

Review Article

Control of Cross Talk between Angiogenesis and Inflammation by Mesenchymal Stem Cells for the Treatment of Ocular Surface Diseases

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Angiogenesis is beneficial in the treatment of ischemic heart disease and peripheral artery disease. However, it facilitates inflammatory cell filtration and inflammation cascade that disrupt the immune and angiogenesis privilege of the avascular cornea, resulting in ocular surface diseases and even vision loss. Although great progress has been achieved, healing of severe ocular surface injury and immunosuppression of corneal transplantation are the most difficult and challenging step in the treatment of ocular surface disorders. Mesenchymal stem cells (MSCs), derived from various adult tissues, are able to differentiate into different cell types such as endothelial cells and fat cells. Although it is still under debate whether MSCs could give rise to functional corneal cells, recent results from different study groups showed that MSCs could improve corneal disease recovery through suppression of inflammation and modulation of immune cells. Thus, MSCs could become a promising tool for ocular surface disorders. In this review, we discussed how angiogenesis and inflammation are orchestrated in the pathogenesis of ocular surface disease. We overviewed and updated the knowledge of MSCs and then summarized the therapeutic potential of MSCs via control of angiogenesis, inflammation, and immune response in the treatment of ocular surface disease.

1. Introduction

Cornea is the transparent front part of the eye. It is composed of epithelium, Bowman's layer, stroma, Descemet's membrane, and endothelium. Limbal stem cells (LSCs) are residing at the basal layer of the limbus and could differentiate into terminal epithelium cells for replacement. In the stage of corneal damage, LSCs could generate epithelial cells for repair [1]. As damage progresses, angiogenesis and lymphangiogenesis in the avascular cornea result in the infiltration of neutrophils and macrophages as well as Th1 cells for further attack. As the pathological process involves regions of corneal limbus, LSCs are lost and dysfunctional and fail to replace the damaged epithelial cells, leading to blindness [2]. In this case, LSCs and corneal transplantation are the most feasible option to improve ocular surface damage and vision. Although the success rate of transplantation is high, graft rejection still occurs resulting from preoperative high-risk factors, postoperative inflammation, angiogenesis,

lymphangiogenesis, and immune response [3–5]. To date, it has been reported that more than 10 million patients have been suffering from corneal blindness in the world [6].

Mesenchymal stem cells (MSCs) are originated from multiple adult tissues such as bone marrow, liver, and adipose tissue. As pluripotent cells, MSCs could differentiate into different cell types [7]. Besides their differentiation potential, MSCs exert immunomodulatory and anti-inflammation effects on the surrounding cells by the release of secreted cytokines [8]. When cocultured with LSCs, MSCs could stimulate LSCs proliferation and growth factor expression *in vitro* [9]. Therefore, MSCs therapy could be a promising approach for ocular surfaces diseases via control of lymphangiogenesis, inflammation, and immune response. In the review, we will first overview the knowledge of MSCs and then focus on how MSCs control the pathological cross talk between lymphangiogenesis and inflammation in the treatment of corneal diseases.

2. Characteristic and Potential of MSCs

2.1. Definition of MSCs. MSCs have been isolated from several adult tissues, including bone marrow, adipose tissue, liver, dental pulp, endometrium, muscle, amniotic fluid, placenta, and umbilical cord blood [10–12]. MSCs have pluripotent or multipotent properties as well as a great potential of differentiating into mesodermal cell lineages (e.g., adipocytes, osteocytes, and chondrocytes) and nonmesodermal cell lineages (e.g., cardiomyocytes, hepatocytes-like cells, neurons, astrocytes, and endothelial cells) both in vivo and in vitro. In addition, it is found that pericytes present in several organs, such as skeletal muscle and pancreas, also express the very same markers used by MSCs [13]. They could share many of the differentiation characteristics of MSCs in vitro [14]. Thus, the perivascular niche can be regarded as a subset of MSCs [13–16]. Due to the lack of specific markers for these cells, the authentic MSCs are difficult to identify. To resolve this problem, the International Society for Cellular Therapy has provided the minimum criteria for defining multipotent MSCs: plastic adherent under standard culture conditions; positive for the expression of CD105, CD73, and CD90 surface markers; absent for the expression of CD11b, CD14, CD19, CD34, CD45, CD79a, and HLA-DR surface markers; and capable of differentiating into osteocytes, adipocytes, and chondrocytes under a specific stimulus in vitro [17].

2.2. Differentiation Ability. MSCs have both endothelial and epithelial tissue coding genes and could be promoted to differentiate into endothelial- or epithelial-like cells both in vitro and in vivo [18–20]. Under specific conditions, MSCs could differentiate into corneal epithelial cells, keratocyte-like cells, and endothelial-like cells to repair damaged corneas [21–25]. However, some lines of evidence found that the replaced cells derived from MSCs do not behave as true tissue cells [26]. This might be due to the inconsistent differentiation protocols and heterogeneity of cell population [27, 28].

2.3. Immunomodulatory and Anti-Inflammation Potential. MSCs are potent regulators of immune response and inflammation. MSCs could be activated by the inflammatory microenvironment through exposure to proinflammatory cytokines, such as interferon- γ (IFN- γ), tumor necrosis factor- α (TNF- α), and effector T cells [29, 30]. In vitro, MSCs interact with innate and acquired immune response and inhibit the proliferation and function of T cells, B cells, dendritic cells, and natural killer cells [31–33]. The regulation of immune cells by MSCs mainly comes from a panel of cytokines secreted by MSCs, including IL-6, IL-10, transforming growth factor- β (TGF- β), metal matrix proteinase (MMP), prostaglandin E2 (PGE2), indoleamine-2,3-dioxygenase (IDO), human leukocyte antigen-G5 (HLA-G), and nitric oxide (NO) [34, 35]. Second, MSCs could decrease the expression levels of Th1 cell factors (IFN- γ and IL-2) and increase the expression levels of Th2 cell factors (IL-4 and IL-10), thereby promoting immune response of naïve CD4+ cells toward the Th2-type response [32]. Third, when cocultured with lymphocytes, MSCs produce PGE2 and

TGF- β to promote regulatory T-cells (Tregs) differentiation and expansion [36, 37]. It is well known that Tregs have the capacity to suppress the proliferation of activated T cells. Therefore, modulation of Tregs has been suggested as the main mechanism of MSCs in maintaining immune tolerance for allografts survival in organ transplantation [38].

The application of MSCs in organ transplantation has been tested in rat and primate transplantation models. MSCs seemed to significantly suppress immune rejection and prolong graft survival in the heart, liver, kidney, pancreas, and other solid organs [39–42]. Following systemic infusion, MSCs could not only migrate to lesion but also be trapped in lungs and other organs [43]. Although it is not fully clear, MSCs homing is regulated via chemokine, chemokine receptors, intracellular signals, adhesion molecules, and proteases, such as stromal cell-derived factor-1 α (SDF-1 α) and C-X-C chemokine receptor type 4 (CXCR4) [44–47]. It is currently under investigation whether improved MSCs homing could bring about better therapeutic effect.

However, MSCs therapy is not always successful and even accelerates allograft rejection after organ transplantation [57]. The controversial results might be influenced by many factors including infusion time and dose, administration mode, and homing efficiency of MSCs.

2.4. Angiogenic Property of MSCs. A growing body of evidence has shown the regulation effect of angiogenesis by MSCs. This effect was mainly attributed to the modulation of angiogenic factors produced by MSCs. For instance, vascular epithelial growth factor (VEGF) and basic fibroblast growth factor (bFGF) both promote the migration and proliferation of vascular endothelial cells [58]. In cardiac ischemia repair, MSCs stimulate neovascularization of infarct tissue through upregulating VEGF to improve cardiac function [59]. The effects may be associated with the role of TLR2 [60]. However, the direct interaction between VEGF and TLR2 in MSCs is not clear. In addition, some reports demonstrated that MSCs could be additional important cells for proangiogenesis to form provisional granulation matrix in the proliferation phase of wound healing [61].

Different from the effect of proneovascularization in ischemic tissues and tumors, MSCs showed an opposite effect on inflammation-related corneal angiogenesis after chemical injury. This action was associated with the significantly smaller mean neovascularized area and a reduced expression level of VEGF [62], which might be attributed to the expression of high level of thrombospondin-1 (TSP-1), which inhibits VEGF [63]. These results suggested that the different microenvironment would modulate different behavior and function of MSCs. Below, we overviewed MSCs applications in ocular surface diseases.

3. MSCs in Ocular Surface Diseases (Table 1)

3.1. Corneal Wound Repair. Corneal injury is caused by thermal injury, alkali or acid burns, and immune or hereditary disorders and leads to corneal inflammation, neovascularization, conjunctivalization, impaired vision, and even

TABLE 1: Experiments of MSCs in ocular surface diseases.

Diseases	Experimental outcomes	The mechanisms	Factors	References
Chemical injury	Reduce corneal opacity	Reduce inflammation and neovascularization	↑ TSG-6	[48]
Chemical injury	Protect the corneal surface	Reduce inflammation and neovascularization Reduce CD4+ cells infiltration	↑ IL-10, IL-6, TSP-1, and TGF- β 1 ↓ IL-2, IFN- γ , and MMP-2	[63]
Alkali burn	Improve wound healing	Enhance the recovery of corneal epithelium Decrease the CNV area	↓ MIP-1 α , TNF- α , and VEGF	[62]
Chemical burn	Restructure damaged corneal surface	Inhibit inflammation and angiogenesis	↓ IL-2 and MMP-2	[71]
Chemical burn	Affect profiling of IL-17-secreting cells	Mainly modulate non-Th17 cells and partially suppress Th17 cells	↓ IL-17	[73]
Corneal allotransplantation	Prolong grafts survival	Inhibit immune response Suppress early inflammation Reduce the activation of APCs	↑ TSG-6	[52]
Corneal allotransplantation	Prolong grafts survival	Prevent T-cells response Regulate the balance of Th1/Th2 to Th2 Increase CD4+CD25+Foxp3+ Treg	↑ IL-10 and IL-4 ↓ IL-2 and IFN- γ	[53]
Corneal allotransplantation	Prolong grafts survival	Reduce NK cells infiltration Increase CD4+ Foxp3+ Treg Suppress peripheral immune response Promote an immunoregulatory milieu	↓ IL-6, IL-1 β , and IFN- γ	[54]
DES	Protect ocular surface	Reduce the CD4+ T cells		[55]
Dry eye secondary to chronic GVHD	Reduce clinical symptoms and improve dry eye scores	Increase the CD8+CD28- T cells Regulate the balance of Th1/Th2 to Th2		[56]

TSG-6: TNF- α -stimulated gene/protein 6; MMP: metal matrix proteinase; CNV: cornea new vessel; TSP-1: thrombospondin-1; TNF- α : tumor necrosis factor- α ; MIP-1 α : macrophage inflammatory protein-1 α ; IFN- γ : interferon- γ ; VEGF: vascular epithelial growth factor; DES: dry eye syndrome; GVHD: graft-versus-host disease; APCs: antigen-presenting cells.

blindness [64]. LSC is an essential cell population for corneal epithelium regeneration and ocular surface reconstruction. Unfortunately, LSC loss and dysfunction in corneal limbus trigger severe inflammation and neovascularization [65]. To be noted, LSC transplantation remains as the effective strategy to treat LSCD but it is challenged by limited donors and allograft rejection [66, 67].

Alternative therapy is to improve resident LSC and corneal epithelial cell (CEP) expansion for repair [10, 68, 69]. In vitro studies showed that MSCs could stimulate LSC and CEP proliferation when they were cocultured. Both systemic and topical administration of MSCs have been shown to accelerate corneal regeneration and healing [48, 49]. In practice, to increase local concentration, MSCs were injected into the injured cornea with a hollow plastic tube [70], through subconjunctival administration of MSCs [62] or through transplantation of MSCs with tissue engineering materials, such as amniotic membrane (AM) [50, 71] and nanofiber scaffolds [72]. The strategy of combining with tissue engineering materials is better for cornea recovery than MSCs or tissue engineering materials used alone [51].

Besides stimulation of LSC proliferation, MSCs injection effectively alleviates inflammation and neovascularization in the injured cornea. MSCs increased the expression of the anti-inflammatory cytokines IL-10, TGF-1, TNF- α -stimulated gene/protein 6 (TSG-6), and the antiangiogenic

factor thrombospondin-1 (TSP-1) and reduced the expression levels of the proinflammatory factors IL-2, IFN- γ , IL-17, macrophage inflammatory protein-1 α (MIP-1 α), and MMP-2 [70, 71, 73, 74]. Ultimately, neutrophil and macrophage infiltration is largely reduced.

3.2. Corneal Transplantation. Corneal transplantation is the most common form of human tissue transplantation. Comparing with other types of organ transplantation, normal-risk corneal transplants have an exceptionally high success rate of up to 80% over 5 years, which is mainly based on the specific immune privilege of cornea including low-level expression of MHC I and MHC II, the lack of indigenous antigen-presenting cells (macrophages or Langerhans cells), the absence of lymphoid and blood vessels, and anterior chamber associated immune deviation (ACAID) [75–77]. However, as risk factors, preexisting lymphangiogenesis and blood vessel together with inflammation promote graft rejection [78].

Currently, a broad range of treatment strategies have been proposed to increase the duration of grafts survival. The leading method for preventing transplant rejection is corticosteroids and immunosuppression such as cyclosporine A, FK-506 [79–81]. However, long-term immunosuppression could produce drug toxicity and potential complications; the used dosages are limited. More recent works have focused

on endothelia transplantation (only endothelia are diseased) and lamellar transplantation (endothelia are not diseased) to reduce immune rejection [82–84].

MSCs administration has been widely tested in corneal transplantation and inconsistent results have been reported. Some studies demonstrated that pretransplant infusion of MSCs was effective to prolong graft survival; meanwhile, MSCs used postoperatively are less effective, especially for kidney and heart transplantation. This might be explained by the fact that preoperative infusion of MSCs modulated Tregs expansion and induced immune tolerance before occurrence of inflammation and immune progress [85–87]. Oh et al. [52] suggested that pretransplant systemic infusion of human MSCs inhibited immune response largely due to suppressing early inflammation caused by surgery and reducing the activation of antigen-presenting cells (APCs) in both cornea and draining lymph nodes (DLNs). The role of MSCs was primarily exerted by secreting the soluble anti-inflammatory protein TSG-6. However, Jia et al. [53] have shown that MSCs prolonged corneal allograft survival time only when injected immediately after the surgery and preoperative administration exerted no significant effect. The cornea immune privilege triggers delayed-type hypersensitivity which may take longer for the corneal allograft to activate an immune response than other types of solid organ transplantation.

By contrast, Oh et al. [88] were the first who examined the immunomodulatory effects of MSCs in corneal transplantation in a pig-to-rat model. Allogeneic rat MSCs were applied topically to corneal grafts for 2 h immediately after transplantation. They observed the increased expression levels of Th2-type cytokine (IL-10) in the rejected grafts from MSC-treated rats and a shift from Th1 to Th2 cell type following MSCs administration. However, MSCs injection failed to prolong pig corneal xenograft survival in rats. Similarly, Fuentes-Julián et al. [89] obtained an insufficient conclusion about adipose-MSCs treatment to prevent corneal grafts rejection. In the study, local or systemic adipose-MSCs administration in rabbit corneal transplantation models at normal- or high-risk rejection does not prolong the graft survival. The used adipose-MSCs lacked immunomodulatory ability on T lymphocytes and immunophenotypical secretion molecules, which may be the reason why the adipose-MSCs destroyed the innate ocular immune privilege and accelerated rejection [89].

All of the above observations demonstrated that MSCs would be a potential therapeutic tool for corneal allograft transplantation and the molecular mechanisms of action need to be further studied. Importantly, many factors including the source of MSCs, the infusion time, the dose of injection, and the mode of administration would influence the best results of MSCs treatment on corneal transplantation. In addition, MSCs in combination with immunomodulatory drugs are an alternative treatment. Intravenous transfusion of MSCs and cyclosporine A (CsA) achieved a synergistic effect on suppressing immune rejection of corneal grafts [53].

3.3. Dry Eye Syndrome. Dry eye syndrome (DES) or keratoconjunctivitis sicca (KCS) is the major ocular surface

disease affecting ranges from 7% to 33% of the worldwide population [90, 91]. Dry eye is a common ocular complication associated with chronic graft-versus-host disease (GVHD) after allogeneic hematopoietic stem cell transplantation, occurring in 60% of patients [92, 93]. DES is characterized by deficiency of the tear film components (lipid, aqueous, and mucin). Although the mechanisms of DES are yet unknown, inflammation in the lacrimal gland and the ocular surface plays a key role in the pathogenesis of the disorder. DES is also an autoimmune disease with immune-mediated destruction in the whole process [94, 95]. Immunohistochemistry of lacrimal gland shows immune cell infiltration and loss of acinar epithelial cells, and the expression of proinflammatory cytokines is increased [96]. Studies have suggested that the proinflammatory factors inhibit neurotransmitter release resulting in insufficient secretion of lacrimal gland [97]. In addition, the lacrimal gland could be involved as a target in several systemic and autoimmune diseases including Sjögren syndrome, sarcoidosis, and chronic GVHD [96]. The current treatment strategies, including tear replacement, anti-inflammatory drugs, and punctual occlusion, often fail to resolve the underlying problems of DES.

Topical application of MSCs could be a safe and available treatment for periocular diseases with immune involvement, such as KCS/DES [98, 99]. Allogeneic adipose-MSCs were implanted around lacrimal glands in dogs with KCS [100]. The implanted cells effectively reduced clinical signs during a 9-month follow-up. Lee et al. [55] also demonstrated that periorbital administration of MSCs could protect the ocular surface in a murine model of DES. In the intraorbital gland and ocular surface, the CD4+ T-cells infiltration was reduced. MSCs suppressed inflammation and increased aqueous tear production [55]. In a recent clinical practice, 22 patients with refractory dry eye secondary to chronic GVHD were treated with MSCs, 12 of whom showed reduced symptoms with improved dry eye scores. The results were accompanied by increasing the number of CD8+CD28–T cells, which suggest that MSCs regulate the balance between Th1 and Th2 [56]. There is a human Phase I/II clinical trial involving allogeneic MSCs treatment for primary Sjögren syndrome (<http://clinicaltrials.gov/ct2/show/NCT00953485>). Based on the double effect between inflammation and immunomodulation, MSCs are a promising source to treat dry eye syndrome.

4. Summary and Perspectives

The ocular immune and angiogenic privileges act as a barrier to protect corneal function. Ingrowth of new blood vessels orchestrates inflammatory cell infiltration leading to inflammation and impaired epithelial cell repair. MSCs have shown therapeutic effect in corneal surface diseases by several lines of mechanisms: inhibition of inflammatory cell infiltration and inflammatory cytokine release, modulation of the switch from Th1 cell type toward Th2 cell type, activation of Treg cells, and stimulation of epithelial cell regeneration (Figure 1). Therefore, MSCs would be a very promising tool in the treatment of corneal diseases. Further studies would focus on increase of MSCs homing efficiency, time, and safety

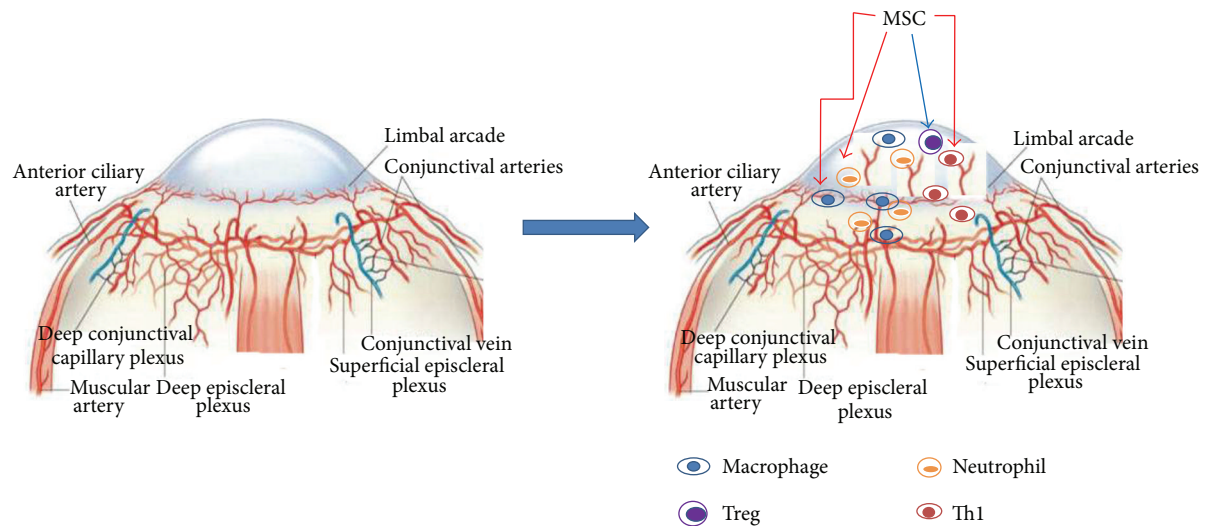


FIGURE 1: Modulation of immune cells and inflammatory cells by MSCs in corneal surface diseases. Cornea is the avascular and transparent front part of the eye, maintained by immune and angiogenesis privilege. In the occurrence of injury and transplantation, ingrowth of blood and lymph vessels into the cornea leads to infiltration of inflammatory cells and Th1 cells, which strengthen the inflammation and damage the cornea structure. MSCs have several protective functions by (1) inhibition of the inflammatory cell infiltration and inflammatory cytokine release, (2) activation of Treg cells for immune tolerance, (3) tuning the transition from Th1 cells toward Th2 cells, and (4) improving epithelium regeneration (not shown).

and MSCs administration, together with evaluation of MSC-based therapies when dealing with ocular surface diseases.

Competing Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

References

- [1] H. S. Dua and A. Azuara-Blanco, "Limbal stem cells of the corneal epithelium," *Survey of Ophthalmology*, vol. 44, no. 5, pp. 415–425, 2000.
- [2] G. Pellegrini, P. Rama, F. Mavilio, and M. De Luca, "Epithelial stem cells in corneal regeneration and epidermal gene therapy," *Journal of Pathology*, vol. 217, no. 2, pp. 217–228, 2009.
- [3] J. B. Jonas, R. M. Rank, and W. M. Budde, "Immunologic graft reactions after allogeneic penetrating keratoplasty," *American Journal of Ophthalmology*, vol. 133, no. 4, pp. 437–443, 2002.
- [4] K. Inoue, S. Amano, T. Oshika, and T. Tsuru, "Risk factors for corneal graft failure and rejection in penetrating keratoplasty," *Acta Ophthalmologica Scandinavica*, vol. 79, no. 3, pp. 251–255, 2001.
- [5] R. W. Thompson Jr., M. O. Price, P. J. Bowers, and F. W. Price Jr., "Long-term graft survival after penetrating keratoplasty," *Ophthalmology*, vol. 110, no. 7, pp. 1396–1402, 2003.
- [6] J. P. Whitcher, M. Srinivasan, and M. P. Upadhyay, "Corneal blindness: a global perspective," *Bulletin of the World Health Organization*, vol. 79, no. 3, pp. 214–221, 2001.
- [7] P. Bianco, P. G. Robey, and P. J. Simmons, "Mesenchymal stem cells: revisiting history, concepts, and assays," *Cell Stem Cell*, vol. 2, no. 4, pp. 313–319, 2008.
- [8] W. R. Otto and N. A. Wright, "Mesenchymal stem cells: from experiment to clinic," *Fibrogenesis and Tissue Repair*, vol. 4, no. 1, article 20, 2011.
- [9] A. Uccelli, L. Moretta, and V. Pistoia, "Mesenchymal stem cells in health and disease," *Nature Reviews Immunology*, vol. 8, no. 9, pp. 726–736, 2008.
- [10] N. Hu, Y.-Y. Zhang, H.-W. Gu, and H.-J. Guan, "Effects of bone marrow mesenchymal stem cells on cell proliferation and growth factor expression of limbal epithelial cells in vitro," *Ophthalmic Research*, vol. 48, no. 2, pp. 82–88, 2012.
- [11] H. K. Salem and C. Thiemeermann, "Mesenchymal stromal cells: current understanding and clinical status," *STEM CELLS*, vol. 28, no. 3, pp. 585–596, 2010.
- [12] M. F. Pittenger, A. M. Mackay, S. C. Beck et al., "Multilineage potential of adult human mesenchymal stem cells," *Science*, vol. 284, no. 5411, pp. 143–147, 1999.
- [13] L. Da Silva Meirelles, P. C. Chagastelles, and N. B. Nardi, "Mesenchymal stem cells reside in virtually all post-natal organs and tissues," *Journal of Cell Science*, vol. 119, no. 11, pp. 2204–2213, 2006.
- [14] M. Crisan, S. Yap, L. Casteilla et al., "A Perivascular origin for mesenchymal stem cells in multiple human organs," *Cell Stem Cell*, vol. 3, no. 3, pp. 301–313, 2008.
- [15] M. Crisan, S. Yap, L. Casteilla et al., "A perivascular origin for mesenchymal stem cells in multiple human organs," *Cell Stem Cell*, vol. 3, no. 3, pp. 301–313, 2008.
- [16] M. Crisan, C.-W. Chen, M. Corselli, G. Andriolo, L. Lazzari, and B. Péault, "Perivascular multipotent progenitor cells in human organs," *Annals of the New York Academy of Sciences*, vol. 1176, pp. 118–123, 2009.
- [17] M. Dominici, K. Le Blanc, I. Mueller et al., "Minimal criteria for defining multipotent mesenchymal stromal cells. The international society for cellular therapy position statement," *Cytotherapy*, vol. 8, no. 4, pp. 315–317, 2006.

- [18] F. Alviano, V. Fossati, C. Marchionni et al., "Term amniotic membrane is a high throughput source for multipotent mesenchymal stem cells with the ability to differentiate into endothelial cells in vitro," *BMC Developmental Biology*, vol. 7, article 11, 2007.
- [19] P. Cipriani, S. Guiducci, I. Miniati et al., "Impairment of endothelial cell differentiation from bone marrow-derived mesenchymal stem cells: new insight into the pathogenesis of systemic sclerosis," *Arthritis and Rheumatism*, vol. 56, no. 6, pp. 1994–2004, 2007.
- [20] V. Păunescu, E. Deak, D. Herman et al., "In vitro differentiation of human mesenchymal stem cells to epithelial lineage," *Journal of Cellular and Molecular Medicine*, vol. 11, no. 3, pp. 502–508, 2007.
- [21] S. Gu, C. Xing, J. Han, M. O. M. Tso, and J. Hong, "Differentiation of rabbit bone marrow mesenchymal stem cells into corneal epithelial cells in vivo and ex vivo," *Molecular Vision*, vol. 15, pp. 99–107, 2009.
- [22] E. M. Martínez-Conesa, E. Espel, M. Reina, and R. P. Casaroli-Marano, "Characterization of ocular surface epithelial and progenitor cell markers in human adipose stromal cells derived from lipoaspirates," *Investigative Ophthalmology & Visual Science*, vol. 53, no. 1, pp. 513–520, 2012.
- [23] H. Liu, J. Zhang, C.-Y. Liu, Y. Hayashi, and W. W.-Y. Kao, "Bone marrow mesenchymal stem cells can differentiate and assume corneal keratocyte phenotype," *Journal of Cellular and Molecular Medicine*, vol. 16, no. 5, pp. 1114–1124, 2012.
- [24] H. Liu, J. Zhang, C.-Y. Liu et al., "Cell therapy of congenital corneal diseases with umbilical mesenchymal stem cells: lumican null mice," *PLoS ONE*, vol. 5, no. 5, Article ID e10707, 2010.
- [25] M. Nakahara, N. Okumura, E. P. Kay et al., "Corneal endothelial expansion promoted by human bone marrow mesenchymal stem cell-derived conditioned medium," *PLoS ONE*, vol. 8, no. 7, Article ID e69009, 2013.
- [26] D. G. Phinney and D. J. Prockop, "Concise review: mesenchymal stem/multipotent stromal cells: the state of transdifferentiation and modes of tissue repair—current views," *STEM CELLS*, vol. 25, no. 11, pp. 2896–2902, 2007.
- [27] J. Galipeau, "The mesenchymal stromal cells dilemma—does a negative phase III trial of random donor mesenchymal stromal cells in steroid-resistant graft-versus-host disease represent a death knell or a bump in the road?" *Cytotherapy*, vol. 15, no. 1, pp. 2–8, 2013.
- [28] J. Lei, D. Hui, W. Huang et al., "Heterogeneity of the biological properties and gene expression profiles of murine bone marrow stromal cells," *International Journal of Biochemistry and Cell Biology*, vol. 45, no. 11, pp. 2431–2443, 2013.
- [29] M. Krampera, L. Cosmi, R. Angeli et al., "Role for interferon- γ in the immunomodulatory activity of human bone marrow mesenchymal stem cells," *Stem Cells*, vol. 24, no. 2, pp. 386–398, 2006.
- [30] G. Ren, J. Su, L. Zhang et al., "Species variation in the mechanisms of mesenchymal stem cell-mediated immunosuppression," *Stem Cells*, vol. 27, no. 8, pp. 1954–1962, 2009.
- [31] X. Zhang, C. Jiao, and S. Zhao, "Role of mesenchymal stem cells in immunological rejection of organ transplantation," *Stem Cell Reviews and Reports*, vol. 5, no. 4, pp. 402–409, 2009.
- [32] S. Aggarwal and M. F. Pittenger, "Human mesenchymal stem cells modulate allogeneic immune cell responses," *Blood*, vol. 105, no. 4, pp. 1815–1822, 2005.
- [33] M. M. Duffy, T. Ritter, R. Ceredig, and M. D. Griffin, "Mesenchymal stem cell effects on T-cell effector pathways," *Stem Cell Research and Therapy*, vol. 2, no. 4, article 34, 2011.
- [34] E. Soleymaninejadian, K. Pramanik, and E. Samadian, "Immunomodulatory properties of mesenchymal stem cells: cytokines and factors," *American Journal of Reproductive Immunology*, vol. 67, no. 1, pp. 1–8, 2012.
- [35] Ê. J. Bassi, D. C. de Almeida, P. M. M. Moraes-Vieira, and N. O. S. Câmara, "Exploring the role of soluble factors associated with immune regulatory properties of mesenchymal stem cells," *Stem Cell Reviews and Reports*, vol. 8, no. 2, pp. 329–342, 2012.
- [36] F. Baratelli, Y. Lin, L. Zhu et al., "Prostaglandin E₂ induces FOXP3 gene expression and T regulatory cell function in human CD4⁺ T cells," *The Journal of Immunology*, vol. 175, no. 3, pp. 1483–1490, 2005.
- [37] K. English, J. M. Ryan, L. Tobin, M. J. Murphy, F. P. Barry, and B. P. Mahon, "Cell contact, prostaglandin E₂ and transforming growth factor beta 1 play non-redundant roles in human mesenchymal stem cell induction of CD4⁺ CD25^{High} forkhead box P3⁺ regulatory T cells," *Clinical & Experimental Immunology*, vol. 156, no. 1, pp. 149–160, 2009.
- [38] F. Casiraghi, N. Perico, and G. Remuzzi, "Mesenchymal stromal cells to promote solid organ transplantation tolerance," *Current Opinion in Organ Transplantation*, vol. 18, no. 1, pp. 51–58, 2013.
- [39] G. D. Wu, J. A. Nolta, Y.-S. Jin et al., "Migration of mesenchymal stem cells to heart allografts during chronic rejection," *Transplantation*, vol. 75, no. 5, pp. 679–685, 2003.
- [40] S. Itakura, S. Asari, J. Rawson et al., "Mesenchymal stem cells facilitate the induction of mixed hematopoietic chimerism and islet allograft tolerance without GVHD in the rat," *American Journal of Transplantation*, vol. 7, no. 2, pp. 336–346, 2007.
- [41] W. Ge, J. Jiang, J. Arp, W. Liu, B. Garcia, and H. Wang, "Regulatory T-cell generation and kidney allograft tolerance induced by mesenchymal stem cells associated with indoleamine 2,3-dioxygenase expression," *Transplantation*, vol. 90, no. 12, pp. 1312–1320, 2010.
- [42] C. D. Wan, R. Cheng, H. B. Wang, and T. Liu, "Immunomodulatory effects of mesenchymal stem cells derived from adipose tissues in a rat orthotopic livertransplantation model," *Hepato-biliary & Pancreatic Diseases International*, vol. 7, no. 1, pp. 29–33, 2008.
- [43] J. Gao, J. E. Dennis, R. F. Muzic, M. Lundberg, and A. I. Caplan, "The dynamic in vivo distribution of bone marrow-derived mesenchymal stem cells after infusion," *Cells Tissues Organs*, vol. 169, no. 1, pp. 12–20, 2001.
- [44] Y. Wu and R. C. Zhao, "The role of chemokines in mesenchymal stem cell homing to myocardium," *Stem Cell Reviews and Reports*, vol. 8, no. 1, pp. 243–250, 2012.
- [45] H. M. Hatch, D. Zheng, M. L. Jorgensen, and B. E. Petersen, "SDF-1 α /CXCR4: a mechanism for hepatic oval cell activation and bone marrow stem cell recruitment to the injured liver of rats," *Cloning and Stem Cells*, vol. 4, no. 4, pp. 339–351, 2002.
- [46] R. F. Wynn, C. A. Hart, C. Corradi-Perini et al., "A small proportion of mesenchymal stem cells strongly expresses functionally active CXCR4 receptor capable of promoting migration to bone marrow," *Blood*, vol. 104, no. 9, pp. 2643–2645, 2004.
- [47] E. Chavakis, C. Urbich, and S. Dimmeler, "Homing and engraftment of progenitor cells: a prerequisite for cell therapy," *Journal of Molecular and Cellular Cardiology*, vol. 45, no. 4, pp. 514–522, 2008.
- [48] J. Ye, K. Yao, and J. C. Kim, "Mesenchymal stem cell transplantation in a rabbit corneal alkali burn model: engraftment and

- involvement in wound healing," *Eye*, vol. 20, no. 4, pp. 482–490, 2006.
- [49] G. W. Roddy, J. Y. Oh, R. H. Lee et al., "Action at a distance: systemically administered adult stem/progenitor cells (MSCs) reduce inflammatory damage to the cornea without engraftment and primarily by secretion of TNF- α stimulated gene/protein 6," *Stem Cells*, vol. 29, no. 10, pp. 1572–1579, 2011.
- [50] S. H. Park, K. W. Kim, Y. S. Chun, and J. C. Kim, "Human mesenchymal stem cells differentiate into keratocyte-like cells in keratocyte-conditioned medium," *Experimental Eye Research*, vol. 101, pp. 16–26, 2012.
- [51] J. Cejkova, P. Trosan, C. Cejka et al., "Suppression of alkali-induced oxidative injury in the cornea by mesenchymal stem cells growing on nanofiber scaffolds and transferred onto the damaged corneal surface," *Experimental Eye Research*, vol. 116, pp. 312–323, 2013.
- [52] J. Y. Oh, R. H. Lee, J. M. Yu et al., "Intravenous mesenchymal stem cells prevented rejection of allogeneic corneal transplants by aborting the early inflammatory response," *Molecular Therapy*, vol. 20, no. 11, pp. 2143–2152, 2012.
- [53] Z. Jia, C. Jiao, S. Zhao et al., "Immunomodulatory effects of mesenchymal stem cells in a rat corneal allograft rejection model," *Experimental Eye Research*, vol. 102, pp. 44–49, 2012.
- [54] O. Treacy, L. O'Flynn, A. E. Ryan et al., "Mesenchymal stem cell therapy promotes corneal allograft survival in rats by local and systemic immunomodulation," *American Journal of Transplantation*, vol. 14, no. 9, pp. 2023–2036, 2014.
- [55] M. J. Lee, A. Y. Ko, J. H. Ko et al., "Mesenchymal stem/stromal cells protect the ocular surface by suppressing inflammation in an experimental dry eye," *Molecular Therapy*, vol. 23, no. 1, pp. 139–146, 2015.
- [56] J. Weng, C. He, P. Lai et al., "Mesenchymal stromal cells treatment attenuates dry eye in patients with chronic graft-versus-host disease," *Molecular Therapy*, vol. 20, no. 12, pp. 2347–2354, 2012.
- [57] S. Inoue, F. C. Popp, G. E. Koehl et al., "Immunomodulatory effects of mesenchymal stem cells in a rat organ transplant model," *Transplantation*, vol. 81, no. 11, pp. 1589–1595, 2006.
- [58] E. Y. Lee, Y. Xia, W.-S. Kim et al., "Hypoxia-enhanced wound-healing function of adipose-derived stem cells: increase in stem cell proliferation and up-regulation of VEGF and bFGF," *Wound Repair and Regeneration*, vol. 17, no. 4, pp. 540–547, 2009.
- [59] T. J. Cashman, V. Gouon-Evans, and K. D. Costa, "Mesenchymal stem cells for cardiac therapy: practical challenges and potential mechanisms," *Stem Cell Reviews and Reports*, vol. 9, no. 3, pp. 254–265, 2013.
- [60] A. M. Abarbanell, Y. Wang, J. L. Herrmann et al., "Toll-like receptor 2 mediates mesenchymal stem cell-associated myocardial recovery and VEGF production following acute ischemia-reperfusion injury," *The American Journal of Physiology—Heart and Circulatory Physiology*, vol. 298, no. 5, pp. H1529–H1536, 2010.
- [61] W. M. Jackson, L. J. Nesti, and R. S. Tuan, "Concise review: clinical translation of wound healing therapies based on mesenchymal stem cells," *Stem Cells Translational Medicine*, vol. 1, no. 1, pp. 44–50, 2012.
- [62] L. Yao, Z.-R. Li, W.-R. Su et al., "Role of mesenchymal stem cells on cornea wound healing induced by acute alkali burn," *PLoS ONE*, vol. 7, no. 2, Article ID e30842, 2012.
- [63] J. Y. Oh, M. K. Kim, M. S. Shin et al., "The anti-inflammatory and anti-angiogenic role of mesenchymal stem cells in corneal wound healing following chemical injury," *Stem Cells*, vol. 26, no. 4, pp. 1047–1055, 2008.
- [64] V. Holan and E. Javorkova, "Mesenchymal stem cells, nanofiber scaffolds and ocular surface reconstruction," *Stem Cell Reviews and Reports*, vol. 9, no. 5, pp. 609–619, 2013.
- [65] H. Reinshagen, C. Auw-Haedrich, R. V. Sorg et al., "Corneal surface reconstruction using adult mesenchymal stem cells in experimental limbal stem cell deficiency in rabbits," *Acta Ophthalmologica*, vol. 89, no. 8, pp. 741–748, 2011.
- [66] L. Liang, H. Sheha, J. Li, and S. C. G. Tseng, "Limbal stem cell transplantation: new progresses and challenges," *Eye*, vol. 23, no. 10, pp. 1946–1953, 2009.
- [67] M. Eslani, A. Baradaran-Rafii, A. Movahedan, and A. R. Djalilian, "The ocular surface chemical burns," *Journal of Ophthalmology*, vol. 2014, Article ID 196827, 9 pages, 2014.
- [68] Y. Lan, S. Kodati, H. S. Lee, M. Omoto, Y. Jin, and S. K. Chauhan, "Kinetics and function of mesenchymal stem cells in corneal injury," *Investigative Ophthalmology and Visual Science*, vol. 53, no. 7, pp. 3638–3644, 2012.
- [69] J. Y. Oh, J. H. Ko, M. K. Kim, and W. R. Wee, "Effects of mesenchymal stem/stromal cells on cultures of corneal epithelial progenitor cells with ethanol injury," *Investigative Ophthalmology & Visual Science*, vol. 55, no. 11, pp. 7628–7635, 2014.
- [70] J. Y. Oh, M. K. Kim, M. S. Shin et al., "The anti-inflammatory and anti-angiogenic role of mesenchymal stem cells in corneal wound healing following chemical injury," *STEM CELLS*, vol. 26, no. 4, pp. 1047–1055, 2008.
- [71] Y. Ma, Y. Xu, Z. Xiao et al., "Reconstruction of chemically burned rat corneal surface by bone marrow-derived human mesenchymal stem cells," *Stem Cells*, vol. 24, no. 2, pp. 315–321, 2006.
- [72] A. Zajicova, K. Pokorna, A. Lencova et al., "Treatment of ocular surface injuries by limbal and mesenchymal stem cells growing on nanofiber scaffolds," *Cell Transplantation*, vol. 19, no. 10, pp. 1281–1290, 2010.
- [73] J. Y. Lee, H. J. Jeong, M. K. Kim, and W. R. Wee, "Bone marrow-derived mesenchymal stem cells affect immunologic profiling of interleukin-17-secreting cells in a chemical burn mouse model," *Korean Journal of Ophthalmology*, vol. 28, no. 3, pp. 246–256, 2014.
- [74] J. Y. Oh, G. W. Roddy, H. Choi et al., "Anti-inflammatory protein TSG-6 reduces inflammatory damage to the cornea following chemical and mechanical injury," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 107, no. 39, pp. 16875–16880, 2010.
- [75] K. Cunnusamy, P. W. Chen, and J. Y. Niederkorn, "Paradigm shifts in the role of CD4+ T cells in keratoplasty," *Discovery Medicine*, vol. 10, no. 54, pp. 452–461, 2010.
- [76] J. Y. Niederkorn and D. F. Larkin, "Immune privilege of corneal allografts," *Ocular Immunology and Inflammation*, vol. 18, no. 3, pp. 162–171, 2010.
- [77] E. Chong and M. R. Dana, "Graft failure IV. Immunologic mechanisms of corneal transplant rejection," *International Ophthalmology*, vol. 28, no. 3, pp. 209–222, 2008.
- [78] J. Y. Niederkorn, "High-risk corneal allografts and why they lose their immune privilege," *Current Opinion in Allergy and Clinical Immunology*, vol. 10, no. 5, pp. 493–497, 2010.
- [79] A. Poon, M. Constantinou, E. Lamoureux, and H. R. Taylor, "Topical Cyclosporin A in the treatment of acute graft rejection: a randomized controlled trial," *Clinical and Experimental Ophthalmology*, vol. 36, no. 5, pp. 415–421, 2008.

- [80] Y. Liu, J. Jiang, H. Xiao et al., "Topical application of FTY720 and cyclosporin A prolong corneal graft survival in mice," *Molecular Vision*, vol. 18, pp. 624–633, 2012.
- [81] J. Yuan, J. J. Zhai, X. Huang, S. Y. Zhou, and J. Q. Chen, "Ocular safety and pharmacokinetics study of FK506 suspension eye drops after corneal transplantation," *Journal of Ocular Pharmacology and Therapeutics*, vol. 28, no. 2, pp. 153–158, 2012.
- [82] A. A. Zaki, M. S. Elalfy, D. G. Said, and H. S. Dua, "Deep anterior lamellar keratoplasty—triple procedure: a useful clinical application of the pre-Descemet's layer (Dua's layer)," *Eye*, vol. 29, no. 3, pp. 323–326, 2015.
- [83] S. M. Pantanelli, A. Herzlich, G. Yeane, and S. T. Ching, "Recurrence of granular corneal dystrophy type I deposits within host stroma after non-Descemet baring anterior lamellar keratoplasty," *Cornea*, vol. 33, no. 12, pp. 1348–1351, 2014.
- [84] G. S. L. Peh, R. W. Beuerman, A. Colman, D. T. Tan, and J. S. Mehta, "Human corneal endothelial cell expansion for corneal endothelium transplantation: an overview," *Transplantation*, vol. 91, no. 8, pp. 811–819, 2011.
- [85] F. Casiraghi, N. Azzollini, M. Todeschini et al., "Localization of mesenchymal stromal cells dictates their immune or proinflammatory effects in kidney transplantation," *American Journal of Transplantation*, vol. 12, no. 9, pp. 2373–2383, 2012.
- [86] M. Krampera, "Mesenchymal stromal cell 'licensing': a multi-step process," *Leukemia*, vol. 25, no. 9, pp. 1408–1414, 2011.
- [87] F. Casiraghi, N. Azzollini, P. Cassis et al., "Pretransplant infusion of mesenchymal stem cells prolongs the survival of a semiallogeneic heart transplant through the generation of regulatory T cells," *Journal of Immunology*, vol. 181, no. 6, pp. 3933–3946, 2008.
- [88] J. Y. Oh, M. K. Kim, J. H. Ko, H. J. Lee, J. H. Lee, and W. R. Wee, "Rat allogeneic mesenchymal stem cells did not prolong the survival of corneal xenograft in a pig-to-rat model," *Veterinary Ophthalmology*, vol. 12, supplement 1, pp. 35–40, 2009.
- [89] S. Fuentes-Julián, F. Arnalich-Montiel, L. Jaumandreu et al., "Adipose-derived mesenchymal stem cell administration does not improve corneal graft survival outcome," *PLoS ONE*, vol. 10, no. 3, Article ID e0117945, 2015.
- [90] "The definition and classification of dry eye disease: report of the definition and classification Subcommittee of the International Dry Eye Workshop (2007)," *The Ocular Surface*, vol. 5, no. 2, pp. 75–92, 2007.
- [91] J. L. Gayton, "Etiology, prevalence, and treatment of dry eye disease," *Clinical Ophthalmology*, vol. 3, no. 1, pp. 405–412, 2009.
- [92] Y. Ogawa and M. Kuwana, "Dry eye as a major complication associated with chronic graft-versus-host disease after hematopoietic stem cell transplantation," *Cornea*, vol. 22, no. 7, pp. S19–S27, 2003.
- [93] Y. Ogawa, S. Okamoto, M. Wakui et al., "Dry eye after haematopoietic stem cell transplantation," *British Journal of Ophthalmology*, vol. 83, no. 10, pp. 1125–1130, 1999.
- [94] M. E. Stern, C. S. Schaumburg, and S. C. Pflugfelder, "Dry eye as a mucosal autoimmune disease," *International Reviews of Immunology*, vol. 32, no. 1, pp. 19–41, 2013.
- [95] W. Stevenson, S. K. Chauhan, and R. Dana, "Dry eye disease: an immune-mediated ocular surface disorder," *Archives of Ophthalmology*, vol. 130, no. 1, pp. 90–100, 2012.
- [96] D. Zoukhri, "Mechanisms involved in injury and repair of the murine lacrimal gland: role of programmed cell death and mesenchymal stem cells," *Ocular Surface*, vol. 8, no. 2, pp. 60–69, 2010.
- [97] D. Zoukhri, R. R. Hodges, D. Byon, and C. L. Kublin, "Role of proinflammatory cytokines in the impaired lacrimation associated with autoimmune xerophthalmia," *Investigative Ophthalmology and Visual Science*, vol. 43, no. 5, pp. 1429–1436, 2002.
- [98] E. Beyazyıldız, F. A. Pınarlı, Ö. Beyazyıldız et al., "Efficacy of topical mesenchymal stem cell therapy in the treatment of experimental dry eye syndrome model," *Stem Cells International*, vol. 2014, Article ID 250230, 9 pages, 2014.
- [99] J. A. Wood, D.-J. Chung, S. A. Park et al., "Periocular and intra-articular injection of canine adipose-derived mesenchymal stem cells: an in vivo imaging and migration study," *Journal of Ocular Pharmacology and Therapeutics*, vol. 28, no. 3, pp. 307–317, 2012.
- [100] A. J. Villatoro, V. Fernández, S. Claros, G. A. Rico-Llanos, J. Becerra, and J. A. Andrades, "Use of adipose-derived mesenchymal stem cells in keratoconjunctivitis sicca in a canine model," *BioMed Research International*, vol. 2015, Article ID 527926, 10 pages, 2015.