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Infectious Diseases of the Central Nervous System

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Prevalence of neurologic disease in cats and the preponderance of cases with no known cause

Neurologic disease is seen commonly in cats. For example, neurologic cases make up approximately 10% of the case load of two separate feline medicine referral clinics (D.A. Gunn-Moore, BVM&S, PhD, Edinburgh University Feline Clinic data, 2004) [1]. There are many well-documented causes of neurologic disorder in cats [2,3], and infectious causes are believed to account for 30% to 45% of cases [1,4]. It is important to realize that a specific cause cannot be identified in 12% to 40% of cases, however. This holds true when looking at clinical cases [2,5,6] and histopathologic data (Table 1) [1,4]. In addition, although there are many known infectious causes (Box 1), a large number of cases (35%-40%) are found to have histopathologic changes suggestive of viral infection (that is not consistent with feline infectious peritonitis [FIP]), but no causal agent can be identified (Table 2) [1,4]. This group becomes even more significant if you consider cats with particular clinical signs. For example, of 30 cats that were investigated for having recurrent seizures, all were found to have structural brain disease and 14 (47%) had nonsuppurative meningoencephalitis suggestive of a viral infection, but no infectious agent could be found [5].

From this, we can see that infectious disease is a common cause of central nervous system (CNS) disorders in cats. In addition, the most common infectious agents are feline coronaviruses (FCoV, which can cause FIP) and some other, as yet unidentified, infectious agent(s), which are probably

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| histopathologic diagnoses in 280 felline neurology cases (University of Bristol 1975–1998) | | | | | |
|--|--------|---------|--|--|--|
| Cause | Number | Percent | | | |
| Inflammatory/infectious | 92 | 32 | | | |
| No abnormalities detected | 51 | 18 | | | |
| Degenerative | 42 | 15 | | | |
| Neoplasia | 38 | 13 | | | |
| Feline dysautonomia | 27 | 9 | | | |
| Feline spongiform encephalopathy (FSE) | 24 | 8 | | | |
| Congenital | 12 | 4 | | | |

| Table 1 | | | | |
|------------------------------|--------------------|----------------------|------------|-----------|
| Histopathologic diagnoses in | 286 feline neurolo | gy cases (University | of Bristol | 1975-1998 |

Data from Bradshaw JM, Pearson GR, Gruffydd-Jones TJ. A retrospective study of 286 cases of neurological disorders of the cat. J Comp Pathol 2004;131:112–20.

viruses [1,5]. Unfortunately, because we cannot identify the cause, we do not know how best to treat these cases, nor do we know how to prevent them. It is therefore essential that we try harder to identify the etiology behind the pathologic findings. This means performing more detailed diagnostics on individual clinical cases, such as looking for potential infectious organisms by serology, cerebrospinal fluid (CSF) IgG quotient, and IgG index [6]; CSF polymerase chain reaction (PCR) assays to detect the infectious organism's DNA or RNA [7]; and immunohistochemistry or PCR on brain samples collected postmortem. In addition, it underlines the need for further experimental investigation into potential pathogens. It is only by adopting a more questioning approach to disease pathogenesis that we can hope to determine what may be causing neurologic disease in many of our pet cats.

Reasons for the increased recognition of infectious central nervous system disease in cats

Improved diagnostics and changing concepts in disease pathogenesis

Over the past 10 years, there has been a dramatic increase in the recognition and understanding of many different infectious diseases and of how they can affect the CNS. Advances in molecular technology have enabled the detection of pathogens within the CNS (eg, by using PCR assays), led to the recognition of new infectious diseases, and expanded our understanding of the etiopathogenesis of these infections.

With the changing understanding of etiopathogenesis, we have had to redefine our concept of "infectious disease." No longer can we think that infectious agents can only cause acute disease that classically fulfills Koch's postulates of cause and effect. For example, the acute disease that is seen when a bacterial infection spreads from otitis media to cause suppurative meningoencephalitis [8]. We now know that some diseases result from chronic insidious infections and that this is particularly true within the relatively protected confines of the CNS. In addition, progressively more

Box 1. Naturally occurring infectious causes of central nervous system disease in domestic cats

Viral

Feline coronavirus (FCoV)^a Feline panleukopenia virus (FPV) Feline immunodeficiency virus (FIV) Feline leukemia virus (FeLV) **Rabies virus** Aujeszky's disease virus Feline herpesvirus-1 (FHV-1) Borna disease virus (BDV) Certain arboviruses (see text) Bacterial Pasturella Staphylococcus Other aerobic organisms Anaerobic organisms Mycobacteria Protozoal **Toxoplasmosis**^a Rickettsial Ehrlichiosis Fungal Cryptococcosis^a Blastomycosis Histoplasmosis Aspergillosis Dematiaceous fungi Parasitic Cuterebra larval myiasis Visceral larva migrans (eg, Toxocara) Sarcocystis Dirofilaria immitis Probable and other Feline spongiform encephalopathy (FSE) Feline polioencephalomyelitis and miscellaneous nonsuppurative (meningo)encephalitides^a

^a Most common causes of encephalitis in cats. Others are sporadic and rare. *Data from* Refs. [8,24,36,37,44,167].

| Ta | ble | 2 |
|----|-----|---|
| | ~ | _ |

Diagnoses in 92 feline neurologic cases found to have central nervous system histopathology consistent with inflammation and/or infection (University of Bristol 1975–1998)

| Cause | Number | Percent | |
|--|--------|---------|--|
| Feline infectious peritonitis (FIP) ^a | 47 | 51 | |
| Viral (non-FIP) ^b | 32 | 35 | |
| Protozoal cysts (eg, toxoplasmosis) | 8 | 9 | |
| Bacterial infection | 3 | 3 | |
| Feline immunodeficiency virus (FIV) | 1 | 1 | |
| Cryptococcosis | 1 | 1 | |

^a One of the cats with FIP was also found to have an incidental nematode larvae (*Toxocara*) within its lateral ventricle.

^b Nonsuppurative meningitis and/or encephalitis was present, but no cause could be found. Five of these cats also had changes consistent with feline spongiform encephalopathy.

Data from Bradshaw JM, Pearson GR, Gruffydd-Jones TJ. A retrospective study of 286 cases of neurological disorders of the cat. J Comp Pathol 2004;131:112–20.

diseases are being identified that, although being associated with the presence of a particular pathogen, require a number of other factors to occur concurrently before disease becomes apparent. For these diseases to develop, there has to be a specific interaction between the infectious organism, host factors (especially genetics affecting the immune system), and the environment.

Unfortunately, establishing a causal relation can be difficult, particularly when the prevalence of the infection is high in the general population but only a few individuals have the necessary factors required for clinical signs to develop. Serologic surveys have been largely responsible for recognizing the role of infectious organisms in this type of disease. After detecting a serologic relation, it is then possible to use more complex molecular biology techniques to detect the pathogen within a particular individual or particular pathologic lesion. It is highly likely that it will be by using this type of approach that the causes for many feline CNS disorders will be found (eg, Borna disease [BD]).

Changing population dynamics

Populations are changing; people are living in progressively larger urban groups, and international travel is now commonplace. This allows for rapid spread of disease, not only among human beings but from human beings to other species. In addition, as the global human population increases, the demand for housing means that previously unexplored habitats are being developed, new pathogens are being exposed, and old pathogens are finding new hosts. There are a number of examples of infections that have crossed between species because of altered population dynamics, and many of them involve feline species. Examples include canine distemper virus (CDV), which is now causing disease in a number of large feline species, particularly lions in the Serengeti [9], but has not yet been detected in domestic cats [10];

106

severe acute respiratory syndrome (SARS), which is caused by a coronavirus that seems to have been passed from civet cats in China to human beings (although the civet cat is not actually a feline species) [11,12]; avian influenza virus (H5N1), which has killed domestic and captive wild felids in Thailand [13]; and West Nile virus (WNV), which is now present in the United States, being spread by mosquitoes to many wild and captive birds as well as to horses and humans beings [14].

WNV is a particularly interesting infection to consider. This is because experimental studies have shown that it is relatively easy to transmit WNV to cats by a mosquito bite or orally via consumption of infected prey [15]. Experimental cats in one study showed only mild nonneurologic signs [15]. This does not mean that WNV cannot cause neurologic disease in cats, however. This is because dogs in the same study remained perfectly healthy [15] but have been shown to develop encephalitis and myocarditis in a separate study [16]. To date, domestic cats tested in New York City have not been found to be seropositive for WNV [17]. Seropositive cats have been identified in the United States, however, and several cases demonstrated seroconversion coincident with neurologic illness (A. Glaser, DVM, PhD, Cornell University Animal Health Diagnostic Laboratory, personal communication, 2004). The potential role that WNV may play in feline neurologic disease requires further investigation.

Increasing demand for inexpensive food

An increasing demand for inexpensive food has resulted in a growing number of food-related infections. In addition to classic types of food poisoning, the transmission of the transmissible spongiform encephalopathies (TSEs) should be considered in this group.

Increasing awareness of zoonotic infections

It is perhaps of some concern that there has been a particular increase in the recognition of zoonotic conditions (diseases that can be spread from animals to people). In fact, three quarters of all emerging human pathogens are zoonotic [18]. Because of this, it is important that we raise our general awareness of this type of disease and monitor closely for any evidence of interspecies transfer of infections.

By studying the genetic relation between pathogens and performing infectivity studies, a number of infectious agents have already been identified that can cross species barriers and raise the possibility of zoonotic infection. Two such diseases are feline spongiform encephalopathy (FSE) and BD. In both cases, the infectious agents can infect a number of mammalian species, including cats and people. In addition, both infections have a poorly understood etiopathogenesis and may result in terminal neurologic disease.

Known causes of central nervous system infection in cats

From the data shown in Tables 1 and 2, we can see that FIP and nonsuppurative encephalitides (also called viral non-FIP encephalitides) are the only two commonly recognized potentially infectious causes of CNS disease in the cat. Other infections, for example, toxoplasmosis, feline leukemia virus (FeLV), feline immunodeficiency virus (FIV), feline panleukopenia (FPV), and fungal and parasitic infections, are seen only rarely. Detailed summaries of the pathogenesis, clinical signs, diagnostic approach, treatment, and neuropathologic findings of these specific infections are readily available [1,4,6,7,19–24]. The rest of this article therefore adds only selected comments in relation to these organisms and focuses on the nonsuppurative encephalitides and the more unusual infections for which information is less readily available.

Feline infectious peritonitis

FIP is the most commonly detected infectious cause of neurologic disease in cats. It accounts for 45% to 50% of all cases associated with inflammatory changes, which equates to 15% to 20% of all feline neurologic cases [1,4]. It is essential to realize just how common this infection is and that although most clinical cases are seen in young pedigree cats, usually with obvious systemic involvement, this is not always the case.

As with many viral infections affecting the CNS, histopathologic examination reveals nonsuppurative meningoencephalomyelitis, with perivascular cuffing and meningeal infiltration with mononuclear cells, gliosis, and variable neuronal degeneration. The inflammation is often pyogranulomatous, is located around the lateral ventricles or in the meninges, or may affect the choroid plexus. In addition, vasculitis or acquired hydrocephalus may be present, and systemic changes are usually apparent [1,25,26].

Feline immunodeficiency virus

FIV can cause neurologic disease as a direct primary neurotropic effect of the virus or via secondary opportunistic infections, such as FIP, toxoplasmosis, or cryptococcosis [27–32]. It is important to note that this virus cannot be excluded on the basis of negative serology. This is because several studies have failed to detect antibody in some virus-positive individuals [27,33], and in one report, approximately 20% of cats naturally infected with FIV were antibody-negative [33]. Although the reason for this is unknown, it has been suggested that some cats were tested early in the course of infection, before the development of an antibody response. These cats are unlikely to show CNS disease. Alternatively, some cats may have been tested late in the disease, once antibody levels had fallen, along with the terminal decline of the immune system [27]. Other factors may also play a role, because some cats fail to produce a detectable antibody response to FIV at any time during infection [33]. Therefore, to confirm that a cat is not infected with FIV, it may be necessary to perform PCR on a blood or CSF sample or PCR or immunohistochemistry on brain sections collected postmortem [32].

Rare or experimental infectious causes of central nervous system disease in cats

Naturally occurring and clinically significant CNS infections have occasionally been seen in cats, resulting from a wide range of organisms. These include feline herpesvirus-1 (FHV-1) [34], feline calicivirus (FCV) [35], dematiaceous fungi (*Cladophialophora bantiana*) [36], and nematodes (eg, *Sarcocystis neurona*) [37].

It is also prudent to consider clinically significant CNS infections that have been produced experimentally in cats. These include *Bartonella henselae* [38], which typically causes cat scratch disease in people [39]; equine herpesvirus-9 [40]; Newcastle disease virus [41]; human poliovirus [42]; and simian cytomegalovirus-related stealth virus (which was taken from a human being with chronic fatigue syndrome) [43].

Much speculation has concerned the arboviruses (the arthropod-borne encephalomyelitis group) and whether or not they may be responsible for significant natural CNS infection in cats [44]. Many of these viruses have been shown to cause natural or experimental infection in cats. For example, natural infections have been seen in cats with BD virus (BDV), which can be subclinical or clinically significant. Antibodies to St. Louis encephalitis virus, Japanese encephalitis virus, yellow fever virus, Tenshaw virus, Snowshoe hare virus, Jamestown Canyon virus, and Powassan virus have been found in free-living cats, indicating previous subclinical infections [45-47]. Experimental infections have been documented with Near Eastern equine encephalitis virus [48,49], Powassan virus [46], and Rift Valley fever virus in kittens [47]. Many of these viruses have a wide geographic distribution, including the United States and Europe, and some are endemically present in wild mammal populations to which free-living cats may become exposed [50]. Although these studies show that cats are potentially at risk of becoming infected with these organisms, their role in causing a significant incidence of naturally arising feline neurologic disease remains to be determined.

Borna disease virus

Epizootiology

Classical BD is a severe neurologic disease that is seen predominantly in horses and sheep in endemic areas of Germany and Switzerland. Natural infections have also been seen in cats and ostriches and, occasionally, in rabbits, cattle, goats, deer, foxes, and dogs [51–59]. Experimentally, BDV can also be transmitted to birds, rodents, and monkeys, and it is likely that the host range includes all warm-blooded animals and birds [52,58,60,61]. The exact geographic distribution of the virus is uncertain, but serologic evidence has documented infection in Europe, the United States, and Asia [62–70].

BD in cats is also known as "staggering disease." It was first described in Sweden [71] and later shown to be caused by BDV [72]. Based on serologic surveys or surveys looking for BDV RNA in peripheral blood samples, it is clear that BDV infection is usually asymptomatic. The prevalence of seropositivity increases steadily with age in cats [72,73] and, interestingly, seems to be higher in cats that are also FIV antibody-positive [68,74]. In the United Kingdom, 6% of cats with no evidence of CNS disease have antibodies against BDV [65], as do 9% of ill cats submitted for FIV, FeLV, and FIP virus testing (D.A. Harbour, PhD, unpublished results from 654 cat blood samples, 1999). In Japan, 13% to 22% of healthy randomly selected cats are seropositive for BDV [62,75]. Antibodies against BDV or BDV RNA are seen most frequently in cats with neurologic disease, however. When cats with undefined neurologic disorders were investigated, 13% were found to be BDV antibody-positive in Germany [76] compared with 35% in the United Kingdom [65] and 67% in Japan [77]. Although most documented cases of feline BD have originated from northern and central Europe, probable cases have been seen in many other countries [65] (see section on nonsuppurative encephalomyelitis of unknown cause). Given the difficulty in making a premortem diagnosis (and even a postmortem diagnosis) and the low index of suspicion, it is likely that BD is underdiagnosed.

Pathogenesis

The source of BDV infection is rarely known. Infected cats usually, but not always, have outdoors access, particularly to rural or woodland areas, however [78–80]. This has led to the suggestion that rodents or wild birds may be viral carriers [80–82]. Natural infections are believed to be transmitted via saliva or nasal secretions [52,73,83].

BDV is a neurotropic RNA virus [84,85] that is genetically stable [86,87]. It is ubiquitously distributed and seems to have many well-adapted species-specific biotypes [54,72,88–90]; in most cases, infection causes little or no sign of disease. It is currently believed that clinical signs only develop when a host is exposed to a particular strain of BDV [57,62,73,91] or is particularly susceptible and mounts an abnormal immune response to the virus (ie, disease seems to result from a T-cell-dependent immune mechanism) [60,91–93].

Clinical signs

Natural BD has been reported in more than 100 cats [54,78–80,94]. It is seen most frequently in male cats, with no particular breed predisposition.

Although a wide age range of cats may be affected (from 5 months to 11 years of age), young adults seem to be most at risk [54,78,79]. Disease is characterized by behavioral and motor disturbances resulting from meningoencephalomyelitis. In experimental infections, clinical signs included protrusion of the third eyelid, behavioral changes, circling, ataxia, and tremors [72]. Natural infections may present with progressive hind limb ataxia, loss of appetite, fever, increased affection toward the owner, unusual staring expression, apparent pain over the sacrum, increased salivation, aggression, an inability to retract claws, seizures, focal or generalized pruritus, hypersensitivity to light and sound, or constipation [76,78,79]. Occasional atypical cases have been seen, for example, causing muscle fasciculation and proprioceptive defects (without evidence of encephalitis) [95]. Disease is usually progressive, and mortality rates are high because affected cats usually warrant euthanasia within a week to 6 months [78,79]. Cats that survive the initial episode may remain chronically infected or may experience recurrent episodes of disease [72]. Although fatal BD is seen most commonly as a rare isolated event, it can occasionally occur as a large outbreak, where as many as 30 to 40 cases may be seen in a week [78].

Diagnosis

Premortem diagnosis is difficult. In most cases, typical clinical signs in a cat from an endemic area result in a presumptive diagnosis of BD. Unfortunately, detection of serum antibodies is not reliable. This is because although raised serum antibodies are present in some cats with BD (\sim 40%) [76], particularly those with acute disease, others may be antibody-negative, particularly if they have subacute or chronic disease [54,62,65,72,79]. Although clinical signs of BD tend to develop at the same time that BDV RNA can be detected within the peripheral blood [89], this does not necessarily reflect the extent of the viral load in the CNS [60], and asymptomatic cats can also be positive [75]. Routine serum biochemistry and hematology are generally unremarkable, although some cats may show a leukopenia, and mild elevations in glucose or alanine aminotransferase (ALT) levels may also occur [72,78]. CSF analysis may show a leukocytosis with mononuclear cells predominating, protein levels may be increased (Table 3) [78], and antibodies to BDV may be detected [92].

Histopathologically, BD usually results in nonsuppurative meningoencephalomyelitis, with neuronophagia, microgliosis, and heavy perivascular cuffing by mononuclear cells [10,72,78]. Occasional cases seem to result in neurologic signs without evidence of associated inflammation [95]. In most cases, lesions are particularly evident in the gray matter of the cerebral hemispheres, the limbic system, and the brain stem [10,54,78]. The cerebellum and spinal cord are less frequently affected [79].

Confirmation that BDV is the cause of this disease has been demonstrated by experimental transmission studies to cats and rabbits [72,79]. In

| Disease | CSF Pressure | CSF Appearance | WBC Count ^a | WBC Type | Total Protein concentration ^b | Albumin | Globulin | CSF Antibodies detectable | Organisms visible |
|---|-------------------|--------------------------|------------------------|-------------------------|---|---------|----------|---------------------------------|----------------------|
| Feline infectious peritonitis (FIP) | WNL or \uparrow | Clear or turbid | +++ (WNL- ++) | PMN-mono- mixed | +++ (WNL-+) | ++ | ++ | Yes | No |
| Other viral encephalitis (eg, Borna disease) ^c | WNL | Clear (turbid) | + (++) | Mono | + (++) | WNL | ? | No | No |
| Protozoal meningoencephalitis (eg, toxoplasmosis) | WNL or \uparrow | Xanthochromic | + (++) | Mixed-PMN, eos, mono | + (++) | + | + | Variable | Rarely |
| Fungal meningoencephalitis (eg, cryptococcosis) | ↑ or viscous | Turbid, xanthochromic | ++ | Mixed-PMN, mono, eos | ++ | ++ | +(+) | Varies | Varies |
| Bacterial meningitis | WNL or \uparrow | Turbid | ++ (+++) | PMN (mixed) | ++ (+++) | ++ | ++ | Varies | Yes (varies) |

Table 3 Infectious disease and typical cerebrospinal fluid changes

Abbreviations: CSF, cerebrospinal fluid; eos, eosinophils predominate; FIP, feline infectious peritonitis; mono, mononuclear cells (ie, lymphocytes, monocytes, macrophages) predominate; PMN, polymorphonuclear cells (neutrophils) predominate; WBC, white blood cell; WNL, within normal limits; \uparrow = increased.

^a Reference range for WBC count = <4 per microliter; +=5-80 per microliter; ++=81-500 per microliter; +++=>500 per microliter.

^b Reference range for total protein concentration = <25 mg/dL; + = 25–100 mg/dL; ++ = 100–300 mg/dL; ++ = >300 mg/dL.

^c Some viral infections cause neuropathologic changes without inflammation, and these may alter the CSF little [43].

Symbols in parentheses indicate less frequently seen variations.

Data from Refs. [4,10,20,72,167].

addition, BDV antigen may be detected by immunohistochemistry or enzyme-linked immunosorbent assay (ELISA) [54], and BDV RNA may be detected by PCR; all three methodologies can be performed on brain samples. Clinical cases are most easily confirmed by detecting BDV RNA within the inflamed areas of the brain using PCR [54,65,72,96,97].

Treatment

There is no specific treatment for BD. Supportive care and corticosteroids may help in some cases. Prednisolone may be given orally at a rate of 1 to 2 mg/kg every 24 hours until clinical signs regress, after which it should be reduced gradually over several weeks or months.

Zoonotic risk

It is currently unclear what role BDV may play in the induction of human disease. Antibodies against BDV, BD viral proteins, and BDV RNA have been found in people in Europe, the United States, and Asia. A higher prevalence of infection is seen in patients with neurologic or psychiatric disorders, particularly schizophrenia and uni- or bipolar disorders [61,70,98–106]. Because the virus has also been detected in clinically normal patients [67,101,107], however, its role in the development of these complex psychiatric disorders has still to be proven [61,63].

The presence of BDV infection in many domestic species as well as evidence of cross-species transfer raises the possibility of zoonotic spread. Although animal species may pose a potential risk to people, finding BDV RNA in blood from normal human blood donors suggests that people may also be at risk from horizontal spread from person to person [64]. Considerably more investigation needs to be performed before the zoonotic potential of BDV can be determined.

Nonsuppurative encephalitides of unknown cause

Introduction and geographic distribution

A number of other nonsuppurative encephalitides have also been described in cats. These seem to comprise a group of diseases that are possibly related, and the histopathologic changes suggest a viral origin. They are geographically widespread and have been reported in Australia [108], the United States [109], Canada [4,5,23], Sweden [10,71,78], Norway [110], Switzerland [44], and the United Kingdom [1,111,112]; other potential cases have been seen as widely distributed as Morocco [113] and Sri Lanka [114]. A similar condition has been found in a number of large cats, including lions and tigers [115–118]. In all cases, the reports state an unknown cause but comment that the histopathologic changes are suggestive of viral infection. Unfortunately, early studies performed few diagnostic inves-

tigations to try to determine the possible cause. Later studies usually assessed for FIV, FeLV, FIP, and *Toxoplasma gondii* and, in some cases, for FHV-1, FCV, FPV, and *Borrelia burgdorferi*. In almost all cases, the cats have been found to be negative for all these agents [1,4,5,10,23,44,112].

Reviewing the data available on these cases suggests that although these diseases generally affect cats of a similar age and sex and cause a range of rather similar clinical signs, they seem to separate into two groups based on histopathologic changes:

- Group I: CNS histopathologic examination reveals nonsuppurative encephalomyelitis [44,78,79,110]
- Group II: CNS histopathology reveals polioencephalomyelitis or polioencephalitis [44,108,109,112,119]

Group I: nonsuppurative encephalomyelitis

Clinical signs

These cats are of a wide age range (from a few months to >18 years), but young adults seem to be overrepresented. They show no sex or breed predisposition. They tend to have an acute duration of illness and typically develop ataxia, nystagmus, seizures, head tremor, anorexia, apathy, fever, and, occasionally, preceding vomiting and diarrhea (although not all cats show all signs) [4,6,10,71,78].

Diagnosis

A premortem diagnosis is typically based on the presence of suggestive clinical signs and typical CSF changes (see Table 3) [4,6]. Some cats show leukopenia or a mildly increased ALT concentration [4,78]. Neuroimaging of the brain may reveal multifocal areas of contrast enhancement suggestive of inflammatory disease.

Histopathologic examination reveals mild to severe nonsuppurative meningoencephalomyelitis characterized by mononuclear perivascular cuffing, inflammatory nodules of lymphocytes and macrophages, and neuronal degeneration. Changes can occur throughout the brain and spinal cord but are most prominent in the thalamocortex and brain stem [4,78]. Lesions may be diffuse or focal [4].

Treatment

There is no curative treatment, but supportive therapies include anticonvulsants to control the seizures and, possibly, corticosteroids to reduce CNS inflammation. Because the disease is often self-limiting, the prognosis is quite good for those cats in which neurologic signs are not too severe [6].

Potential causes

Interestingly, when blood samples from affected Swedish and Austrian cats [10,71,78,79] were retrospectively assessed by serologic testing for the organisms listed previously, they were found to be negative, as were their brain sections when assessed by immunohistochemistry for T gondii, CDV, FHV-1, tick-borne encephalitis, and Aujeszky's disease virus [10,79]. After finding that the cats were seropositive for BDV and then performing experimental transmission studies, however, subsequent publications from the same authors determined that these cases had actually been caused by BDV [72,79]. It has therefore been suggested that BDV may be responsible for more of these cases. Although this remains a possibility, it is also possible that there are a number of other previously unrecognized viruses. For this reason, it is important that these cases be fully studied before any causal relation can be proven (for diagnostic methods, see section on prevalence of neurological disease in cats and the preponderance of cases with no known cause) [6,7]. In addition, because BDV can be found in the CNS of some clinically normal individuals [65], its presence, per se, within the CNS of a cat showing neurologic disease does not prove that it is the cause of the disorder.

Group II: polioencephalomyelitis or polioencephalitis

Clinical signs

These diseases also tend to affect younger cats (from a few months old to middle-aged), and there is no sex or breed predisposition. Affected individuals tend to have a subacute to chronic course that may last for months. Partial recovery may be seen in some cats, which can go on to live for many years [44,109,112]. Disease is most commonly sporadic [44,109,119]; however, there are also reports describing what seems to be the chronic form of this condition within large groups of research cats in the United Kingdom [111,112].

Cats with polioencephalomyelitis or polioencephalitis present with problems of locomotion, including ataxia, paresis, and depressed postural reaction in all four limbs. Affected cats may occasionally show hyperesthesia or even lower motor neuron signs (muscle atrophy and decreased tendon reflexes). They can also show tremors, pupillary abnormalities, defective vision, nystagmus, and seizures [44,109,111,112,119]. When seizures occur, they do so as episodes of multiple short seizures [109]. Affected cats rarely show any other signs of systemic disease [109].

Diagnosis

A premortem diagnosis is typically based on the presence of suggestive clinical signs and typical CSF changes (see Table 3) [4,6]. Some affected cats may be leukopenic or anemic (with myeloid hypoplasia) [109].

Early on in the disease, histopathologic examination reveals disseminated inflammatory lesions in the brain and spinal cord, with the spinal cord and

medulla oblongata being most severely affected. The changes are those of polioencephalomyelitis or polioencephalitis. The lesions consist of perivascular mononuclear cuffing, gliosis, and neuronal degeneration, with the latter being most obvious in the ventral horns of the spinal cord [44,109,119].

In chronic cases, little inflammation remains. There is extensive neuronal loss and intense astrogliosis, however, particularly in the spinal cord. Wallerian degeneration arises secondary to the neuronal damage and is particularly evident in the lateral and ventral columns, where it may resemble a primary degenerative disorder [112,119]. In addition, some cats have multifocal areas of Purkinje cell degeneration and gliosis in the cerebellar cortex [109,112]. The changes are similar to those seen in human [120] and porcine poliomyelitis [121].

Treatment

There is no curative treatment. Supportive therapies are as previously discussed and include anticonvulsants to control seizures and, possibly, corticosteroids to reduce CNS inflammation. The prognosis can be good for those cats in which neurologic signs are not too severe.

Potential causes

The cause remains unknown. Genetic and nutritional causes seem unlikely [112], and a viral cause has been suggested by most authors [109,112,119]. Transmission studies have only been attempted occasionally and have been unsuccessful [108]. A number of infectious agents are known to be able to cause poliomyelitis or demyelination in cats. Some of these, including rabies virus, Aujeszky's disease virus, and Newcastle disease virus, have been ruled out on the basis of somewhat differing pathologic findings [109]. It has also been suggested that the condition may be an unusual manifestation of FPV infection [109,112]. Although FPV classically causes cerebellar hypoplasia in kittens, with degeneration of the germinal and Purkinje cells [122,123], it has occasionally been associated with inflammatory lesions within the brain, leukodystrophic lesions, neuronal degeneration, and gliosis of the spinal cord gray matter or spinal cord demyelination [124]. In addition, in FPV-vaccinated cats, FPV infection can still cause leukopenia and nonregenerative anemia [125]. Interestingly, 2 of 33 cases of clinical BD were seen in kittens that came from litters in which the rest of the litter had died of FPV [78]. Other suggested causes include FeLV [126] and arborviruses (see section on rare or experimental infectious causes of CNS disease in cats).

Although the pattern of disease is clinically and histopathologically distinct from that known to be caused by BDV, the author can find no evidence that this possibility has been investigated. Interestingly, the clinical signs and histopathologic findings of the chronic cases are somewhat similar to those of a case report in which the disease was attributed to an unusual case of BDV. It was an interesting case because it was found to have massive neuronal infection with BDV but lacked inflammatory change [95]. This poses an intriguing question as to whether this case was incorrectly attributed to BDV (with the BDV representing a striking but clinically insignificant finding), while also raising the possibility that most of the rest of the cases could actually be attributable to a second variant of BDV infection.

Central nervous system neuropathologic findings without inflammatory changes

Based on the histopathologic changes seen in the chronic cases of polioencephalomyelitis, it is important to realize that viral infections do not necessarily need to be associated with inflammatory changes within the CNS to cause clinically significant neurologic disease.

Neuropathologic changes, without inflammation, may be seen for a number of reasons. It may be that the inflammatory phase has passed and has been missed. Alternately, some viruses can induce direct neuropathic effects while hiding from the immune system within neurons or glial cells (eg, FIV [28]; CDV [127]; human stealth viruses [43]; many herpes viruses, such as chickenpox [128] and varicella zoster virus [129]). In addition, some perinatal infections can result in lasting CNS infection without the development of antibodies (circulating within the blood stream or within the CNS) or encephalitis, but they can still be associated with neurologic disease [95]. This has been seen with BDV infection of some rodents, where infected individuals develop subtle behavioral changes and defects in memory and learning [130,131]. It is therefore possible that BDV infection or other viral infections could result in a number of different disease patterns, depending on differences in viral pathogenicity as well as on as yet unidentified host-specific factors.

Realizing that viral infections can cause CNS disease without obvious inflammatory change and recognizing that the changes can consist mainly of degenerative changes raise the possibility that a viral cause may be responsible for an even higher proportion of CNS disease in cats. Because approximately 35% of feline neurology cases are currently found to result from infectious or inflammatory causes and approximately 15% are degenerative, this could suggest that up to approximately 50% of all feline neurologic disease may potentially be caused by CNS infection (see Table 1) [1].

Unusual patterns of seizure activity

When seizures have been reported as part of the clinical syndromes described previously, the seizure pattern is rather striking; they occur in episodes of multiple short seizures [109]. When looking at studies focusing

on all causes of recurrent seizures in cats, it is fascinating to see that they all result from structural brain disease and that the most common cause (\sim 50%) seems to be a result of nonsuppurative encephalitides [4,5,23]. Although the seizure pattern is similar, the cats in these studies were presented primarily for seizures rather than for ataxia, and despite the onset of seizures being rather dramatic, the prognosis was reasonably good [5].

Whether or not the same virus (or group of viruses) results in all forms of seizure-associated nonsuppurative encephalitides remains to be proven. Interestingly, experimental studies have shown that this pattern of seizure results from lesions in the periaqueductal gray matter of the midbrain [120]. It is therefore possible that it may simply be that this area is preferentially targeted by a number of different infectious agents in cats.

Feline spongiform encephalopathy

Epizootiology

FSE was first recognized in 1990 during the bovine spongiform encephalopathy (BSE) epidemic in the United Kingdom [132-134]. FSE is one of a group of naturally occurring TSEs. TSEs occur in many mammalian species [135], including scrapie in sheep and goats; BSE in cattle and captive exotic ungulates [136,137]; FSE in domestic cats and captive exotic feline species, including the cheetah [137-139], puma [137,140], and lion (A.L. Meredith, MA, VetMB, MRCVS, University of Edinburgh, personal communication, 1999); chronic wasting disease (CWD) of deer and elk [141]; transmissible mink encephalopathy (TME) in mink [142]; and Creutzfeldt-Jakob disease (CJD), variant CJD (vCJD), Gerstmann-Sträussler-Scheinker disease, and kuru in human beings [143]. Experimentally, TSEs can be transmitted to an even wider range of species, including rodents and nonhuman primates [144-146]. Although the widespread interest in TSEs developed only fairly recently, associated with the BSE epidemic and the recognition of vCJD, this type of disease is far from new. Historical records show that scrapie was first recognized approximately 300 years ago [147].

TSEs have been seen throughout the world. Although scrapie and human TSEs have a widespread distribution, BSE has been seen mainly in Europe, particularly in the United Kingdom. The situation is similar with FSE, with almost all cases having been seen in Britain; occasional cases have been seen in animals that had previously lived in the United Kingdom [148] or been fed on tissue from British cattle [135–141].

To understand FSE, it is necessary to know how BSE is believed to have originated. BSE was first reported in 1987 [149]. It is believed to have resulted from the inclusion of scrapie-infected sheep carcasses into feedstuffs for cattle [150]. This resulted in a change to the agent's pathogenicity, making it more infectious to cattle (and cats). Cattle succumbing to BSE were then included in cattle feed, thereby amplifying the transmission and

spreading the infection [150,151]. Once this epidemiologic pattern had been determined, the feeding of meat and bonemeal to ruminants was banned. Since then, the incidence of BSE first plateaued and then fell [152,153].

The agent responsible for FSE is believed to be the same as for BSE [146]. It probably entered the feline population of the United Kingdom in contaminated pet food, and the temporal distribution of cases supports this hypothesis. Since its recognition in 1990, approximately 90 cases of FSE have been confirmed, mostly between 1990 and 1994 (J.W. Wilesmith, BVSc, PhD, J. Spriopoulos, DVM, PhD, Veterinary Laboratories Agency, Weybridge, UK, personal communication on confirmed cases up to the end of 2001, 2004) (Fig. 1). In addition, retrospective study of brain tissue from cats with neurologic disease failed to find cases of FSE before 1990 [1,154]. Although, like BSE, the peak of FSE seems to have passed, occasional cases are still seen (see Fig. 1) [153]. Because few domestic cats are subject to routine postmortem examination, it is likely that the total number of FSE cases has been underestimated.

Most TSEs, like FSE and BSE, seem to be transmitted by ingestion. Although maternal and even genetic transmission may occur in some species [135,143,155], there is no evidence of it occurring in cats or cattle [150,152].

Pathogenesis

The TSE agents are unlike any other microorganisms. All TSE diseases are characterized by the accumulation of an abnormal isoform of a hostcoded protein, the prion protein (PrP). PrP is found in all animals; it is a cell surface glycoprotein of unknown significance. Although the PrP isolated from normal individuals (PrPc) and the PrP isolated from TSE-infected individuals (PrP-res) have the same amino acid sequence and secondary structure, PrPc is totally degraded by proteinase K, whereas PrP-res resists digestion. Once present, PrP-res is believed to induce additional copies of



Fig. 1. Graph showing the incidence of feline spongiform encephalopathy cases in cats by year of onset of clinical signs (cases confirmed to mid 2004; J.W. Wilesmith, BVSc, PhD, J. Spriopoulos, DVM, PhD, Veterinary Laboratories Agency, Weybridge, UK, personal communication).

itself by interacting with normal PrPc. In doing this, PrP-res acts as an infectious agent [156]. Once the host-coded PrPc has been transformed to PrP-res, it accumulates in fibrils (scrapie-associated fibrils [SAFs]), and this eventually leads to disease [135]. Because the process is slow, however, all TSEs have prolonged incubation periods. More detailed information on TSE pathogenesis is reviewed elsewhere [157–159]. Because PrP is host-coded, the accumulation of SAFs induces no immune response [160,161].

Clinical signs

FSE shows no breed predisposition, and cats from all types of households have been affected. There seems to be a slight male predisposition [154]. The mean age at onset is approximately 5 to 7 years (range: 2–12 years) [132–134,148].

FSE is characterized by progressive behavioral and motor disturbances. Affected cats present with progressive hind limb ataxia; increased aggression or affection; hyperesthesia to touch, sound, or light; altered grooming patterns; increased salivation; dilated pupils with an unusual staring expression; polyphagia or polydipsia; abnormal head posture; muscle fasciculations; or an inability to retract their claws [132–134,148,154]. Behavioral changes have usually been noted first, followed by progressive locomotor dysfunction. The cats tend to show ataxia, with dysmetria or hypermetria, which often leads to an erratic crouching gait [134,148]. They also show an inability to judge distances. The disease is generally progressive, warranting euthanasia within 8 to 12 weeks of the onset of clinical signs [134].

Diagnosis

Premortem diagnosis is rarely possible. Although clinical signs may be suggestive of FSE, and nonspecific tests, such as electroencephalography (EEG) or MRI, may indicate the presence of diffuse CNS disease, specific tests are currently lacking. Significant abnormalities have not been detected on serum biochemistry, hematology, or CSF analysis [154]. Diagnosis of FSE is usually made by histopathologic examination of the brain (formalin-fixed tissue) and ultrastructural detection of SAFs in brain extracts (fresh-frozen brain or spinal cord) [162]. After euthanasia, any animal suspected of having FSE should have a full postmortem examination, which should be performed by a trained veterinary pathologist.

Pathologic changes are confined to the CNS and consist of variable degrees of neurophil vacuolation, vacuolation of the neuronal parenchyma, and an astrocytic response. Changes are particularly evident in the gray matter of the thalamus, basal ganglia, and cerebral and cerebellar cortices. More advanced cases may show neuronal loss and more striking gliosis. There are no inflammatory changes. Fibrils analogous to SAFs can be seen on electron microscopy [134,163,164].

Treatment

There is no effective treatment for FSE.

Zoonotic risk

Although it is generally difficult to transmit a TSE agent from one species to another by mouth, BSE seems to have been transmitted naturally, not only to cats, captive exotic felids, and captive exotic ungulates but to human beings, in the form of vCJD [137,165]. Thankfully, with the introduction of strict laws regulating the slaughter and rendering of ruminants and the overall decline in the incidence of BSE, the possibility of the BSE agent continuing to be included in the food chain is extremely small. Because the incubation period is long and variable, however, we are likely to continue to see new cases of vCJD in the United Kingdom for a few years yet to come. That said, the increasing prevalence of CWD in deer and elk in the United States is of concern, and the potential for this TSE to be transmitted to human beings (or cats) is still to be determined [141].

It is unlikely that cats present a zoonotic risk. This is because the disease is now extremely rare (it was never common) and the likelihood of FSEinfected brain or spinal cord entering the human food chain is almost nonexistent. Although there has been one case of CJD and FSE occurring within the same household, the strain of TSE with which both individuals were affected seems to have been a variant more typically associated with spontaneous CJD rather than with BSE [166], and even that diagnosis has been questioned. The method of transmission in this case is not known.

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