

Increasing Age Affected Polymorphonuclear Neutrophils in Prognosis of *Mycoplasma pneumoniae* Pneumonia

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Purpose: It is well known that age is related to the incidence of *Mycoplasma pneumoniae* pneumonia (MPP), and how age and other factors contribute to MPP remains unclear. In this study, we investigate how age affects the prognosis of MPP.

Patients and Methods: A total number of 1875 hospitalized children with pneumonia were enrolled in this study, including 52 children with refractory *M. pneumoniae* pneumonia (RMPP) and 298 children with non-RMPP. We used multiple logistic regression analysis to further identify the risk factors of RMPP, and found that age and polymorphonuclear neutrophils (PMNs) count were the key independent risk factors for the occurrence of RMPP. In order to improve specificity, 4.5 years old was taken as the cut-off value. Then, according to the cut-off value of age, 76 participants were recruited and divided into four groups: <4.5y MPP group, ≥4.5y MPP group, <4.5y health control (<4.5yHC) and ≥4.5y HC group. We explored the diverse functions of primary PMNs from children of different ages with MPP at cellular level. Besides, we studied the relationship between lung injury and PMNs in mice model with MPP of different ages.

Results: We found that the age and PMNs count of RMPP group were significantly higher than those of the non-RMPP group. Importantly, there is a linear correlation between the age of patients with RMPP and the percentage of PMNs. Further analysis showed that elderly patients infected with *M. pneumoniae* had more active PMNs function. Meanwhile, proteomics showed that children with *M. pneumoniae* infection in different age groups have differences in PMNs apoptosis, nicotinamide adenine dinucleotide phosphate, mitochondrial function and oxidative stress. Finally, we found that age is also involved in the pathogenesis of mouse model with MPP.

Conclusion: We speculate that age may contribute to the development of RMPP.

Keywords: refractory *Mycoplasma pneumoniae* pneumonia, *Mycoplasma pneumoniae*, polymorphonuclear neutrophils, reactive oxygen species, neutrophil extracellular traps

Introduction

Mycoplasma pneumoniae pneumonia (MPP) accounts for 10–40% of community-acquired pneumonia cases in hospitalized children, most of which occur over 5 years old.¹ In recent years, more and more cases of *M. pneumoniae* (MP) infection are manifested as refractory *Mycoplasma pneumoniae* pneumonia (RMPP), followed by pulmonary embolism, plastic bronchitis, necrotizing pneumonia, lung abscess and other extrapulmonary complications.^{2,3} According to reports, the main population of fulminant MP infection is young people without serious

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underlying diseases, and they required early administration of steroids as cellular immunosuppressants.⁴ These studies indicate that MP infection can trigger excessive lung and systemic inflammation in children with RMPP or fulminant *Mycoplasma pneumoniae* pneumonia.

MP infection is more common in children aged 5 years or older than children under 5 years old in the United States.¹ In northern China, the peak age of MPP is between 6 and 10 years.⁵ Zhang et al reported that the peak age of patients is much older than that of Non-RMPP patients in East China.⁶ Bi et al regarded age as one of the predictors of RMPP.⁷ These studies indicate that age may be the main factor associated with MP infection, but the underlying mechanism remains unclear. Previous studies have shown that age may have an impact on the function of polymorphonuclear neutrophils (PMNs). It has been observed that premature infants migrate fewer PMNs than full-term newborns, and the migration rate is slower than that of young adults.⁸ In addition, Peniche et al demonstrated that middle-aged mice exhibit significant defects in the recruitment of PMNs to the colon.⁹ There are also data supporting that PMNs from the elderly group have significantly reduced targeting accuracy¹⁰ and Fc-mediated phagocytosis.¹¹ Further studies have shown that epigenetic changes related to aging in PMNs, such as changes in DNA methylation, may affect cell function through modification of gene expression.¹² However, whether age affects the function of PMNs after MP infection remains unclear.

In order to investigate how age affects the prognosis of MPP, in this study, we firstly looked for the differences in PMNs and age between RMPP and non-RMPP group, and then we explored the diverse functions of PMNs from children of different age with MPP at cellular level; besides, we studied the lung injury in mice model with MPP of different ages.

Materials and Methods

Ethics Statements

All animals' experiments were approved by the Institutional Animal Care and Use Committee of the Nanjing Medical University at Nanjing (Approval number IACUC-1905057) and followed the Guidelines for the Ethical Review of Laboratory Animal Welfare People's Republic of China National Standard (GB/T 35892–2018).¹³ The BALB/c mice were kept in a pathogen-free environment and fed ad lib. The clinical study (Protocol

number 201703058) was consented by the Ethics Committee of the Children's Hospital of Nanjing Medical University in compliance with the Declaration of Helsinki. Informed consents were obtained from the parents of all patients included in this study.

Population

This study included patients with MPP who were hospitalized at the Department of Respiratory Medicine, Children's Hospital of Nanjing Medical University from September 1, 2016 to March 31, 2017. The criteria for diagnosis of MPP are provided in [Supplement 1](#). RMPP was defined as MPP cases whose chest X-ray or low-dose multi-slice spiral CT scan showed clinical and radiological deteriorations, such as malabsorption, atelectasis, pleural effusion, lung abscess, and lung necrosis, despite the appropriate macrolide antibiotic therapy for 7 days or longer.¹⁴ Patients whose chest imaging showed clear absorption after 7 days of treatment were assigned into non-RMPP groups. Patients with immune dysfunction, congenital ciliary dysplasia, cystic fibrosis, bronchiectasis, diffuse interstitial lung disease, and bronchopulmonary dysplasia were excluded from the study.

Isolation and Culture of PMNs from Peripheral Blood in vitro

Seventy-six participants were recruited and we divided them into 4 groups: <4.5y healthy control (<4.5yHC), ≥4.5y HC, <4.5y MPP group (MPP patients under 4.5 years old) and ≥4.5y MPP group (MPP patients greater than or equal to 4.5 years old). All subjects were recruited from March 2018 to November 2019. Peripheral blood samples of 76 participants at admission were collected in centrifuge tubes containing ethylenediaminetetraacetic acid (EDTA) solution to prevent blood clotting. The detailed medical information about 76 participants was provided in [Supplement 2 \(Tables S1 and S2\)](#). Among them, 46 children of the MPP groups met the diagnostic and exclusive criteria for MPP. The sample was then centrifuged at 3000 rpm for 10 minutes at 4°C. The plasma was collected and stored at –80°C. The cell components were used to separate PMNs by Ficoll₂ Hypaque density gradient centrifugation. PMNs were divided into control group (MP-) and MP stimulation group (MP+), and they were cultured in the medium of PMNs containing 10%

fetal calf serum in a humidified atmosphere containing 5% CO₂ at 37°C. MP stimulation group was treated with 10⁸ CFU of M129 ([Supplement 3](#)). This standard MP strain was provided by Professor Zhimin Chen from Zhejiang Children's Hospital.

Plasma Concentration of Inflammatory Factors

The levels of plasma interleukin-8 (IL-8) and granulocyte colony stimulating factor (G-CSF) were measured by using Enzyme-Linked Immunosorbent Assay (ELISA) (RayBio, America) according to the manufacturer's instructions.

PMNs Oxidative Stress and Generation of NETs

The intracellular reactive oxygen molecules (ROS) levels were determined by flow cytometry (DCFH-DA, Beyotime). In brief, the cell culture supernatants were collected and subjected to centrifugation under 3000 rev/min for 10 min at 4°C after 30 minutes of incubation. The level of myeloperoxidase (MPO) in the supernatant was detected by colorimetry (Sigma-Aldrich). Quantification of neutrophil extracellular traps (NETs) in the culture supernatants of PMN was determined by PicoGreen staining (Invitrogen) after 4 hours of incubation with cells.

Proteomic Analysis of PMNs

Eleven samples of PMNs were analyzed using label-free proteomics, including 7 PMNs specimens of different ages with MPP and 4 healthy children. Raw data files were acquired from triple TOF 5600. Protein identification was performed using the Andromeda search engine integrated with maxquant. Data are filtered to a 1% false-discovery rate (FDR). Each protein contains at least one unique peptide. While *P* value <0.05 and fold-changes >1.5 were considered significant using Welch's *t*-test. In total, 2985 proteins and 19,016 peptides were identified.

Model of BALB/c Mice with MPP

The mice aged 3–4 and 7–8 weeks old were provided by the Animal Experimental Center of Nanjing Medical University. Two independent trials of the experiments were performed. Six BALB/c mice were used for per group and randomly allocated to the different groups. Mice were anaesthetized with 1% pentobarbital Sodium, endotracheally infected with 1×10⁸ CFU of M129 MP strain in 10 μL PMNs culture medium ([Supplement 4](#)).

Histopathological Staining

Mice were infected with MP for 72 hours before euthanasia and their lungs were harvested. The right inferior lobe was fixed in 4% paraformaldehyde, followed by dehydration with ethanol and embedding in paraffin. Paraffin-embedded lung tissues were sectioned at 8 μm and processed for H&E and immunohistochemical staining. Immunohistochemistry was performed to detect PMNs infiltration using anti-Ly6G antibody (Abcam, America).

Preparation and Cytological Examination of Bronchoalveolar Lavage Fluid

Mice were infected with MP for 72 hours before euthanasia. Bronchoalveolar lavage fluid (BALF) was obtained by irrigation with 1 mL of PBS using a blunted needle inserted into the trachea after a tracheotomy. The BALF was centrifuged at 12000 rpm and 4°C for 10 minutes and stored at –80°C for further analysis. Cell components were used for measuring the total number of cytology and percentage of PMNs using Diff-Quik Stain (Solarbio, China).

Detection of Pulmonary Vascular Permeability of the Lung Injury

The residual left lung in the above experiment was weighed, and then dried in a 65°C incubator for 24 hours. The wet-to-dry weight ratio was calculated after 24 hours. Pulmonary vascular permeability was assessed by the ratio of protein quantity of alveolar lavage fluid to serum. Proteins in serum samples from mice were isolated. Protein concentration in the supernatant of 10% bronchoalveolar lavage fluid (BALF) and mice serum were determined using bicinchoninic acid (BCA) Protein Assay kit (Beyotime, China) according to the manufacturer's instructions.

Preparation of Lung Homogenate

Following euthanasia, the right lungs were dissociated from the pulmonary hilum. The inferior lobes of the right lung were separated and placed in a pre-frozen PBS balance salt. The 10% lung tissue homogenate was prepared by the fully automatic freeze-mixed ball mill and was centrifuged at 4°C 12000 rpm for 15 minutes, and the supernatant was aliquoted and stored at –80 °C until use.

Measurement of Surrogate Markers

Malondialdehyde (MDA) and MPO in 5% lung tissue homogenate were determined by thiobarbituric acid (TBA) colorimetric method (JIAN CHENG TECHNOLOGY, China) and ELISA (Fcmacs, America), respectively. NETs of BALF were examined with the PicoGreen fluorescent dye (Invitrogen, America).

Data Analysis

Statistical analyses were performed by using SPSS software, version 17.0 (IBM, Armonk, NY, USA). Data with normal distribution were expressed as mean±standard deviation, and compared using with the independent sample *t*-test and paired-samples *t*-test. The Levene homogeneity variance was analyzed for quantitative data of multiple groups. One-way ANOVA was performed on data with multiple groups before application of *t*-test for one-to-one comparison. The qualitative data were compared using Chi-square test. Linear correlation of quantitative data was analyzed using with the Pearson correlation analysis, and nonlinear correlation was analyzed using with the restricted cubic spline method. Statistical significance was defined as $P < 0.05$.

Results

Age and PMNs Counts Were Critical Independent Risk Factors for the Occurrence of RMPP

Age is reported to be associated with the incidence and adverse outcomes of MPP. To determine the optimal cut-off value of age for assessing the incidence of RMPP,

a total number of 1875 hospitalized children with pneumonia were enrolled in this study, including 298 children with Non-RMPP and 52 children with RMPP. The demographic and clinical data of these patients are presented in [Table 1](#). There were significant differences between non-RMPP group and RMPP group in age distribution and duration of fever. As for laboratory findings, there was no significant difference in gender, peripheral WBC counts, HB, PLT, ALT, AST and LDH between the two groups as well. However, we found that peripheral PMNs count and CRP in the RMPP group were significantly higher than those in the non-RMPP group. We then use multiple logistic regression analysis to further identify the risk factors of RMPP, and find that age and PMNs count are the key independent risk factors for the occurrence of RMPP. In order to improve specificity, 4.5 years old was taken as the cut-off value. The diagnostic sensitivity and specificity at the cut-off of 4.5 years old were 61.54% and 65.77%, respectively. The area under the ROC curve (AUC) was 0.68 ([Figure 1A](#)). The correlation coefficient between age and the percentage of PMNs in cases with RMPP was found to be 0.45 ([Figure 1B](#)). The age distribution of RMPP group is presented in [Figure 1C](#). As presented in [Table 2](#), the odds ratio (OR) values of each variable were calculated and evaluated. The incidence of RMPP in children aged 4.5~9 years was 4.22 times higher than that in children under 4.5 years old, and OR value was 1.90 to 9.38 (95% CI) by adjusted WBC, the count and percentage of PMNs, duration days of fever, and levels of HB, PLT, AST, ALT, LDH and CRP. Collectively, these data suggested that age affected the

Table 1 Demographic Data and Clinical and Laboratory Characteristics of Non-RMPP and RMPP Patients

	Non-RMPP (n=298)	RMPP (n=52)	P
Age, years	3.83±2.46	5.17±1.96	0.00
Gender (male/female)	298 (158/140)	52 (28/24)	0.56
Fever days	4.56±3.44	6.75±4.53	0.00
WBC×10 ⁹ /L	9.03±3.79	9.57±4.34	0.41
PMNs (%)	54.23±16.07	62.40±16.48	0.00
PMNs×10 ⁹ /L	5.05±3.04	6.09±3.32	0.03
HB (g/L)	123.23±11.44	126.42±7.92	0.05
CRP, mg/L	11.36±7.06	19.97±23.95	0.00
PLT×10 ⁹ /L	254.90±99.47	286.44±115.48	0.07
AST,U/L	35.15±16.31	33.15±34.37	0.68
ALT, U/L	15.67±11.65	25.91±38.59	0.06
LDH,U/L	365.02±172.72	382.72±249.2	0.62

Note: Data was presented as mean ± standard deviation.

Abbreviations: RMPP, refractory *Mycoplasma pneumoniae* pneumonia; WBC, white blood cells; PMNs, polymorphonuclear neutrophils; HB, hemoglobin; PLT, platelet; CRP, C-reactive protein; AST, glutamic oxaloacetic transaminase; ALT, glutamic-pyruvic transaminase; LDH, L-lactate dehydrogenase.

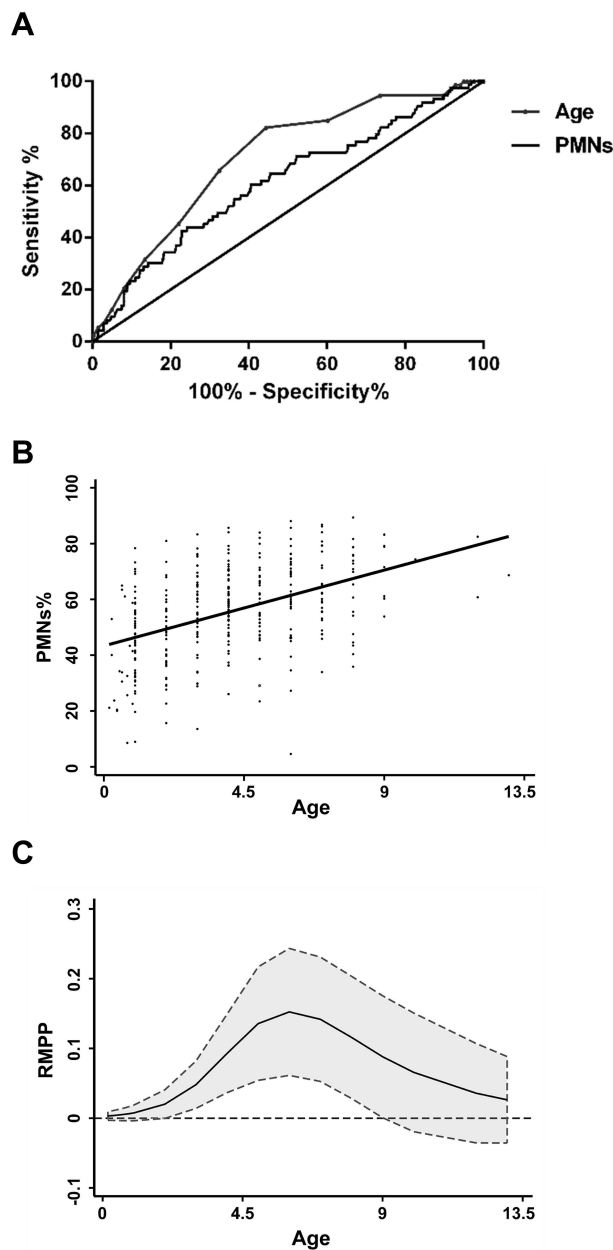


Figure 1 Age and PMNs associated with RMPP in children. **(A)** The areas under curves (AUC) of age and PMNs were 0.68 and 0.61, respectively. A cut-off of 4.5 years of age produced a sensitivity and specificity of 61.54% and 65.77%, respectively. **(B)** The PMN percentage of MPP patients in different age. The linear correlation coefficient was performed between age and PMNs ($P < 0.01$). **(C)** The solid line showed age distribution of odds ratio (OR) values for the occurrence of RMPP. The dashed lines indicated the 95% confidence intervals of OR.

percentage of PMNs in peripheral blood, and that age ≥ 4.5 years could partially predict the development of RMPP.

Age is Correlated with the Counts and Function of PMNs in Children with MPP

G-CSF can stimulate the proliferation and differentiation of granulocyte precursor cells in the bone marrow. To determine

Table 2 OR for RMPP by Various Factors

	OR	P
Age		
<4.5y	Ref	
4.5–9y	4.22[1.90–9.38]	0.00
>9y	0.91[0.16–5.13]	0.91
WBC	1.15[0.84–1.57]	0.40
PMNs%	1.02[0.96–1.08]	0.51
PMNs	0.93[0.56–1.53]	0.77
Fever day	1.16[1.06–1.27]	0.00
HB	1.02[0.99–1.06]	0.20
PLT	1.00[0.99–1.00]	0.02
AST	1.04[1.01–1.07]	0.01
ALT	0.94[0.90–0.98]	0.00
LDH	1.00[1.00–1.00]	0.40
CRP	0.94[0.90–0.99]	0.01

Notes: The incidence of RMPP in children aged greater than or equal to 4.5 to 9 was 4.16 times higher than that of children less than 4.5 years by adjusted gender, WBC, PMNs percentage, PMNs count, fever days, HB, PLT, AST, ALT, LDH and CRP. A nonlinear logistic regression analysis was performed using the restricted cubic splines method.

Abbreviations: RMPP, refractory Mycoplasma pneumoniae pneumonia; WBC, white blood cells; PMNs%, PMNs percentage; PMNs, polymorphonuclear neutrophils; HB, hemoglobin; PLT, platelet; AST, glutamic oxaloacetic transaminase; ALT, glutamic-pyruvic transaminase; LDH, L-lactate dehydrogenase; CRP, C-reactive protein.

the influence of age on PMNs counts in peripheral blood, serum inflammatory factors were detected in children with MPP. The MPP patients were classified into $<4.5y$ MPP group and $\geq 4.5y$ MPP group at a cut-off of 4.5 years of age. As documented in Table 3, it was found that PMNs counts were significantly higher in $\geq 4.5y$ MPP group than in $<4.5y$ MPP group. As shown in Figure 2A, the level of G-CSF was significantly increased in $\geq 4.5y$ MPP group compared with $<4.5y$ MPP or HC group. Proinflammatory cytokine IL-8 act as recruiter of PMNs infiltrating into the infection sites. A similar pattern was observed for both G-CSF and IL-8 levels in our study (Figure 2A).

To characterize the effect of age on activated PMNs function after MP infection, we separated and cultured primary PMNs in vitro. Compared with HC or MPP group, intracellular ROS fluorescence intensity (Figure 2B) in PMNs stimulated by MP, MPO (Figure 2C) and NETs (Figure 2D) in the culture supernatants of PMN at different post-infection time were significantly increased. The levels of ROS, MPO and NETs of PMNs in $\geq 4.5y$ MPP group were significantly higher than those of $< 4.5y$ age group with or without MP stimulation. These data suggest that age acts as a factor in inducing different activations of PMNs in response to MP.

Table 3 Comparison of Clinical Characteristics of Non-RMPP in Different Age Groups

	<4.5 y MPP (n=23)	≥4.5y MPP (n=23)	P
Age, years	2.78±1.00	6.00±1.91	0.00
Gender (male/female)	23 (12/11)	23 (13/10)	0.09
Fever days	6.14±3.79	9.44±5.36	0.04
WBC×10 ⁹ /L	9.11±3.06	9.24±3.12	0.91
PMNs (%)	63.50±15.68	76.77±9.86	0.02
PMNs×10 ⁹ /L	5.70±2.07	7.63±2.41	0.02
HB (g/L)	126.22±8.63	124.56±7.07	0.59
CRP, mg/L	13.68±9.32	13.78±9.47	0.98
PLT×10 ⁹ /L	257.03±88.51	267.89±6.05	0.73
AST,U/L	36.46±17.80	40.33±25.46	0.59
ALT, U/L	16.08±12.62	34.00±59.07	0.39
LDH,U/L	357.59±161.54	554.67±271.20	0.001

Note: Data was presented as mean ± standard deviation or number (percentage).
Abbreviations: MPP, Mycoplasma pneumoniae pneumonia; WBC, white blood cells; PMNs, polymorphonuclear neutrophils; HB, hemoglobin; PLT, platelet; CRP, C-reactive protein; AST, glutamic oxaloacetic transaminase; ALT, glutamic-pyruvic transaminase; LDH, L-lactate dehydrogenase.

PMNs Responded Diversely to MP Infection in Children of Different Ages

In order to further understand the functional differences of PMNs in children of different ages after MP stimulation, eleven PMNs specimens, including 4 cases of ≥4.5y MPP (A9, A21, A30 and A31), 3 cases of <4.5y MPP (A13, A28 and A37) and 4 cases of HC group were analyzed by label free proteomics approach. A total of 2684 proteins were identified, of which 46 proteins were classified as differentially accumulated proteins (DAPs), including 8 up-regulated DAPs (G3V126, Q6N093, MOQXF7, AOA024QZCO, E5KNY5, J3QSX6, Q8ND71 and Q9NZ08) and 38 down-regulate DAPs (W8QEY1, Q8BFZ3, V9HWO4, Q53Y51, P20292, AOA14OVJY7, C9J8E1, Q9NXV6 and so on) (Figure 3A and B). DAPs have been found to be associated with apoptosis, phagosome, nitrogen metabolism and leukocyte transendothelial migration, etc. (Figure 3C). In addition to chemotaxis and phagocytosis, PMNs function scores, such as apoptosis, nicotinamide adenine dinucleotide phosphate (NADPH), mitochondrion and oxidative stress, statistically increased in older MPP cases (Figure 3D), which indicate that there are multiple functional differences of PMNs after MP infection in children of different age.

Age Affected the Consequence of Mice with Experimental MPP

To further clarify the correlation between the infiltration and activation of PMNs and lung injury in mice of different ages, 3-week-old weaned and 8-week-old puberty male BALB/c mice were selected according to age conversion of human and the lifespan of mice.¹⁵ The model of BALB/c mice with MPP is established by intratracheal instillation with MP at 72 h post infection. Histopathological sections of the lung tissues stained with HE and showed that the alveolar septums were thickened obviously, and a large number of inflammatory cells were infiltrated in peribronchial, perivascular and vascular lumens (Figure 4A), which proved that the MPP mice model was established successfully.

Compared to 3-week-old mice group with MPP, the total number of cells (Figure 4B) and percentage of PMNs (Figure 4C) in BALF were significantly increased, and increasing numbers of pulmonary infiltrative PMNs were observed in histopathological sections in 8-week-old mice group with MPP by immunohistochemical staining using Anti-Ly6g antibody (Figure 4A). Meanwhile, the level of MDA (Figure 4E), MPO (Figure 4G) and production of NETs (Figure 4H) in lung homogenate were significantly higher in MP infected group of 8-week-old mice. Finally, the indicators related to lung injury, including the ratio of protein quantity of BALF to serum (Figure 4D), wet-to-dry weight ratio of lung tissue (Figure 4F) of 8-week-old mice MPP model, were significantly higher than those of 3-week-old mice group infected with MP. These data suggest that age-related aberrant infiltration and activation of PMNs in mice lung tissue lead to different degrees of pulmonary damage after MP infection.

Discussion

MP is the most common pathogenic microorganism in children over 5 years old in the America,^{1,16} it can also affect infants and young children during epidemic years in Denmark.¹⁷ In recent years, the onset age of MPP has shown a downtrend in Eastern China. However, RMPP mainly occurs in school-age children.¹⁸ Our findings indicate that age is a key factor in the prognosis of MPP, which is consistent with previous reports.

This study presents novel data on the effect of age on the number of circulating PMNs after pulmonary MP infection. Lymphocytes are the most abundant leukocyte type in the circulation of children aged from 4–6 days to

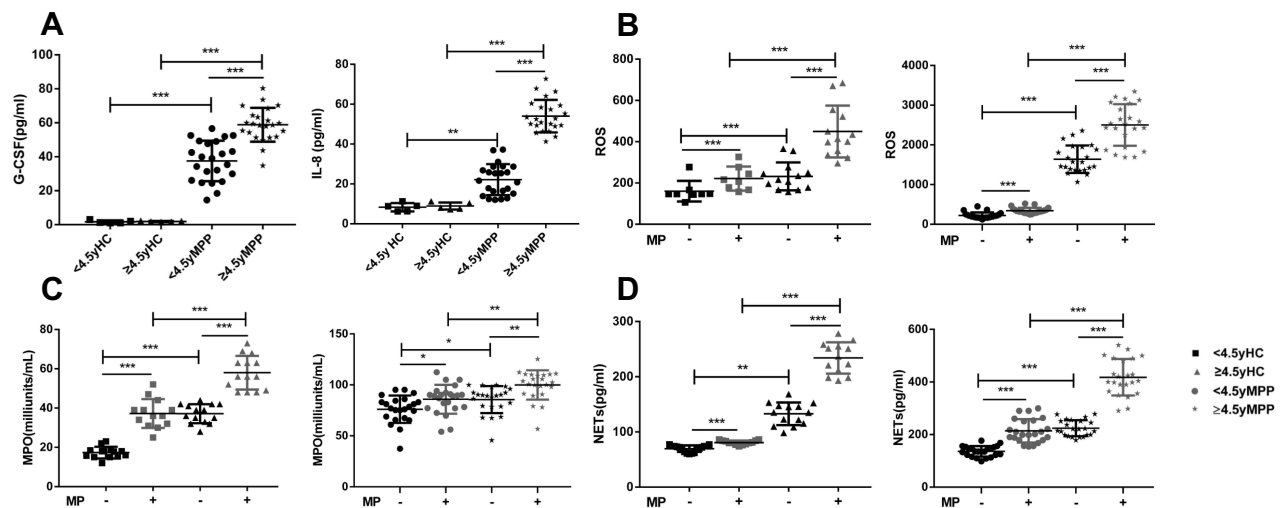


Figure 2 Comparisons of the inflammation indicators and PMNs function in vitro among the health control, <math><4.5y</math> MPP group and $\ge 4.5y$ MPP group. **(A)** The quantity of PMNs from peripheral blood and pulmonary infiltration influenced by G-CSF and IL-8 respectively. ($n = 5-23$). **(B)** Inter-cellular ROS fluorescence intensity in PMNs were detected at 30 minutes with or without 10^8 CFU MP infection ($n = 15$). **(C)** The levels of MPO in culture supernatants of PMNs, isolated from HC or MPP cases co-cultured with or without 10^8 CFU of MP for 1 hour ($n = 15$). **(D)** NETs production by PMNs after 4 hours post infection ($n = 15$). Data was presented as mean \pm standard deviation. *** $P < 0.001$, ** $P < 0.01$, * $P < 0.05$.

4–6 years, the percentage of PMNs continue to increase with age and gradually reach adult level after 4–6 years of age. Since G-CSF drives myeloid progenitors to bias differentiation towards the granulocytic lineage and regulates production of neutrophils by controlling their release into blood,¹⁹ so we may speculate that age affects the amounts of PMNs in acute inflammation after MP infection by increasing G-CSF.

In this study, we found that age can also affect the activation function of PMNs stimulated by MP. Human PMNs were capable of triggering superoxide and MPO releasing.²⁰ PMNs separated from the elder cases showed enhanced respiratory burst and NETs production. Therefore, it is suggested that age has an impact on the activation of PMNs through promoting the secretion of IL-8, and may be related to the adverse consequences of MPP.

It is well known that NETs formation occurs via NADPH oxidase (NOX)-dependent and mitochondria NOX-independent pathways. The suppression of mitochondrial oxidative stress reduced NETs release from PMNs of aged mice.²¹ Functional analysis of PMNs implies that the enhancement of mitochondrial function in elder MPP patients could be related to the increase in injury NETs and oxidative stress injury.

Recently, a large amount of studies focused on abnormal cellular immunity in children with MPP. Macrophages are implicated in the clearance of MP.²² A decreased percentage of macrophages and an increased

percentage of PMNs in bronchoalveolar lavage fluid were observed in children with MPP,²³ as well as RMPP.²⁴ Recruitment of PMNs into the lung tissue is more associated with the severity and progression in mice with MPP.²⁵ In mice model with MPP, increased PMNs in airspaces and interstitium mediate microvascular damage, pulmonary edema, leading to lung injury. Meanwhile, reduced pulmonary infiltration of PMNs decreased lung injury in immunosuppressed mice with MPP. Therefore, PMNs are deemed to play an important role in the development of MPP. Inhibition of pulmonary PMNs infiltration could reduce lung injury. In our study, we focused on the effects of age on the mobilizing function of PMNs stimulated by MP. In pulmonary infection, activated PMNs eliminate invasive pathogens by producing abundant ROS/nitrogen species, concomitant delivery of MPO and other antimicrobial enzymes into the phagosome.²⁶ MPO represents one of the most abundant protein enzymes that participates in immune defense through the formation of microbicidal reactive oxidants and diffusible radical species.²⁷ However, overproduction of MPO will lead to oxidative stress and tissue damage when they exceed the limits of local antioxidant defense protection. Long-term exposure to a high concentration of ROS can lead to inflammatory injury, which may be associated with cardiovascular diseases,²⁸ neurological diseases,²⁹ cancer and aging.³⁰ Our study confirmed that age affects the “oxidative burst” of PMNs after MP

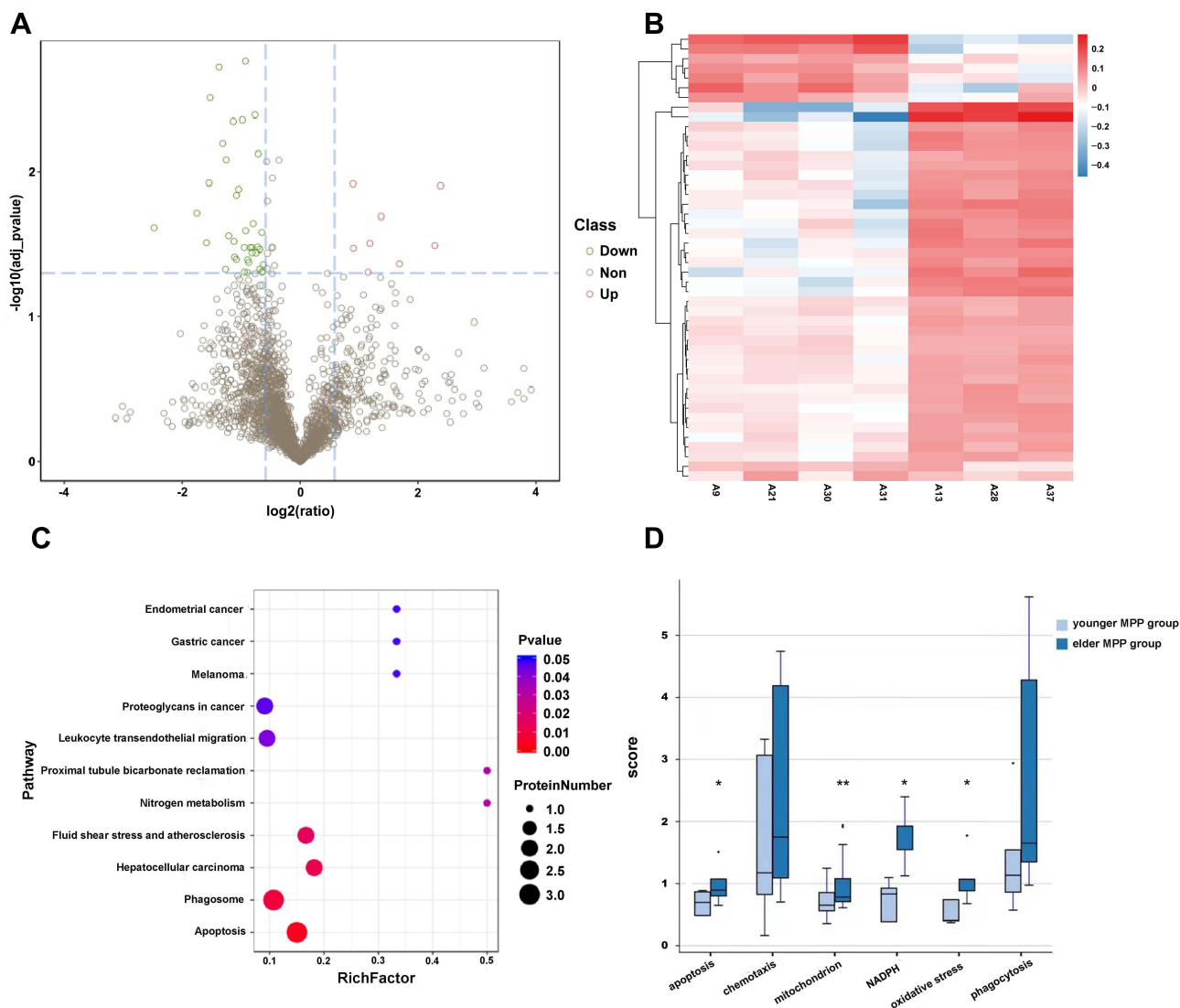


Figure 3 Proteomics analysis of PMNs from peripheral blood of children of different ages with MPP. **(A)** Volcano map. The red dots indicate significantly up-regulated proteins and the green ones indicate down-regulated proteins using the criteria of fold change > 1.5 and *P*-value < 0.05. **(B)** Heat map. Hierarchical Cluster to cluster the differential proteins. The PMNs specimen of A13, A28 and A37 were from MPP patients under 4.5 years old, and A9, A21, A30 and A31 were from MPP cases greater than or equal to 4.5y. **(C)** Bubble chart. This figure shows the metabolic pathway in which the differential protein is significantly enriched. The X-axis enrichment Factor (RichFactor) is the number of differential proteins annotated to the Pathway. (n=3–4). **(D)** PMNs function score. MPP cases greater than or equal to 4.5y was elder MPP group, and MPP patients under 4.5 years old was younger MPP group.

infection, and eventually contributes to different degrees of lung injury.

Recent studies have shown that NETs are another mechanism for PMNs to eliminate pathogens by forming physical barriers and its chemical bactericidal effects, which depends on the transport of MPO from granules to nucleus³¹ and the release of mitochondrial DNA triggered by ROS.³² Meanwhile, NETs play a critical role against fungal infection,³³ and respiratory syncytial virus³⁴ in vitro. Furthermore, NETs are thought to be associated with the pathological damage of severe influenza virus infection. Excessive NETs can also damage normal tissues, leading to

thrombosis,³⁵ acute respiratory distress syndrome³⁶ and so on. In this study, age was found to be related to PMNs' production of NETs stimulated by MP in human and mice. Excessive NETs release results in MP-induced pulmonary tissue damage. Therefore, NETs clearance might be helpful in attenuating pathological injury of MPP.

Conclusion

This study identified that age is associated with the prognosis of MPP by increasing PMNs' counts and activation in children under 9 years old, which could explain the difference in age sensitivity to MP infection. New therapeutic strategies

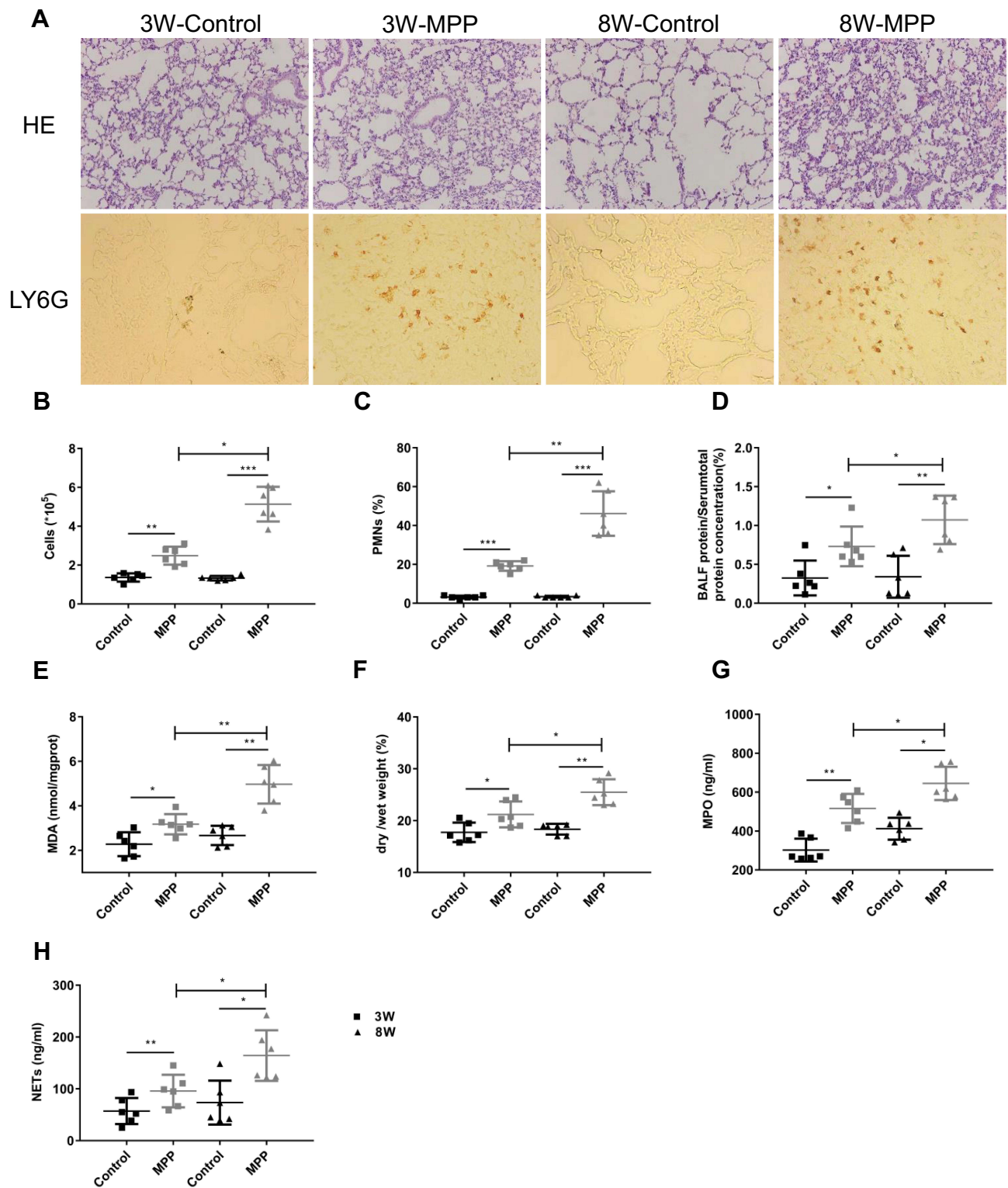


Figure 4 Elder mice had more severe lung injury during pulmonary MP infection compared to younger mice. **(A)** HE staining of lungs sections from BALB/c mice at 72 hours after infection with MP ($\times 100$). Immunohistochemical staining of PMNs using anti-Ly6G antibody in the lung sections from BALB/c mice with MPP ($\times 400$). **(B)** Cytological count and percentage of PMNs **(C)** in alveolar lavage fluid of mice infected with MP after 72 hours ($n=6$). **(D)** The ratio of protein quantity of alveolar lavage fluid to serum. **(E)** Levels of MDA. **(F)** The ratio of wet-to-dry weight ratio of lung tissue. MPO **(G)** and NETs **(H)** in lung homogenate of mice after 72 hours post infection. The levels of wet-to-dry weight ratio, MDA, MPO and NETs were detected in 6 uninfected and infected BALB/c mice were used for this experiments. $***P<0.001$, $**P<0.01$, $*P<0.05$.

that inhibit neutrophil recruitment and target neutrophil function can improve the adverse outcome of RMPP.

Abbreviations

RMPP, refractory *Mycoplasma pneumoniae* pneumonia; MPP, *Mycoplasma pneumoniae* pneumonia; PMNs, polymorphonuclear neutrophils; HC, healthy control; IL-8, interleukin-8; G-CSF, granulocyte colony stimulating factor; AUC, areas under curves; OR, odds ratio; ROS, reactive oxygen species; MPO, Myeloperoxidase; NETs, neutrophil extracellular traps; MDA, malondialdehyde; MP-, without *Mycoplasma pneumoniae* stimulation; MP+, *Mycoplasma pneumoniae* stimulation.

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Disclosure

The authors report no conflicts of interest in this work.

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