

High *EVII* Expression due to *NRIP1/EVII* Fusion in Therapy-related Acute Myeloid Leukemia: Description of the First Pediatric Case

Mariella D'Angiò¹, Grazia Fazio¹, Andrea Grioni^{1,2}, Sonia Palamini¹, Simona Sala¹, Marta Galbiati¹, Andrea Biondi^{1,3}, Adriana Balduzzi³, Carmelo Rizzari³, Giovanni Cazzaniga^{1,4}

Correspondence: Giovanni Cazzaniga (e-mail: gianni.cazzaniga@hsgerardo.org).

Disruption of chromosome 3 at band 3q26 has been well-documented in acute myeloid leukemia (AML), chronic myeloid leukemia (CML) and myelodysplastic syndromes (MDS).¹ Clinically, 3q26.2 rearrangements correlate with elevated platelet counts, marked hyperplasia with dysplasia of megakaryocytes, aggressive clinical course and unfavorable prognosis with conventional therapy.^{2,3} Chromosome 3q26 abnormalities have been reported to activate the aberrant expression of the human *Ecotropic Virus Integration site-1* gene (*EVII* known as *MECOM-MDS1* and *EVII* complex locus), a transcriptional factor with an essential role in proliferation and maintenance of hematopoietic stem cells.^{4,5} Although the exact role of *EVII* in leukemogenesis is not completely known, a recent study revealed that it may be involved in leukemic cell proliferation and apoptosis via the regulation of miR-9 promoter methylation, thus suggesting the possible role of hypomethylating agents in *EVII* over-expressed leukaemia.⁶⁻⁸ *EVII* up regulation occurs in approximately 8% to 10% of human adult AML and, strikingly, up to 27% of pediatric *KMT2A* (*MLL*) rearranged leukemia cases.^{9,10} The most frequent abnormalities resulting in inappropriate activation of *EVII* are the inversion *inv(3)(q21q26)* and the translocation *t(3;3)(q21;q26.2)*. However, *EVII* can be rearranged with a variety of other partner genes [1q41 (*DUSP10*), 7q21 (*CDK6*), 7q34 (*TCRB*), 12p34 (*ETV6*), 21q22 (*RUNX1*)].^{3,11-12} Its increased expression can also be detected in cytogenetically normal AML, in aberrant cytogenetic subgroups, such as

monosomy 7 and 11q23/*MLL* translocations, as well as in patients with cryptic 3q26 rearrangements.^{3,12} In particular, the cryptic *t(3;21)(q26;q11)* rearrangement, resulting in *NRIP1/EVII* fusion, has been identified using FISH analyses in 9 adults, 4 AML and 5 MDS so far.¹² All patients showed a high *EVII* expression and an adverse prognosis with a median overall survival (OS) of 9.4 months.

Here we report the first pediatric case of therapy-related AML (t-AML) carrying a cryptic *t(3;21)(q26;q11)* rearrangement and *EVII* over-expression.

A previously healthy 4-year old child had been referred to our institution for cervical lymphadenopathy, peripheral cytopenia and spleen enlargement; peripheral blood (PB) smear and bone marrow (BM) aspiration revealed infiltration of lymphoid blast cells, cytogenetic analysis showed normal karyotype. The child was diagnosed with B-lineage acute lymphoblastic leukemia (ALL), treated according to AIEOP-BFM ALL2009 protocol and allocated to the high-risk arm, due to an insufficient molecular response [<https://clinicaltrials.gov/ct2/show/study/NCT01117441>].

Eight years after disease onset and 6 years after elective discontinuation of treatment, the patient experienced fatigue and her cell blood count (CBC) showed anemia, thrombocytopenia and circulating blasts (hemoglobin 8.8 g/dL, platelets $68 \times 10^9/L$, white blood cell count $8.8 \times 10^9/L$); morphological analyses detected 79% of undifferentiated blasts in PB and 60% in BM of undetermined FAB (French American British) classification (Fig. 1A). Cells were positive for CD45, CD34, CD117, HLA-DR, CD13, CD33, CD123, and MPO (Fig. 1B), consistent with a diagnosis of AML. Cytogenetic analysis, carried out on BM blasts by standard techniques and evaluated by Q banding, showed a complex karyotype with numerical and structural abnormalities (44,XX,-2,-3,del(5)(q22q35),-6,-7,del21(21)(q11),+2mar[19]/46,XX[1]) (Fig. 1C), thus including monosomy 7, known to be associated with poor prognosis.¹³ Target capture RNA NGS sequencing performed by TruSight Pancancer panel (Illumina, San Diego, CA, USA)¹⁴ revealed a fusion involving the *NRIP1* (exon 2) and *EVII* (exon 2) genes, subsequently confirmed by RT-PCR and Sanger sequencing (Fig. 2A, B). *EVII* expression was evaluated by Real-time quantitative PCR (RQ-PCR) (Light Cycler 480 System Roche Diagnostics) and calculated by the comparative cycle time (DDCt) method with *GUS* as the housekeeping gene. Over-expression of *EVII* was found in the patient's BM compared to four healthy donors and three pediatric AML (2 de novo and 1 t-AML) negative for 3q26

¹Centro Ricerca Tettamanti, Pediatrics, Fondazione MBBM/San Gerardo Hospital, University of Milano-Bicocca, University of Milano-Bicocca, Monza, Italy

²Faculty of Science, National Centre for Biomolecular Research, Masaryk University, Brno, Czech Republic

³Pediatrics, Fondazione MBBM/San Gerardo Hospital, University of Milano-Bicocca, Italy

⁴Genetics, University of Milano-Bicocca, Monza, Italy

Mariella D'Angiò and Grazia Fazio contributed equally to the manuscript.

The authors have no conflicts of interest to disclose.

Copyright © 2020 the Author(s). Published by Wolters Kluwer Health, Inc. on behalf of the European Hematology Association. This is an open access article distributed under the terms of the Creative Commons Attribution-Non Commercial-No Derivatives License 4.0 (CCBY-NC-ND), where it is permissible to download and share the work provided it is properly cited. The work cannot be changed in any way or used commercially without permission from the journal.

HemaSphere (2020) 4:5(e471). <http://dx.doi.org/10.1097/HS9.0000000000000471>.

Received: 25 May 2020 / Accepted: 16 June 2020

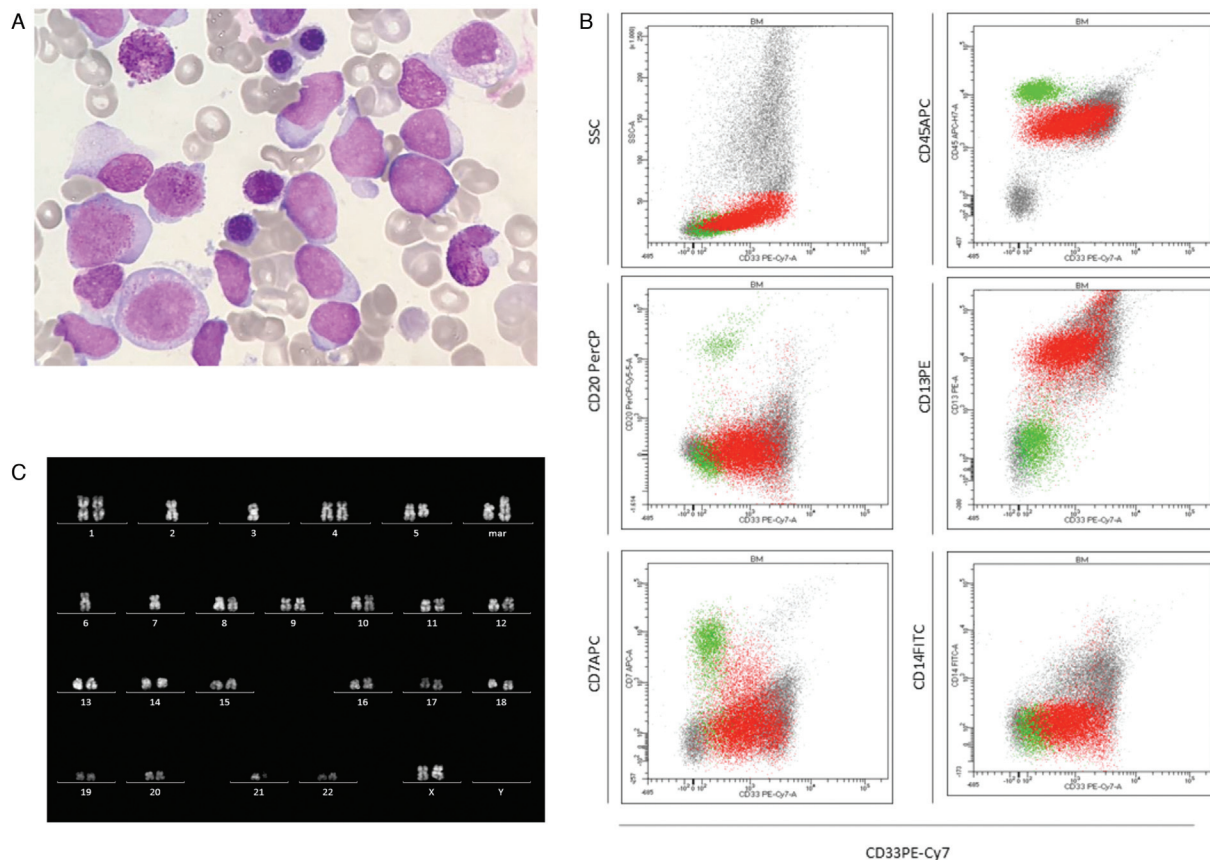


Figure 1. Morphology, immunophenotype and cytogenetic analyses of t-AML diagnosis. A. Image of bone marrow smear showing undifferentiated blast cells. B. Cytometry analysis is consistent with AML diagnosis. C. Karyotype analysis: G banding consistent with complex karyotype with numerical and structural abnormalities.

rearrangements, whose BM were analyzed as controls (Fig. 2C). In order to exclude a clonal relationship between the ALL and AML diseases, the *IG/TR* rearrangements detected at ALL onset were evaluated on AML blasts; similarly, the *NRIP1/EV11* fusion transcript, detected on AML blasts, was tested on ALL cells (Fig. 2A); both analysis were negative, revealing that the two leukemias have no clonal relation. Furthermore, to exclude an underlying Li Fraumeni syndrome associated to leukemia recurrence, *TP53* sequencing was performed by NGS, and neither mutations nor copy number variation (CNV) were detected.

Conventional AML treatment was given, consisting of three sequential high-dose chemotherapy blocks (fludarabine, cytarabine and liposomal doxorubicin; mitoxantrone, etoposide and cytarabine; cladribine and cytarabine) usually used for AML; a progressive reduction of BM blast percentage was obtained but no CR was ever reached. A haploidentical hematopoietic stem cell transplantation (HSCT), after a treosulfan-based conditioning (treosulfan, fludarabine and thiotepa), was performed during post-chemotherapy pancytopenia 5 months after the diagnosis of AML. Ninety days after transplantation 2% atypical undifferentiated blasts and mixed chimerism (donor $\leq 95\%$) were detected in her BM. Neither a donor lymphocyte infusion (DLI) nor a dose of gentuzumab ozogamicin plus standard dose cytarabine were able to control the disease. Based on the potential efficacy of hypomethylating agents in *EV11* over-expressed leukemia,^{6–8} one cycle of azacitidine was given without any clinical or hematological benefits. The patient died due to disease progression 7 months after HSCT.

The case here described is, to the best of our knowledge, the first reported pediatric patient affected by *NRIP1/EV11* fusion t-AML and *EV11* over-expression. Furthermore, it may be considered exceptional, due to the lowest reported frequency of pediatric tAML and of the *NRIP1/EV11* fusion transcript.

In fact, up to now, the diagnosis of t-AML has been reported in only 0.5% of children and young adults previously treated for cancer¹⁵ and *NRIP1/EV11* fusion has been described exclusively in 0.9% of adult AML-MDS patients.¹²

Consistent with our case, monosomy 7 has been frequently reported as associated to *EV11* rearrangements,¹² and most adult cases have similarly shown a very aggressive clinical course as well as an inadequate response to conventional therapy.

Recent evidences showed that *EV11* may regulate the abilities of methyltransferase and acetyltransferase to modify gene expression via epigenetic mechanism; in particular, the down regulation of miR-9 may be involved in leukaemogenesis.⁸ These findings, worthy of further investigation, indicate the possibility of using hypomethylating agents in AML/MDS patients with *NRIP1/EV11* fusion transcript and/or *EV11* over-expression refractory to chemotherapy.

In patients with de novo or t-AML partially refractory to chemotherapy, it could be worth to search *EV11* rearrangements, even if cytogenetically cryptic, and/or to investigate *EV11* expression in order to consider including hypomethylating agents early in the treatment course.

Furthermore, with the aim to improve the poor prognosis reported in both *EV11* rearranged and/or over-expressed de novo

7. Wanquet A, Prebet T, Berthon C, et al. Azacitidine treatment for patients with myelodysplastic syndrome and acute myeloid leukemia with chromosome 3q abnormalities. *Am J Hematol*. 2015;90:859–863.
8. Mittal N, Li L, Sheng Y, et al. A critical role of epigenetic inactivation of miR-9 in *EVI1* high pediatric AML. *Mol Cancer*. 2019;18:1–6.
9. Glass C, Wilson M, Gonzalez R, et al. The role of *EVI1* in myeloid malignancies. *Blood Cells Mol Dis*. 2014;53:67–76.
10. Bindels EM, Havermans M, Lugthart S, et al. *EVI1* is critical for the pathogenesis of a subset of *MLL-AF9*, rearranged AMLs. *Blood*. 2012;119:5838–5849.
11. Bobadilla D, Enriquez EL, Alvarez G, et al. An interphase fluorescence in situ hybridisation assay for the detection of 3q26.2/*EVI1* rearrangements in myeloid malignancies. *Br J Haematol*. 2007;136:806–813.
12. Haferlach C, Bacher U, Grossmann V, et al. Three novel cytogenetically cryptic *EVI1* rearrangements associated with increased *EVI1* expression and poor prognosis identified in 27 acute myeloid leukemia cases. *Genes Chromosomes Cancer*. 2012;51:1079–1085.
13. Canaani J, Labopin M, Itälä-Remes M, et al. Prognostic significance of recurring chromosomal abnormalities in transplanted patients with acute myeloid leukemia. *Leukemia*. 2019;33:1944–1952.
14. Grioni A, Fazio G, Rigamonti S, et al. A Simple RNA target capture NGS strategy for fusion genes assessment in the diagnostics of pediatric B-cell acute lymphoblastic leukemia. *HemaSphere*. 2019;3:1–9.
15. Brown CA, Youlden DR, Aitken JF, et al. Therapy-related acute myeloid leukemia following treatment for cancer in childhood: a population-based registry study. *Pediatr Blood Cancer*. 2018;65:1–7.