

Visible Light-Driven Reductive Azaarylation of Coumarin-3-carboxylic Acids

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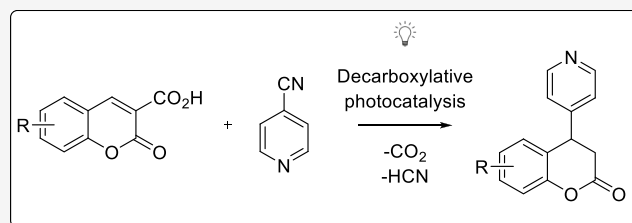


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ABSTRACT: In the manuscript, reductive and decarboxylative azaarylation of coumarin-3-carboxylic acids is described. It utilizes the photocatalytic activation of (cyano)azaarenes in the presence of *fac*-Ir(ppy)₃ as a photocatalyst. The methodology is versatile and provides access to biologically relevant 4-substituted-chroman-2-ones. Visible light, photoredox catalyst, base, anhydrous solvent, and inert atmosphere constitute key parameters for the success of the described strategy. The developed methodology involves a wide range of coumarin-3-carboxylic acids as well as (cyano)-azaarenes.



INTRODUCTION

Chroman-2-one, pyridine, and their derivatives constitute privileged structural motifs present in various natural products.¹ Representative examples of both groups of compounds are shown in Scheme 1. Although these compounds are abundant in nature, synthetic methods for their preparation are of importance.² In this context, it is worth noting that pyridine is the second most frequent nitrogen-containing heterocyclic scaffold that is present in 62 U.S. FDA approved drugs displaying a wide range of biological activities.³ Thus, the pyridine skeleton often serves as a “privileged” scaffold in drug design and discovery. Moreover, it is also a versatile building block utilized for the synthesis of chiral ligands applied in asymmetric catalysis.⁴ As a consequence, a lot of effort has been devoted toward the development of methods for the synthesis of pyridine derivatives.⁵ Recently, radical-based pyridylation reactions have attracted much attention, providing a flexible approach to pyridine derivatives by the application of photocatalysis. These strategies benefit from good functional group tolerance, procedural simplicity, and mild reaction conditions.⁶

The addition of free radicals to electron-deficient olefins is known as Giese reaction (Scheme 2).⁷ Recent advancements in this field arise from the development of photo-mediated methods allowing for the free-radical formation under mild and nontoxic conditions.⁸

A decarboxylative Michael reaction based on nucleophilic addition to carboxylic-acid-activated olefins followed by a decarboxylation reaction constitutes a powerful synthetic tool.⁹ Recently, we described the first photocatalytic, doubly decarboxylative Giese reaction applicable to a wide range of carboxylic acids.¹⁰ Coumarin-3-carboxylic acids **1** constitute useful acceptors in this reaction, opening access to biologically relevant chroman-2-ones **3**.¹¹ Given the interesting properties

of coumarin and pyridine derivatives, the task of development of synthetic routes leading to hybrid molecules bearing both structural motifs was undertaken. Notably, the synthesis of hybrid molecules containing more than one biologically active unit constitutes an important approach in modern drug design.¹²

Herein, we present our studies on the development of decarboxylative reductive arylation of coumarin-3-carboxylic acids. (Cyano)azaarenes were applied as nucleophiles in the Giese-type transformation. This methodology benefits from mild reaction conditions and a broad scope of substrates.

RESULTS AND DISCUSSION

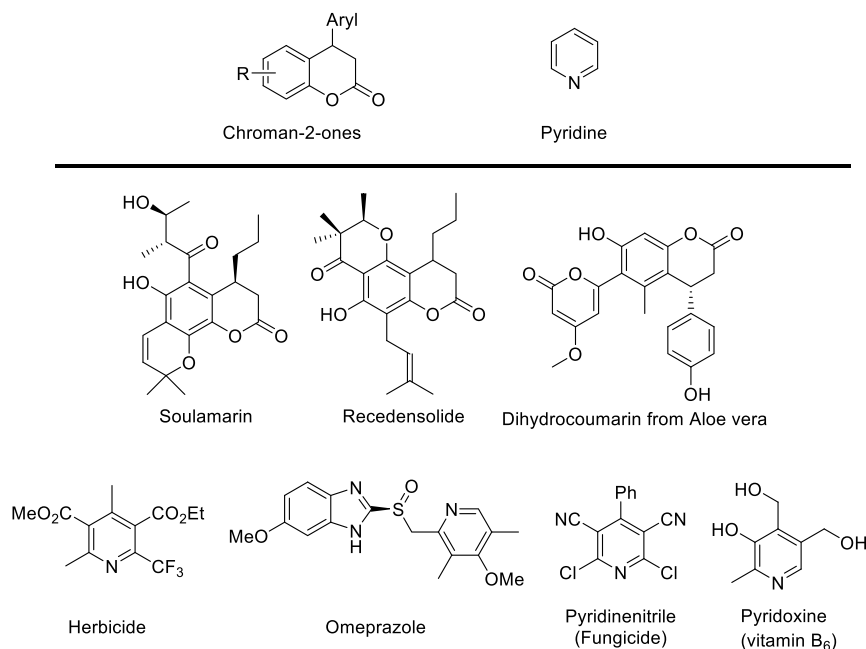
Initially, reactions between cyanopyridine **2a** and coumarin derivatives **1** bearing either no or various activating groups in the 3-position were performed (Table 1, entries 1–4). Experiments were performed in acetonitrile in the presence of **4a** as a photocatalyst and triethylamine as a base under irradiation with blue light and an inert atmosphere at room temperature. When simple coumarin **5a** was used, no reaction was observed. Therefore, EWG-activated coumarin derivatives **1b–e** were tested. Surprisingly, derivatives **5b–d** displayed no reactivity under these conditions. To our delight, the incorporation of the carboxylic acid moiety into the structure of coumarin **1a** resulted in the formation of the desired product **3aa**, indicating the crucial role of the carboxylic-acid-

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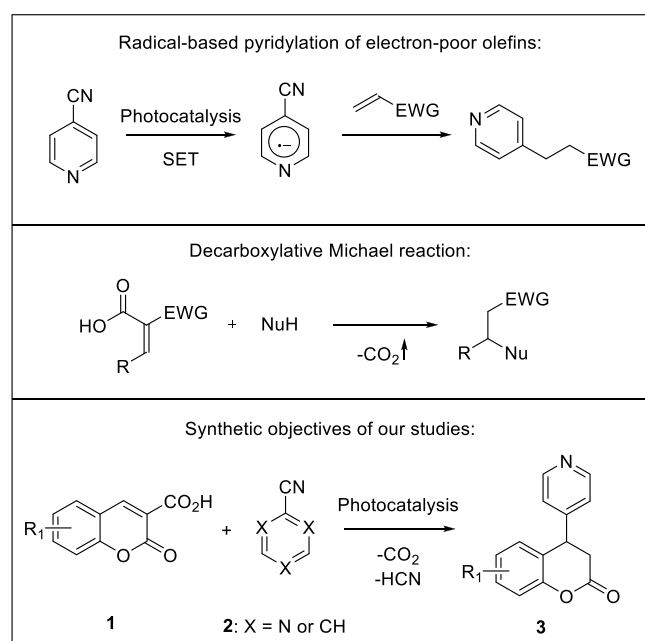
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Scheme 1. Importance of Chroman-2-one and Pyridine Derivatives



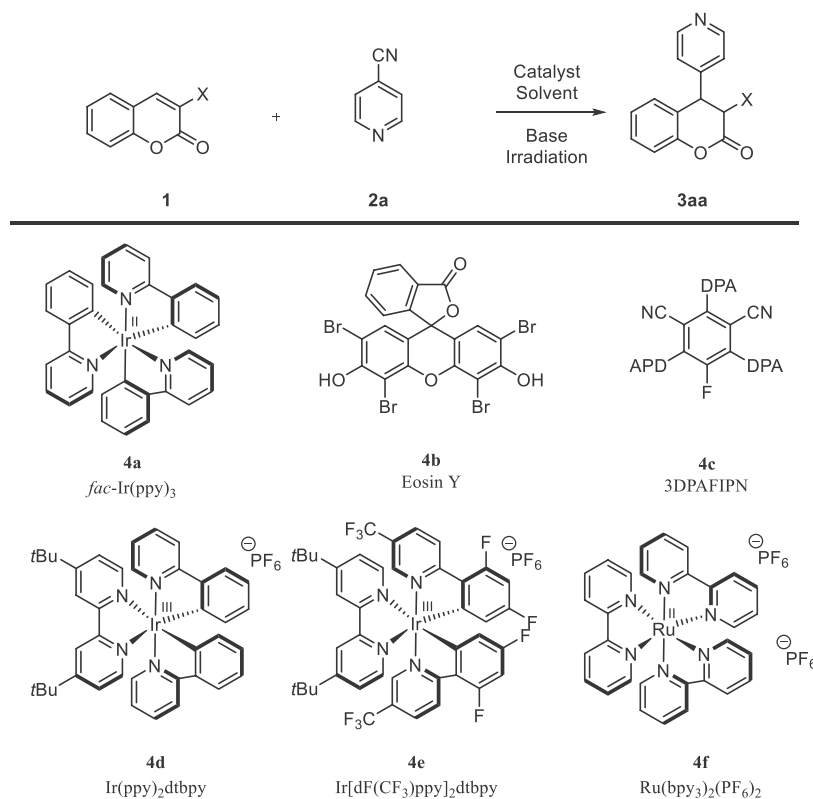
Scheme 2. Importance of Decarboxylative Approaches in Organic Synthesis and the Objectives of Our Study



group activation in the devised reactivity (Table 1, entry 5). Further optimization studies were performed using coumarin-3-carboxylic acid **1a** and 4-cyanopyridine **2a** as model substrates (Table 1, entries 6–22). In the first part of the optimization studies, the catalytic activity of six different photoredox catalysts was tested (with the irradiation with the light source of suitable wavelength) (Table 1, entries 5–10). When Eosin Y **4b** was used, the formation of target product **3aa** was not observed (Table 1, entry 6). Catalysts **4a** and **4c–f** provided the desired reactivity (Table 1, entries 5 and 7–10, respectively) with the best results obtained in the presence of catalysts **4a** (Table 1, entry 5). In the course of further studies, the amount of 4-cyanopyridine **2a** was tested. It was shown

that the reaction with a 3-fold excess of **2a** gave the product **3aa** with 49% yield (Table 1, entry 11). Further increasing the amount of 4-cyanopyridine **2a** did not improve the result. In the next step of optimization studies, the effect of the solvent on the reaction outcome was evaluated (Table 1, entries 11–16). The use of different solvents ensured the product formation; however, the best result was obtained when dimethyl sulfoxide was employed (Table 1, entry 13). During further investigations, the amount of catalyst **4a** was studied (Table 1, entries 16–18). It proved possible to be lowered to 3 mol % without a significant change of the result (Table 1, entry 17). Furthermore, the effect of base on the reaction outcome was evaluated (Table 1, entries 19–21). When DABCO was used, product **3aa** was not formed (Table 1, entry 20) and the application of DIPEA and *N*-methyl morpholine resulted in diminished yields (Table 1, entries 19 and 21). In the course of further studies, control experiments were performed (Table 1, entries 24–26). The use of stoichiometric amount of Et₃N yielded the product **3aa** with low yield (25%) (Table 1, entry 24). The reaction did not proceed in the absence of photoredox catalysts (Table 1, entry 25). A similar effect was observed when the transformation was attempted in the dark (Table 1, entry 26), thus confirming the crucial effect of photocatalyst and the source of light on the reaction outcome. Notably, the optimized reaction proved readily scalable to a 2 mmol scale and the product **3aa** was obtained with a high yield (Table 1, entry 27). Finally, the experiment in the presence of TEMPO was carried out and no reaction was observed, thus confirming the radical nature of the developed reaction (Table 1, entry 28).

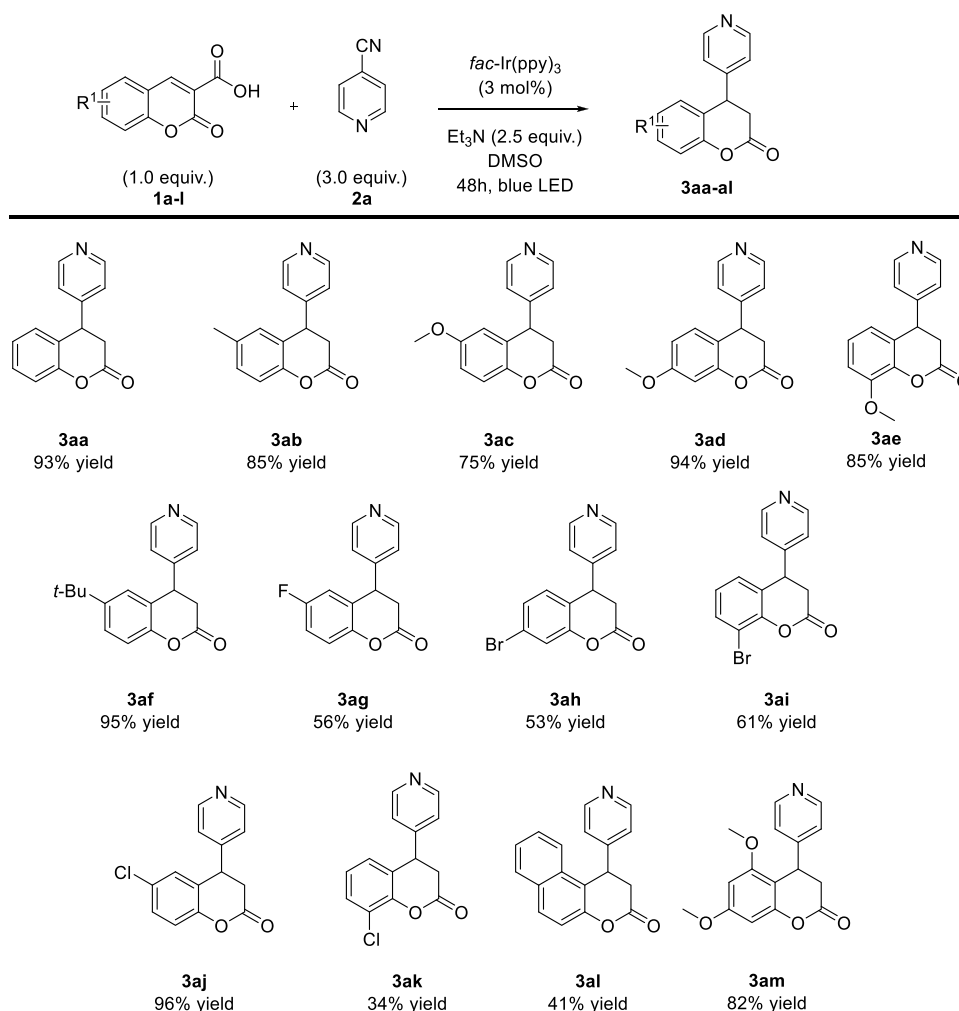
With the optimized reaction conditions in hand (Table 1, entry 23), the scope of the developed methodology was evaluated (Schemes 3 and 4). Initially, various coumarin-3-carboxylic acids **1a–m** were tested in the reaction (Scheme 3). Acids **1b–f** bearing electron-donating groups on the aromatic ring provided products **3ab–af** with very good yields. For the coumarin carboxylic acid **1a** with a *t*-butyl substituent at the 6-position of the aromatic ring, the yield was the highest despite

Table 1. Visible Light-Driven Reductive Azaarylation of Coumarin 1a and Its Derivatives 5a–d: Optimization studies^a

entry	catalyst	X	solvent	base	catalyst [mol %]	yield [%]
1 ^b	4a	H (5a)	CH ₃ CN	Et ₃ N	10	
2 ^b	4a	CN (5b)	CH ₃ CN	Et ₃ N	10	
3 ^b	4a	CO ₂ Et (5c)	CH ₃ CN	Et ₃ N	10	
4 ^b	4a	C(O)Ph (5d)	CH ₃ CN	Et ₃ N	10	
5 ^b	4a	CO ₂ H (1a)	CH ₃ CN	Et ₃ N	10	30
6 ^c	4b	CO ₂ H (1a)	CH ₃ CN	Et ₃ N	10	
7 ^b	4c	CO ₂ H (1a)	CH ₃ CN	Et ₃ N	10	21
8 ^b	4d	CO ₂ H (1a)	CH ₃ CN	Et ₃ N	10	14
9 ^b	4e	CO ₂ H (1a)	CH ₃ CN	Et ₃ N	10	24
10 ^b	4f	CO ₂ H (1a)	CH ₃ CN	Et ₃ N	10	12
11 ^{b,d}	4a	CO ₂ H (1a)	CH ₃ CN	Et ₃ N	10	49
12 ^{b,d}	4a	CO ₂ H (1a)	CH ₂ Cl ₂	Et ₃ N	10	27
13 ^{b,d}	4a	CO ₂ H (1a)	DMSO	Et ₃ N	10	61
14 ^{b,d}	4a	CO ₂ H (1a)	DMF	Et ₃ N	10	15
15 ^{b,d}	4a	CO ₂ H (1a)	CH ₃ OH	Et ₃ N	10	26
16 ^{b,d}	4a	CO ₂ H (1a)	DMSO	Et ₃ N	5	67
17 ^{b,d}	4a	CO ₂ H (1a)	DMSO	Et ₃ N	3	68
18 ^{b,d}	4a	CO ₂ H (1a)	DMSO	Et ₃ N	1	47
19 ^{b,d}	4a	CO ₂ H (1a)	DMSO	DIPEA	3	42
20 ^{b,d}	4a	CO ₂ H (1a)	DMSO	DABCO	3	
21 ^{b,d}	4a	CO ₂ H (1a)	DMSO	NMM	3	49
22 ^{b,d,e}	4a	CO ₂ H (1a)	DMSO	Et ₃ N	3	81
23 ^{b,d,e,f}	4a	CO ₂ H (1a)	DMSO	Et ₃ N	3	93
24 ^{b,d,e,f,g}	4a	CO ₂ H (1a)	DMSO	Et ₃ N	3	25
25 ^{d,e,f}	4a	CO ₂ H (1a)	DMSO	Et ₃ N		
26 ^{b,d,e,f,h}	4a	CO ₂ H (1a)	DMSO	Et ₃ N	3	
27 ^{b,d,e,f,i}	4a	CO ₂ H (1a)	DMSO	Et ₃ N	3	74 (333 mg)
28 ^j	4a	CO ₂ H (1a)	DMSO	Et ₃ N	3	

^aAll reactions were performed in a 0.1 mmol scale using 1a or 5 (1.0 equiv) and 2a (2.0 equiv) in the presence of the corresponding photoredox catalyst 4 (10 mol %) and the corresponding base (2.5 equiv) in the solvent (1 mL) for 24 h at room temperature. ^bReaction performed under irradiation with blue light. ^cReaction performed under irradiation with green light. ^dReaction performed using 2a (3 equiv). ^eReaction performed for 48 h. ^fReaction performed in DMSO (3 mL). ^gReaction performed using Et₃N (1 equiv). ^hReaction performed in the dark. ⁱReaction performed at a 2 mmol scale. ^jReaction performed in the presence of TEMPO (1 equiv).

Scheme 3. Visible Light-Driven Reductive Arylation of Coumarin-3-carboxylic Acids 1: Scope of Coumarin-3-carboxylic Acids 1



the presence of a bulky *t*-butyl substituent. In the course of further studies, it was found that substrates **1** bearing electron-withdrawing groups delivered products **3** in diminished yields. Short reoptimization studies indicated that modification of a previously developed procedure (involving dropwise addition of coumarin carboxylic acids **1g–m** in dry DMSO (1 mL) over 2 h to the reaction mixture, see general procedure for details) enabled the improvement of the results. Dropwise addition of coumarin carboxylic acids **1g–m** suppressed its decomposition over reaction time. Under these conditions, the reaction using coumarins **1g–k** bearing fluorine, bromine, or chlorine atoms at various positions provided the corresponding products **3g–k** in moderate to high yields. It is only in the case of coumarin **1k** with a chlorine substituent in the 8-position of the aromatic ring that the yield of the reaction dropped to 34%. Similar results were observed for doubly substituted coumarin **1l**. In this context, it is worth noting that coumarin **1l** was not effective in the previous decarboxylative reactions performed by our group.^{9d,10} What is also worth emphasizing is that the reaction with doubly substituted coumarin **1m** with two methoxy substituents in the aromatic ring provided the desired product **3am** with very good yield.

Subsequently, the scope of the methodology with regard to different (cyano)azaarenes **2a–c** was evaluated (Scheme 4). It was demonstrated that the developed protocol worked well for

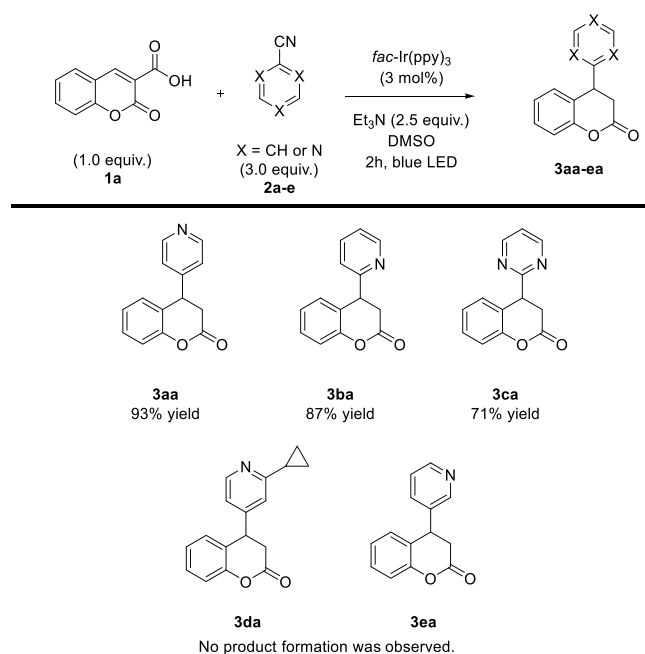
4- and 2-substituted pyridines **2a** and **2b** as well as pyrimidine-2-carbonitrile **2c** to give target products **3aa–3ca** with very good yields. Disappointingly, no product formation was observed when cyanopyridines **2d** and **2e** were employed under optimized reaction conditions.

The postulated mechanism of the developed methodology begins with the blue light-driven excitation of the photocatalyst **4b** (Scheme 5). Then, the electron transfer from the triethylamine to the photocatalyst takes place. Fluorescence quenching and cyclic voltammetry experiments confirmed the lack of quenching in the case of acids **1a** as well as cyanopyridine **2a** (for details, see the Supporting Information). Subsequently, the reduced Ir-catalyst acts as a reductant of the (cyano)azaarene **2a** to give **7**. The newly formed radical **7** undergoes the decarboxylative Giese-type reaction with the acceptor **8** to give **9** that undergoes hydrogen atom transfer to give **10**. Two separate processes transform **10** into **3aa**: (1) rearomatization of the pyridine ring via dehydrocyanation and (2) decarboxylative protonation to afford **3aa** as the final product.

CONCLUSIONS

In conclusion, we have developed a decarboxylative photocatalytic reductive arylation of coumarin-3-carboxylic acids **1** that represents a unique application of free-carboxylic-acid-

Scheme 4. Visible Light-Driven Reductive Arylation of Coumarin-3-carboxylic Acids 1: Reaction Involving Cyanoheteroaromatic Derivatives 2a–2c



activated olefins in radical transformations. The reactions between coumarin-3-carboxylic acids **1a–m** and (cyano)azaarenes **2a–c** were realized under photocatalytic activation in the presence of only 3 mol % of $\text{fac-Ir}(\text{ppy})_3$. The methodology proved versatile, leading to biologically relevant 4-substituted-chroman-2-ones **3aa–ca** in good to high yields under mild reaction conditions.

EXPERIMENTAL SECTION

General Information. NMR spectra were acquired on a Bruker Ultra Shield 700 instrument, running at 700 MHz for ^1H and 176 MHz for ^{13}C . Chemical shifts (δ) are reported in ppm relative to

residual solvent signals (CDCl_3 : 7.26 ppm for ^1H NMR, 77.16 ppm for ^{13}C NMR). Mass spectra were recorded on a Bruker Maxis Impact spectrometer using electrospray (ES+) ionization (referenced to the mass of the charged species). Analytical thin layer chromatography (TLC) was performed using pre-coated aluminum-backed plates (Merck Kieselgel 60 F254) and visualized by ultraviolet irradiation. Unless otherwise noted, analytical-grade solvents and commercially available reagents were used without further purification. For flash chromatography (FC), silica gel (w/ Ca, ~0.1%, 230–400 mesh), green LED (50 W, $\lambda = 525$ nm), and blue LED (50 W, $\lambda = 456$ nm) were purchased from commercial supplier Kessil LED Photoreactor Lightning. Fluorescence measurements were performed using a Varian Cary Eclipse spectrofluorometer equipped with a thermostatted cell holder. Coumarin-3-carboxylic acids **1b–k** were synthesized according to the literature procedure.¹³ Catalyst **4c** was synthesized according to the literature procedure.¹⁴

6-Methyl-2-oxo-2H-chromene-3-carboxylic Acid (1b). Compound **1b** was synthesized according to the literature procedure¹³ as a white solid in 75% yield (153.0 mg). Analytical data were in accordance with the literature.

6-Methoxy-2-oxo-2H-chromene-3-carboxylic Acid (1c). Compound **1c** was synthesized according to the literature procedure¹³ as a white solid in 82% yield (180.4 mg). Analytical data were in accordance with the literature.

7-Methoxy-2-oxo-2H-chromene-3-carboxylic Acid (1d). Compound **1d** was synthesized according to the literature procedure¹³ as a white solid in 64% yield (140.8 mg). Analytical data were in accordance with the literature.

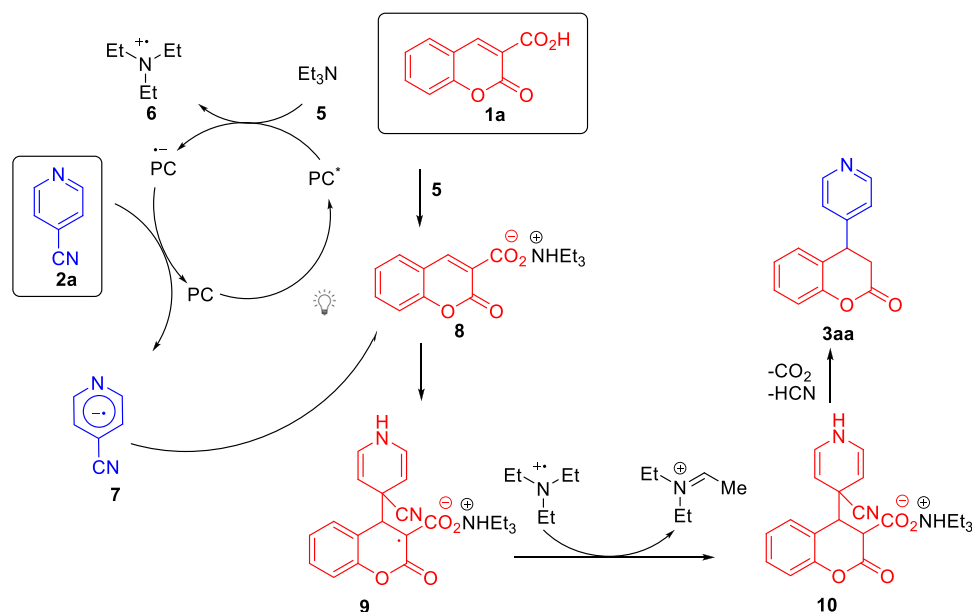
8-Methoxy-2-oxo-2H-chromene-3-carboxylic Acid (1e). Compound **1e** was synthesized according to the literature procedure¹³ as a white solid in 72% yield (158.4 mg). Analytical data were in accordance with the literature.

6-(tert-Butyl)-2-oxo-2H-chromene-3-carboxylic Acid (1f). Compound **1f** was synthesized according to the literature procedure¹³ as a white solid in 89% yield (218.9 mg). Analytical data were in accordance with the literature.

6-Fluoro-2-oxo-2H-chromene-3-carboxylic Acid (1g). Compound **1g** was synthesized according to the literature procedure¹³ as a white solid in 84% yield (174.7 mg). Analytical data were in accordance with the literature.

7-Bromo-2-oxo-2H-chromene-3-carboxylic Acid (1h). Compound **1h** was synthesized according to the literature procedure¹³

Scheme 5. Visible Light-Driven Reductive Arylation of Coumarin-3-carboxylic Acids 1: Reaction Mechanism



as a yellow solid in 62% yield (166.8 mg). Analytical data were in accordance with the literature.

8-Bromo-2-oxo-2H-chromene-3-carboxylic Acid (1i). Compound **1i** was synthesized according to the literature procedure¹³ as a yellow solid in 54% yield (145.3 mg). Analytical data were in accordance with the literature.

6-Chloro-2-oxo-2H-chromene-3-carboxylic Acid (1j). Compound **1j** was synthesized according to the literature procedure¹³ as a yellow solid in 89% yield (199.8 mg). Analytical data were in accordance with the literature.

8-Chloro-2-oxo-2H-chromene-3-carboxylic Acid (1k). Compound **1k** was synthesized according to the literature procedure¹³ as a yellow solid in 72% yield (161.6 mg). Analytical data were in accordance with the literature.

3-Oxo-3H-benzof[f]-chromene-2-carboxylic Acid (1l). Compound **1l** was synthesized according to the literature procedure¹³ as a yellow solid in 67% yield (160.9 mg). Analytical data were in accordance with the literature.

5,7-Dimethoxy-2-oxo-2H-chromene-3-carboxylic Acid (1m). Compound **1m** was synthesized according to the literature procedure¹³ as a white solid in 56% yield (140.1 mg). Analytical data were in accordance with the literature.

General Procedure for the Synthesis of Substituted 4-(Pyridin-4-yl)chroman-2-ones (3aa–3af). In a 10 mL Schlenk tube, coumarin-3-carboxylic acids **1a–f** (0.1 mmol, 1.0 equiv), 4-cyanopyridyne (0.3 mmol, 3.0 equiv), Et₃N (0.25 mmol, 2.5 equiv), and catalyst *fac*-Ir(ppy)₃ (3 mol %) were dissolved in dry DMSO (3 mL). The reaction mixture was degassed and filled three times with argon. Subsequently, the mixture was irradiated with blue LED for 48 h at room temperature. Next, the reaction was quenched with saturated solution of NaHCO₃ (5 mL), extracted with CH₂Cl₂ (3 × 10 mL), and washed with brine (5 mL). The organic phase was dried over MgSO₄ and concentrated under reduced pressure. The crude product was purified by silica gel chromatography (*n*-hexane:ethyl acetate, 2:1) to provide the desired products **3aa–af**.

4-(Pyridin-4-yl)chroman-2-one (3aa).¹⁵ The pure product was isolated by flash chromatography on silica gel (*n*-hexane/ethyl acetate, 2:1) as a yellow oil in 93% yield (20.9 mg). ¹H NMR (700 MHz, chloroform-*d*) δ 8.64–8.54 (m, 2H), 7.37–7.34 (m, 1H), 7.17 (dd, *J* = 8.2, 1.2 Hz, 1H), 7.14 (td, *J* = 7.5, 1.2 Hz, 1H), 7.10 (ddd, *J* = 4.4, 1.6, 0.6 Hz, 2H), 7.01 (dt, *J* = 7.5, 1.1 Hz, 1H), 4.34 (t, *J* = 6.5 Hz, 1H), 3.12 (dd, *J* = 16.0, 6.1 Hz, 1H), 3.04 (dd, *J* = 16.0, 6.8 Hz, 1H). ¹³C {¹H} NMR (176 MHz, CDCl₃) δ 166.7, 151.9, 150.7 (2C), 149.4, 129.6, 128.3, 125.1, 123.9, 122.7 (2C), 117.6, 40.2, 36.3. HRMS (ESI-TOF) *m/z* [M + H⁺] calculated for C₁₄H₁₂NO₂⁺: 226.0863, found: 226.0864.

6-Methyl-4-(pyridin-4-yl)chroman-2-one (3ab). The pure product was isolated by flash chromatography on silica gel (*n*-hexane/ethyl acetate, 2:1) as a brown oil in 85% yield (20.3 mg). ¹H NMR (700 MHz, chloroform-*d*) δ 8.59–8.56 (m, 2H), 7.13 (dd, *J* = 8.4, 2.1 Hz, 1H), 7.09–7.07 (m, 2H), 7.04 (d, *J* = 8.4 Hz, 1H), 6.79 (d, *J* = 2.1 Hz, 1H), 4.28 (t, *J* = 6.4 Hz, 1H), 3.08 (dd, *J* = 16.0, 6.2 Hz, 1H), 3.00 (dd, *J* = 16.0, 6.5 Hz, 1H), 2.61 (s, 1H), 2.28 (s, 3H). ¹³C {¹H} NMR (176 MHz, CDCl₃) δ 167.0, 150.7 (2C), 149.8, 149.5, 134.8, 130.1, 128.6, 123.5, 122.7 (2C), 117.3, 40.3, 36.4, 20.9. HRMS (ESI-TOF) *m/z* [M + H⁺] calculated for C₁₅H₁₄NO₂⁺: 240.1019, found: 240.1016.

6-Methoxy-4-(pyridin-4-yl)chroman-2-one (3ac). The pure product was isolated by flash chromatography on silica gel (*n*-hexane/ethyl acetate, 2:1) as a yellow oil in 75% yield (19.1 mg). ¹H NMR (700 MHz, chloroform-*d*) δ 8.60–8.55 (m, 2H), 7.11–7.08 (m, 3H), 6.87 (ddd, *J* = 8.9, 3.0, 0.5 Hz, 1H), 6.51 (dd, *J* = 3.0, 0.8 Hz, 1H), 4.28 (t, *J* = 6.4 Hz, 1H), 3.73 (s, 3H), 3.08 (dd, *J* = 16.0, 6.1 Hz, 1H), 3.00 (dd, *J* = 16.1, 6.7 Hz, 1H). ¹³C {¹H} NMR (176 MHz, CDCl₃) δ 166.9, 156.7, 150.6 (2C), 149.4, 145.8, 124.7, 122.8 (2C), 118.4, 114.5, 113.5, 55.8, 40.6, 36.3. HRMS (ESI-TOF) *m/z* [M + H⁺] calculated for C₁₅H₁₄NO₃⁺: 256.0968, found: 256.0968.

7-Methoxy-4-(pyridin-4-yl)chroman-2-one (3ad). The pure product was isolated by flash chromatography on silica gel (*n*-hexane/ethyl acetate, 2:1) as a pale yellow oil in 94% yield (24.0 mg). ¹H NMR

(700 MHz, chloroform-*d*) δ 8.57 (d, *J* = 5.0 Hz, 2H), 7.09–7.07 (m, 2H), 6.89 (dd, *J* = 8.4, 0.8 Hz, 1H), 6.70 (d, *J* = 2.5 Hz, 1H), 6.67 (dd, *J* = 8.4, 2.6 Hz, 1H), 4.27 (t, *J* = 6.4 Hz, 1H), 3.81 (s, 3H), 3.09 (dd, *J* = 15.9, 6.2 Hz, 1H), 3.00 (dd, *J* = 15.9, 6.7 Hz, 1H). ¹³C {¹H} NMR (176 MHz, CDCl₃) δ 166.7, 160.6, 152.7, 150.5 (2C), 150.1, 128.9, 122.8 (2C), 115.6, 111.3, 103.0, 55.8, 39.7, 36.6. HRMS (ESI-TOF) *m/z* [M + H⁺] calculated for C₁₅H₁₄NO₃⁺: 256.0968, found: 256.0964.

8-Methoxy-4-(pyridin-4-yl)chroman-2-one (3ae). The pure product was isolated by flash chromatography on silica gel (*n*-hexane/ethyl acetate, 2:1) as a pale yellow oil in 85% yield (21.7 mg). ¹H NMR (700 MHz, chloroform-*d*) δ 8.57 (d, *J* = 5.0 Hz, 2H), 7.12–7.02 (m, 3H), 6.94 (dd, *J* = 8.2, 1.3 Hz, 1H), 6.59 (ddd, *J* = 7.7, 1.4, 0.8 Hz, 1H), 4.32 (t, *J* = 6.3 Hz, 1H), 3.92 (s, 3H), 3.14–3.07 (m, 1H), 3.03 (dd, *J* = 15.9, 6.3 Hz, 1H). ¹³C {¹H} NMR (176 MHz, CDCl₃) δ 166.1, 150.7 (2C), 149.3, 148.2, 141.2, 125.0, 124.9, 122.7 (2C), 119.6, 112.1, 56.3, 40.5, 36.1. HRMS (ESI-TOF) *m/z* [M + H⁺] calculated for C₁₅H₁₄NO₃⁺: 256.0968, found: 256.0971.

6-tert-Butyl-4-(pyridin-4-yl)chroman-2-one (3af). The pure product was isolated by flash chromatography on silica gel (*n*-hexane/ethyl acetate, 2:1) as a yellow oil in 95% yield (26.7 mg). ¹H NMR (700 MHz, chloroform-*d*) δ 8.58 (d, *J* = 5.1 Hz, 2H), 7.36 (dd, *J* = 8.6, 2.4 Hz, 1H), 7.10–7.08 (m, 3H), 7.02 (d, *J* = 2.4 Hz, 1H), 4.31 (t, *J* = 6.1 Hz, 1H), 3.10 (dd, *J* = 15.9, 6.2 Hz, 1H), 3.02 (dd, *J* = 15.9, 6.0 Hz, 1H), 1.25 (s, 9H). ¹³C {¹H} NMR (176 MHz, CDCl₃) δ 167.0, 150.7 (2C), 149.7, 149.7, 148.3, 126.6, 125.2, 122.9, 122.7 (2C), 117.1, 40.6, 36.6, 34.6, 31.5 (3C). HRMS (ESI-TOF) *m/z* [M + H⁺] calculated for C₁₈H₂₀NO₂⁺: 282.1489, found: 282.1491.

General Procedure for the Synthesis of Substituted 4-(Pyridin-4-yl)chroman-2-one (3ag–am). In a 10 mL Schlenk tube, 4-cyanopyridyne **2a** (0.3 mmol, 3.0 equiv), Et₃N (0.25 mmol, 2.5 equiv), and catalyst *fac*-Ir(ppy)₃ (3 mol %) were dissolved in dry DMSO (2 mL). The reaction mixture was degassed and filled three times with argon. The mixture was irradiated with blue LED for 2 h at room temperature. Subsequently, coumarin-3-carboxylic acids **1g–m** (0.1 mmol, 1.0 equiv) in dry DMSO (1 mL) was added dropwise over 2 h and stirred for additional 48 h. Next, the reaction was quenched with saturated solution of NaHCO₃ (5 mL), extracted with CH₂Cl₂ (3 × 10 mL), and washed with brine (5 mL). The organic phase was dried over MgSO₄ and concentrated under reduced pressure. The crude product was purified by silica gel chromatography (*n*-hexane:ethyl acetate, 2:1) to provide the desired products **3ag–am**.

6-Fluoro-4-(pyridin-4-yl)chroman-2-one (3ag). The pure product was isolated by flash chromatography on silica gel (*n*-hexane/ethyl acetate, 2:1) as a yellow oil in 56% yield (13.6 mg). ¹H NMR (700 MHz, chloroform-*d*) δ 8.62–8.60 (m, 2H), 7.13 (dd, *J* = 9.0, 4.6 Hz, 1H), 7.10–7.08 (m, 2H), 7.04 (ddd, *J* = 9.0, 7.7, 2.9 Hz, 1H), 6.70 (ddd, *J* = 8.2, 2.9, 0.8 Hz, 1H), 4.31 (t, *J* = 6.7 Hz, 1H), 3.09 (dd, *J* = 16.0, 6.1 Hz, 1H), 3.02 (dd, *J* = 16.0, 7.3 Hz, 1H). ¹³C {¹H} NMR (176 MHz, chloroform-*d*) δ 166.3, 159.3 (d, *J* = 245.2 Hz), 150.9 (2C), 148.5, 147.9 (d, *J* = 2.8 Hz), 125.6 (d, *J* = 7.6 Hz), 122.7 (2C), 119.0 (d, *J* = 8.3 Hz), 116.4 (d, *J* = 23.5 Hz), 114.9 (d, *J* = 24.4 Hz), 40.3, 35.9. HRMS (ESI-TOF) *m/z* [M + H⁺] calculated for C₁₄H₁₁NO₂F⁺: 244.0768, found: 244.0769.

7-Bromo-4-(pyridin-4-yl)chroman-2-one (3ah). The pure product was isolated by flash chromatography on silica gel (*n*-hexane/ethyl acetate, 2:1) as a pale yellow oil in 53% yield (16.1 mg). ¹H NMR (700 MHz, chloroform-*d*) δ 8.61–8.60 (m, 2H), 7.34 (d, *J* = 2.0 Hz, 1H), 7.28–7.25 (m, 1H), 7.09–7.07 (m, 2H), 6.88 (d, *J* = 8.1 Hz, 1H), 4.29 (t, *J* = 6.6 Hz, 1H), 3.10 (dd, *J* = 16.0, 6.1 Hz, 1H), 3.02 (dd, *J* = 16.0, 7.0 Hz, 1H). ¹³C {¹H} NMR (176 MHz, CDCl₃) δ 165.8, 152.4, 150.9 (2C), 148.7, 129.5, 128.2, 123.0, 122.6 (2C), 122.6, 126.0, 77.2, 40.0, 36.1. HRMS (ESI-TOF) *m/z* [M + H⁺] calculated for C₁₄H₁₁BrNO₂⁺: 303.9968, found: 303.9971.

8-Bromo-4-(pyridin-4-yl)chroman-2-one (3ai). The pure product was isolated by flash chromatography on silica gel (*n*-hexane/ethyl acetate, 2:1) as a pale yellow oil in 61% yield (18.5 mg). ¹H NMR (700 MHz, chloroform-*d*) δ 8.62–8.58 (m, 2H), 7.59 (ddd, *J* = 8.0, 1.6, 0.5 Hz, 1H), 7.08 (ddd, *J* = 4.4, 1.7, 0.6 Hz, 2H), 7.01 (t, *J* = 7.6 Hz, 1H), 6.95 (ddd, *J* = 7.6, 1.6, 0.8 Hz, 1H), 4.35 (t, *J* = 6.5 Hz, 1H),

3.12 (dd, $J = 15.9, 6.1$ Hz, 1H), 3.07 (dd, $J = 15.9, 6.9$ Hz, 1H). ^{13}C { ^1H } NMR (176 MHz, CDCl_3) δ 165.4, 150.9 (2C), 148.9, 148.6, 133.5, 127.4, 125.8, 125.7, 122.6 (2C), 111.5, 40.6, 36.0. HRMS (ESI-TOF) m/z [$\text{M} + \text{H}^+$] calculated for $\text{C}_{14}\text{H}_{11}\text{NO}_2\text{Br}^+$: 303.9968, found: 303.9973.

6-Chloro-4-(pyridin-4-yl)chroman-2-one (3aj). The pure product was isolated by flash chromatography on silica gel (*n*-hexane/ethyl acetate, 2:1) as a pale yellow oil in 96% yield (24.9 mg). ^1H NMR (700 MHz, chloroform-*d*) δ 8.63–8.61 (m, 2H), 7.32 (ddd, $J = 8.7, 2.5, 0.6$ Hz, 1H), 7.11 (d, $J = 8.7$ Hz, 1H), 7.09 (ddd, $J = 4.4, 1.6, 0.6$ Hz, 2H), 6.98 (dd, $J = 2.5, 0.8$ Hz, 1H), 4.31 (t, $J = 6.6$ Hz, 1H), 3.10 (dd, $J = 16.0, 6.1$ Hz, 1H), 3.03 (dd, $J = 16.0, 7.1$ Hz, 1H). ^{13}C { ^1H } NMR (176 MHz, CDCl_3) δ 166.0, 150.9 (2C), 150.4, 148.4, 130.3, 129.7, 128.2, 125.6, 122.6 (2C), 119.0, 40.2, 35.9. HRMS (ESI-TOF) m/z [$\text{M} + \text{H}^+$] calculated for $\text{C}_{14}\text{H}_{11}\text{ClNO}_2^+$: 260.0473, found: 260.0471.

8-Chloro-4-(pyridin-4-yl)chroman-2-one (3ak). The pure product was isolated by flash chromatography on silica gel (*n*-hexane/ethyl acetate, 2:1) as a pale yellow oil in 34% yield (8.8 mg). ^1H NMR (700 MHz, chloroform-*d*) δ 8.60 (d, $J = 5.0$ Hz, 2H), 7.42 (d, $J = 8.0$ Hz, 1H), 7.07 (dd, $J = 21.2, 6.4$ Hz, 3H), 6.91 (d, $J = 7.7$ Hz, 1H), 4.36 (t, $J = 6.6$ Hz, 1H), 3.12 (dd, $J = 15.9, 6.1$ Hz, 1H), 3.07 (dd, $J = 15.9, 6.9$ Hz, 1H). ^{13}C { ^1H } NMR (176 MHz, CDCl_3) δ 165.4, 150.8 (2C), 148.7, 147.8, 130.5, 126.6, 125.8, 125.2, 122.8, 122.7 (2C), 40.6, 36.0. HRMS (ESI-TOF) m/z [$\text{M} + \text{H}^+$] calculated for $\text{C}_{14}\text{H}_{11}\text{ClNO}_2^+$: 260.0473, found: 260.0475.

1,2-Dihydro-3H-benzof[*j*]-1-(pyridin-4-yl)chromen-3-one (3al).¹⁶ The pure product was isolated by flash chromatography on silica gel (*n*-hexane/ethyl acetate, 2:1) as a pale yellow oil in 41% yield (11.3 mg). ^1H NMR (700 MHz, chloroform-*d*) δ 8.53–8.51 (m, 2H), 7.92 (d, $J = 9.1$ Hz, 1H), 7.90–7.89 (m, 1H), 7.71 (dd, $J = 8.5, 0.9$ Hz, 1H), 7.51 (ddd, $J = 8.5, 6.8, 1.4$ Hz, 1H), 7.48 (ddd, $J = 8.0, 6.8, 1.6$ Hz, 1H), 7.37 (d, $J = 9.0$ Hz, 1H), 7.06 (ddd, $J = 4.4, 1.6, 0.6$ Hz, 2H), 4.94 (d, $J = 6.7$ Hz, 1H), 3.27 (dd, $J = 16.0, 7.3$ Hz, 1H), 3.18 (dd, $J = 16.0, 1.8$ Hz, 1H). ^{13}C { ^1H } NMR (176 MHz, CDCl_3) δ 166.3, 150.8 (2C), 150.1, 149.4, 131.3, 130.9, 130.7, 129.1, 128.0, 125.7, 122.7, 122.3 (2C), 117.7, 116.0, 37.1, 36.6. HRMS (ESI-TOF) m/z [$\text{M} + \text{H}^+$] calculated for $\text{C}_{18}\text{H}_{14}\text{NO}_2^+$: 276.1019, found: 276.1023.

5,7-Dimethoxy-4-(pyridin-4-yl)chroman-2-one (3am). The pure product was isolated by flash chromatography on silica gel (*n*-hexane/ethyl acetate, 2:1) as a pale yellow oil in 82% yield (23.4 mg). ^1H NMR (700 MHz, chloroform-*d*) δ 8.63–8.29 (m, 2H), 7.17–6.90 (m, 2H), 6.31 (d, $J = 2.3$ Hz, 1H), 6.28 (d, $J = 2.3$ Hz, 1H), 4.52 (dd, $J = 6.8, 2.4$ Hz, 1H), 3.80 (s, 3H), 3.75 (s, 3H), 3.08–2.96 (m, 2H). ^{13}C { ^1H } NMR (176 MHz, CDCl_3) δ 166.9, 161.3, 157.5, 153.2, 150.6, 150.3 (3C), 122.2, 104.4, 95.3, 94.3, 56.0, 55.7, 36.0, 34.1. HRMS (ESI-TOF) m/z [$\text{M} + \text{H}^+$] calculated for $\text{C}_{16}\text{H}_{16}\text{NO}_4^+$: 286.1074, found: 286.1068.

General Procedure for the Synthesis of 4-Substituted-chroman-2-one 3ba and 3ca. In a 10 mL Schlenk tube, Et_3N (0.25 mmol, 2.5 equiv) and catalyst *fac*-Ir(ppy)₃ (3 mol %) were dissolved in dry DMSO (1 mL). The reaction mixture was degassed and filled three times with argon. The mixture was irradiated with blue LED at room temperature. Coumarin-3-carboxylic acid **1a** (0.1 mmol, 1.0 equiv) in dry DMSO (1 mL) and cyanoarenes **2a–c** (0.3 mmol, 3.0 equiv) in dry DMSO (1 mL) were added dropwise over 2 h and stirred for additional 48 h. Next, the reaction was quenched with saturated solution of NaHCO_3 (5 mL), extracted with CH_2Cl_2 (3 \times 10 mL), and washed with brine (5 mL). The organic phase was dried over MgSO_4 and concentrated under reduced pressure. The crude product was purified by silica gel chromatography (*n*-hexane:ethyl acetate, 5:1) to provide the desired products **3ba** and **3ca**.

4-(Pyridin-2-yl)chroman-2-one (3ba). The pure product was isolated by flash chromatography on silica gel (*n*-hexane/ethyl acetate, 10:1) as a pale yellow oil in 87% yield (19.8 mg). ^1H NMR (700 MHz, chloroform-*d*) δ 8.57 (ddd, $J = 4.8, 1.9, 1.0$ Hz, 1H), 7.62 (td, $J = 7.7, 1.9$ Hz, 1H), 7.29–7.27 (m, 1H), 7.17 (ddd, $J = 7.6, 4.8, 1.1$ Hz, 1H), 7.14–7.10 (m, 2H), 7.10–7.06 (m, 2H), 4.43 (dd, $J = 6.3, 5.1$ Hz, 1H), 3.31 (dd, $J = 16.0, 5.1$ Hz, 1H), 3.04 (dd, $J =$

$16.0, 6.3$ Hz, 1H). ^{13}C { ^1H } NMR (176 MHz, CDCl_3) δ 167.9, 160.1, 151.8, 150.2, 137.1, 129.1, 128.3, 124.6 (2C), 122.6, 122.0, 117.6, 43.0, 34.8. HRMS (ESI-TOF) m/z [$\text{M} + \text{H}^+$] calculated for $\text{C}_{14}\text{H}_{12}\text{NO}_2^+$: 226.0863, found: 226.0864.

4-(Pyrimidin-2-yl)chroman-2-one (3ca). The pure product was isolated by flash chromatography on silica gel (*n*-hexane/ethyl acetate, 10:1) as a yellow oil in 71% yield (16.0 mg). ^1H NMR (700 MHz, chloroform-*d*) δ 8.67 (d, $J = 4.9$ Hz, 2H), 7.34 (ddd, $J = 7.9, 1.6, 0.7$ Hz, 1H), 7.28–7.25 (m, 1H), 7.16 (t, $J = 4.9$ Hz, 1H), 7.10–7.07 (m, 2H), 4.59 (dd, $J = 6.5, 3.2$ Hz, 1H), 3.29 (dd, $J = 16.1, 3.2$ Hz, 1H), 3.07 (dd, $J = 16.1, 6.5$ Hz, 1H). ^{13}C { ^1H } NMR (176 MHz, CDCl_3) δ 169.8, 167.7, 157.7 (2C), 151.6, 129.3, 128.7, 124.6, 123.3, 119.7, 117.6, 44.5, 33.6. HRMS (ESI-TOF) m/z [$\text{M} + \text{H}^+$] calculated for $\text{C}_{13}\text{H}_{11}\text{N}_2\text{O}_2^+$: 227.0815, found: 227.0818.

General Procedure for the Synthesis of 4-(Pyridin-4-yl)chroman-2-one (3aa) in a 2 mmol Scale.¹⁵ In a 50 mL Schlenk tube, coumarin-3-carboxylic acid **1a** (380.3 mg, 2.0 mmol, 1.0 equiv), 4-cyanopyridyne **2a** (624.7 mg, 6.0 mmol, 3.0 equiv), Et_3N (506.0 mg, 5.0 mmol, 2.5 equiv), and catalyst *fac*-Ir(ppy)₃ (39.3 mg, 3 mol %) were dissolved in dry DMSO (20 mL). The reaction mixture was degassed and filled three times with argon. Subsequently, the mixture was irradiated with blue LED for 48 h at room temperature. Next, the reaction was quenched with saturated solution of NaHCO_3 (50 mL), extracted with CH_2Cl_2 (3 \times 75 mL), and washed with brine (50 mL). The organic phase was dried over MgSO_4 and concentrated under reduced pressure. The crude product **3aa** was purified by silica gel chromatography (*n*-hexane:ethyl acetate, 2:1) to provide the desired product **3aa** as a yellow oil in 74% yield (333 mg).

■ ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acs.joc.2c00683>.

Cyclic voltammetry, fluorescence quenching, photochemical reaction setup, and copies of ^1H and ^{13}C NMR spectra (PDF)

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Notes

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REFERENCES

- (1) (a) Kamat, D. P.; Tilve, S. G.; Kamat, V. P.; Kirtany, J. K. Syntheses and Biological Activities of Chroman-2-ones. *Org. Prep. Proc. Int.* **2015**, *47*, 1. (b) Okamoto, T.; Kobayashi, T.; Yoshida, S. Chemical aspects of coumarin compounds for the prevention of hepatocellular carcinomas. *Curr. Med. Chem.* **2005**, *5*, 47. (c) Asai, F.; Iinuma, M.; Tanaka, T.; Mizuno, M. Complex flavonoids in farinose exudate from *Pityrogramma calomelanos*. *Phytochemistry* **1991**, *30*, 3091. (d) Ee, G. C. L. S.; Mah, S. H.; Teh, S. S.; Rahmani, M.; Go, R.; Taufiq-Yap, Y. H. Soullamarin, a New Coumarin from Stem Bark of *Calophyllum soulattri*. *Molecules* **2011**, *16*, 9721.
- (2) (a) Andersen, Ø. M.; Markham, K. R. *Flavonoids: Chemistry, Biochemistry and Applications*; CRC, Taylor & Francis, Boca Raton, FL, 2006; (b) Kamat, D. P.; Tilve, S. G.; Kamat, V. P.; Kirtany, J. K. Syntheses and Biological Activities of Chroman-2-ones. A Review. *Org. Prep. Proc. Int.* **2015**, *47*, 1–79. (c) Masters, K.-S.; Bräse, S. Xanthenes from Fungi, Lichens, and Bacteria: The Natural Products and Their Synthesis. *Chem. Rev.* **2012**, *112*, 3717–3776. (d) Zhao, D.-L.; Shao, C.-L.; Gan, L.-S.; Wang, M.; Wang, C.-Y. Chromone Derivatives from a Sponge-Derived Strain of the Fungus *Corynespora cassicola*. *J. Nat. Prod.* **2015**, *78*, 286–293. (e) Fu, P.; Wang, S.; Hong, K.; Li, X.; Liu, P.; Wang, Y.; Zhu, W. Cytotoxic Bipyridines from the Marine-Derived Actinomycete *Actinoalloteichus cyanogriseus* WHI-2216-6. *J. Nat. Prod.* **2011**, *74*, 1751–1756. (f) Olbe, L.; Carlsson, E.; Lindberg, P. A proton-pump inhibitor expedition: the case histories of omeprazole and esomeprazole. *Nat. Rev. Drug Discovery* **2003**, *2*, 132–139. For selected reviews, see: (g) Nibbs, A. E.; Scheidt, K. A. Asymmetric Methods for the Synthesis of Flavanones, Chromanones, and Azaflavanones. *Eur. J. Org. Chem.* **2012**, *2012*, 449. (h) McDonald, B. R.; Scheidt, K. A. Pyranone Natural Products as Inspirations for Catalytic Reaction Discovery and Development. *Acc. Chem. Res.* **2015**, *48*, 1172.
- (3) (a) Moffett, R. B. Central Nervous System Depressants. VII. 1-pyridyl Coumarins. *J. Med. Chem.* **1964**, *7*, 446–449. (b) Vitaku, E.; Smith, D. T.; Njardarson, J. T. Analysis of the structural diversity, substitution patterns, and frequency of nitrogen heterocycles among U.S. FDA approved pharmaceuticals. *J. Med. Chem.* **2014**, *57*, 10257–10274. (c) Shtyrlin, N. V.; Pavelyev, R. S.; Pugachev, M. V.; Syssoeva, L. P.; Musin, R. Z.; Shtyrlin, Y. G. Synthesis of novel 6-substituted sulfur-containing derivatives of pyridoxine. *Tetrahedron Lett.* **2012**, *53*, 3967–3970. (d) Reynolds, R. D. Bioavailability of vitamin B-6 from plant foods. *Am. J. Clin. Nutr.* **1988**, *48*, 863–867. (e) Gandhi, P. T.; Athmarama, T. N.; Arunkumar, G. R. Novel nicotine analogues with potential anti-mycobacterial activity. *Bioorg. Med. Chem.* **2016**, *24*, 1637–1647.
- (4) For pyridine ligands, see: Chelucci, G. Metal-complexes of optically active amino- and imino-based pyridine ligands in asymmetric catalysis. *Coord. Chem. Rev.* **2013**, *257*, 1887.
- (5) (a) Minisci, F.; Vismara, E.; Fontana, F. Recent developments of free-radical substitutions of heteroaromatic bases. *Heterocycles* **1989**, *28*, 489–519. (b) Schlosser, M.; Mongin, F. Pyridine elaboration through organometallic intermediates: regiochemical control and completeness. *Chem. Soc. Rev.* **2007**, *36*, 1161–1172. (c) Bull, J. A.; Mousseau, J. J.; Pelletier, G.; Charette, A. B. Synthesis of Pyridine and Dihydropyridine Derivatives by Regio- and Stereoselective Addition to N-Activated Pyridines. *Chem. Rev.* **2012**, *112*, 2642–2713.
- (6) For selected examples of the radical-based pyridylation reactions, see: (a) Betori, R. C.; Scheidt, K. A. Reductive Arylation of Arylidene Malonates Using Photoredox Catalysis. *ACS Catal.* **2019**, *9*, 10350–10357. (b) Gao, L.; Wang, G.; Chen, H.; Cao, J.; Su, X.; Liu, X.; Yang, M.; Cheng, X.; Li, S. Metal-free reductive coupling of aliphatic aldehydes/ketones with 4-cyanopyridines: expanded scope and mechanistic studies. *Org. Chem. Front.* **2020**, *7*, 2744–2751. (c) Seiple, I. B.; Su, S.; Rodriguez, R. A.; Gianatassio, R.; Fujiwara, Y.; Sobel, A. L.; Baran, P. S. Direct C–H Arylation of Electron-Deficient Heterocycles with Arylboronic Acids. *J. Am. Chem. Soc.* **2010**, *132*, 13194–13196. (d) Zhang, S.; Li, L.; Li, J.; Shi, J.; Xu, K.; Gao, W.; Zong, L.; Li, G.; Findlater, M. Electrochemical Arylation of Aldehydes, Ketones, and Alcohols: from Cathodic Reduction to Convergent Paired Electrolysis. *Angew. Chem., Int. Ed.* **2021**, *60*, 7275–7282. (e) Zhang, S.; Li, L.; Li, X.; Zhang, J.; Xu, K.; Li, G.; Findlater, M. Electroreductive 4-Pyridylation of Electron-deficient Alkenes with Assistance of Ni(acac)₂. *Org. Lett.* **2020**, *22*, 3570–3575. (f) Shen, J.; Zhang, Y.; Yua, Y.; Wang, M. Metal-free visible-light-induced photoredox-catalyzed intermolecular pyridylation/phosphinylation of alkenes. *Org. Chem. Front.* **2021**, *8*, 901–907.
- (7) (a) Giese, B.; González-Gómez, J. A.; Witzel, T. The Scope of Radical CC-Coupling by the “Tin Method”. *Angew. Chem., Int. Ed.* **1984**, *23*, 69. (b) Rostoll-Berenguer, J.; Blay, G.; Pedro, J. R.; Vila, C. Photocatalytic Giese Addition of 1,4-Dihydroquinoxalin-2-ones to Electron-Poor Alkenes Using Visible Light. *Org. Lett.* **2020**, *22*, 8012.
- (8) (a) Parida, S. K.; Mandal, T.; Das, S.; Hota, S. K.; De Sarkar, S.; Murarka, S. Single Electron Transfer-Induced Redox Processes Involving N-(Acyloxy)phthalimides. *ACS Catal.* **2021**, *11*, 1640. (b) Dong, Y.; Ji, P.; Zhang, Y.; Wang, C.; Meng, X.; Wang, W. Organophotoredox-Catalyzed Formation of Alkyl-Aryl And -Alkyl C-S/Se Bonds from Coupling of Redox-Active Esters with Thio/Selenosulfonates. *Org. Lett.* **2020**, *22*, 9562. (c) Saget, T.; König, B. Photocatalytic Synthesis of Polycyclic Indolones. *Chem. – Eur. J.* **2020**, *26*, 7004. (d) Zheng, Ch.; Wang, G.-Z.; Shang, R. Catalyst-free Decarboxylation and Decarboxylative Giese Additions of Alkyl Carboxylates through Photoactivation of Electron Donor-Acceptor Complex. *Adv. Synth. Catal.* **2019**, *361*, 4500. Buzzetti, L.; Crisenza, G. E. M.; Melchiorre, P. Mechanistic Studies in Photocatalysis. *Angew. Chem., Int. Ed.* **2019**, *58*, 3730–3747. (e) Vega-Peñalosa, A.; Javier Mateos, J.; Companyó, X.; Escudero-Casao, M.; Dell’Amico, L. A Rational Approach to Organo-Photocatalysis: Novel Designs and Structure-Property Relationships. *Angew. Chem., Int. Ed.* **2021**, *133*, 1096–1111. (f) Bortolato, T.; Cuadros, S.; Simionato, G.; Dell’Amico, L. The advent and development of organophotoredox catalysis. *Chem. Commun.* **2022**, *58*, 1263–1283.
- (9) For reviews on decarboxylative strategies, see: (a) Pan, Y.; Tan, C.-H. Catalytic Decarboxylative Reactions: Biomimetic Approaches Inspired by Polyketide Biosynthesis. *Synthesis* **2011**, 2044. (b) Wang, Z.-L. Recent Advances in Catalytic Asymmetric Decarboxylative Addition Reactions. *Adv. Synth. Catal.* **2013**, *355*, 2745. (c) Nakamura, S. Catalytic enantioselective decarboxylative reactions using organo-catalysts. *Org. Biomol. Chem.* **2014**, *12*, 394. (d) Bojanowski, J.; Albrecht, A. Carboxylic-Acid-Activated Olefins in Decarboxylative Reactions. *Asian J. Org. Chem.* **2019**, *8*, 746.
- (10) Moczulski, M.; Kowalska, E.; Kuśmierk, E.; Albrecht, L.; Albrecht, A. Visible-light synthesis of 4-substituted-chroman-2-ones and 2-substituted-chroman-4-ones via doubly decarboxylative Giese reaction. *RSC Adv.* **2021**, *11*, 27782–27786.
- (11) (a) Xu, L.; Shao, Z.; Wang, L.; Xiao, J. Tandem sp³ C-H Functionalization/Decarboxylation of 2-Alkylazaarenes with Coumarin-3-carboxylic Acids. *Org. Lett.* **2014**, *16*, 796. (b) Han, F.; Xun, S.; Jia, L.; Zhang, Y.; Zou, L.; Hu, X. Traceless-Activation Strategy for Rh-Catalyzed Csp²–H Arylation of Coumarins. *Org. Lett.* **2019**, *21*, 5907.
- (12) (a) Fayed, E. A.; Sabour, R.; Harras, M. F.; Mehany, A. B. M. Design, synthesis, biological evaluation and molecular modeling of new coumarin derivatives as potent anticancer agents. *Med. Chem. Res.* **2019**, *28*, 1284–1297. (b) Feng, Z.; Yu, Y.; Yang, X.; Zhong, D.; Song, D.; Yang, H.; Chen, X.; Zhou, G.; Wu, Z. Isomers of Coumarin-Based Cyclometalated Ir(III) Complexes with Easily Tuned Phosphorescent Color and Features for Highly Efficient Organic Light-Emitting Diodes. *Inorg. Chem.* **2019**, *58*, 7393–7408.
- (13) Song, A.; Wang, X.; Lam, K. S. A convenient synthesis of coumarin-3-carboxylic acids via Knoevenagel condensation of Mel-

drum's acid with ortho-hydroxyaryl aldehydes or ketones. *Tetrahedron Lett.* **2003**, *44*, 1755.

(14) Luo, J.; Zhang, J. Donor–Acceptor Fluorophores for Visible-Light-Promoted Organic Synthesis: Photoredox/Ni Dual Catalytic C(sp³)–C(sp²) Cross-Coupling. *ACS Catal.* **2016**, *6*, 873–877.

(15) Hoshikawa, T.; Inoue, M. Photoinduced direct 4-pyridination of C(sp³)–H Bonds. *Chem. Sci.* **2013**, *4*, 3118–3123.

(16) Das, P.; Ray, S.; Bhanja, P.; Bhaumik, A.; Mukhopadhyay, C. Serendipitous Observation of Liquid-Phase Size Selectivity inside a Mesoporous Silica Nanoreactor in the Reaction of Chromene with Formic Acid. *ChemCatChem* **2018**, *10*, 2260–2270.