



Draft Genome Sequence of *Bacillus vallismortis* Strain BL01, Isolated from *Artemisia lerchiana* Web. Roots

^(b) Vladimir K. Chebotar,^a Maria S. Gancheva,^{a,b} Elena P. Chizhevskaya,^a Oksana V. Keleinikova,^a Maria E. Baganova,^a Alexander N. Zaplatkin,^a Veronika N. Pishchik^{a,c}

^aAll-Russia Research Institute for Agricultural Microbiology, St. Petersburg, Pushkin, Russia
^bDepartment of Genetics and Biotechnology, Faculty of Biology, Saint Petersburg State University, St. Petersburg, Russia
^cAgrophysical Scientific Research Institute, St. Petersburg, Russia

ABSTRACT Some strains of *Bacillus vallismortis* have been reported to be efficient plant-growth-promoting bacteria as well as inducers of systemic resistance. Here, we report the draft genome sequence of *Bacillus vallismortis* strain BL01, isolated from the roots of *Artemisia lerchiana* Web.

acillus vallismortis can produce metabolites with strong growth inhibition activity against phytopathogenic fungi (1-3) and bacteria (4) and can increase plant protection against viruses (5). Also, Bacillus vallismortis is able to produce growth-regulating substances that increase yields (3, 5, 6). B. vallismortis strain BL01 was isolated from the roots of Artemisia lerchiana Web. from the Astrakhan region, Russia (46.519511°N, 47.915334°E). Plant roots were disinfected with tap water for 30 s followed by 70% ethanol for 5 min and then 15% H₂O₂ for 10 min and sterile water for 2 min 5 times and subsequently crushed with a mortar and pestle under sterile conditions. Aliquots of 100 mL of the resulting plant juices were plated onto a 1/20 dilution of tryptic soy agar (TSA; Difco Laboratories, MI, USA) plates. The sterility check consisted of aliquots of water from the last rinsing that were plated onto 1/20 TSA according to methods reported previously (7). Plates were incubated at 28°C for 3 days. Strain BL01 was grown in LB medium at 30°C overnight. The total cellular DNA was isolated from a single colony using the cetyltrimethylammonium bromide (CTAB)-NaCl method (8). Paired-end reads were generated using the Nextera DNA Flex kit (Illumina, USA). The complete genome was sequenced using the Illumina HiSeq 2500 technology. A total of 415,271 reads were generated, with an average length of 280 bp. FastQC v0.11.9 (9) was used to assess the quality of the reads. Trimmomatic v0.39 (10) was used for trimming low-quality sequences. The trimmed reads were classified using Kaiju v1.8.2 against the RefSeq genomes under maximum exact match (MEM) mode to maximize exact matches (11). The reads were de novo assembled using SPAdes v3.14.1 (12) with the "-careful" option. The nucleotide compositions of contigs and scaffolds were determined using Seqtk v1.3-r106 (https://github.com/lh3/seqtk). The contigs were mapped to the Bacillus vallismortis strain Bac111 (GenBank accession number CP033052.1) genome by CONTIGuator v2.7 (13). Default parameters were used for all software unless otherwise specified. Contigs were assembled into a single scaffold using Ragtag v2.1.0 (gaps filled with N's) (14). Evaluation of the genome assembly was performed with Quast v5.1.0 (15) and BUSCO v5.2.2 (bacillales_odb10) (16). The final assembly contained 4,115,091 bp in 1 scaffold, and the average GC content of the assembly was 44.01%. The BUSCO results recovered 99.8% of the Bacillales database. Genome annotation was performed using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP-6.1) (17). A total of 4,029 protein-coding sequences (CDSs) and 80 tRNA genes were predicted. The complete genome sequence of Bacillus vallismortis BL01 will contribute to revealing the role of Bacillus vallismortis in plant growth and physiology.

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Address correspondence to Vladimir K. Chebotar, vladchebotar@rambler.ru. The authors declare no conflict of interest. **Received** 4 July 2022

Accepted 15 August 2022 Published 17 October 2022 **Data availability.** All data are available in the National Center for Biotechnology Information database under BioProject accession number PRJNA809498. The raw reads were deposited in the Sequence Read Archive under accession number SRR18107534. The GenBank nucleotide sequence accession number is CP092751.

ACKNOWLEDGMENTS

The article was made with the support of the Ministry of Science and Higher Education of the Russian Federation in accordance with agreement number 075-15-2021-1055, 28 September 2021, on providing a grant in the form of subsidies from the Federal budget of the Russian Federation. The grant was provided for the implementation of the project Mobilization of the Genetic Resources of Microorganisms on the Basis of the Russian Collection of Agricultural Microorganisms (RCAM) at the All-Russia Research Institute for Agricultural Microbiology (ARRIAM) According to the Network Principle of Organization.

REFERENCES

- Zhao Z, Wang Q, Wang K, Brian K, Liu C, Gu Y. 2010. Study of the antifungal activity of Bacillus vallismortis ZZ185 in vitro and identification of its antifungal components. Bioresour Technol 101:292–297. https://doi.org/ 10.1016/j.biortech.2009.07.071.
- Castaldi S, Petrillo C, Donadio G, Piaz FD, Cimmino A, Masi M, Evidente A, Isticato R. 2021. Plant growth promotion function of Bacillus sp. strains isolated from salt-pan rhizosphere and their biocontrol potential against Macrophomina phaseolina. Int J Mol Sci 22:3324. https://doi.org/10.3390/ ijms22073324.
- Park K-S, Paul D, Yeh W-H. 2006. Bacillus vallismortis EXTN-1-mediated growth promotion and disease suppression in rice. Plant Pathol J 22: 278–282. https://doi.org/10.5423/PPJ.2006.22.3.278.
- Noh SW, Seo R, Park JK, Manir MM, Park K, Sang MK, Moon SS, Jung HW. 2017. Cyclic dipeptides from Bacillus vallismortis BS07 require key components of plant immunity to induce disease resistance in Arabidopsis against Pseudomonas infection. Plant Pathol J 33:402–409.
- Park K-S, Paul D, Ryu K-R, Kim E-Y, Kim Y-K. 2006. Bacillus vallismortis strain EXTN-1 mediated systemic resistance against potato virus Y and X in the field. Plant Pathol J 22:360–363. https://doi.org/10.5423/PPJ.2006 .22.4.360.
- Park J-W, Kotnala B, Kim J-W, Lee S-W, Park K. 2013. Systemic resistance and growth promotion of chili pepper induced by an antibiotic producing Bacillus vallismortis strain BS07. Biol Control 65:246–257. https://doi.org/ 10.1016/j.biocontrol.2013.02.002.
- Malfanova N, Kamilova F, Validov S, Shcherbakov A, Chebotar V, Tikhonovich I, Lugtenberg B. 2011. Characterization of *Bacillus subtilis* HC8, a novel plantbeneficial endophytic strain from giant hogweed. Microb Biotechnol 4: 523–532. https://doi.org/10.1111/j.1751-7915.2011.00253.x.
- Wilson K. 2001. Preparation of genomic DNA from bacteria. Curr Protoc Mol Biol Chapter 2:Unit 2.4. https://doi.org/10.1002/0471142727.mb0204s56.

- Andrews S. 2010. FastQC: a quality control tool for high throughput sequence data. http://www.bioinformatics.babraham.ac.uk/projects/fastqc.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120. https://doi.org/10 .1093/bioinformatics/btu170.
- Menzel P, Ng K, Krogh A. 2016. Fast and sensitive taxonomic classification for metagenomics with Kaiju. Nat Commun 7:11257. https://doi.org/10 .1038/ncomms11257.
- 12. Prjibelski A, Antipov D, Meleshko D, Lapidus A, Korobeynikov A. 2020. Using SPAdes de novo assembler. Curr Protoc Bioinformatics 70:e102. https://doi.org/10.1002/cpbi.102.
- Galardini M, Biondi EG, Bazzicalupo M, Mengoni A. 2011. CONTIGuator: a bacterial genomes finishing tool for structural insights on draft genomes. Source Code Biol Med 6:11. https://doi.org/10.1186/1751-0473-6-11.
- Alonge M, Soyk S, Ramakrishnan S, Wang X, Goodwin S, Sedlazeck FJ, Lippman ZB, Schatz MC. 2019. RaGOO: fast and accurate reference-guided scaffolding of draft genomes. Genome Biol 20:224. https://doi.org/10.1186/ s13059-019-1829-6.
- Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. Bioinformatics 29:1072–1075. https:// doi.org/10.1093/bioinformatics/btt086.
- Manni M, Berkeley MR, Seppey M, Simão FA, Zdobnov EM. 2021. BUSCO update: novel and streamlined workflows along with broader and deeper phylogenetic coverage for scoring of eukaryotic, prokaryotic, and viral genomes. Mol Biol Evol 38:4647–4654. https://doi.org/10.1093/molbev/msab199.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI prokaryotic genome annotation pipeline. Nucleic Acids Res 44:6614–6624. https://doi.org/ 10.1093/nar/gkw569.