

SHORT COMMUNICATION



Neutrophil extracellular trap release driven by bacterial motility: Relevance to cystic fibrosis lung disease

Balázs Rada

Department of Infectious Diseases, College of Veterinary Medicine, The University of Georgia, Athens, GA, USA

ABSTRACT

Neutrophil extracellular trap (NET) formation represents a unique effector function of neutrophils (PMN). The mechanism of NET release in response to bacteria is largely unknown. We studied the process by which *Pseudomonas aeruginosa*, an opportunistic pathogen, interacts with primary PMNs, and found that flagellar swimming motility of the bacterium is essential for inducing NET extrusion. Cystic fibrosis (CF) lung disease is associated with *P. aeruginosa* infection and PMN-dominated inflammation. Although NETs are abundant in CF airways, the main factors triggering NET release in CF remain unclear. Our study implicates that motile *P. aeruginosa* is a strong NET-inducer in CF. In early stages of CF lung disease flagellated, motile isolates of *P. aeruginosa* are characteristic and their interactions with PMNs could lead to NET formation. In chronic CF, *P. aeruginosa* down-regulates its flagellum expression to avoid recognition by the immune system and forms biofilms. Flagellated bacteria, however, are released from biofilms and could interact with PMNs to form NETs. Although flagellated forms likely represent only a small fraction of the total *P. aeruginosa* load in chronic CF, NET release induced by them could have a significant impact on inflammation and lung function since flagellated forms trigger the most robust response of the immune system including PMNs.

Overall, we speculate that NET formation driven by motile *P. aeruginosa* could be a novel, significant contributor to pathogenesis at both, early and late stages of CF lung disease.

ARTICLE HISTORY

Received 16 December 2016
Revised 10 February 2017
Accepted 13 February 2017

KEYWORDS

cystic fibrosis; flagellum;
motility; NETs; neutrophil;
Pseudomonas aeruginosa



CF airways contain large numbers of PMNs. CF sputum PMN counts, levels of extracellular DNA, myeloperoxidase and human neutrophil elastase all correlate with CF lung disease severity.¹⁻⁶ PMNs are the clinically most important leukocyte in chronic CF airways and PMN-mediated inflammation contributes to lung disease. Extracellular DNA is derived from the host,⁷ mainly PMNs.^{5,7,8} Although PMNs were thought to die by necrosis in CF airways, recently other mechanisms have been proposed. NET formation provides an attractive alternative explanation since PMNs simultaneously release lung-damaging extracellular DNA and primary granule components by forming NETs, and NETs are abundant in the airways of adult CF patients. We and others reported robust NET release from human PMNs induced by laboratory strains and CF clinical isolates of *P. aeruginosa*.⁹⁻¹⁴ Since NETs could mediate the release of lung-damaging PMN cargo but could also be important in fighting pathogens, their exact, potentially complex, role in CF airway disease remains to be elucidated (Fig. 1).

Flagellar motility drives *P. aeruginosa*-induced NET release

In our current study, we found that early exponential growth phase cultures of *P. aeruginosa* elicited the most robust NET release and presence of a functional flagellum was essential for this process.¹⁴ Immotile bacterial mutants without flagellum or with nonfunctional flagellum are weak NET-inducers.¹⁴ Forced contact of immotile *P. aeruginosa* with PMNs restored their ability to trigger maximal NET extrusion.¹⁴ *P. aeruginosa* flagellin alone was unable to induce NET release.¹⁴ In a genetic complementation study we found that both, *motAB* and *motCD* loci of *P. aeruginosa* flagellar motor genes are needed for maximal NET induction in human PMNs.¹⁴ Thus, we identified flagellar swimming motility as a novel microbial factor crucial to PMN activation and NET formation.

Flagellated *P. aeruginosa* in CF

Although it is undocumented whether NETs are present at early stages of CF lung disease, bacterial motility-

CONTACT Balázs Rada,  radab@uga.edu  College of Veterinary Medicine, Department of Infectious Diseases, University of Georgia, 501 D.W. Brooks Drive, Athens, GA 30602, USA.

© 2017 Balázs Rada. Published with license by Taylor & Francis.

This is an Open Access article distributed under the terms of the Creative Commons Attribution-NonCommercial-NoDerivatives License (<http://creativecommons.org/licenses/by-nc-nd/4.0/>), which permits non-commercial re-use, distribution, and reproduction in any medium, provided the original work is properly cited, and is not altered, transformed, or built upon in any way.

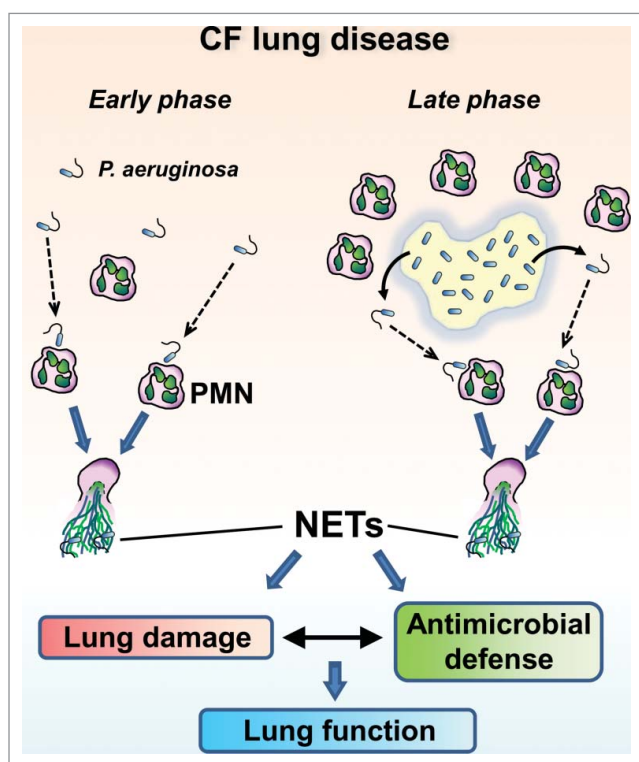


Figure 1. Potential contributions of motile *P. aeruginosa*-driven NET formation to CF lung disease.

fueled NET formation likely occurs at this initial phase because early CF clinical isolates of *P. aeruginosa* typically express flagellum^{15,16} and PMNs are also present¹⁷ (Fig. 1). The interaction of PMNs with early forms of *P. aeruginosa* must be critical to determine later progression of CF lung disease. Important questions to be answered are why NETs released at this early stage would be incapable of clearing *P. aeruginosa* infection and instead drive bacterial adaption toward an aflagellated, biofilm-forming phenotype.

Over the course of CF lung disease, *P. aeruginosa* down-regulates its flagellum expression.^{15,16,18-20} In chronic CF airways, *P. aeruginosa* mainly exist in 3-dimensional, “suspension biofilms” also called non-attached aggregates (Fig. 1).²¹⁻²³ These suspension biofilms surrounded by PMNs represent the characteristic clinical picture in chronic CF airways.²¹⁻²⁴ Biofilms are dynamic structures, and motile, flagellated bacteria likely break free from biofilms in chronic CF, possibly interacting with PMNs (Fig. 1). This is supported by recent data showing that *P. aeruginosa* flagellin is detected in sputa of chronic CF patients.²⁵ PMNs phagocytosing planktonic *P. aeruginosa* have also been observed in chronic CF.^{26,27} Nonmucoid revertant cells of *P. aeruginosa* have also been documented in chronic CF airways.^{15,18} A minor population of flagellated *P. aeruginosa* in

chronic CF airways could also have been marginalized so far because this topic has not been intensely investigated yet, and this population is hard to study and could have been overlooked in the presence of much more abundant biofilm-bound bacteria accumulating well-characterized mutations (*mucaA*, *lasR*).^{28,29} Conclusions with respect to the population structure of *P. aeruginosa* in CF have largely been generalized based on results obtained on single bacterial isolates, although high levels of phenotypic diversity among *P. aeruginosa* isolates within individual CF patients have already been noted.³⁰ A small population of flagellated *P. aeruginosa* could be found in CF airways while most *P. aeruginosa* are present in form of alginate-producing, elastase-negative bacteria. *MucaA* mutations drive the mucoid, biofilm-forming phenotype, *lasR* mutations contribute to PMN recruitment²⁸ while outbreaks of flagellated bacteria from biofilms could be mainly responsible for PMN activation and NET release.¹⁴ PMNs quickly and easily recognize motile, flagellated forms of *P. aeruginosa* and launch their robust effector mechanisms including NET release in response to them.¹⁴ On the other hand, bacterial biofilms likely provide a much weaker stimulus for PMN activation. Therefore, the way motile *P. aeruginosa* interacts with PMNs, the cell type representing most of cells found in chronic CF airways, is likely an important factor in influencing the progression of CF lung disease despite the fact that planktonic forms of *P. aeruginosa* are outnumbered by those found in biofilms (Fig. 1).

Overall, we speculate that *P. aeruginosa* motility-driven PMN activation has clinical relevance not only at initial but also later stages of CF airway disease.¹⁴

Disclosure of potential conflicts of interest

No potential conflicts of interest were disclosed.

References

- [1] Carlsson M, Eriksson L, Erwander I, Wieslander J, Segelmark M. Pseudomonas-induced lung damage in cystic fibrosis correlates to bactericidal-permeability increasing protein (BPI)-autoantibodies. *Clin Exp Rheumatol* 2003; 21:S95-100; PMID:14740434
- [2] Garner HP, Phillips JR, Herron JG, Severson SJ, Milla CE, Regelman WE. Peroxidase activity within circulating neutrophils correlates with pulmonary phenotype in cystic fibrosis. *J Lab Clin Med* 2004; 144:127-33; PMID:15454881; <http://dx.doi.org/10.1016/j.lab.2004.04.010>
- [3] Kim JS, Okamoto K, Rubin BK. Pulmonary function is negatively correlated with sputum inflammatory markers and cough clearability in subjects with cystic fibrosis but not those with chronic bronchitis. *Chest* 2006; 129:1148-54; PMID:16685004; <http://dx.doi.org/10.1378/chest.129.5.1148>

- [4] Gibson RL, Emerson J, Mayer-Hamblett N, Burns JL, McNamara S, Accurso FJ, Konstan MW, Chatfield BA, Retsch-Bogart G, Waltz DA, et al. Duration of treatment effect after tobramycin solution for inhalation in young children with cystic fibrosis. *Pediatr Pulmonol* 2007;42:610-23; PMID:17534969; <http://dx.doi.org/10.1002/ppul.20625>
- [5] Regelman WE, Siefferman CM, Herron JM, Elliott GR, Clawson CC, Gray BH. Sputum peroxidase activity correlates with the severity of lung disease in cystic fibrosis. *Pediatr Pulmonol* 1995; 19:1-9; PMID:7675551; <http://dx.doi.org/10.1002/ppul.1950190102>
- [6] Mayer-Hamblett N, Kronmal RA, Gibson RL, Rosenfeld M, Retsch-Bogart G, Treggiari MM, Burns JL, Khan U, Ramsey BW, Investigators E. Initial *Pseudomonas aeruginosa* treatment failure is associated with exacerbations in cystic fibrosis. *Pediatr Pulmonol* 2012; 47:125-34; PMID:21830317; <http://dx.doi.org/10.1002/ppul.21525>
- [7] Lethem MI, James SL, Marriott C, Burke JF. The origin of DNA associated with mucus glycoproteins in cystic fibrosis sputum. *Eur Resp J* 1990; 3:19-23; PMID:2107097
- [8] Sagel SD, Wagner BD, Anthony MM, Emmett P, Zemanick ET. Sputum biomarkers of inflammation and lung function decline in children with cystic fibrosis. *Am J Respir Crit Care Med* 2012; 186:857-65; PMID:22904182; <http://dx.doi.org/10.1164/rccm.201203-0507OC>
- [9] Rada B, Jendrysik MA, Pang L, Hayes CP, Yoo DG, Park JJ, Moskowitz SM, Malech HL, Leto TL. Pyocyanin-enhanced neutrophil extracellular trap formation requires the NADPH oxidase. *PloS one* 2013; 8: e54205; PMID:23342104; <http://dx.doi.org/10.1371/journal.pone.0054205>
- [10] Yoo DG, Floyd M, Winn M, Moskowitz SM, Rada B. NET formation induced by *Pseudomonas aeruginosa* cystic fibrosis isolates measured as release of myeloperoxidase-DNA and neutrophil elastase-DNA complexes. *Immunol Lett* 2014; 160:186-94; PMID:24670966; <http://dx.doi.org/10.1016/j.imlet.2014.03.003>
- [11] Yoo DG, Winn M, Pang L, Moskowitz SM, Malech HL, Leto TL, Rada B. Release of cystic fibrosis airway inflammatory markers from *Pseudomonas aeruginosa*-stimulated human neutrophils involves NADPH oxidase-dependent extracellular DNA trap formation. *Journal of immunology* 2014; 192:4728-38; <http://dx.doi.org/10.4049/jimmunol.1301589>
- [12] Parker H, Dragnow M, Hampton MB, Kettle AJ, Winterbourn CC. Requirements for NADPH oxidase and myeloperoxidase in neutrophil extracellular trap formation differ depending on the stimulus. *J Leukoc Biol* 2012; 92:841-9; PMID:22802447; <http://dx.doi.org/10.1189/jlb.1211601>
- [13] Young RL, Malcolm KC, Kret JE, Caceres SM, Poch KR, Nichols DP, Taylor-Cousar JL, Saavedra MT, Randell SH, Vasil ML, et al. Neutrophil extracellular trap (NET)-mediated killing of *Pseudomonas aeruginosa*: evidence of acquired resistance within the CF airway, independent of CFTR. *PloS one* 2011; 6:e23637; PMID:21909403; <http://dx.doi.org/10.1371/journal.pone.0023637>
- [14] Floyd M, Winn M, Cullen C, Sil P, Chassaing B, Yoo DG, Gewirtz AT, Goldberg JB, McCarter LL, Rada B. Swimming Motility Mediates the Formation of Neutrophil Extracellular Traps Induced by Flagellated *Pseudomonas aeruginosa*. *PLoS Pathog* 2016; 12: e1005987; PMID:27855208; <http://dx.doi.org/10.1371/journal.ppat.1005987>
- [15] Mahenthalingam E, Campbell ME, Speert DP. Nonmotility and phagocytic resistance of *Pseudomonas aeruginosa* isolates from chronically colonized patients with cystic fibrosis. *Infect Immun* 1994; 62:596-605; PMID:8300217
- [16] Wolfgang MC, Jyot J, Goodman AL, Ramphal R, Lory S. *Pseudomonas aeruginosa* regulates flagellin expression as part of a global response to airway fluid from cystic fibrosis patients. *Proc Natl Acad Sci U S A* 2004; 101:6664-8; PMID:15084751; <http://dx.doi.org/10.1073/pnas.0307553101>
- [17] Sly PD, Gangell CL, Chen L, Ware RS, Ranganathan S, Mott LS, Murray CP, Stick SM, Investigators AC. Risk factors for bronchiectasis in children with cystic fibrosis. *N Engl J Med* 2013; 368:1963-70; PMID:23692169; <http://dx.doi.org/10.1056/NEJMoal1301725>
- [18] Luzar MA, Thomassen MJ, Montie TC. Flagella and motility alterations in *Pseudomonas aeruginosa* strains from patients with cystic fibrosis: relationship to patient clinical condition. *Infect Immun* 1985; 50:577-82; PMID:3932214
- [19] Mahenthalingam E, Speert DP. Nonopsonic phagocytosis of *Pseudomonas aeruginosa* by macrophages and polymorphonuclear leukocytes requires the presence of the bacterial flagellum. *Infect Immun* 1995; 63:4519-23; PMID:7591095
- [20] Boyce JR, Miller RV. Motility as a selective force in the reversion of cystic fibrosis-associated mucoid *Pseudomonas aeruginosa* to the nonmucoid phenotype in culture. *Infect Immun* 1982; 37:840-4; PMID:6811443
- [21] Pritt B, O'Brien L, Winn W. Mucoid *Pseudomonas* in cystic fibrosis. *Am J Clin Pathol* 2007; 128:32-34; PMID:17580270; <http://dx.doi.org/10.1309/KJRPC7DD5TR9NTDM>
- [22] Caceres SM, Malcolm KC, Taylor-Cousar JL, Nichols DP, Saavedra MT, Bratton DL, Moskowitz SM, Burns JL, Nick JA. Enhanced in vitro formation and antibiotic resistance of nonattached *Pseudomonas aeruginosa* aggregates through incorporation of neutrophil products. *Antimicrob Agents Chemother* 2014; 58:6851-60; PMID:25182651; <http://dx.doi.org/10.1128/AAC.03514-14>
- [23] Bjarnsholt T, Alhede M, Alhede M, Eickhardt-Sorensen SR, Moser C, Kuhl M, Jensen PO, Hoiby N. The in vivo biofilm. *Trends Microbiol* 2013; 21:466-74; PMID:23827084; <http://dx.doi.org/10.1016/j.tim.2013.06.002>
- [24] Kragh KN, Alhede M, Jensen PO, Moser C, Scheike T, Jacobsen CS, Seier Poulsen S, Eickhardt-Sorensen SR, Trostrup H, Christoffersen L, et al. Polymorphonuclear leukocytes restrict growth of *Pseudomonas aeruginosa* in the lungs of cystic fibrosis patients. *Infect Immun* 2014; 82:4477-86; PMID:25114118; <http://dx.doi.org/10.1128/IAI.01969-14>
- [25] Balloy V, Thevenot G, Bienvenu T, Morand P, Corvol H, Clement A, Ramphal R, Hubert D, Chignard M. Flagellin concentrations in expectorations from cystic fibrosis patients. *BMC Pulm Med* 2014; 14:100; PMID:24909229; <http://dx.doi.org/10.1186/1471-2466-14-100>
- [26] Williams BJ, Dehnhostel J, Blackwell TS. *Pseudomonas aeruginosa*: host defence in lung diseases. *Respirology* 2010; 15:1037-56; PMID:20723140; <http://dx.doi.org/10.1111/j.1440-1843.2010.01819.x>

- [27] Hoiby N, Ciofu O, Bjarnsholt T. *Pseudomonas aeruginosa* biofilms in cystic fibrosis. *Future Microbiol* 2010; 5:1663-74; PMID:21133688; <http://dx.doi.org/10.2217/fmb.10.125>
- [28] LaFayette SL, Houle D, Beaudoin T, Wojewodka G, Radzioch D, Hoffman LR, Burns JL, Dandekar AA, Smalley NE, Chandler JR, et al. Cystic fibrosis-adapted *Pseudomonas aeruginosa* quorum sensing *lasR* mutants cause hyperinflammatory responses. *Sci Adv* 2015; 1(6); pii: e1500199; PMID:26457326; <http://dx.doi.org/10.1126/sciadv.1500199>
- [29] Hogardt M, Heesemann J. 2013. Microevolution of *Pseudomonas aeruginosa* to a chronic pathogen of the cystic fibrosis lung. *Curr Top Microbiol Immunol* 2015; 358:91-118; PMID:22311171
- [30] Folkesson A, Jelsbak L, Yang L, Johansen HK, Ciofu O, Hoiby N, Molin S. Adaptation of *Pseudomonas aeruginosa* to the cystic fibrosis airway: an evolutionary perspective. *Nat Rev Microbiol* 2012; 10:841-51; PMID:23147702; <http://dx.doi.org/10.1038/nrmicro.2907>