dsg3-CAART, Dsg3 chimeric autoantibody receptor T cell; IL, interleukin; EAE, experimental autoimmune encephalomyelitis; DAS, disease activity score; NLRP3, nucleotide-binding domain, leucine-rich repeat family, pyrin domain-containing 3; BPDAI, bullous pemphigoid disease area index; ECP, eosinophil cationic protein; SYK, spleen tyrosine kinase; FcyR, Fc gamma receptor; LTB4, leukotriene B4.

NCT number	Disease	Interventions	Target	Allocation	Masking	Phase	Status
NCT01930175	PV	VAY736	BAFF-R	Randomized	Double blind	2	Active, not recruiting
NCT01920477	PV	Ofatumumab	CD20	Randomized	Double blind	3	Completed
NCT03334058	PV	ARGX-113	FcRn	Single group	None	2	Recruiting
NCT02704429	PV or PF	PRN1008	BTK	Single group	None	2	Recruiting
NCT03762265	PV or PF	PRN1008	BTK	Randomized	Quadruple blind	3	Recruiting
NCT02383589	PV	Rituximab MMF	CD20 IMPDH	Randomized	Double blind	3	Active, not recruiting
NCT03239470	PV or PF	Poly Tregs	Immune tolerance	Non- randomized	None	1	Recruiting
NCT02828163	PV	PRP	Wound healing	Randomized	Double blind	3	Recruiting
NCT00784589	PV or PF	Rituximab	CD20	Randomized	None	3	Completed
NCT03075904	PV or PF	SYNT001	FcRn	Non-randomized	None	1/2	Completed

TABLE 1 | Current clinical trials in pemphigus.

PV, pemphigus vulgaris; PF, pemphigus foliaceus; BAFF-R, B cell activating factor of the tumor necrosis factor family receptor; FcRn, neonatal Fc receptor; BTK, Bruton's tyrosine kinase; MMF, mycophenolate mofetil; IMPDH, inosine 5[']-monophosphate dehydrogenase; Tregs, regulatory T cells; PRP, Platelet rich plasma.

TABLE 2	Current c	linical trials	in bullous	pemphigoid.

NCT number	Interventions	Target	Allocation	Masking	Phase	Status
NCT03099538	Ixekizumab	IL-17	Single group	None	2	Recruiting
NCT01408550	NPB-1	FcRn	Randomized	Double blind	3	Completed
NCT03286582	AC-203	Inflammasome	Randomized	None	2	Recruiting
NCT02226146	Bertilimumab	eotaxin	Single group	None	2	Completed
NCT00525616	Rituximab	CD20	Single group	None	3	Completed

No clinical trials are being performed for any other pemphigoid diseases. FcRn, neonatal Fc receptor.

to severe PV patients with a history of at least failure of tapered steroid therapy. Participants received a stable dose of prednisone/prednisolone from a minimum of 20 mg/day up to a maximum of 120 mg/day or 1.5 mg/kg/day for 2 weeks prior to randomization. Thirty-five patients with PV were randomized to receive either of atumumab or a placebo. Of atumumab at 40 mg was subcutaneously injected at week 0 and week 4. From week 8, subjects were subcutaneously administered of atumumab 20 mg every 4 weeks through week 56. The primary endpoints were the time to sustained CR on minimal steroid therapy (prednisone/prednisolone dose to <10 mg/day) and the duration of CR on minimal steroid therapy. The results of this study have thus far not been reported.

Similar to these two trials, another double-blind, randomized, phase 3 clinical trial of adjuvant rituximab vs. mycophenolate mofetil (MMF) as a therapy for pemphigus is ongoing (NCT02383589). In that study, 135 PV patients who are receiving standard systemic steroid treatment (oral prednisone 60–120 mg/day or equivalent) will randomly receive either (i) MMF and a rituximab-matched placebo or (ii) an MMF-matched placebo and rituximab. MMF is orally administered twice daily from day 1 to week 56. The initial dose of MMF is 500 mg, and the dose will be titrated to achieve a goal of 1,000 mg. Rituximab is administered at a dose of 1,000 mg intravenously on days 1, 15, 168, and 182. The primary endpoint is the proportion of patients who achieve sustained CR.

VAY736 in Pemphigus

B cell activating factor of the tumor necrosis factor family (BAFF) is a crucial cytokine for regulating B cell development in mice and humans (27, 28). BAFF functions by binding to the BAFF receptor (BAFF-R), B cell maturation antigen, and tumor necrosis factor receptor superfamily member (29). Although at physiological concentrations BAFF cannot rescue B cell apoptosis due to a strong B cell death signal, which is transduced via the B cell receptor (BCR) stimulated by autoantigens, at higher concentrations, BAFF causes the survival of autoreactive B cells, which contributes to the pathogenesis of autoimmune diseases (27, 30, 31). For example, elevated BAFF serum levels have been detected in various autoimmune diseases such as RA, systemic lupus erythematosus, Sjögren's syndrome, and systemic sclerosis (32-34). Taken together, BAFF is a likely therapeutic target for the treatment of autoimmune diseases. Regarding clinical translation, the anti-BAFF antibody belimumab was licensed for the treatment of systemic lupus erythematosus in 2011 (35). However, clinical trials of belimumab have not been conducted in pemphigus thus far. Alternatively, with regard to the inhibition of BAFF, its receptor can be blocked to achieve similar results. BAFF-R signaling drives B cell differentiation, proliferation and survival (36). VAY736 is a novel, defucosylated, human IgG1 monoclonal antibody targeting BAFF-R, providing both enhanced antibody-dependent cellular cytotoxicity-mediated depletion of B cells and the blockade of BAFF. To investigate the safety, tolerability and efficacy of VAY736 in PV, a randomized, placebo-controlled, double-blind,



FIGURE 1 Schematic representation of the pathophysiology and new therapeutic targets of pemphigus and pemphigoid diseases. The pathophysiology of pemphigus and pemphigoid diseases consists of the following three phases: (1) CD4+T cells promote autoreactive B cell activation, proliferation, and differentiation to plasma cells that produce pathogenic autoantibodies. (2) Circulating pathogenic antibodies are transferred to the dermal epidermal junction or intracellular space of the epidermis. Neonatal Fc receptor (FcRn) plays a role in prolonging the half-life of IgG antibodies during this phase. (3) After the binding of pathogenic autoantibodies to target molecules, pro-inflammatory cells such as granulocytes and macrophages are recruited to the immune complex in lesional skin by chemokines (e.g., eotaxin). Then, granulocytes elicit reactive oxygen species (ROS), elastases, and proteases, resulting in tissue damage such as blisters and/or erythema, which are clinical symptoms in pemphigoid diseases but not in pemphigus diseases. Cytokines [e.g., interleukin (IL)-1beta and IL-18] and Th17 polarization are thought to enhance local inflammation. During the antibody production phase, rituximab and ofatumumab deplete autoreactive B cells to prevent their differentiation to plasma cells. PolyTregs act on CD4+T cells, and VAY736 and PRN1008 act on autoreactive B cells, resulting in less activation of autoreactive B cells. In the transcytosis phase, intravenous immunoglobulin (IVIg), SYNT001, and ARGX-113 saturate FcRn, contributing to the shortened half-life of pathogenic autoantibodies. In the effector phase, ixekizumab restores Th17 polarization and suppresses inflammatory augmentation. AC-203 modulates cytokines such as IL-1beta and IL-18, contributing to decreased inflammation. The inhibition of eotaxin with bertilimumab ameliorates the recruitment of eosinophils to local inflammation sites in pemphigoid disease, especially bullous pemphigoid. Platelet-rich plasma (PRP) is thought to promote wound healing in erosi

phase 2 clinical trial is currently ongoing (**NCT01930175**). In this trial, 16 mild-moderate PV patients will randomly receive either intravenous VAY736 or a placebo once. The primary endpoint is the efficacy of single cycle VAY736 administration in reducing Pemphigus Disease Area Index (PDAI) scores at week 12 compared to at the baseline. No results of this trial are currently available.

Anti-neonatal Fc Receptor (FcRn) in Pemphigus (SYNT001, ARGX-113)

FcRn plays an essential role in regulating host circulating IgG levels (37). FcRn protects IgG from intracellular digestion, leading to the prolongation of its half-life. This "IgG recycling system" is essential for host defense. However, it also maintains the concentration of circulating pathogenic IgG in various autoimmune diseases, including AIBD. Specifically, FcRndeficient mice do not (or to a lesser extent) develop experimental AIBD after the injection of AIBD-inducing antibodies (10). Interestingly, saturation of the FcRn by the administration of high-dose human IgG (IVIg) reduces the pathogenic effects in antibody-transfer AIBD models. Thus, FcRn inhibition is one possible mode of action of IVIg therapy in AIBD (38, 39). Based on these observations, anti-FcRn targeting treatments have been developed. Currently, the following 2 clinical trials targeting FcRn are being conducted in pemphigus: SYNT001 (NCT03075904) and ARGX-113 (NCT03334058). SYNT001 is a humanized, deimmunized IgG4 monoclonal antibody that blocks the binding of FcRn to the Fc portion of IgG (40). ARGX-113 is a human IgG1-derived Fc-modified fragment with increased affinity for FcRn that reduces the circulating IgG concentration (41). In addition, both SYNT001 and ARGX-113 do not alter serum levels of albumin. As both trials are ongoing, no results have been published so far. Regarding NCT03075904, 16 PV or PF patients will be sequentially assigned to receive three different doses of intravenous SYNT001 weekly for either 5 or 14 weeks, and the primary endpoint is the count and percentage of adverse events (time frame; days 0-112 or days 0-175). At the 2018 pre-International Investigative Dermatology meeting in Orlando, the first results were reported for SYNT001. The infusion of SYNT001 in human subjects resulted in a rapid lowering of the circulating levels of IgG (mean total IgG reduction of 56% by day 30) with good safety and tolerability. Furthermore, 5 of the 7 subjects showed a reduction in disease activity by day 42 (42). Regarding NCT03334058, 12 newly diagnosed or relapsed PV patients will receive ARGX-113 intravenously. The primary endpoints are safety and tolerability up to 17 weeks.

PRN1008 in Pemphigus

B cell receptor signaling is a key player in B cell development and function. Bruton's tyrosine kinase (BTK) belongs to the Tec family of nonreceptor tyrosine kinases, and it is a vital component of B cell receptor signaling (43). BTK is predominantly expressed by B-lymphocytes from the pre-B cell stage to the mature B cell stage (44, 45). Based on the crucial role of BTK in B cell function, it has been identified as a potential target for the treatment of autoimmune disorders. Several studies have shown that ibrutinib, which is one of the BTK inhibitors under development, binds to BTK with high affinity, leading to the inhibition of B cell receptor signaling and resulting in the reduction of B cell activation involved in autoimmunity (46). In pemphigus treatment, a previous case report showed that ibrutinib improved clinical cutaneous lesions of paraneoplastic pemphigus complicated with chronic lymphocytic leukemia (47). However, there have been no case reports or series investigating the effects of this BTK inhibitor in PNP, PV and/or PF. PRN1008 is another BTK inhibitor that was evaluated in a phase 1 clinical trial that enrolled 80 healthy volunteers (Australian New Zealand Clinical Trials Registry No. ACTRN12614000359639). In that study, PRN1008 was considered safe. Regarding the pharmacokinetics and pharmacodynamics, BTK occupancy of more than 90% was observed within 4h after dosing in both the single and multiple dose regimens and was closely correlated with the maximum plasma concentration (48). Based on the promising results from the phase 1 trial, an open-label, singlearmed, phase 2 clinical trial of PRN1008 for PV treatment has been conducted (NCT02704429). In this trial, 27 pemphigus patients (including PV and PF) received PRN1008 orally for 12 weeks with a 12-week follow-up period. The primary efficacy endpoint was the initial control of disease activity during the first 4 weeks of therapy, during which new lesions cease to form and existing lesions begin to heal, without the need for prednisoneequivalent corticosteroid doses >0.5 mg/kg/day. More than 50% of patients have achieved control of disease activity within 4 weeks of starting PRN1008 thus far³. Furthermore, Principia has recently initiated a global, randomized, double-blind, placebocontrolled, pivotal phase 3 clinical trial (NCT03762265) in approximately 120 PV or PF patients to evaluate PRN1008 vs. a placebo. The primary endpoint is the proportion of participants who are in CR from week \leq 29 to 37 with a prednisone dose of \leq 5 mg/day.

Polyclonal Regulatory T Cells (PolyTregs) in Pemphigus

Among the T cell subtypes, regulatory T cells (Tregs) play an important role in regulating the immune system and preventing autoimmune disease development. Based on findings in animal models, including AIBD (49, 50), it is hypothesized that naturally occurring Tregs may be utilized for the treatment of autoimmune diseases and potentially replace the use of chronic immunosuppressive therapies that are associated with multiple adverse effects. A clinical trial of Treg adoptive therapy was started by treating graft vs. host diseases (GVHD) with expanded allogeneic Tregs (51). There has been a small study demonstrating the safe administration of autologous Tregs with decreased disease activity in patients with insulin-dependent diabetes (52). Subsequently, Brunstein et al. reported that HLA-matched umbilical cord blood-derived Tregs decreased the incidence of GVHD after double umbilical cord blood transplantation (53), indicating the potential efficacy and safety profile of the passive transfer of autologous Tregs in humans. The application of Tregs for lupus, cancer and organ transplantation

³https://ir.principiabio.com/news-releases/news-release-details/principiabiopharma-reports-positive-prn1008-phase-2-top-line

has been addressed (54). The suppressive effects of Tregs have been reported by studies using an active PV mouse model (55). A recent study indicated that Treg induction via the anti-CD28 antibody reduces pathogenic IgG directing desmoglein (Dsg) 3 in the HLA-DRB1*04:02- transgenic PV mouse model (56). Although Tregs are expected to improve PV symptoms, there are no case reports or case series of autologous Treg injections in patients with PV to date. To evaluate the effects of Tregs on the manifestation of PV, a nonrandomized, open-label, phase 1 clinical trial is ongoing (NCT03239470). In this trial, 12 PV or PF patients will receive one infusion of autologous expanded Tregs (CD4+CD127lo/negCD25+) at one of the following doses: either 2.5 x 10^8 poly Tregs or 10×10^8 poly Tregs. The primary endpoint is the number of significant grade 3 or higher adverse events by week 52. The results from this study have thus far not been reported.

Comparison of Injections of Steroids to Autologous Platelet-Rich Plasma (PRP) in Oral Erosions in PV

Oral erosive lesions are a major hallmark of PV. These oral lesions cause severe pain, resulting in problems with eating and drinking (57). To improve oral lesions in PV, adjuvant topical or intralesional steroids are used, and based on evidence from case reports, the treatment is effective (58, 59). To date, however, no controlled clinical trial of the treatment of oral lesions in PV has been performed. PRP, which is concentrated plasma derived from autologous whole blood, is believed to promote wound healing (60). In a case report series, El-Komy et al. reported that six of seven PV patients showed improvement of their oral PDAI scores after PRP intralesional injection (61). To evaluate the effects of PRP in PV, an open-label, doseescalation, multicenter phase 1 trial using autologous PRP has been conducted in adults with active PV (NCT02828163). This clinical trial was designed to compare the effects of PRP to those of intralesional steroid injection. Eleven PV patients received a 10 mg/mL triamcinolone injection on one side of the oral mucosa and a 1-mL PRP injection on the other side every 2 weeks for 3 months. The primary endpoint was the improvement of oral PV lesions in 3 months. Nine out of 11 participants completed the protocol, and 7 (78%) of those 9 patients showed improvement in oral PDAI and/or pain scores at the PRP injection sites. Although PRP resulted in clinical improvement, as in the previous study, there were no significant differences between PRP and intralesional steroid injections (62). Thus, autologous PRP might be used for the treatment of resistant oral erosions in pemphigus patients when intralesional steroid injection is contraindicated.

Future Potential Clinical Trials in Pemphigus

B cell depletion therapy using an anti-CD20 monoclonal antibody leads to short-term remission in the majority of pemphigus patients; however, we often observe relapsing disease after treatment. Although disease remission is related to the depletion of circulating Dsg3-specific B cells, the expansion of the same pathogenic B cell clone is observed during relapsing disease (63). Therefore, to maintain complete remission in PV, targeted removal of anti-Dsg3 memory B cells is essential. Recently, chimeric antigen receptor (CAR) technology was developed and resulted in novel treatments that led to the prolonged remission of refractory B cell leukemia and lymphoma. Adapted from that strategy, Dsg3 chimeric autoantibody receptor T cell (Dsg3-CAART) therapy has been reported to result in serological and histological improvements in experimental pemphigus mice without detectable off-target toxicity (15). A phase 1 clinical trial of Dsg3-CAART in PV patients is planned to investigate its safety and therapeutic potential.

PEMPHIGOID

Ixekizumab in BP

Th17 cells were first identified by their production of interleukin (IL)-17 (64, 65). A previous study indicated that the transfer of IL-17-producing Th17 cells into healthy SJL/J mice induced experimental autoimmune encephalomyelitis an animal model of MS (66). The prominent role of the Th17 polarization of T cells, as well as that of IL-17, in inflammation and autoimmunity was recognized after this observation. In addition to EAE, the Th17-IL-17 axis is also important in the pathogenesis of RA (67-69). Notably, neutralizing anti-IL-17 antibodies significantly decreased joint inflammation, cartilage destruction, and bone erosion in a collagen-immunized arthritis mouse model. Several recent studies suggest that Th17 cells and their cytokines also play roles in the pathogenesis of AIBD, including BP (70). More specifically, Le Jan et al. reported that IL-17A expression was significantly higher in both serum and lesional skin in BP patients than in normal healthy individuals (71). Furthermore, Chakievska et al. reported that IL-17A -/- mice were protected from the development of experimental BP induced by the transfer of anti-COL17 IgG (72). Taken together, these results indicate that IL-17A is a potential target for the treatment of BP. Ixekizumab is a recombinant fully humanized monoclonal antibody targeting IL-17 and is used for the treatment of psoriasis (73). However, there are neither case reports nor clinical studies reporting the effect of ixekizumab or other IL-17-targeting compounds on BP. To determine the efficacy of ixekizumab for the treatment of BP, an open-label, single-group phase 2 clinical trial is currently ongoing (NCT03099538). In this trial, twelve BP patients will receive subcutaneous injections of 160 mg ixekizumab on day 0, and then they will receive 80 mg of ixekizumab every 2 weeks until week 12. The primary endpoint is the median time from the start of treatment to the cessation of blister formation during the 12 weeks of therapy. There is no available report of the results of this trial to date.

NPB-01 in BP Unresponsive to Corticosteroids

With regard to pemphigus, the efficacy and safety of IVIg in refractory BP patients has been suggested by case reports and case report series (74–76). Based on these observations, several guidelines and consensus statements recommend the use of IVIg for refractory BP (77–79). Recently, Sasaoka

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et al. reported that IVIg reduced serum IL-6 concentrations, circulating pathogenic IgG levels, and skin inflammation disease activity in an active BP mouse model (80). Kamaguchi et al. reported that an anti-idiotypic antibody in IVIg significantly reduced the BP180 depletion of cultured keratinocytes stimulated with BP pathogenic IgG (81). To evaluate the effectiveness of IVIg in BP, a multicenter, randomized, double-blind, placebocontrolled, phase 3 clinical trial was conducted (NCT01408550). In this trial, 56 BP patients were parallelly assigned to receive either IVIg or a placebo. The IVIg group received an intravenous drip infusion of human IgG at 400 mg/kg/day for 5 consecutive days. The primary endpoint was the disease activity score (DAS) of skin lesions on day 15. The results of that study have recently been published (82). The DAS on day 15 was evaluated as the primary endpoint. Although the DAS on day 15 in the IVIg group (19.8 \pm 22.2) was 12.5 points lower than that in the placebo group (32.3 \pm 31.5), the difference between the groups was not significant (p = 0.089). However, a post hoc analysis of covariance using the DAS on day 1 as a covariate showed a significant difference between the IVIg and placebo groups (p = 0.041). Regarding the safety of IVIg in BP, the incidence of adverse drug reactions was 37.9% (n = 11/29) in the IVIg group vs. 18.5% (n = 5/27) in the placebo group. No significant difference in the incidence of adverse drug reactions was observed between the IVIg and placebo groups (p = 0.143). No patients in either group experienced any severe adverse drug reactions. Therefore, this report suggested that IVIg is a beneficial treatment modality for refractory BP cases treated with a moderate dose of systemic steroids.

Topical AC-203 in BP

The nucleotide-binding domain, leucine-rich repeat family, pyrin domain-containing 3 (NLRP3) inflammasome regulates the activation of caspase-1 and the release of pro-inflammatory cytokines such as IL-1 β and IL-18 in macrophages (83–87). It is an essential innate immune sensor that is activated in response to various damage-associated molecular patterns (83, 84). Previous reports have suggested that dysregulation of the inflammasome attenuates several chronic diseases, including autoimmune diseases such as EAE (88) and systemic lupus erythematosus (89, 90). In addition, polymorphisms in both NLRP3 and caspase-1 recruitment domain-8 genes led to increased IL-1ß production and are related to disease susceptibility and severity in RA (91). A recent study demonstrated that the expression levels of NLRP3 and inflammasome components are significantly higher in peripheral blood mononuclear cells from BP patients than in those from healthy donors (92). Furthermore, higher NLRP3 levels were positively correlated with BP disease activity. Although the precise mechanism by which NLRP3 contributes to pathogenesis in BP is still unclear, pharmacological modulation of the inflammasome pathway could be a novel therapeutic strategy in BP. However, in a mouse model of inflammatory EBA, caspase-1/11-deficient mice developed clinical disease manifestations identical to those of the wild-type controls after the injection of autoantibodies against type VII collagen (93).

To evaluate the therapeutic efficacy of caspase-1 inhibition in BP, AC-203, which is a topical ointment formulation of a modulator of the inflammasome and IL-1 β pathways, was developed. To investigate the effects of AC-203 on BP, a randomized, open-label phase 2 clinical trial is currently conducted (**NCT03286582**). Forty BP patients received either topical AC-203 or clobetasol 0.05% topical ointment twice a day (time frame is 10 weeks). The primary endpoint is the incidence of adverse effects during the treatment period. No results of this trial are currently available.

Bertilimumab in BP

While the presence of eosinophils in the dermal infiltrate is one of the histological hallmarks of BP, their contribution to the pathogenesis of BP remains to be determined (94–97). Recent data, however, support the notion that eosinophils promote subepidermal blistering in BP (98). In line with the presence of eosinophils in the dermal infiltrate in BP, elevated expression levels of IL-5, eotaxin, and eosinophil colony-stimulating factor were detected in BP blister fluids (99). Furthermore, matrix metallopeptidase 9 is secreted by eosinophils in BP lesional skin (100), eosinophil cationic protein (ECP) is detected at the dermal-epidermal junction of lesional skin, and serum concentrations of ECP correlate with BP disease activity (101, 102). Based on these findings, eotaxin, which is released upon eosinophil activation (103), has emerged as a therapeutic target for the treatment of BP.

Bertilimumab is a fully human monoclonal antibody targeting eotaxin-1 (104). Therefore, the administration of bertilimumab is expected to impair eosinophil infiltration into the skin in BP. This hypothesis has been recently tested in an open-label, single arm, phase 2 clinical trial (NCT02226146). Eleven moderate to severe BP patients received 10 mg/kg of bertilimumab intravenously on days 0, 14, and 28 with approximately 13 weeks of followup. The primary endpoint was safety, including the incidence of adverse effects. The results of the study were presented at the 2018 American Academy of Dermatology annual meeting. Of the 11 subjects enrolled, 2 patients withdrew consent and 9 received bertilimumab. Bertilimumab was well tolerated in all 9 subjects, and no drug-associated serious adverse events were reported⁴ Preliminary analyses indicated that the subjects had an 81% decline in their Bullous Pemphigoid Disease Area Index (BPDAI) scores. Based on these results, bertilimumab has been granted fast track designation for the treatment of BP⁵.

Rituximab in BP

Rituximab has a well-documented efficacy in treating pemphigus (24). However, controlled clinical trials in pemphigoid diseases, including BP, are missing. Several case reports and case report series reported good outcomes of rituximab treatment for BP. Although the efficiency of rituximab for the treatment of BP varied among the studies, the overall CR rates were

⁴https://www.immunepharma.com/what-we-do/bertilimumab

⁵https://ir.immunepharma.com/press-releases/detail/168/bertilimumab-granted-fast-track-designation-for-the

60–70% and the relapse rates were approximately 20% in BP patients receiving rituximab therapy (105, 106). To evaluate the efficacy and safety of rituximab for the treatment of BP, an open-label, prospective, phase 3 clinical trial was conducted (**NCT00525616**). Eighteen BP patients received two intravenous injections of 1,000 mg of rituximab at 15-day intervals. The primary endpoint was clinical and biological disease control for up to 2 years. To date, no results of this trial are available.

Future Potential Clinical Trials in Pemphigoid

Spleen tyrosine kinase (SYK) is a nonreceptor cytoplasmic enzyme that is mainly expressed in hematopoietic cells and plays a pivotal role in regulating cellular responses to extracellular pathogens or antigen-immunoglobulin complexes (107). Because SYK acts downstream of activating Fc gamma receptor (FcyR), it has been considered a candidate drug target for antibodyinduced diseases, such as pemphigoid diseases. In fact, a previous study has shown that SYK inhibition led to less activation of neutrophils stimulated with immune complexes and prevented the development of skin lesions in a preclinical EBA mouse model (108). In lesional skin in pemphigoid diseases, both eicosanoid leukotriene B4 (LTB4) and complement factor C5, the precursor of anaphylatoxin C5a, are found; however, their significance in the pathogenesis of pemphigoid diseases still needs to be elucidated. A previous preclinical mouse model of pemphigoid disease indicated a critical role of LTB4 in the recruitment of polymorph nuclear cells to the dermal epidermal junction (109). It is suggested that LTB4 may closely interact with C5a in the regulation of skin inflammation. Thus, inhibiting these two factors individually or in parallel might be effective for the treatment of pemphigoid diseases. These newly identified therapeutic targets should be addressed in future clinical trials.

REFERENCES

- Kasperkiewicz M, Ellebrecht CT, Takahashi H, Yamagami J, Zillikens D, Payne AS, et al. Pemphigus. *Nat Rev Dis Primers*. (2017) 3:17026. doi: 10.1038/nrdp.2017.26
- Schmidt E, Zillikens D. Pemphigoid diseases. Lancet. (2013) 381:320–32. doi: 10.1016/S0140-6736(12)61140-4
- van Beek N, Zillikens D, Schmidt E. Diagnosis of autoimmune bullous diseases. J Dtsch Dermatol Ges. (2018) 16:1077–91. doi: 10.1111/ddg.13637
- Kasperkiewicz M, Zillikens D, Schmidt E. Pemphigoid diseases: pathogenesis, diagnosis, and treatment. *Autoimmunity*. (2012) 45:55–70. doi: 10.3109/08916934.2011.606447
- Otten JV, Hashimoto T, Hertl M, Payne AS, Sitaru C. Molecular diagnosis in autoimmune skin blistering conditions. *Curr Mol Med.* (2014) 14:69–95. doi: 10.2174/15665240113136660079
- Joly P, Roujeau JC, Benichou J, Picard C, Dreno B, Delaporte E, et al. A comparison of oral and topical corticosteroids in patients with bullous pemphigoid. N Engl J Med. (2002) 346:321–7. doi: 10.1056/NEJMoa011592
- Buchman AL. Side effects of corticosteroid therapy. J Clin Gastroenterol. (2001) 33:289–94.
- Kridin K, Sagi SZ, Bergman R. Mortality and cause of death in patients with pemphigus. *Acta Derm Venereol.* (2017) 97:607–11. doi: 10.2340/00015555-2611

CONCLUSIONS

The clinical trials discussed here, which include several trials investigating novel therapeutic targets, demonstrate that translational research in pemphigus and pemphigoid is a fast-growing field. We thus expect that several novel treatments will be shortly available for the treatment of pemphigus and pemphigoid patients. Given the high, and thus far unmet, medical need in this field (110), this is highly encouraging and will hopefully improve the quality of life of the affected patients. In addition to the compounds and targets described here, several new targets have been recently identified in preclinical model systems, such as PDE4 or PI3K δ inhibitors (111, 112). Hence, the preclinical pipeline is well developed and will contribute to the growing number of clinical trials in pemphigus and pemphigoid.

AUTHOR CONTRIBUTIONS

KI and RL wrote the manuscript. KB prepared the figure. All authors read, commented on, and approved the final version of the manuscript.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fimmu. 2019.00978/full#supplementary-material

- Cai SC, Allen JC, Lim YL, Chua SH, Tan SH, Tang MB. Mortality of bullous pemphigoid in Singapore: risk factors and causes of death in 359 patients seen at the National Skin Centre. *Br J Dermatol.* (2014) 170:1319–26. doi: 10.1111/bjd.12806
- Ludwig RJ, Vanhoorelbeke K, Leypoldt F, Kaya Z, Bieber K, McLachlan SM, et al. Mechanisms of autoantibody-induced pathology. *Front Immunol.* (2017) 8:603. doi: 10.3389/fimmu.2017.00603
- Liu Y, Li L, Xia Y. BP180 is critical in the autoimmunity of bullous pemphigoid. *Front Immunol.* (2017) 8:1752. doi: 10.3389/fimmu.2017. 01752
- Spindler V, Rotzer V, Dehner C, Kempf B, Gliem M, Radeva M, et al. Peptide-mediated desmoglein 3 crosslinking prevents pemphigus vulgaris autoantibody-induced skin blistering. J Clin Invest. (2013) 123:800–11. doi: 10.1172/JCI60139
- Dehner C, Rotzer V, Waschke J, Spindler V. A desmoplakin point mutation with enhanced keratin association ameliorates pemphigus vulgaris autoantibody-mediated loss of cell cohesion. *Am J Pathol.* (2014) 184:2528– 36. doi: 10.1016/j.ajpath.2014.05.016
- Mao X, Cho MJT, Ellebrecht CT, Mukherjee EM, Payne AS. Stat3 regulates desmoglein 3 transcription in epithelial keratinocytes. *JCI Insight*. (2017) 2:92253. doi: 10.1172/jci.insight.92253
- 15. Ellebrecht CT, Bhoj VG, Nace A, Choi EJ, Mao X, Cho MJ, et al. Reengineering chimeric antigen receptor T cells for targeted

therapy of autoimmune disease. *Science*. (2016) 353:179–84. doi: 10.1126/science.aaf6756

- Lee J, Werth VP, Hall RP 3rd, Eming R, Fairley JA, Fajgenbaum DC, et al. Perspective from the 5th international pemphigus and pemphigoid foundation scientific conference. *Front Med.* (2018) 5:306. doi: 10.3389/fmed.2018.00306
- Coiffier B, Haioun C, Ketterer N, Engert A, Tilly H, Ma D, et al. Rituximab (anti-CD20 monoclonal antibody) for the treatment of patients with relapsing or refractory aggressive lymphoma: a multicenter phase II study. *Blood.* (1998) 92:1927–32.
- Herrmann G, Hunzelmann N, Engert A. Treatment of pemphigus vulgaris with anti-CD20 monoclonal antibody (rituximab). Br J Dermatol. (2003) 148:602–3. doi: 10.1046/j.1365-2133.2003.05209_10.x
- Cooper HL, Healy E, Theaker JM, Friedmann PS. Treatment of resistant pemphigus vulgaris with an anti-CD20 monoclonal antibody (Rituximab). *Clin Exp Dermatol.* (2003) 28:366–8. doi: 10.1046/j.1365-2230.2003.01283.x
- Goebeler M, Herzog S, Brocker EB, Zillikens D. Rapid response of treatmentresistant pemphigus foliaceus to the anti-CD20 antibody rituximab. Br J Dermatol. (2003) 149:899–901. doi: 10.1046/j.1365-2133.2003.05580.x
- Salopek TG, Logsetty S, Tredget EE. Anti-CD20 chimeric monoclonal antibody (rituximab) for the treatment of recalcitrant, life-threatening pemphigus vulgaris with implications in the pathogenesis of the disorder. *J Am Acad Dermatol.* (2002) 47:785–8. doi: 10.1067/mjd.2002.126273
- 22. Leshem YA, Hodak E, David M, Anhalt GJ, Mimouni D. Successful treatment of pemphigus with biweekly 1-g infusions of rituximab: a retrospective study of 47 patients. *J Am Acad Dermatol.* (2013) 68:404–11. doi: 10.1016/j.jaad.2012.08.010
- Joly P, Mouquet H, Roujeau JC, D'Incan M, Gilbert D, Jacquot S, et al. A single cycle of rituximab for the treatment of severe pemphigus. N Engl J Med. (2007) 357:545–52. doi: 10.1056/NEJMoa067752
- Joly P, Maho-Vaillant M, Prost-Squarcioni C, Hebert V, Houivet E, Calbo S, et al. First-line rituximab combined with short-term prednisone versus prednisone alone for the treatment of pemphigus (Ritux 3): a prospective, multicentre, parallel-group, open-label randomised trial. *Lancet.* (2017) 389:2031–40. doi: 10.1016/S0140-6736(17)30070-3
- Castillo J, Milani C, Mendez-Allwood D. Ofatumumab, a second-generation anti-CD20 monoclonal antibody, for the treatment of lymphoproliferative and autoimmune disorders. *Expert Opin Investig Drugs*. (2009) 18:491–500. doi: 10.1517/13543780902832679
- 26. Lin TS. Ofatumumab: a novel monoclonal anti-CD20 antibody. *Pharmgenomics Pers Med.* (2010) 3:51–9.
- 27. Mackay F, Browning JL. BAFF: a fundamental survival factor for B cells. *Nat Rev Immunol.* (2002) 2:465–75. doi: 10.1038/nri844
- Ng LG, Sutherland AP, Newton R, Qian F, Cachero TG, Scott ML, et al. B cell-activating factor belonging to the TNF family (BAFF)-R is the principal BAFF receptor facilitating BAFF costimulation of circulating T and B cells. J Immunol. (2004) 173:807–17. doi: 10.4049/jimmunol.173.2.807
- Salzer U, Jennings S, Grimbacher B. To switch or not to switch-the opposing roles of TACI in terminal B cell differentiation. *Eur J Immunol.* (2007) 37:17–20. doi: 10.1002/eji.200636914
- Lesley R, Xu Y, Kalled SL, Hess DM, Schwab SR, Shu HB, et al. Reduced competitiveness of autoantigen-engaged B cells due to increased dependence on BAFF. *Immunity*. (2004) 20:441–453. doi: 10.1016/S1074-7613(04)00079-2
- Mackay F, Woodcock SA, Lawton P, Ambrose C, Baetscher M, Schneider P, et al. Mice transgenic for BAFF develop lymphocytic disorders along with autoimmune manifestations. *J Exp Med.* (1999) 190:1697–710.
- Matsushita T, Hasegawa M, Matsushita Y, Echigo T, Wayaku T, Horikawa M, et al. Elevated serum BAFF levels in patients with localized scleroderma in contrast to other organ-specific autoimmune diseases. *Exp Dermatol.* (2007) 16:87–93. doi: 10.1111/j.1600-0625.2006.00485.x
- Groom J, Kalled SL, Cutler AH, Olson C, Woodcock SA, Schneider P, et al. Association of BAFF/BLyS overexpression and altered B cell differentiation with Sjogren's syndrome. J Clin Invest. (2002) 109:59–68. doi: 10.1172/JCI14121
- 34. Cheema GS, Roschke V, Hilbert DM, Stohl W. Elevated serum B lymphocyte stimulator levels in patients with systemicimmune-

based rheumatic diseases. Arthritis Rheum. (2001) 44:1313-9. doi: 10.1002/1529-0131(200106)44:6<1313::AID-ART223>3.0.CO;2-S

- 35. Navarra SV, Guzman RM, Gallacher AE, Hall S, Levy RA, Jimenez RE, et al. Efficacy and safety of belimumab in patients with active systemic lupus erythematosus: a randomised, placebo-controlled, phase 3 trial. *Lancet*. (2011) 377:721–31. doi: 10.1016/S0140-6736(10)61354-2
- Dörner T, Posch M, Wagner F, Hüser A, Fischer T, Mooney L, et al. THU0313 Double-blind, randomized study of VAY736 single dose treatment in patients with primary sjögren's syndrome (PSS). *Ann Rheumatic Dis.* (2016) 75(Suppl. 2):300–1. doi: 10.1136/annrheumdis-2016-eular.5840
- Yoshida M, Kobayashi K, Kuo TT, Bry L, Glickman JN, Claypool SM, et al. Neonatal Fc receptor for IgG regulates mucosal immune responses to luminal bacteria. J Clin Invest. (2006) 116:2142–2151. doi: 10.1172/JCI27821
- Iwata H, Vorobyev A, Koga H, Recke A, Zillikens D, Prost-Squarcioni C, et al. Meta-analysis of the clinical and immunopathological characteristics and treatment outcomes in epidermolysis bullosa acquisita patients. Orphanet J Rare Dis. (2018) 13:153. doi: 10.1186/s13023-018-0896-1
- Ishii N, Hashimoto T, Zillikens D, Ludwig RJ. High-dose intravenous immunoglobulin (IVIG) therapy in autoimmune skin blistering diseases. *Clin Rev Allergy Immunol.* (2010) 38:186–195. doi: 10.1007/s12016-009-8153-y
- Blumberg L, Jumphries JE, Lasseter KC, Blumberg RS. SYNT001: a humanized IgG4 monoclonal antibody that disrupts the interaction of FcRn and IgG for the treatment of IgG-mediated autoimmune diseases. *Blood*. (2017) 130(Suppl. 1):3483.
- Ulrichts P, Guglietta A, Dreier T, van Bragt T, Hanssens V, Hofman E, et al. Neonatal Fc receptor antagonist efgartigimod safely and sustainably reduces IgGs in humans. J Clin Invest. (2018) 128:4372–86. doi: 10.1172/JCI97911
- Werth VP CD, Blumberg L, Humphries J, Blumberg R, Hall R. FcRn blockade with SYNT001 for the treatment of pemphigus. J Invest Dermatol. (2018) 138:S92. doi: 10.1016/j.jid.2018.03.546
- Satterthwaite AB, Li Z, Witte ON. Btk function in B cell development and response. Semin Immunol. (1998) 10:309–16. doi: 10.1006/smim. 1998.0123
- 44. de Weers M, Verschuren MC, Kraakman ME, Mensink RG, Schuurman RK, van Dongen JJ, et al. The Bruton's tyrosine kinase gene is expressed throughout B cell differentiation, from early precursor B cell stages preceding immunoglobulin gene rearrangement up to mature B cell stages. *Eur J Immunol.* (1993) 23:3109–14. doi: 10.1002/eji.1830231210
- 45. Smith CI, Islam TC, Mattsson PT, Mohamed AJ, Nore BF, Vihinen M. The Tec family of cytoplasmic tyrosine kinases: mammalian Btk, Bmx, Itk, Tec, Txk and homologs in other species. *Bioessays*. (2001) 23:436–46. doi: 10.1002/bies.1062
- 46. Hutcheson J, Vanarsa K, Bashmakov A, Grewal S, Sajitharan D, Chang BY, et al. Modulating proximal cell signaling by targeting Btk ameliorates humoral autoimmunity and end-organ disease in murine lupus. *Arthritis Res Ther.* (2012) 14:R243. doi: 10.1186/ar4086
- 47. Lee A, Sandhu S, Imlay-Gillespie L, Mulligan S, Shumack S. Successful use of Bruton's kinase inhibitor, ibrutinib, to control paraneoplastic pemphigus in a patient with paraneoplastic autoimmune multiorgan syndrome and chronic lymphocytic leukaemia. *Australas J Dermatol.* (2017) 58:e240–2. doi: 10.1111/ajd.12615
- Smith PF, Krishnarajah J, Nunn PA, Hill RJ, Karr D, Tam D, et al. A phase I trial of PRN1008, a novel reversible covalent inhibitor of Bruton's tyrosine kinase, in healthy volunteers. *Br J Clin Pharmacol.* (2017) 83:2367–76. doi: 10.1111/bcp.13351
- Haeberle S, Wei X, Bieber K, Goletz S, Ludwig RJ, Schmidt E, et al. Regulatory T-cell deficiency leads to pathogenic bullous pemphigoid antigen 230 autoantibody and autoimmune bullous disease. J Allergy Clin Immunol. (2018) 142:1831–42.e1837. doi: 10.1016/j.jaci.2018.04.006
- Bieber K, Sun S, Witte M, Kasprick A, Beltsiou F, Behnen M, et al. Regulatory T cells suppress inflammation and blistering in pemphigoid diseases. *Front Immunol.* (2017) 8:1628. doi: 10.3389/fimmu.2017.01628
- Trzonkowski P, Bieniaszewska M, Juscinska J, Dobyszuk A, Krzystyniak A, Marek N, et al. First-in-man clinical results of the treatment of patients with graft versus host disease with human *ex vivo* expanded CD4+CD25+CD127- T regulatory cells. *Clin Immunol.* (2009) 133:22–6. doi: 10.1016/j.clim.2009.06.001

- Bluestone JA, Buckner JH, Fitch M, Gitelman SE, Gupta S, Hellerstein MK, et al. Type 1 diabetes immunotherapy using polyclonal regulatory T cells. *Sci Transl Med.* (2015) 7:315ra189. doi: 10.1126/scitranslmed. aad4134
- Brunstein CG, Miller JS, Cao Q, McKenna DH, Hippen KL, Curtsinger J, et al. Infusion of *ex vivo* expanded T regulatory cells in adults transplanted with umbilical cord blood: safety profile and detection kinetics. *Blood*. (2011) 117:1061–70. doi: 10.1182/blood-2010-07-293795
- Zwang NA, Leventhal JR. Cell therapy in kidney transplantation: focus on regulatory T cells. J Am Soc Nephrol. (2017) 28:1960–72. doi: 10.1681/ASN.2016111206
- Yokoyama T, Matsuda S, Takae Y, Wada N, Nishikawa T, Amagai M, et al. Antigen-independent development of Foxp3+ regulatory T cells suppressing autoantibody production in experimental pemphigus vulgaris. *Int Immunol.* (2011) 23:365–73. doi: 10.1093/intimm/dxr020
- 56. Schmidt T, Willenborg S, Hunig T, Deeg CA, Sonderstrup G, Hertl M, et al. Induction of T regulatory cells by the superagonistic anti-CD28 antibody D665 leads to decreased pathogenic IgG autoantibodies against desmoglein 3 in a HLA-transgenic mouse model of pemphigus vulgaris. *Exp Dermatol.* (2016) 25:293–8. doi: 10.1111/exd.12919
- 57. Zand N, Mansouri P, Fateh M, Ataie-Fashtami L, Rezaee Khiabanloo S, Safar F, et al. Relieving pain in oral lesions of pemphigus vulgaris using the non-ablative, non-thermal, CO2 laser therapy (NTCLT): preliminary results of a novel approach. J Lasers Med Sci. (2017) 8:7–12. doi: 10.15171/jlms.2017.02
- Mignogna MD, Fortuna G, Leuci S, Adamo D, Dell'Aversana Orabona G, Ruoppo E. Adjuvant triamcinolone acetonide injections in oro-pharyngeal pemphigus vulgaris. J Eur Acad Dermatol Venereol. (2010) 24:1157–65. doi: 10.1111/j.1468-3083.2010.03610.x
- Bystryn JC, Rudolph JL. Pemphigus. Lancet. (2005) 366:61–73. doi: 10.1016/S0140-6736(05)66829-8
- Chicharro-Alcantara D, Rubio-Zaragoza M, Damia-Gimenez E, Carrillo-Poveda JM, Cuervo-Serrato B, Pelaez-Gorrea P, et al. Platelet rich plasma: new insights for cutaneous wound healing management. *J Funct Biomater*. (2018) 9:10. doi: 10.3390/jfb9010010
- EL-Komy MH, Hassan AS, Abdel Raheem HM, Doss SS, EL-Kaliouby M, Saleh NA, et al. Platelet-rich plasma for resistant oral erosions of pemphigus vulgaris: a pilot study. *Wound Repair Regen.* (2015) 23:953–5. doi: 10.1111/wrr.12363
- El-Komy MHM, Saleh NA, Saleh MA. Autologous platelet-rich plasma and triamcinolone acetonide intralesional injection in the treatment of oral erosions of pemphigus vulgaris: a pilot study. *Arch Dermatol Res.* (2018) 310:375–81. doi: 10.1007/s00403-018-1824-x
- Hammers CM, Chen J, Lin C, Kacir S, Siegel DL, Payne AS, et al. Persistence of anti-desmoglein 3 IgG(+) B-cell clones in pemphigus patients over years. *J Invest Dermatol.* (2015) 135:742–9. doi: 10.1038/jid.2014.291
- Miossec P, Korn T, Kuchroo VK. Interleukin-17 and type 17 helper T cells. N Engl J Med. (2009) 361:888–98. doi: 10.1056/NEJMra0707449
- 65. Volpe E, Servant N, Zollinger R, Bogiatzi SI, Hupe P, Barillot E, et al. A critical function for transforming growth factor-beta, interleukin 23 and proinflammatory cytokines in driving and modulating human T(H)-17 responses. *Nat Immunol.* (2008) 9:650–7. doi: 10.1038/ni.1613
- Langrish CL, Chen Y, Blumenschein WM, Mattson J, Basham B, Sedgwick JD, et al. IL-23 drives a pathogenic T cell population that induces autoimmune inflammation. *J Exp Med.* (2005) 201:233–40. doi: 10.1084/jem.20041257
- Yang J, Sundrud MS, Skepner J, Yamagata T. Targeting Th17 cells in autoimmune diseases. *Trends Pharmacol Sci.* (2014) 35:493–500. doi: 10.1016/j.tips.2014.07.006
- Takahashi H, Tsuji H, Hashimoto Y, Ishida-Yamamoto A, Iizuka H. Serum cytokines and growth factor levels in Japanese patients with psoriasis. *Clin Exp Dermatol.* (2010) 35:645–9. doi: 10.1111/j.1365-2230.2009.03704.x
- Lubberts E, Joosten LA, van de Loo FA, Schwarzenberger P, Kolls J, van den Berg WB. Overexpression of IL-17 in the knee joint of collagen type II immunized mice promotes collagen arthritis and aggravates joint destruction. *Inflamm Res.* (2002) 51:102–4. doi: 10.1007/BF02684010
- Toosi S, Bystryn JC. Potential role of interleukin-17 in the pathogenesis of bullous pemphigoid. *Med Hypotheses*. (2010) 74:727–8. doi: 10.1016/j.mehy.2009.10.038

- Le Jan S, Plee J, Vallerand D, Dupont A, Delanez E, Durlach A, et al. Innate immune cell-produced IL-17 sustains inflammation in bullous pemphigoid. *J Invest Dermatol.* (2014) 134:2908–17. doi: 10.1038/jid.2014.263
- Chakievska L, Holtsche MM, Kunstner A, Goletz S, Petersen BS, Thaci D, et al. IL-17A is functionally relevant and a potential therapeutic target in bullous pemphigoid. *J Autoimmun.* (2018) 96:104–12. doi: 10.1016/j.jaut.2018.09.003
- Gordon KB, Blauvelt A, Papp KA, Langley RG, Luger T, Ohtsuki M, et al. Phase 3 trials of ixekizumab in moderate-to-severe plaque psoriasis. N Engl J Med. (2016) 375:345–56. doi: 10.1056/NEJMoa1512711
- Engineer L, Ahmed AR. Role of intravenous immunoglobulin in the treatment of bullous pemphigoid: analysis of current data. J Am Acad Dermatol. (2001) 44:83–8. doi: 10.1067/mjd.2000.112288
- Jolles S. A review of high-dose intravenous immunoglobulin (hdIVIg) in the treatment of the autoimmune blistering disorders. *Clin Exp Dermatol.* (2001) 26:127–31. doi: 10.1046/j.1365-2230.2001.00779.x
- Ahmed AR. Intravenous immunoglobulin therapy for patients with bullous pemphigoid unresponsive to conventional immunosuppressive treatment. J Am Acad Dermatol. (2001) 45:825–35. doi: 10.1067/mjd.2001. 116337
- 77. Venning VA, Taghipour K, Mohd Mustapa MF, Highet AS, Kirtschig G. British association of dermatologists' guidelines for the management of bullous pemphigoid 2012. Br J Dermatol. (2012) 167:1200–14. doi: 10.1111/bjd.12072
- Mydlarski PR, Ho V, Shear NH. Canadian consensus statement on the use of intravenous immunoglobulin therapy in dermatology. *J Cutan Med Surg.* (2006) 10:205–21. doi: 10.2310/7750.2006.00048
- Ahmed AR, Dahl MV. Consensus statement on the use of intravenous immunoglobulin therapy in the treatment of autoimmune mucocutaneous blistering diseases. *Arch Dermatol.* (2003) 139:1051–9. doi: 10.1001/archderm.139.8.1051
- Sasaoka T, Ujiie H, Nishie W, Iwata H, Ishikawa M, Higashino H, et al. Intravenous IgG reduces pathogenic autoantibodies, serum IL-6 levels, and disease severity in experimental bullous pemphigoid models. J Invest Dermatol. (2018) 138:1260–7. doi: 10.1016/j.jid.2018.01.005
- Kamaguchi M, Iwata H, Mori Y, Toyonaga E, Ujiie H, Kitagawa Y, et al. Antiidiotypic antibodies against BP-IgG prevent type XVII collagen depletion. *Front Immunol.* (2017) 8:1669. doi: 10.3389/fimmu.2017.01669
- Amagai M, Ikeda S, Hashimoto T, Mizuashi M, Fujisawa A, Ihn H, et al. A randomized double-blind trial of intravenous immunoglobulin for bullous pemphigoid. *J Dermatol Sci.* (2017) 85:77–84. doi: 10.1016/j.jdermsci.2016.11.003
- Martinon F, Mayor A, Tschopp J. The inflammasomes: guardians of the body. *Annu Rev Immunol.* (2009) 27:229–65. doi: 10.1146/annurev.immunol.021908.132715
- Lamkanfi M, Dixit VM. Mechanisms and functions of inflammasomes. *Cell*. (2014) 157:1013–22. doi: 10.1016/j.cell.2014.04.007
- Wen H, Miao EA, Ting JP. Mechanisms of NOD-like receptorassociated inflammasome activation. *Immunity*. (2013) 39:432–41. doi: 10.1016/j.immuni.2013.08.037
- Latz E, Xiao TS, Stutz A. Activation and regulation of the inflammasomes. Nat Rev Immunol. (2013) 13:397–411. doi: 10.1038/nri3452
- Franchi L, Eigenbrod T, Munoz-Planillo R, Nunez G. The inflammasome: a caspase-1-activation platform that regulates immune responses and disease pathogenesis. *Nat Immunol.* (2009) 10:241–7. doi: 10.1038/ni.1703
- Shaw PJ, Lukens JR, Burns S, Chi H, McGargill MA, Kanneganti TD. Cutting edge: critical role for PYCARD/ASC in the development of experimental autoimmune encephalomyelitis. *J Immunol.* (2010) 184:4610– 4. doi: 10.4049/jimmunol.1000217
- Kahlenberg JM, Kaplan MJ. The inflammasome and lupus: another innate immune mechanism contributing to disease pathogenesis? *Curr Opin Rheumatol.* (2014) 26:475–81. doi: 10.1097/BOR.0000000000 00088
- Duan Z, Zhang Y, Zeng Z, Pan F. Comment on "Inflammasome activation of IL-18 results in endothelial progenitor cell dysfunction in systemic lupus erythematosus". *J Immunol.* (2012) 189:499; author reply 499–500. doi: 10.4049/jimmunol.1290031

- Kastbom A, Verma D, Eriksson P, Skogh T, Wingren G, Soderkvist P. Genetic variation in proteins of the cryopyrin inflammasome influences susceptibility and severity of rheumatoid arthritis (the Swedish TIRA project). *Rheumatology*. (2008) 47:415–7. doi: 10.1093/rheumatology/ kem372
- Fang H, Shao S, Cao T, Lei J, Dang E, Zhang J, et al. Increased expression of NLRP3 inflammasome components and interleukin-18 in patients with bullous pemphigoid. *J Dermatol Sci.* (2016) 83:116–23. doi: 10.1016/j.jdermsci.2016.04.009
- 93. Sadeghi H, Lockmann A, Hund AC, Samavedam UK, Pipi E, Vafia K, et al. Caspase-1-independent IL-1 release mediates blister formation in autoantibody-induced tissue injury through modulation of endothelial adhesion molecules. *J Immunol.* (2015) 194:3656–63. doi: 10.4049/jimmunol.1402688
- Amber KT, Murrell DF, Schmidt E, Joly P, Borradori L. Autoimmune subepidermal bullous diseases of the skin and mucosae: clinical features, diagnosis, and management. *Clin Rev Allergy Immunol.* (2018) 54:26–51. doi: 10.1007/s12016-017-8633-4
- Hashimoto T, Ohzono A, Teye K, Numata S, Hiroyasu S, Tsuruta D, et al. Detection of IgE autoantibodies to BP180 and BP230 and their relationship to clinical features in bullous pemphigoid. *Br J Dermatol.* (2017) 177:141–51. doi: 10.1111/bjd.15114
- van Beek N, Luttmann N, Huebner F, Recke A, Karl I, Schulze FS, et al. Correlation of serum levels of IgE autoantibodies against BP180 with bullous pemphigoid disease activity. *JAMA Dermatol.* (2017) 153:30–8. doi: 10.1001/jamadermatol.2016.3357
- Amber KT, Valdebran M, Kridin K, Grando SA. The role of eosinophils in bullous pemphigoid: a developing model of eosinophil pathogenicity in mucocutaneous disease. *Front Med.* (2018) 5:201. doi: 10.3389/fmed.2018.00201
- Lin L, Hwang BJ, Culton DA, Li N, Burette S, Koller BH, et al. Eosinophils mediate tissue injury in the autoimmune skin disease bullous pemphigoid. J Invest Dermatol. (2018) 138:1032–43. doi: 10.1016/j.jid.2017.11.031
- Wakugawa M, Nakamura K, Hino H, Toyama K, Hattori N, Okochi H, et al. Elevated levels of eotaxin and interleukin-5 in blister fluid of bullous pemphigoid: correlation with tissue eosinophilia. *Br J Dermatol.* (2000) 143:112–6. doi: 10.1046/j.1365-2133.2000.03599.x
- 100. Stahle-Backdahl M, Inoue M, Guidice GJ, Parks WC. 92-kD gelatinase is produced by eosinophils at the site of blister formation in bullous pemphigoid and cleaves the extracellular domain of recombinant 180kD bullous pemphigoid autoantigen. J Clin Invest. (1994) 93:2022–30. doi: 10.1172/JCI117196
- 101. Borrego L, Maynard B, Peterson EA, George T, Iglesias L, Peters MS, et al. Deposition of eosinophil granule proteins precedes blister formation in bullous pemphigoid. Comparison with neutrophil and mast cell granule proteins. Am J Pathol. (1996) 148:897–909.
- 102. Bieber K, Ernst AL, Tukaj S, Holtsche MM, Schmidt E, Zillikens D, et al. Analysis of serum markers of cellular immune activation in patients with bullous pemphigoid. *Exp Dermatol.* (2017) 26:1248–52. doi: 10.1111/exd.13382

- Honda K, Chihara J. Eosinophil activation by eotaxin-eotaxin primes the production of reactive oxygen species from eosinophils. *Allergy*. (1999) 54:1262–9.
- 104. Ding C, Li J, Zhang X. Bertilimumab Cambridge antibody technology group. *Curr Opin Investig Drugs*. (2004) 5:1213–8.
- 105. Amber KT, Maglie R, Solimani F, Eming R, Hertl M. Targeted therapies for autoimmune bullous diseases: current status. *Drugs*. (2018) 78:1527–48. doi: 10.1007/s40265-018-0976-5
- Lamberts A, Euverman HI, Terra JB, Jonkman MF, Horvath B. Effectiveness and safety of rituximab in recalcitrant pemphigoid diseases. *Front Immunol.* (2018) 9:248. doi: 10.3389/fimmu.2018.00248
- Geahlen RL. Getting Syk: spleen tyrosine kinase as a therapeutic target. Trends Pharmacol Sci. (2014) 35:414–22. doi: 10.1016/j.tips.2014.05.007
- 108. Samavedam UK, Mitschker N, Kasprick A, Bieber K, Schmidt E, Laskay T, et al. Whole-genome expression profiling in skin reveals SYK as a key regulator of inflammation in experimental epidermolysis bullosa acquisita. *Front Immunol.* (2018) 9:249. doi: 10.3389/fimmu.2018.00249
- 109. Sezin T, Krajewski M, Wutkowski A, Mousavi S, Chakievska L, Bieber K, et al. The leukotriene B4 and its receptor BLT1 act as critical drivers of neutrophil recruitment in murine bullous pemphigoid-like epidermolysis bullosa acquisita. *J Invest Dermatol.* (2017) 137:1104–13. doi: 10.1016/j.jid.2016.12.021
- 110. Lamberts A, Yale M, Grando SA, Horvath B, Zillikens D, Jonkman MF. Unmet needs in pemphigoid diseases: an international survey amongst patients, clinicians and researchers. *Acta Derm Venereol.* (2018) 99:224–5. doi: 10.2340/00015555-3052
- 111. Koga H, Kasprick A, Lopez R, Auli M, Pont M, Godessart N, et al. Therapeutic effect of a novel phosphatidylinositol-3-kinase delta inhibitor in experimental epidermolysis bullosa acquisita. *Front Immunol.* (2018) 9:1558. doi: 10.3389/fimmu.2018.01558
- 112. Koga H, Recke A, Vidarsson G, Pas HH, Jonkman MF, Hashimoto T, et al. PDE4 inhibition as potential treatment of epidermolysis bullosa acquisita. J Invest Dermatol. (2016) 136:2211–20. doi: 10.1016/j.jid.2016.06.619

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