

# The Role and Therapeutic Potential of Macropinocytosis in Cancer

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Qiu Z, Liu W, Zhu Q, Ke K, Zhu Q, Jin W, Yu S, Yang Z, Li L, Sun X, Ren S, Liu Y, Zhu Z, Zeng J, Huang X, Huang Y, Wei L, Ma M, Lu J, Chen X, Mou Y, Xie T and Sui X (2022) The Role and Therapeutic Potential of Macropinocytosis in Cancer. Front. Pharmacol. 13:919819. doi: 10.3389/fphar.2022.919819 Macropinocytosis, a unique endocytosis pathway characterized by nonspecific internalization, has a vital role in the uptake of extracellular substances and antigen presentation. It is known to have dual effects on cancer cells, depending on cancer type and certain microenvironmental conditions. It helps cancer cells survive in nutrientdeficient environments, enhances resistance to anticancer drugs, and promotes invasion and metastasis. Conversely, overexpression of the RAS gene alongside drug treatment can lead to methuosis, a novel mode of cell death. The survival and proliferation of cancer cells is closely related to macropinocytosis in the tumor microenvironment (TME), but identifying how these cells interface with the TME is crucial for creating drugs that can limit cancer progression and metastasis. Substantial progress has been made in recent years on designing anticancer therapies that utilize the effects of macropinocytosis. Both the induction and inhibition of macropinocytosis are useful strategies for combating cancer cells. This article systematically reviews the general mechanisms of macropinocytosis, its specific functions in tumor cells, its occurrence in nontumor cells in the TME, and its application in tumor therapies. The aim is to elucidate the role and therapeutic potential of macropinocytosis in cancer treatment.

Keywords: macropinocytosis, promoting cancer growth, tumor microenvironment, methuosis, anticancer therapies

# INTRODUCTION

The development of new anticancer strategies is aided by understanding the molecular mechanisms that afford growth and survival advantages to cancer cells (Zhao et al., 2020). One such adaptive strategy of cancer cells to meet their metabolic needs is the ability to develop alternative nutrient access pathways (Recouvreux and Commisso, 2017). For example, macropinocytosis can enhance the survival, metastasis, and therapeutic resistance of cancer cells (Jayashankar and Edinger, 2020). This ability is essential to cancer cells, and anticancer strategies can be designed around the targeting of macropinocytosis. Macropinocytosis-mediated tumor growth and drug resistance have recently become hot topics in cancer research. This review describes the macropinocytotic mechanisms that affect cancerous growth, resistance, and metastasis. Most significantly, the abnormal function of macropinocytosis is shown to induce methuosis, a new model of cell death. Secondly, the impacts of

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microenvironmental components on tumor cell growth *via* macropinocytosis are described. Thirdly, anticancer strategies targeting macropinocytosis are reviewed. These include the delivery of chemotherapeutic drugs *via* macropinocytosis to improve their efficacy, the utilization of macropinocytosis inhibitors, and the induction of methuosis. Finally, future research directions are discussed.

## DEFINITION AND ORIGIN OF MACROPINOCYTOSIS

Macropinocytosis is clathrin- and caveolin-independent endocytosis (Puccini et al., 2022). This morphological phenomenon-the internalization of extracellular fluids by cytoplasmic membrane folding (King and Kay, 2019)-was first described in the 1930s. It can occur spontaneously or be activated by growth factors, chemokines, or Toll-like receptors, although both cases require the presence of critical signaling hubs. Macropinocytosis signaling pathways form robust submembranous actin networks that induce membrane ruffling and ultimately lead to macropinosome closure (Marques et al., 2017). Macropinocytosis facilitates the largescale, nonselective internalization of solute molecules in various cell types, including immune cells, fibroblasts, and cancer cells (Hetzenecker et al., 2016; Lin et al., 2020). It enables immune cells to absorb a vast number of pathogens, facilitating immune surveillance and response shaping (Canton, 2018). In solid

tumors, macropinocytosis is also a metabolic adaptation in response to nutrient starvation and metabolic stresses. For example, amino acids such as glutamine are essential for cell growth. In RAS-mutant cancers, macropinocytosis enables cancer cells to absorb extracellular proteins which they degrade to amino acids to fuel cell survival and proliferation (Commisso et al., 2013).

Although macropinocytosis varies between different cell types, the same molecular machinery is utilized. The main steps include cytoplasmic membrane shrinkage, formation of macropinocytic cups, generation and maturation of macropinosomes, macropinosome-lysosome fusion, recycling, and degradation (Figure 1) (Song et al., 2020). Macropinocytosis is initiated by cup-shaped extensions of the plasma membrane, often known as circular ruffles, that are driven by actin polymerization (Ha et al., 2016; Palm, 2019). The formation of macropinosomes thus involves cytoskeleton remodeling. Merlin deficiency was shown to prime the membrane:cytoskeleton interface for epidermal growth factor (EGF)-induced macropinocytosis by increasing cortical ezrin (Chiasson-MacKenzie et al., 2018). The small GTPases activation of was implicated in macropinocytosis by an insertional mutant (Williams et al., 2019). Phosphoinositides (PIs), which transport specific proteins to the endocytic vesicles at a specific time, are also essential for macropinocytosis (Schink et al., 2021).

Loss of liquid or membrane make vesicles more concentrated. Increasing osmotic pressure can cause macropinosomes to lose more surface area than bulk during fission (Hamilton et al., 2007).



to metastasize. The stimulation of shear stress promotes ARAP1/Arf1 trafficking to support cancer cell migration.

Therefore, the concentration stage is likely to be fundamental to the maturation of macropinosomes. In addition, small GTPases and PIs regulate macropinosome maturation (Egami et al., 2014).

After maturation, macropinosomes undergo circulation and digestion processes. Their rapid volume loss is mediated by a twopore channel that transports sodium ions (Freeman et al., 2020). This process reduces the hydrostatic pressure in the macropinosome and facilitates the recycling pathways that return the endocytosed lipids and proteins to the plasma membrane (King and Smythe, 2020). The WASH (WASP and SCAR homologs) complex and retromer sorting complex (RSC) interact with the retromer complex at both early and late phases of macropinosome maturation to mediate recycling (Buckley et al., 2016). In addition, RAS-induced macropinocytosis is regulated by the translocation of vacuolar ATPase (v-ATPase) from intracellular membranes to the plasma membrane (Ramirez et al., 2019).

TRPML1 is a downstream effector of PIKfyve which partially regulates vacuole size (Krishna et al., 2016). It is a lysosome Ca<sup>2+</sup> channel regulated by PI(3,5)P2. Lysosomes can modify macropinosomes isolated from TRPML1 silencing cells, but they do not fuse with each other (Dayam et al., 2015). PIKfyve and PI(3,5)P2 activate TRPML1 to induce macropinosome–lysosome fusion. Melanoma cells lacking

TRPML1 exhibit lower survival, proliferation, tumor development, and macropinocytosis (Kasitinon et al., 2019). The filamentous septin GTPases connect preferentially with mature macropinosomes and localize to their fusion sites with macropinosomes in a phosphatidylinositol 3.5bisphosphate-dependent manner. Septin deficiency leads to large clusters of docked macropinosomes that persist for longer and undergo fewer fusion events (Dolat and Spiliotis, 2016). This indicates that PIKfyve, TRPML1, and septin are involved in macropinosome degradation.

# MACROPINOCYTOSIS AND TUMOR CELLS

# Benefits of Macropinocytosis for Cancer Cells

Macropinocytosis Promotes Cell Growth and Survival Cancer cells can acquire extracellular nutrients from nutrientpoor environments via macropinocytosis (Figure 2). This process depends on the activation of the RAS gene, growth factor receptors (GFRs), and other signal pathways (Hesketh et al., 2020; Hobbs et al., 2020). KRAS-transformed pancreatic cancer cells activate macropinocytosis to absorb and degrade extracellular fluids in order to survive (Hou et al., 2021). Membrane localization of trafficking syndecan-1 (SDC1) has been shown to be an essential mechanism for promoting macropinocytosis and sustaining the uncontrolled growth of oncogenic KRAS-driven pancreatic ductal adenocarcinoma (PDAC). SDC1 generates a signaling complex that activates Rac1 GTPase, which in turn stimulates PAK1 and activates macropinocytosis (Yao et al., 2019). Nonsmall cell lung cancer (NSCLC) exploits Rac-driven macropinocytosis of extracellular protein as an adaptive mechanism to survive glucose deprivation. Macropinocytosis is enhanced in these cells and is regulated via Rac-Pak signaling activation by phosphoinositide 3-kinase (PI3K) (Hodakoski et al., 2019).

Epidermal growth factor receptor (EGFR) enhances macropinocytosis during glutamine starvation by activating the serine-threonine kinase PAK1, which directly orchestrates the required alterations in membrane and cytoskeletal dynamics (Weerasekara et al., 2019). Lee et al. demonstrated that regional amino acid deficiency was associated with increased levels of macropinocytosis. This was mediated by nutritional stress-induced potentiation of EGFR signaling, which controlled the extent of macropinocytosis in PDAC *via* PAK1 activation (Lee et al., 2019).

The mammalian target of rapamycin (mTOR), a cellular pathway coordinating nutrient sensing and adaptive responses, has been implicated in macropinocytosis (Cang et al., 2013). mTOR inhibition elevated lysosomal catabolism of extracellular proteins and promoted cancer cell survival under nutrientdepleted conditions (Palm et al., 2015). In melanoma cells, TRPML1 is essential to inhibit the MAPK pathway and mTORC1 signaling. Macropinocytosis is reduced in TRPML1deficient melanoma cells but is partially or totally restored by mTORC1 inhibition. TRPML1 expression is increased in melanoma cells compared to melanocytes, which suppresses MAPK and mTORC1 signaling, maintains macropinocytosis, and prevents proteotoxic stress (Kasitinon et al., 2019).

Xiao et al. demonstrated that Rab GTPases significantly increased in chRCC specimens and induced membrane trafficking and vesicle formation. Furthermore, the phospholipase C gamma 2 (PLCG2)/inositol 1,4,5trisphosphate (IP3)/Ca2b/protein kinase C (PKC) pathway regulated the activation of macropinocytosis and promoted chRCC cell proliferation under nutrient stress (Xiao et al., 2020). Thus, the oncogenic KRAS gene, GFRs, mTORC1, and other factors are highly important for macropinocytosiscancer cell survival nutrient-deficient mediated in environments. These studies inform the development of effective anticancer agents.

### Macropinocytosis and Therapeutic Resistance

Therapeutic resistance is a significant challenge to effective cancer treatment, especially in patients with aggressive cancers in which mutations that drive macropinocytosis are widespread. Hypoxia and nutrient deprivation are common in rapidly growing malignant tumors due to limited blood supply. This results in necrotic cell death in the core of solid tumors (Lee et al., 2018). The cancer cells ingest the necrotic cell debris *via* macropinocytosis, utilizing the amino acids, fatty acids, sugars, and nucleotides for biosynthesis, which makes them resistant to therapies that target anabolic pathways (Jayashankar and Edinger, 2020).

Collagen type I (Col I) induces EGFR tyrosine kinase inhibitor (EGFR-TKI) resistance by mTOR activation *via* Akt and ERK1/2 independent pathways (Yamazaki et al., 2018). 5-(N-ethyl-N-isopropyl) amiloride (EIPA) is a macropinocytosis inhibitor. EGFR-TKI resistance developed in EGFR-mutated lung cancer cells by Col I absorption *via* macropinocytosis, while macropinocytosis inhibition reduced Col I absorption in PC-9 cells and restored their EGFR-TKI sensitivity (Yamazaki et al., 2020).

The regulation of microtubule affinity-regulating kinase 3 (MARK3), a kinase regulator of cytoskeleton dynamics, is involved in macropinocytosis. Changes in KRAS (KRAS<sup>\*</sup>) are found in virtually all PDAC cases and are linked to tumor survival. Hou et al. identified a deubiquitinase USP21-driven mechanism that was resistant to KRAS<sup>\*</sup> treatment. USP21 promotes KRAS<sup>\*</sup>-independent cancer development *via* the mTOR signaling pathway by MARK3-mediated up-regulation of macropinocytosis, which helps to fulfil the anabolic requirements (Hou et al., 2021).

Inhibition of autophagy can limit the progression of existing tumors and increase responsiveness to cancer treatment (Amaravadi et al., 2019). However, when PDAC is treated with autophagy inhibitors, the cancer cells take up exogenous proteins *via* macropinocytosis induced by NRF2, which provides the metabolites and energy required to develop therapeutic resistance (Su et al., 2021).

A recent study found that macropinocytosis reduces sensitivity to mTOR inhibition by restoring AKT phosphorylation at serine 473, and leading to cancer cell proliferation. This macropinocytosis-mediated resistance was eliminated when mTOR and lysosomal degradation of absorbed protein were inhibited (Michalopoulou et al., 2020). Thus, macropinocytosis allows cancer cells to develop resistance to antitumor drug therapies, making them ineffective. However, macropinocytosis inhibition is a potential therapy to combat this resistance.

### Macropinocytosis and Metastasis

Systemic metastasis of cancers is the main prelude to mortality in cancer patients. The detection of metastasis using clinical imaging methods usually indicates a poor prognosis (Klein, 2020; Qin et al., 2020). Metastasis is a major challenge because of its complexity. It is associated with the development of epithelial-mesenchymal transition (EMT) in a wide range of cancers (Jiao et al., 2020; Fuh et al., 2021). Extracellular ATP (eATP) can trigger EMT in cancer cells via purinoceptor signals activated by P2X7. It can also activate production of matrix metallopeptidases (MMPs) and promote lung cancer cell shedding, EMT, migration, and invasion. This induction happens outside the cell but the functions internalized are into the cell bv macropinocytosis. During this process, the enhancing transcription factors Snail and Slug (and probably others) may promote EMT (Cao et al., 2019).

The TAM receptor AXL and its ligand, GAS6, have been linked to the development and metastasis of various malignancies (Falcone et al., 2020). Zdalik-Bielecka et al. demonstrated that GAS6-AXL signaling induced PI3K- and Rac1-dependent, actindriven cytoskeletal rearrangements and macropinocytosis that jointly promoted cancer invasion and metastasis (Zdzalik-Bielecka et al., 2021).

Actomyosin is recruited to the cell front of immature dendritic cells to produce macropinosomes. As a result, these cells are unaffected by hydraulic resistance (HR) because macropinocytosis allows fluid to pass through them. This mechanism enhances their ability to expand into other body spaces and may also enhance the ability of cancer cells to invade and metastasize (Moreau et al., 2019).

Circular dorsal ruffles (CDRs) are ring-shaped membrane structures that are rich in F-actin. Shear pressure promotes CDR development through the integrin-connected kinase (ILK)-ADP-ribosylation factors (ARAP1/Arf1) cascade. Shear stress can induce the establishment of intracellular macropinocytosis after CDR depolymerization, promoting ARAP1/Arf1 trafficking from macropinocytosis to early endosome marker Rab5 to facilitate cancer cell migration (Qin et al., 2021).

Yao et al. showed that aborting macropinocytosis by deletion of the HSPG receptor SDC1 might reduce lung metastatic potential and make pancreatic cancer cells less clonogenic (Yao et al., 2019). Choi et al. found that the number of metastatic nodules was dramatically reduced when RAS3 cells pretreated with EIPA were delivered intravenously into immunedeficient mice (Choi et al., 2021). Inhibiting macropinocytosis function is likely to significantly slow down metastasis and reduce cancer-related deaths.

# Negative Impacts of Dysregulated Macropinocytosis on Cancer Cells

Cancer cells sustain their vitality through macropinocytosis when the nutrient and growth factors in the tumor microenvironment (TME) change. However, hyperstimulation of macropinocytosis by active RAS expression or some medicines can cause cell death, also known as methuosis. Methuosis occurs when cells accumulate too much fluid through uncontrolled macropinocytosis (Figure 3). In the normal macropinocytosis process, folds in the plasma membrane (PM) produce macropinocytic cups that close to form macropinosomes inside the cell. Circulating receptors transport some macropinosomes back to the PM (Toh et al., 2019), while other macropinosomes fuse with lysosomes and are cleaved (Maltese and Overmeyer, 2015).

Liquid uptake via macropinocytosis is enhanced during methuosis. Instead of shrinking, the vesicles swell, do not acidify, remain nonfunctional, and merge with each other rather than with lysosomes. This creates large vacuoles and catastrophic cell swelling that result in bursting of the PM, cell lysis, and death (Ritter et al., 2021). When activated Ras enters cancer cells, it causes the formation of large phase-lucent cytoplasmic vacuoles, leading to methuosis (Dendo et al., 2018). Endosome morphological changes associated with Rasinduced methuosis are caused by Rac1 downstream activation and Arf6 reciprocal inactivation (Bhanot et al., 2010). Activating receptor tyrosine kinase TrkA the overstimulates macropinocytosis in non-Ras altered medulloblastoma brain tumors, leading to catastrophic membrane disintegration and methuosis (Li et al., 2010). The molecular basis of this uncontrolled TrkA-driven macropinocytosis is the inhibition of RhoB that relieves actin stress fibers, and FRS2-scaf folded Src and H-Ras activation of RhoA that stimulates actin reorganization and the formation of lamellipodia (Li et al., 2016).

# MACROPINOCYTOSIS AND THE TUMOR MICROENVIRONMENT

# Macropinocytosis and Cancer-Associated Fibroblasts (CAFs)

CAFs are a prominent cellular component of the TME in most solid tumors and macropinocytosis in CAFs correlates with cancer growth (**Figure 4**). Zhang et al. demonstrated that glutamine deprivation triggered macropinocytosis in pancreatic CAFs. They discovered that stromal macropinocytosis was aided by increased cytosolic  $Ca^{2+}$  and was dependent on ARHGEF2 and CaMKK2-AMPK signaling. This macropinocytosis had two purposes: as a source of intracellular amino acids to maintain the functionality of the CAF cells, and to feed the PDAC cells with secreted amino acids that helped them avoid the negative effects of nutritional stress in the TME (Zhang Y. et al., 2021).

The Ras inhibitor, RASAL3 (epigenetically silenced in human prostatic CAFs) results in oncogenic Ras activity that promotes macropinocytosis-mediated glutamine synthesis. In prostate cancer, stromal glutamine acts as a source of energy *via* 



anaplerosis and as a facilitator of neuroendocrine differentiation. The suppression of macropinocytosis and glutamine transport has anticancer effects in an orthotopic xenograft model (Mishra et al., 2018). In addition, Zhao et al. found that CAF-derived exosomes (CDEs) increased glycolysis and glutamine-dependent reductive carboxylation in cancer cells by inhibiting mitochondrial oxidative phosphorylation. Using <sup>13</sup>C-isotope labeling, they showed that exosomes delivered amino acids to cancer cells in a manner similar to macropinocytosis (Zhao et al., 2016).

# Macropinocytosis and the Cancer Immune Response

Dendritic cells trigger an immune response by presenting a polypeptide-MHC complex on their surface. T cells recognize and are activated by this complex. The ability of dendritic cells to efficiently present such complexes is partly due to their ability to capture antigens through macropinocytosis (Liu and Roche, 2015). The normal physiological functioning of macrophages, B cells, and T cells is inseparable from macropinocytosis (Redka et al., 2018; Lin et al., 2020). Under amino acid (AA) replete conditions, macropinocytosis enables extracellular AA to enter an endolysosomal compartment in T cells, allowing mTORC1 to remain activated and stimulating T cell proliferation (Charpentier et al., 2020). The relationship between macropinocytosis and immune cells in the TME is illustrated in **Figure 4**. Neutral lipids accumulating in lung neutrophils were transferred to metastatic tumor cells *via* a macropinocytosis-lysosome pathway. This enhanced the survival and proliferation of the tumor cells, while pharmacological suppression of macropinocytosis significantly reduced metastatic colonization by breast tumor cells (Li et al., 2020).

Treatment resistance in PDAC is partly attributable to responses including the general infiltration of immunosuppressive macrophages (M2), whilst type M1



macrophages inhibit tumor growth (Avila-Ponce de Leon et al., 2021). Macrophages ingest nanoparticle albumin-bound (nab)paclitaxel primarily through macropinocytosis, which is sufficient to cause macrophage M1 polarization. This nab-paclitaxelmediated M1 activation may result in positive feedback signaling, increasing drug uptake and strengthening M1activating effects in both an autocrine and paracrine manner (Cullis et al., 2017). Further investigation is required into the role of macropinocytosis in immune cells in cancer.

# Macropinocytosis and Other Cell Types in Cancer

Macropinocytosis in other cell types has been connected to cancer survival in the TME (**Figure 4**). Bone marrow-derived mesenchymal stem cells (BMSCs), a component of the TME, play an essential role in cancer (Zhang et al., 2019). Transforming growth factor  $\beta$  receptor II (TGF $\beta$ RII), with its complex functions in cancer progression, is often mutated in cancers. Exosomes produced by PC12 cells can enter BMSCs *via* macropinocytosis and transfer microRNAs into these cells. miR-21 can reduce the expression of TGFRII and tropomyosin-1 (TPM1) in BMSCs. As a result, tumor exosomes may regulate cancer progression by lowering TGFRII and TPM1 levels in normal cells (Tian et al., 2014). The communication between multiple myeloma (MM) cells and BMSCs is pivotal in MM cell development, proliferation, and treatment resistance. Macropinocytosis was partially involved in the uptake of exosomes from MM cells, while

exosomes containing miR-146a could be transported into BMSCs. The secretion of various cytokines and chemokines (including CXCL1, IL6, IL-8, IP-10, MCP-1, and CCL-5) increased after miR-146a was overexpressed in BMSCs, leading to increased MM cell survival and migration (De Veirman et al., 2016; Zheng et al., 2019).

Ma et al. analyzed transmission electron micrographs of endothelial cells in pancreatic cancer (PC) and found several extended projections on the luminal surface that resembled macropinocytic filopodia, suggesting the presence of characteristics conducive to macropinocytosis. PC endothelial cells may be the primary location in the microvasculature net where nutritional exchange occurs, and macropinocytosis may facilitate the exchange of nutrients or waste that promotes cancer growth (Ma et al., 2020).

# ANTICANCER THERAPIES TARGETING MACROPINOCYTOSIS

# Exploitation of Macropinocytosis in Cancer Treatment

### The Use of Macropinocytosis-Inducing Carriers

Despite breakthroughs in cancer therapies and the development of personalized medicine, chemoresistance remains a significant factor contributing to cancer mortality (Chern and Tai, 2020). However, macropinocytosis can increase the transport of anticancer agents into cancer cell lines, rendering



tamoxifen etc. can have synergistic antitumor effects.

chemotherapeutic drugs more effective against drug-resistant cells (Muley et al., 2020). Studies have focused on combining substances that induce macropinocytosis with anticancer agents that are not dependent on macropinocytosis. The efficacy of anticancer drugs is improved by the formation of such conjugates. Extracellular vesicles (EVs) (including exosomes) play a crucial role in cell information communication and have potential as drug carriers (Costa Verdera et al., 2017). Nakase et al. described a technique for inducing active macropinocytosis to increase cellular EV uptake using arginine-rich, cell-penetrating peptide (CPP)-modified EVs. The macropinocytosis pathway was activated after the arginine-rich CPPs on the EV membrane were modified, with the number of arginine residues affecting cellular EV absorption efficiency. Subsequently, anticancer activity was achieved using ribosome-inactivating, protein saporin-encapsulated EVs modified with hexadeca-arginine (R16) peptide (Nakase et al., 2017).

Exosomes produced by normal fibroblast-like mesenchymal cells are designed to contain siRNA or shRNA specific to oncogenic KRAS<sup>G12D</sup> (iExosomes), a frequent pancreatic cancer mutation. iExosomes exhibit higher efficacy than liposomes for targeting oncogenic KRAS. This is dependent on

CD47 and aided by macropinocytosis. Multiple pancreatic cancer mice models have been treated with iExosomes, inhibiting tumor growth and improving survival rates (Kamerkar et al., 2017).

Neutral red (NR) is known for its ability to enter cells by a process similar to macropinocytosis, while histone deacetylase (HDAC) plays a vital role in the transcriptional regulation of tumor suppressor genes and oncogenes (Cui et al., 2021). The anticancer effects of HDAC inhibitors such as trans-cinnamic acid (TCA) are known (Kim and Bae, 2011; Zhu et al., 2016). Zhu et al. demonstrated that cinnamoyl phenazine composed of NR and TCA entered cancer cells *via* macropinocytosis and exhibited anticancer effects in pancreatic carcinoma xenografts (Zhu et al., 2017).

Albumin accumulates in certain tumor cells *via* macropinocytosis, so albumin-coupled cytotoxic drugs can be designed to carry anticancer agents (Lagiewka et al., 2021). Nab-paclitaxel, a widely applied clinical nanomedicine, is a nano-albumin-bound drug that leverages the therapeutic potential of macropinocytosis (Adkins et al., 2021).

### The Use of Macropinocytosis-Inducing Drugs

In these previous studies the active medication was coated with, or coupled to, a molecule that caused tumor cells to undergo macropinocytosis. However, a more straightforward option is to use a macropinocytosis inducer to directly increase macropinocytotic extracellular drug absorption by cancer cells (**Figure 5**). The IGF1R kinase inhibitor AXL1717 enhanced the uptake and efficacy of nab-paclitaxel by mimicking glucose deprivation and promoting macropinocytosis *via* APMK, a cellular nutrient sensor (Li et al., 2021). KRAS-enhanced macropinocytosis and reduced FcRn-mediated recycling make pancreatic cancer cells sensitive to albumin-conjugated drugs, such that the drugs are more likely to accumulate in cancer cells (Liu et al., 2019).

Torin1-mediated mTORC1/2 inhibition increases macropinocytosis, whereas verteporfin-induced YAP inhibition causes apoptosis. Torin1-induced macropinocytosis promoted the absorption of verteporfin, boosting its pro-apoptotic properties in breast cancer cells (Dai et al., 2021). Song et al. demonstrated that silmitasertib (formerly CX-4945) caused macropinocytosis in oral squamous cell carcinoma, which could boost cisplatin intracellular uptake and promote apoptosis (Song et al., 2021).

Tubeimoside-1 (TBM1) is a low-toxicity triterpenoid saponin derived from *Bolbostemma paniculatum*, a traditional Chinese herb. It efficiently trafficked dextrans into heterotopic xenografts *via* macropinocytosis. High performance liquid chromatography analysis showed that 5-fluorouracil (5-FU) content dramatically increased in tumor tissues pretreated with TMB1 (Gong et al., 2018). Drugs that induce macropinocytosis when combined with anticancer medications eliminate the need for time-consuming nanoparticle or conjugate syntheses.

# Disrupting Macropinocytosis Suppresses Cancer Cells

### Inhibitors of Macropinocytosis

Given the benefits of macropinocytosis for tumor growth, inhibition of macropinocytosis can impact tumor survival and drug resistance (**Figure 5**) (Hussein et al., 2021). Cytochalasin D, an inhibitor of actin polymerization, suppresses membrane ruffling and macropinocytosis induction in fibroblast cells (Stockinger et al., 2006; Sipos et al., 2019). The PI3K inhibitors wortmannin and LY294002 inhibit the closure of macropinosomes in fibroblasts and macrophages by suppressing the PI3K signaling pathway (Amyere et al., 2000; Araki et al., 2007). The sodium/hydrogen exchange inhibitors amiloride, HOE-694, and EIPA negate EGF-induced macropinocytosis by decreasing the sub-membranous pH (Koivusalo et al., 2010). Bafilomycin A1, a v-ATPase inhibitor that affects lysosomal pH, also prevents macropinocytosis (Kitazawa et al., 2017).

# Synergistic Antitumor Effects of Macropinocytosis Inhibition

As an energy access pathway, macropinocytosis can influence proliferation, metastasis, and metabolic alterations in cancer. However, it is only one cancerous metabolic pathway and there are other survival strategies under nutrient-poor conditions. Consequently, synergistic antitumor effects are possible when macropinocytosis inhibition is combined with anticancer drugs (**Figure 5**). L-asparaginase, a critical component of acute lymphoblastic leukemia therapy, catabolizes L-asparagine to L-aspartate, causing extracellular asparagine to be depleted (Chan et al., 2014). Hanada et al. reported that asparagine depletion (following a combination of asparagine synthetase deletion and L-asparaginase therapy) combined with EIPA treatment suppressed tumor growth in KRAS-mutant CRC cells (Hanada et al., 2021).

Su et al. discovered that the ULK1/2 inhibitor MRT68921 and EIPA on their own had only minor effects on KC6141, 1444 and 1334 tumor growth. However, when combined, the two medicines had a significant influence on growth and lowered intracellular ATP and amino acids (Su et al., 2021). In poorly perfused locations where nutrients are scarce, necrotic cell debris ingested via macropinocytosis can support tumor cell anabolism (Kim et al., 2018). In the presence of standard-of-care chemotherapies that target biosynthetic pathways, the nutrients obtained from necrocytosis may allow cells to proliferate. Therefore, when used in conjunction with regular chemotherapy, macropinocytosis inhibitors have the potential to improve survival rates in patients with potentially fatal or aggressive malignancies (Puccini et al., 2022). A recent study found that EIPA combined with tamoxifen had a synergistic cytotoxic effect in MCF-7 breast cancer cells. EIPA also enhanced the curative effects of cisplatin, doxorubicin, and 5-FU on breast cancer, but the relationship with macropinocytosis inhibition needs further research (Rolver et al., 2020). Unlike the induction of macropinocytosis, which increases antineoplastic drug absorption, macropinocytosis inhibitors can offer antitumor therapies that disrupt the metabolic activity of cancer cells (Table 1).

### Methuosis-Inducing Compounds

Although various anticancer agents are clinically available, the occurrence of intrinsic or acquired resistance is a major obstacle to the use of chemotherapy drugs and targeted therapies. Drug resistance in cancer cells is intimately linked to various mechanisms including increased rates of drug efflux, changes in drug metabolism, and drug target mutations (Holohan et al., 2013; Wang et al., 2020). Many chemotherapeutic agents function by triggering intrinsic apoptosis, so it is reasonable to seek alternative cell death pathways (such as methuosis) to target apoptosis-resistant tumor cells. Various natural products or their derivatives, in addition to synthetic drugs, can induce methuosis (**Figure 3**). These provide a basis for further clinical treatments.

### Natural Products and Derivatives

Many medications derived from natural products or their derivatives have been recognized as potential phytochemical cancer therapies that utilize methuosis (**Table 2**). Induction of methuosis in cancer cells is one of the advantages of natural product antitumor therapies, and these research studies are aiding the development of new anticancer strategies.

Epimedokoreanin C (EKC) is a pentenylated flavonoid extracted from the Chinese herbal medicine epimedium. Liu et al. describe how EKC reduces cell viability and causes

#### TABLE 1 | Examples of use of macropinocytosis in cancer therapy.

Drug	Effect on macropinocytosis	Macropinocytosis target medicine	Type of cancer	Mechanism	Reference
Torin1	Promotion	Verteporfin	Breast cancer	Torin1-induced macropinocytosis led to increased verteporfin intake	Dai et al. (2021)
Silmitasertib	Promotion	Cisplatin	Oral squamous cell carcinoma	Silmitasertib-induced macropinocytosis led to increased cisplatin intake	Yang et al. (2019)
Tubeimoside-1	Promotion	5-FU	Colorectal cancer	Tubeimoside-1-induced macropinocytosis led to increased 5-FU intake	Gong et al. (2018)
Cytochalasin D	Inhibition	None	RAS-driven cancer	Inhibition of actin polymerization	Sipos et al. (2019), Stockinger et al. (2006)
Wortmannin and LY294002	Inhibition	None	RAS-driven cancer	Inhibition of PI3K signaling pathway	Amyere et al. (2000), Araki et al. (2007)
EIPA	Inhibition	MRT68921	Pancreatic ductal adenocarcinoma etc.	Tumor starvation	Su et al. (2021)
Bafilomycin A1	Inhibition	None	RAS-driven cancer	Altered lysosomal pH	Kitazawa et al. (2017)

#### TABLE 2 | Methuosis-inducing compounds.

Drug	Origin	Type of cancer	Mechanism	Reference
EKC	Pentenylated flavonoid isolated from the Chinese herbal medicine epimedium	NCI-H292 and A549 lung cancer cells	Rac1 and Arf6-regulated macropinosomes fused, forming large vacuoles resulting in methuosis	Liu et al. (2021)
IBC	Naturally occurring chalcone compound	Leukemia cells	Vacuolar-type H <sup>+</sup> -ATPase and AKT involved in methuosis-like death	Yang et al. (2019)
UA	Pentacyclic triterpenoid carboxylic acid	HeLa cells etc.	Excessive vacuoles	Sun et al. (2017)
PD	Extracted from the root of <i>Platycodon</i> grandiflorum	A549 and MCF7 cells	Excessive vacuoles	Jeon et al. (2019)
JB	Natural sphingolipid	HGC-27 cells	Excessive vacuoles	Cingolani et al. (2017)
EPS	The infructescence of <i>Platycarya Strobilacea Sieb. et Zucc.</i>	Nasopharyngeal carcinoma cells	Transcription factor c-FOS and related genes may regulate methuosis	Liu et al. (2020)
Spiropachysine A	Extracted from <i>Pachysandra axillaries</i> Franch. var. styiosa (Dunn) M. Cheng	MHCC-97H cells	Ras/Rac1 signaling partially involved in methuosis	Fang et al. (2022)
TBM1	Triterpenoid saponin derived from Bolbostemma paniculatum	SW480 cells	Excessive vacuoles	Gong et al. (2018)
METH	Psychostimulant drug	Neuroblastoma cells	Activation of Rac/Rac1 induced defective lysosomal function	Nara et al. (2012)
MIPP and MOMIPP	Indole-based chalcones	U251 glioblastoma cells	Disruption of Rab5 GTPase	Robinson et al. (2012)
F14512	Polyamine-modified topoisomerase II inhibitor	A549 cells	Excessive vacuoles	Brel et al. (2011)
Compound 13	4,6-disubstituted azaindole	MDA-MB-231 etc.	Excessive vacuoles	Huang et al. (2018)
Vacquinol-1	Quinine derivative	Glioblastoma cells	Excessive vacuoles	Ahlstedt et al. (2018)
CX-4945 and CX-	Protein kinase CK2 inhibitors	HepG2 cells	Uncontrolled Rac1 activation promoted vesicle	D'Amore et al.
Compound V6	S-triazine compound	U87 glioblastoma cells	Formation of H-bonds with <sup>273</sup> R and <sup>276</sup> Y in vimentin rod domain	Zhang L. et al. (2021)

significant vacuolation in human lung cancer cells. Rac1 and Arf6 regulation was partially involved in this methuosis. Furthermore, the effects of doxorubicin and etoposide were enhanced by EKC, suggesting that EKC may be a beneficial complement to conventional chemotherapy (Liu et al., 2021). Isobavachalcone (IBC), a naturally occurring chalcone, is extracted from traditional Chinese medicines such as *Psoralea corylifolia L*. (Kuete and Sandjo, 2012). It selectively causes non-apoptotic, methuosis-like cell death in leukemia cells. Two unique features of methuosis are the activation of vacuolar-type  $H^+$ -ATPase and AKT (Yang et al., 2019).

Ursolic acid (UA), a pentacyclic triterpenoid carboxylic acid extracted from Chinese herbal medicines, exhibits significant anticancer activity by triggering p53-mediated cell cycle arrest and apoptosis (Yin et al., 2018). Sun et al. described the discovery of a UA-derived small molecule (compound 17) that caused cancer cell death *via* hyperstimulation of macropinocytosis. Cell death halted when stimulation was inhibited by the macropinocytosis inhibitor amiloride, simultaneous with methuosis (Sun et al., 2017). Platycodin D (PD), extracted from the root of *Platycodon grandiflorum*, enhances human immunity and is one of the main saponins used against tumors (Zhao et al., 2015). Jeon et al. proposed that PD-induced cell death with extreme vacuolation constituted methuosis (Jeon et al., 2019).

Jaspine B (JB), a natural sphingolipid, is isolated from marine sponges (Canals et al., 2009). It caused atypical cell death of HGC-27 cells characterized by cytoplasmic vacuoles developing in a time- and dose-dependent manner. Vacuoles seemed to form as a result of macropinocytosis, causing cytoplasmic disruption. JB joins the list of substances that cause cytoplasmic vacuolation and methuosis (Cingolani et al., 2017). An ethanolic extract of the infructescence of *Platycarya strobilacea Sieb. et Zucc.* (EPS) demonstrated potent antinasopharyngeal cancer properties *in vitro*. The production of CNE1 and CNE2 cell fusion and vacuoles, the disruption of lysosomal vesicle transit, and the activation of methuosis were all linked to the anticancer effects of EPS (Liu et al., 2020).

Spiropachysine A is a traditional Chinese steroidal alkaloid extract that is extensively used to improve blood circulation and reduce pain and inflammation. Its antitumor activity is mediated through methuosis and is not dependent on classical cell death signaling pathways. Activation of the Rac1 signaling pathway appears to be required for methuosis (Fang et al., 2022). TBM1 causes the rapid production of a large number of vesicular-like structures. The primary mechanism of TBM1-induced vacuolization is macropinocytosis, which leads to methuosis.

Thus, it is evident that natural products and their derivatives have broad application in clinical antitumor therapies mediated *via* methuosis (Gong et al., 2018).

### Synthetic Compounds

Non-natural compounds can also induce methuosis (**Table 2**). Methamphetamine (METH) primarily alters the integrity of dopamine terminal neurons and interferes with nerve conduction (Abbruscato and Trippier, 2018). When neuroblastoma cells were cultured in METH they lost their activity after macropinocytosis was activated by Ras and Rac1 (Nara et al., 2012).

Researchers reported a chalcone-like molecule, MIPP, that promoted methuosis in glioblastoma and other cancer cells. A derivative, MOMIPP, induced methuosis at low concentrations (Robinson et al., 2012). The JNK signaling pathway was involved in methuosis triggered by MOMIPP (Li et al., 2019). F14512, a polyamine-modified topoisomerase II inhibitor, is considered an effective anticancer drug candidate (Barret et al., 2008). Brel et al. showed that its anticancer effects were related to the accumulation of numerous vacuoles, as seen during methuosis (Brel et al., 2011).

The azaindole-based compound 13 caused MDA-MB-231 cells to vacuolate and die. It was cytotoxic towards a wide range of cancer cell lines, including MDA-MB-231, A375, HCT116, and MCF-7, with cell death being mediated by

methuosis (Huang et al., 2018). Vacquinol-1, a quinolone derivative, induced glioblastoma multiforme cell death by methuosis and has promising therapeutic potential. Cell death was counter-regulated by exogenous ATP, which activated the transient receptor potential cation channel, subfamily M, member 7 (TRPM7) (Ahlstedt et al., 2018).

CK2, a pleiotropic enzyme deeply involved in cell proliferation and apoptosis, is considered a promising therapeutic agent against cancer (Manni et al., 2017; Fasolato et al., 2021). In cholangiocarcinoma and other cancer cell lines, CX-4945 treatment stimulated the formation of cytosolic vacuoles. Extracellular fluid at neutral pH was seen in the vacuoles, which is characteristic of methuosis (Lertsuwan et al., 2018). CX-5011, a CK2 inhibitor related to CX-4945, acts as a CK2independent methuosis inducer with four times the potency of its parent compound and the ability to promote the creation of larger cytosolic vacuoles at low micromolar doses. The pharmacological suppression of small GTPase Rac1, siRNA-mediated downregulation of Rac1, and overexpression of the dominantnegative mutant form of Rac1 (Rac1 T17N) both reduced CX-5011's ability to induce methuosis (D'Amore et al., 2020).

Vimentin is involved in the membrane trafficking and transporting of endolysosomal vesicles (Margiotta and Bucci, 2016). Zhang et al. created a variety of novel s-triazine compounds that caused methuosis in various cancer cells. For example, compound V6 exhibited striking methuotic properties and vimentin was its specific target protein. According to molecular docking analysis, V6 can form hydrogen bonds with vimentin at <sup>273</sup>R and <sup>276</sup>Y in its rod domain (Zhang L. et al., 2021). The molecular mechanisms involved in drug-induced methuosis remain unclear and require further study. Meanwhile, other methuosis inducers remain to be discovered.

# **CONCLUSION AND PERSPECTIVES**

Tumor growth and drug resistance mediated by macropinocytosis have recently become hot topics in cancer tumor research, as illustrated by the many studies of the mechanisms, regulation, and applications of macropinocytosis. Understanding the mechanisms of macropinocytosis and its effects on tumor cells benefits the development of new therapeutic strategies. Studying drugs that promote macropinocytosis helps to improve the targeting and efficacy of macropinocytosis-dependent chemotherapeutic drugs. Anticancer medications can be combined with carriers or drugs that induce macropinocytosis. Meanwhile, inhibition of macropinocytosis cuts off an alternative pathway of cancer cell metabolism, reverses tumor resistance, and increases tumor sensitivity to anticancer drugs.

Methuosis inducers provide an alternative pathway to cancer cell death beyond programmed apoptosis and have powerful anticancer potential. Screening natural products for methuosisinducing drugs is a viable strategy, but the mechanisms connecting cell death with extreme vacuolation require greater research.

Macropinocytosis in Cancer

Understanding the relationship between tumor cells and macropinocytosis in the TME is crucial to developing antitumor strategies. Macropinocytosis in the TME promotes survival tumor and blocking microenvironmental macropinocytosis can provide additional benefits to antitumor therapies. However, most studies are concerned with macropinocytosis in cancer cells and few address macropinocytosis related to the microenvironment. In addition, macropinocytosis inhibitors inhibit the growth of tumor cells but also impede the supportive effects that the TME provides to cancer cells. Cullis et al. confirmed that drugs affected microenvironmental components through macropinocytosis and provided strategies for tumor treatment (Cullis et al., 2017). The TME is extremely complex and the mechanisms by which macropinocytosis affects the survival of cancer cells need to be elucidated. The extent to which macropinocytosis in the TME affects tumor survival, metastasis, and therapy resistance remains unclear.

Challenges and problems remain in this area. For example, not all cancer cells exhibit the phenomenon of macropinocytosis, which may limit the application of treatment methods that target macropinocytosis. Means of identifying macropinocytosis in cells are also limited. Inhibitors (e.g., EIPA and cytochalasin D) have been routinely used to demonstrate macropinocytosis, while detecting the uptake of dextran, a macropinocytosis marker, is the most commonly

### REFERENCES

- Abbruscato, T. J., and Trippier, P. C. (2018). DARK Classics in Chemical Neuroscience: Methamphetamine. ACS Chem. Neurosci. 9, 2373–2378. doi:10.1021/acschemneuro.8b00123
- Adkins, D., Ley, J., Atiq, O., Powell, S., Spanos, W. C., Gitau, M., et al. (2021). Nanoparticle Albumin-Bound Paclitaxel with Cetuximab and Carboplatin as First-Line Therapy for Recurrent or Metastatic Head and Neck Cancer: A Single-Arm, Multicenter, Phase 2 Trial. Oral Oncol. 115, 105173. doi:10.1016/j. oraloncology.2020.105173
- Ahlstedt, J., Förnvik, K., Zolfaghari, S., Kwak, D., Hammarström, L. G. J., Ernfors, P., et al. (2018). Evaluating Vacquinol-1 in Rats Carrying Glioblastoma Models RG2 and NS1. Oncotarget 9, 8391–8399. doi:10.18632/oncotarget.23842
- Amaravadi, R. K., Kimmelman, A. C., and Debnath, J. (2019). Targeting Autophagy in Cancer: Recent Advances and Future Directions. *Cancer Discov.* 9, 1167–1181. doi:10.1158/2159-8290.CD-19-0292
- Amyere, M., Payrastre, B., Krause, U., P., V. D. S.Van Der Smissen, A., and Courtoy, P. J. (2000). Constitutive Macropinocytosis in Oncogene-Transformed Fibroblasts Depends on Sequential Permanent Activation of Phosphoinositide 3-kinase and Phospholipase C. *Mol. Biol. Cell.* 11, 3453–3467. doi:10.1091/mbc.11.10.3453
- Araki, N., Egami, Y., Watanabe, Y., and Hatae, T. (2007). Phosphoinositide Metabolism during Membrane Ruffling and Macropinosome Formation in EGF-Stimulated A431 Cells. *Exp. Cell. Res.* 313, 1496–1507. doi:10.1016/j.yexcr.2007.02.012
- Avila-Ponce de León, U., Vázquez-Jiménez, A., Matadamas-Guzman, M., Pelayo, R., and Resendis-Antonio, O. (2021). Transcriptional and Microenvironmental Landscape of Macrophage Transition in Cancer: A Boolean Analysis. Front. Immunol. 12, 642842. doi:10.3389/fimmu.2021.642842
- Barret, J. M., Kruczynski, A., Vispé, S., Annereau, J. P., Brel, V., Guminski, Y., et al. (2008). F14512, a Potent Antitumor Agent Targeting Topoisomerase II Vectored into Cancer Cells via the Polyamine Transport System. *Cancer Res.* 68, 9845–9853. doi:10.1158/0008-5472.CAN-08-2748
- Bhanot, H., Young, A. M., Overmeyer, J. H., and Maltese, W. A. (2010). Induction of Nonapoptotic Cell Death by Activated Ras Requires Inverse Regulation of

used method. However, neither method specifically identifies macropinocytosis and researchers lack real-time, quantitative detection options in macropinocytosis research. Further research is required into techniques for effectively regulating macropinocytosis and related mechanisms. This review of the field highlights promising new methods and strategies for tumor treatment.

## **AUTHOR CONTRIBUTIONS**

YM, TX, and XS (23rd author) contributed to conception and design of the study. ZQ wrote the first draft of the manuscript. WL, QZ (3rd author), KK, QZ (5th author), WJ, SY, ZY, LL, XS (10th author), SR, YL, ZZ, JZ, XH, YH, LW, MM, JL, and XC wrote sections of the manuscript. All authors contributed to manuscript revision, read, and approved the submitted version.

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Rac1 and Arf6. Mol. Cancer Res. 8, 1358-1374. doi:10.1158/1541-7786.MCR-10-0090

- Brel, V., Annereau, J. P., Vispé, S., Kruczynski, A., Bailly, C., and Guilbaud, N. (2011). Cytotoxicity and Cell Death Mechanisms Induced by the Polyamine-Vectorized Anti-cancer Drug F14512 Targeting Topoisomerase II. *Biochem. Pharmacol.* 82, 1843–1852. doi:10.1016/j.bcp.2011.08.028
- Buckley, C. M., Gopaldass, N., Bosmani, C., Johnston, S. A., Soldati, T., Insall, R. H., et al. (2016). WASH Drives Early Recycling from Macropinosomes and Phagosomes to Maintain Surface Phagocytic Receptors. *Proc. Natl. Acad. Sci. U. S. A.* 113, E5906–E5915. doi:10.1073/pnas.1524532113
- Canals, D., Mormeneo, D., Fabriàs, G., Llebaria, A., Casas, J., and Delgado, A. (2009). Synthesis and Biological Properties of Pachastrissamine (Jaspine B) and Diastereoisomeric Jaspines. *Bioorg Med. Chem.* 17, 235–241. doi:10.1016/j.bmc. 2008.11.026
- Cang, C., Zhou, Y., Navarro, B., Seo, Y. J., Aranda, K., Shi, L., et al. (2013). mTOR Regulates Lysosomal ATP-Sensitive Two-Pore Na(+) Channels to Adapt to Metabolic State. *Cell*. 152, 778–790. doi:10.1016/j.cell.2013.01.023
- Canton, J. (2018). Macropinocytosis: New Insights into its Underappreciated Role in Innate Immune Cell Surveillance. Front. Immunol. 9, 2286. doi:10.3389/ fimmu.2018.02286
- Cao, Y., Wang, X., Li, Y., Evers, M., Zhang, H., and Chen, X. (2019). Extracellular and Macropinocytosis Internalized ATP Work Together to Induce Epithelial-Mesenchymal Transition and Other Early Metastatic Activities in Lung Cancer. *Cancer Cell. Int.* 19, 254. doi:10.1186/s12935-019-0973-0
- Chan, W. K., Lorenzi, P. L., Anishkin, A., Purwaha, P., Rogers, D. M., Sukharev, S., et al. (2014). The Glutaminase Activity of L-Asparaginase Is Not Required for Anticancer Activity against ASNS-Negative Cells. *Blood* 123, 3596–3606. doi:10.1182/blood-2013-10-535112
- Charpentier, J. C., Chen, D., Lapinski, P. E., Turner, J., Grigorova, I., Swanson, J. A., et al. (2020). Macropinocytosis Drives T Cell Growth by Sustaining the Activation of mTORC1. *Nat. Commun.* 11, 180. doi:10.1038/s41467-019-13997-3
- Chern, Y. J., and Tai, I. T. (2020). Adaptive Response of Resistant Cancer Cells to Chemotherapy. *Cancer Biol. Med.* 17, 842–863. doi:10.20892/j.issn.2095-3941. 2020.0005

- Chiasson-MacKenzie, C., Morris, Z. S., Liu, C. H., Bradford, W. B., Koorman, T., and McClatchey, A. I. (2018). Merlin/ERM Proteins Regulate Growth Factor-Induced Macropinocytosis and Receptor Recycling by Organizing the Plasma Membrane:cytoskeleton Interface. *Genes. Dev.* 32, 1201–1214. doi:10.1101/gad. 317354.118
- Choi, D., Montermini, L., Meehan, B., Lazaris, A., Metrakos, P., and Rak, J. (2021). Oncogenic RAS Drives the CRAF-dependent Extracellular Vesicle Uptake Mechanism Coupled with Metastasis. *J. Extracell. Vesicles* 10, e12091. doi:10.1002/jev2.12091
- Cingolani, F., Simbari, F., Abad, J. L., Casasampere, M., Fabrias, G., Futerman, A. H., et al. (2017). Jaspine B Induces Nonapoptotic Cell Death in Gastric Cancer Cells Independently of its Inhibition of Ceramide Synthase. J. Lipid Res. 58, 1500–1513. doi:10.1194/jlr.M072611
- Commisso, C., Davidson, S. M., Soydaner-Azeloglu, R. G., Parker, S. J., Kamphorst, J. J., Hackett, S., et al. (2013). Macropinocytosis of Protein Is an Amino Acid Supply Route in Ras-Transformed Cells. *Nature* 497, 633–637. doi:10.1038/ nature12138
- Costa Verdera, H., Gitz-Francois, J. J., Schiffelers, R. M., and Vader, P. (2017). Cellular Uptake of Extracellular Vesicles Is Mediated by Clathrin-independent Endocytosis and Macropinocytosis. *J. Control Release* 266, 100–108. doi:10. 1016/j.jconrel.2017.09.019
- Cui, Y., Cai, J., Wang, W., and Wang, S. (2021). Regulatory Effects of Histone Deacetylase Inhibitors on Myeloid-Derived Suppressor Cells. *Front. Immunol.* 12, 690207. doi:10.3389/fimmu.2021.690207
- Cullis, J., Siolas, D., Avanzi, A., Barui, S., Maitra, A., and Bar-Sagi, D. (2017). Macropinocytosis of Nab-Paclitaxel Drives Macrophage Activation in Pancreatic Cancer. *Cancer Immunol. Res.* 5, 182–190. doi:10.1158/2326-6066.CIR-16-0125
- D'Amore, C., Moro, E., Borgo, C., Itami, K., Hirota, T., Pinna, L. A., et al. (2020). "Janus" Efficacy of CX-5011: CK2 Inhibition and Methuosis Induction by Independent Mechanisms. *Biochim. Biophys. Acta Mol. Cell. Res.* 1867, 118807. doi:10.1016/j.bbamcr.2020.118807
- Dai, M., Yan, G., Wang, N., Daliah, G., Edick, A. M., Poulet, S., et al. (2021). In Vivo genome-wide CRISPR Screen Reveals Breast Cancer Vulnerabilities and Synergistic mTOR/Hippo Targeted Combination Therapy. *Nat. Commun.* 12, 3055. doi:10.1038/s41467-021-23316-4
- Dayam, R. M., Saric, A., Shilliday, R. E., and Botelho, R. J. (2015). The Phosphoinositide-Gated Lysosomal Ca(2+) Channel, TRPML1, Is Required for Phagosome Maturation. *Traffic* 16, 1010–1026. doi:10.1111/tra.12303
- De Veirman, K., Wang, J., Xu, S., Leleu, X., Himpe, E., Maes, K., et al. (2016). Induction of miR-146a by Multiple Myeloma Cells in Mesenchymal Stromal Cells Stimulates Their Pro-tumoral Activity. *Cancer Lett.* 377, 17–24. doi:10. 1016/j.canlet.2016.04.024
- Dendo, K., Yugawa, T., Nakahara, T., Ohno, S. I., Goshima, N., Arakawa, H., et al. (2018). Induction of Non-apoptotic Programmed Cell Death by Oncogenic RAS in Human Epithelial Cells and its Suppression by MYC Overexpression. *Carcinogenesis* 39, 202–213. doi:10.1093/carcin/bgx124
- Dolat, L., and Spiliotis, E. T. (2016). Septins Promote Macropinosome Maturation and Traffic to the Lysosome by Facilitating Membrane Fusion. J. Cell. Biol. 214, 517–527. doi:10.1083/jcb.201603030
- Egami, Y., Taguchi, T., Maekawa, M., Arai, H., and Araki, N. (2014). Small GTPases and Phosphoinositides in the Regulatory Mechanisms of Macropinosome Formation and Maturation. *Front. Physiol.* 5, 374. doi:10. 3389/fphys.2014.00374
- Falcone, I., Conciatori, F., Bazzichetto, C., Bria, E., Carbognin, L., Malaguti, P., et al. (2020). AXL Receptor in Breast Cancer: Molecular Involvement and Therapeutic Limitations. *Int. J. Mol. Sci.* 21, 8419. doi:10.3390/ijms21228419
- Fang, Y., Zhong, T., Yang, L., Luo, F., Li, Q., Wang, D., et al. (2022). Spiropachysine A Suppresses Hepatocellular Carcinoma Proliferation by Inducing Methuosis In Vitro and In Vivo. *Phytomedicine* 102, 154151. doi:10.1016/j.phymed.2022. 154151
- Fasolato, L., Magro, M., Cozza, G., Sbarra, F., Molinari, S., Novelli, E., et al. (2021). An Iron Shield to Protect Epigallocatehin-3-Gallate from Degradation: Multifunctional Self-Assembled Iron Oxide Nanocarrier Enhances Protein Kinase CK2 Intracellular Targeting and Inhibition. *Pharmaceutics* 13, 1266. doi:10.3390/pharmaceutics13081266
- Freeman, S. A., Uderhardt, S., Saric, A., Collins, R. F., Buckley, C. M., Mylvaganam, S., et al. (2020). Lipid-gated Monovalent Ion Fluxes Regulate Endocytic Traffic

and Support Immune Surveillance. *Science* 367, 301–305. doi:10.1126/science. aaw9544

- Fuh, K. F., Shepherd, R. D., Withell, J. S., Kooistra, B. K., and Rinker, K. D. (2021). Fluid Flow Exposure Promotes Epithelial-To-Mesenchymal Transition and Adhesion of Breast Cancer Cells to Endothelial Cells. *Breast Cancer Res.* 23, 97. doi:10.1186/s13058-021-01473-0
- Gong, X., Sun, R., Gao, Z., Han, W., Liu, Y., Zhao, L., et al. (2018). Tubeimoside 1 Acts as a Chemotherapeutic Synergist via Stimulating Macropinocytosis. *Front. Pharmacol.* 9, 1044. doi:10.3389/fphar.2018.01044
- Ha, K. D., Bidlingmaier, S. M., and Liu, B. (2016). Macropinocytosis Exploitation by Cancers and Cancer Therapeutics. *Front. Physiol.* 7, 381. doi:10.3389/fphys. 2016.00381
- Hamilton, N., Kerr, M. C., Burrage, K., and Teasdale, R. D. (2007). Analyzing Real-Time Video Microscopy: The Dynamics and Geometry of Vesicles and Tubules in Endocytosis. *Curr. Protoc. Cell. Biol.* 35. doi:10.1002/0471143030.cb0416s35
- Hanada, K., Kawada, K., Nishikawa, G., Toda, K., Maekawa, H., Nishikawa, Y., et al. (2021). Dual Blockade of Macropinocytosis and Asparagine Bioavailability Shows Synergistic Anti-tumor Effects on KRAS-Mutant Colorectal Cancer. *Cancer Lett.* 522, 129–141. doi:10.1016/j.canlet.2021.09.023
- Hesketh, G. G., Papazotos, F., Pawling, J., Rajendran, D., Knight, J. D. R., Martinez, S., et al. (2020). The GATOR-Rag GTPase Pathway Inhibits mTORC1 Activation by Lysosome-Derived Amino Acids. *Science* 370 (6514), 351–356. doi:10.1126/science.aaz0863
- Hetzenecker, S., Helenius, A., and Krzyzaniak, M. A. (2016). HCMV Induces Macropinocytosis for Host Cell Entry in Fibroblasts. *Traffic* 17, 351–368. doi:10.1111/tra.12355
- Hobbs, G. A., Baker, N. M., Miermont, A. M., Thurman, R. D., Pierobon, M., Tran, T. H., et al. (2020). Atypical KRASG12R Mutant Is Impaired in PI3K Signaling and Macropinocytosis in Pancreatic Cancer. *Cancer Discov.* 10, 104–123. doi:10.1158/2159-8290.CD-19-1006
- Hodakoski, C., Hopkins, B. D., Zhang, G., Su, T., Cheng, Z., Morris, R., et al. (2019). Rac-Mediated Macropinocytosis of Extracellular Protein Promotes Glucose Independence in Non-small Cell Lung Cancer. *Cancers (Basel)* 11, 37. doi:10. 3390/cancers11010037
- Holohan, C., Van Schaeybroeck, S., Longley, D. B., and Johnston, P. G. (2013). Cancer Drug Resistance: an Evolving Paradigm. *Nat. Rev. Cancer* 13, 714–726. doi:10.1038/nrc3599
- Hou, P., Ma, X., Yang, Z., Zhang, Q., Wu, C. J., Li, J., et al. (2021). USP21 Deubiquitinase Elevates Macropinocytosis to Enable Oncogenic KRAS Bypass in Pancreatic Cancer. *Genes. Dev.* 35, 1327–1332. doi:10.1101/gad.348787.121
- Huang, W., Sun, X., Li, Y., He, Z., Li, L., Deng, Z., et al. (2018). Discovery and Identification of Small Molecules as Methuosis Inducers with In Vivo Antitumor Activities. J. Med. Chem. 61, 5424–5434. doi:10.1021/acs. jmedchem.8b00753
- Hussein, N. A., Malla, S., Pasternak, M. A., Terrero, D., Brown, N. G., Ashby, C. R., Jr., et al. (2021). The Role of Endolysosomal Trafficking in Anticancer Drug Resistance. *Drug Resist Updat* 57, 100769. doi:10.1016/j.drup.2021.100769
- Jayashankar, V., and Edinger, A. L. (2020). Macropinocytosis Confers Resistance to Therapies Targeting Cancer Anabolism. Nat. Commun. 11, 1121. doi:10.1038/ s41467-020-14928-3
- Jeon, D., Kim, S. W., and Kim, H. S. (2019). Platycodin D, a Bioactive Component of Platycodon Grandiflorum, Induces Cancer Cell Death Associated with Extreme Vacuolation. *Anim. Cells Syst. Seoul.* 23, 118–127. doi:10.1080/ 19768354.2019.1588163
- Jiao, K., Zhen, J., Wu, M., Teng, M., Yang, K., Zhou, Q., et al. (2020). 27-Hydroxycholesterol-induced EndMT Acts via STAT3 Signaling to Promote Breast Cancer Cell Migration by Altering the Tumor Microenvironment. *Cancer Biol. Med.* 17, 88–100. doi:10.20892/j.issn.2095-3941.2019.0262
- Kamerkar, S., LeBleu, V. S., Sugimoto, H., Yang, S., Ruivo, C. F., Melo, S. A., et al. (2017). Exosomes Facilitate Therapeutic Targeting of Oncogenic KRAS in Pancreatic Cancer. *Nature* 546, 498–503. doi:10.1038/nature22341
- Kasitinon, S. Y., Eskiocak, U., Martin, M., Bezwada, D., Khivansara, V., Tasdogan, A., et al. (2019). TRPML1 Promotes Protein Homeostasis in Melanoma Cells by Negatively Regulating MAPK and mTORC1 Signaling. *Cell. Rep.* 28, 2293–e9. doi:10.1016/j.celrep.2019.07.086
- Kim, H. J., and Bae, S. C. (2011). Histone Deacetylase Inhibitors: Molecular Mechanisms of Action and Clinical Trials as Anti-cancer Drugs. Am. J. Transl. Res. 3, 166–179.

- Kim, S. M., Nguyen, T. T., Ravi, A., Kubiniok, P., Finicle, B. T., Jayashankar, V., et al. (2018). PTEN Deficiency and AMPK Activation Promote Nutrient Scavenging and Anabolism in Prostate Cancer Cells. *Cancer Discov.* 8, 866–883. doi:10.1158/2159-8290.CD-17-1215
- King, J. S., and Kay, R. R. (2019). The Origins and Evolution of Macropinocytosis. *Philos. Trans. R. Soc. Lond B Biol. Sci.* 374, 20180158. doi:10.1098/rstb.2018. 0158
- King, J. S., and Smythe, E. (2020). Water Loss Regulates Cell and Vesicle Volume. Science 367, 246–247. doi:10.1126/science.aba3623
- Kitazawa, S., Nishizawa, S., Nakagawa, H., Funata, M., Nishimura, K., Soga, T., et al. (2017). Cancer with Low Cathepsin D Levels Is Susceptible to Vacuolar (H+)-ATPase Inhibition. *Cancer Sci.* 108, 1185–1193. doi:10.1111/cas.13240
- Klein, C. A. (2020). Cancer Progression and the Invisible Phase of Metastatic Colonization. Nat. Rev. Cancer 20, 681–694. doi:10.1038/s41568-020-00300-6
- Koivusalo, M., Welch, C., Hayashi, H., Scott, C. C., Kim, M., Alexander, T., et al. (2010). Amiloride Inhibits Macropinocytosis by Lowering Submembranous pH and Preventing Rac1 and Cdc42 Signaling. *J. Cell. Biol.* 188, 547–563. doi:10. 1083/jcb.200908086
- Krishna, S., Palm, W., Lee, Y., Yang, W., Bandyopadhyay, U., Xu, H., et al. (2016). PIKfyve Regulates Vacuole Maturation and Nutrient Recovery Following Engulfment. *Dev. Cell.* 38, 536–547. doi:10.1016/j.devcel.2016.08.001
- Kuete, V., and Sandjo, L. P. (2012). Isobavachalcone: an Overview. Chin. J. Integr. Med. 18, 543–547. doi:10.1007/s11655-012-1142-7
- Łagiewka, J., Girek, T., and Ciesielski, W. (2021). Cyclodextrins-Peptides/Proteins Conjugates: Synthesis, Properties and Applications. *Polymers* 13, 1759. doi:10. 3390/polym13111759
- Lee, S. W., Zhang, Y., Jung, M., Cruz, N., Alas, B., and Commisso, C. (2019). EGFRpak Signaling Selectively Regulates Glutamine Deprivation-Induced Macropinocytosis. Dev. Cell. 50, 381–e5. doi:10.1016/j.devcel.2019.05.043
- Lee, S. Y., Ju, M. K., Jeon, H. M., Jeong, E. K., Lee, Y. J., Kim, C. H., et al. (2018). Regulation of Tumor Progression by Programmed Necrosis. Oxid. Med. Cell. Longev. 2018, 3537471. doi:10.1155/2018/3537471
- Lertsuwan, J., Lertsuwan, K., Sawasdichai, A., Tasnawijitwong, N., Lee, K. Y., Kitchen, P., et al. (2018). CX-4945 Induces Methuosis in Cholangiocarcinoma Cell Lines by a CK2-independent Mechanism. *Cancers (Basel)* 10, 283. doi:10. 3390/cancers10090283
- Li, C., Macdonald, J. I., Hryciw, T., and Meakin, S. O. (2010). Nerve Growth Factor Activation of the TrkA Receptor Induces Cell Death, by Macropinocytosis, in Medulloblastoma Daoy Cells. J. Neurochem. 112, 882–899. doi:10.1111/j.1471-4159.2009.06507.x
- Li, C., MacDonald, J. I., Talebian, A., Leuenberger, J., Seah, C., Pasternak, S. H., et al. (2016). Unravelling the Mechanism of TrkA-Induced Cell Death by Macropinocytosis in Medulloblastoma Daoy Cells. *Mol. Cell. Biol.* 36, 2596–2611. doi:10.1128/MCB.00255-16
- Li, P., Lu, M., Shi, J., Gong, Z., Hua, L., Li, Q., et al. (2020). Lung Mesenchymal Cells Elicit Lipid Storage in Neutrophils that Fuel Breast Cancer Lung Metastasis. *Nat. Immunol.* 21, 1444–1455. doi:10.1038/s41590-020-0783-5
- Li, R., Ng, T. S. C., Wang, S. J., Prytyskach, M., Rodell, C. B., Mikula, H., et al. (2021). Therapeutically Reprogrammed Nutrient Signalling Enhances Nanoparticulate Albumin Bound Drug Uptake and Efficacy in KRAS-Mutant Cancer. *Nat. Nanotechnol.* 16, 830–839. doi:10.1038/s41565-021-00897-1
- Li, Z., Mbah, N. E., Overmeyer, J. H., Sarver, J. G., George, S., Trabbic, C. J., et al. (2019). The JNK Signaling Pathway Plays a Key Role in Methuosis (Nonapoptotic Cell Death) Induced by MOMIPP in Glioblastoma. *BMC Cancer* 19, 77. doi:10.1186/s12885-019-5288-y
- Lin, X. P., Mintern, J. D., and Gleeson, P. A. (2020). Macropinocytosis in Different Cell Types: Similarities and Differences. *Membr. (Basel)* 10, 177. doi:10.3390/ membranes10080177
- Liu, H., Sun, M., Liu, Z., Kong, C., Kong, W., Ye, J., et al. (2019). KRAS-Enhanced Macropinocytosis and Reduced FcRn-Mediated Recycling Sensitize Pancreatic Cancer to Albumin-Conjugated Drugs. J. Control Release 296, 40–53. doi:10. 1016/j.jconrel.2019.01.014
- Liu, J., Ying, M., Wu, B., and Fu, C. (2020). Ethanol Extract of the Infructescence of Platycarya strobilacea Sieb. et Zucc. Induces Methuosis of Human Nasopharyngeal Carcinoma Cells. *Evid. Based Complement. Altern. Med.* 2020, 2760979. doi:10.1155/2020/2760979

- Liu, X., Wang, S., Zheng, H., Liu, Q., Shen, T., Wang, X., et al. (2021). Epimedokoreanin C, a Prenylated Flavonoid Isolated from Epimedium Koreanum, Induces Non-apoptotic Cell Death with the Characteristics of Methuosis in Lung Cancer Cells. Am. J. Cancer Res. 11, 3496–3514.
- Liu, Z., and Roche, P. A. (2015). Macropinocytosis in Phagocytes: Regulation of MHC Class-II-Restricted Antigen Presentation in Dendritic Cells. Front. Physiol. 6, 1. doi:10.3389/fphys.2015.00001
- Ma, L., Han, X., Gu, J., Li, J., Lou, W., Jin, C., et al. (2020). The Physiological Characteristics of the Basal Microvilli Microvessels in Pancreatic Cancers. *Cancer Med.* 9, 5535–5545. doi:10.1002/cam4.3177
- Maltese, W. A., and Overmeyer, J. H. (2015). Non-apoptotic Cell Death Associated with Perturbations of Macropinocytosis. *Front. Physiol.* 6, 38. doi:10.3389/ fphys.2015.00038
- Manni, S., Carrino, M., and Piazza, F. (2017). Role of Protein Kinases CK1α and CK2 in Multiple Myeloma: Regulation of Pivotal Survival and Stress-Managing Pathways. J. Hematol. Oncol. 10, 157. doi:10.1186/s13045-017-0529-5
- Margiotta, A., and Bucci, C. (2016). Role of Intermediate Filaments in Vesicular Traffic. Cells 5, 20. doi:10.3390/cells5020020
- Marques, P. E., Grinstein, S., and Freeman, S. A. (2017). SnapShot: Macropinocytosis. Cell. 169, 766–e1. doi:10.1016/j.cell.2017.04.031
- Michalopoulou, E., Auciello, F. R., Bulusu, V., Strachan, D., Campbell, A. D., Tait-Mulder, J., et al. (2020). Macropinocytosis Renders a Subset of Pancreatic Tumor Cells Resistant to mTOR Inhibition. *Cell. Rep.* 30, 2729–e4. doi:10.1016/ j.celrep.2020.01.080
- Mishra, R., Haldar, S., Placencio, V., Madhav, A., Rohena-Rivera, K., Agarwal, P., et al. (2018). Stromal Epigenetic Alterations Drive Metabolic and Neuroendocrine Prostate Cancer Reprogramming. J. Clin. Investig. 128, 4472–4484. doi:10.1172/JCI99397
- Moreau, H. D., Blanch-Mercader, C., Attia, R., Maurin, M., Alraies, Z., Sanséau, D., et al. (2019). Macropinocytosis Overcomes Directional Bias in Dendritic Cells Due to Hydraulic Resistance and Facilitates Space Exploration. *Dev. Cell.* 49, 171–e5. doi:10.1016/j.devcel.2019.03.024
- Muley, H., Fadó, R., Rodríguez-Rodríguez, R., and Casals, N. (2020). Drug Uptake-Based Chemoresistance in Breast Cancer Treatment. *Biochem. Pharmacol.* 177, 113959. doi:10.1016/j.bcp.2020.113959
- Nakase, I., Noguchi, K., Aoki, A., Takatani-Nakase, T., Fujii, I., and Futaki, S. (2017). Arginine-rich Cell-Penetrating Peptide-Modified Extracellular Vesicles for Active Macropinocytosis Induction and Efficient Intracellular Delivery. *Sci. Rep.* 7, 1991. doi:10.1038/s41598-017-02014-6
- Nara, A., Aki, T., Funakoshi, T., Unuma, K., and Uemura, K. (2012). Hyperstimulation of Macropinocytosis Leads to Lysosomal Dysfunction during Exposure to Methamphetamine in SH-Sy5y Cells. *Brain Res.* 1466, 1–14. doi:10.1016/j.brainres.2012.05.017
- Palm, W. (2019). Metabolic Functions of Macropinocytosis. Philos. Trans. R. Soc. Lond B Biol. Sci. 374, 20180285. doi:10.1098/rstb.2018.0285
- Palm, W., Park, Y., Wright, K., Pavlova, N. N., Tuveson, D. A., and Thompson, C. B. (2015). The Utilization of Extracellular Proteins as Nutrients Is Suppressed by mTORC1. *Cell.* 162, 259–270. doi:10.1016/j.cell.2015.06.017
- Puccini, J., Badgley, M. A., and Bar-Sagi, D. (2022). Exploiting Cancer's Drinking Problem: Regulation and Therapeutic Potential of Macropinocytosis. *Trends Cancer* 8, 54–64. doi:10.1016/j.trecan.2021.09.004
- Qin, T., Liu, Z., Wang, J., Xia, J., Liu, S., Jia, Y., et al. (2020). Anlotinib Suppresses Lymphangiogenesis and Lymphatic Metastasis in Lung Adenocarcinoma through a Process Potentially Involving VEGFR-3 Signaling. *Cancer Biol. Med.* 17, 753–767. doi:10.20892/j.issn.2095-3941.2020.0024
- Qin, X., Zhang, Y., He, Y., Chen, K., Zhang, Y., Li, P., et al. (2021). Shear Stress Triggered Circular Dorsal Ruffles Formation to Facilitate Cancer Cell Migration. Arch. Biochem. Biophys. 709, 108967. doi:10.1016/j.abb.2021.108967
- Ramirez, C., Hauser, A. D., Vucic, E. A., and Bar-Sagi, D. (2019). Plasma Membrane V-ATPase Controls Oncogenic RAS-Induced Macropinocytosis. *Nature* 576, 477–481. doi:10.1038/s41586-019-1831-x
- Recouvreux, M. V., and Commisso, C. (2017). Macropinocytosis: A Metabolic Adaptation to Nutrient Stress in Cancer. Front. Endocrinol. (Lausanne) 8, 261. doi:10.3389/fendo.2017.00261
- Redka, D. S., Gütschow, M., Grinstein, S., and Canton, J. (2018). Differential Ability of Proinflammatory and Anti-inflammatory Macrophages to Perform Macropinocytosis. *Mol. Biol. Cell.* 29, 53–65. doi:10.1091/mbc.E17-06-0419

- Ritter, M., Bresgen, N., and Kerschbaum, H. H. (2021). From Pinocytosis to Methuosis-Fluid Consumption as a Risk Factor for Cell Death. *Front. Cell. Dev.*
- Biol. 9, 651982. doi:10.3389/fcell.2021.651982
  Robinson, M. W., Overmeyer, J. H., Young, A. M., Erhardt, P. W., and Maltese, W. A. (2012). Synthesis and Evaluation of Indole-Based Chalcones as Inducers of Methuosis, a Novel Type of Nonapoptotic Cell Death. J. Med. Chem. 55, 1940–1956. doi:10.1021/jm201006x
- Rolver, M. G., Elingaard-Larsen, L. O., Andersen, A. P., Counillon, L., and Pedersen, S. F. (2020). Pyrazine Ring-Based Na+/H+ Exchanger (NHE) Inhibitors Potently Inhibit Cancer Cell Growth in 3D Culture, Independent of NHE1. Sci. Rep. 10, 5800. doi:10.1038/s41598-020-62430-z
- Schink, K. O., Tan, K. W., Spangenberg, H., Martorana, D., Sneeggen, M., Stévenin, V., et al. (2021). The Phosphoinositide Coincidence Detector Phafin2 Promotes Macropinocytosis by Coordinating Actin Organisation at Forming Macropinosomes. *Nat. Commun.* 12, 6577. doi:10.1038/s41467-021-26775-x
- Sipos, A., Kim, K. J., Sioutas, C., and Crandall, E. D. (2019). Evidence for Nanoparticle-Induced Lysosomal Dysfunction in Lung Adenocarcinoma (A549) Cells. Int. J. Mol. Sci. 20, 5253. doi:10.3390/ijms20215253
- Song, S., Xia, X., Qi, J., Hu, X., Chen, Q., Liu, J., et al. (2021). Silmitasertib-induced Macropinocytosis Promoting DDP Intracellular Uptake to Enhance Cell Apoptosis in Oral Squamous Cell Carcinoma. *Drug Deliv.* 28, 2480–2494. doi:10.1080/10717544.2021.2000677
- Song, S., Zhang, Y., Ding, T., Ji, N., and Zhao, H. (2020). The Dual Role of Macropinocytosis in Cancers: Promoting Growth and Inducing Methuosis to Participate in Anticancer Therapies as Targets. *Front. Oncol.* 10, 570108. doi:10. 3389/fonc.2020.570108
- Stockinger, W., Zhang, S. C., Trivedi, V., Jarzylo, L. A., Shieh, E. C., Lane, W. S., et al. (2006). Differential Requirements for Actin Polymerization, Calmodulin, and Ca2+ Define Distinct Stages of Lysosome/phagosome Targeting. *Mol. Biol. Cell.* 17, 1697–1710. doi:10.1091/mbc.e05-12-1140
- Su, H., Yang, F., Fu, R., Li, X., French, R., Mose, E., et al. (2021). Cancer Cells Escape Autophagy Inhibition via NRF2-Induced Macropinocytosis. *Cancer Cell*. 39, 678–e11. doi:10.1016/j.ccell.2021.02.016
- Sun, L., Li, B., Su, X., Chen, G., Li, Y., Yu, L., et al. (2017). An Ursolic Acid Derived Small Molecule Triggers Cancer Cell Death through Hyperstimulation of Macropinocytosis. J. Med. Chem. 60, 6638–6648. doi:10.1021/acs.jmedchem.7b00592
- Tian, T., Zhu, Y. L., Zhou, Y. Y., Liang, G. F., Wang, Y. Y., Hu, F. H., et al. (2014). Exosome Uptake through Clathrin-Mediated Endocytosis and Macropinocytosis and Mediating miR-21 Delivery. J. Biol. Chem. 289, 22258–22267. doi:10.1074/jbc.M114.588046
- Toh, W. H., Louber, J., Mahmoud, I. S., Chia, J., Bass, G. T., Dower, S. K., et al. (2019). FcRn Mediates Fast Recycling of Endocytosed Albumin and IgG from Early Macropinosomes in Primary Macrophages. J. Cell. Sci. 133, jcs235416. doi:10.1242/jcs.235416
- Wang, L., Mo, S., Li, X., He, Y., and Yang, J. (2020). Single-cell RNA-Seq Reveals the Immune Escape and Drug Resistance Mechanisms of Mantle Cell Lymphoma. *Cancer Biol. Med.* 17, 726–739. doi:10.20892/j.issn.2095-3941. 2020.0073
- Weerasekara, V. K., Patra, K. C., and Bardeesy, N. (2019). EGFR Pathway Links Amino Acid Levels and Induction of Macropinocytosis. *Dev. Cell.* 50, 261–263. doi:10.1016/j.devcel.2019.07.020
- Williams, T. D., Paschke, P. I., and Kay, R. R. (2019). Function of Small GTPases in Dictyostelium Macropinocytosis. *Philos. Trans. R. Soc. Lond B Biol. Sci.* 374, 20180150. doi:10.1098/rstb.2018.0150
- Xiao, Y., Rabien, A., Buschow, R., Amtislavskiy, V., Busch, J., Kilic, E., et al. (2020). Endocytosis-Mediated Replenishment of Amino Acids Favors Cancer Cell Proliferation and Survival in Chromophobe Renal Cell Carcinoma. *Cancer Res.* 80, 5491–5501. doi:10.1158/0008-5472.CAN-20-1998
- Yamazaki, S., Higuchi, Y., Ishibashi, M., Hashimoto, H., Yasunaga, M., Matsumura, Y., et al. (2018). Collagen Type I Induces EGFR-TKI Resistance in EGFR-Mutated Cancer Cells by mTOR Activation through Akt-independent Pathway. *Cancer Sci.* 109, 2063–2073. doi:10.1111/cas.13624
- Yamazaki, S., Su, Y., Maruyama, A., Makinoshima, H., Suzuki, J., Tsuboi, M., et al. (2020). Uptake of Collagen Type I via Macropinocytosis Cause mTOR Activation and Anti-cancer Drug Resistance. *Biochem. Biophys. Res. Commun.* 526, 191–198. doi:10.1016/j.bbrc.2020.03.067

- Yang, L., Song, L., Zhao, S., Ma, C., Wu, D., and Wu, Y. L. (2019). Isobavachalcone Reveals Novel Characteristics of Methuosis-like Cell Death in Leukemia Cells. *Chem. Biol. Interact.* 304, 131–138. doi:10.1016/j.cbi.2019.03.011
- Yao, W., Rose, J. L., Wang, W., Seth, S., Jiang, H., Taguchi, A., et al. (2019). Syndecan 1 Is a Critical Mediator of Macropinocytosis in Pancreatic Cancer. *Nature* 568, 410–414. doi:10.1038/s41586-019-1062-1
- Yin, R., Li, T., Tian, J. X., Xi, P., and Liu, R. H. (2018). Ursolic Acid, a Potential Anticancer Compound for Breast Cancer Therapy. *Crit. Rev. Food Sci. Nutr.* 58, 568–574. doi:10.1080/10408398.2016.1203755
- Zdżalik-Bielecka, D., Poświata, A., Kozik, K., Jastrzębski, K., Schink, K. O., Brewińska-Olchowik, M., et al. (2021). The GAS6-AXL Signaling Pathway Triggers Actin Remodeling that Drives Membrane Ruffling, Macropinocytosis, and Cancer-Cell Invasion. *Proc. Natl. Acad. Sci. U.S.A.* 118, e2024596118. doi:10.1073/pnas.2024596118
- Zhang, L., Qu, Z., Wu, J., Yao, S., Zhang, Q., Zhang, T., et al. (2021). SARs of a Novel Series of S-Triazine Compounds Targeting Vimentin to Induce Methuotic Phenotype. *Eur. J. Med. Chem.* 214, 113188. doi:10.1016/j.ejmech. 2021.113188
- Zhang, X., Sai, B., Wang, F., Wang, L., Wang, Y., Zheng, L., et al. (2019). Hypoxic BMSC-Derived Exosomal miRNAs Promote Metastasis of Lung Cancer Cells via STAT3-Induced EMT. *Mol. Cancer* 18, 40. doi:10.1186/s12943-019-0959-5
- Zhang, Y., Recouvreux, M. V., Jung, M., Galenkamp, K. M. O., Li, Y., Zagnitko, O., et al. (2021). Macropinocytosis in Cancer-Associated Fibroblasts Is Dependent on CaMKK2/ARHGEF2 Signaling and Functions to Support Tumor and Stromal Cell Fitness. *Cancer Discov.* 11, 1808–1825. doi:10.1158/2159-8290. CD-20-0119
- Zhao, H., Yang, L., Baddour, J., Achreja, A., Bernard, V., Moss, T., et al. (2016). Tumor Microenvironment Derived Exosomes Pleiotropically Modulate Cancer Cell Metabolism. *Elife* 5, e10250. doi:10.7554/eLife.10250
- Zhao, R., Chen, M., Jiang, Z., Zhao, F., Xi, B., Zhang, X., et al. (2015). Platycodin-D Induced Autophagy in Non-small Cell Lung Cancer Cells via PI3K/Akt/mTOR and MAPK Signaling Pathways. J. Cancer 6, 623–631. doi:10.7150/jca.11291
- Zhao, X., Yongchun, Z., Qian, H., Sanhui, G., Jie, L., Hong, Y., et al. (2020). Identification of a Potential Tumor Suppressor Gene, UBL3, in Non-small Cell Lung Cancer. *Cancer Biol. Med.* 17, 76–87. doi:10.20892/j.issn.2095-3941.2019. 0279
- Zheng, Y., Tu, C., Zhang, J., and Wang, J. (2019). Inhibition of Multiple Myelomaderived E-xosomes U-ptake S-uppresses the F-unctional R-esponse in B-one M-arrow S-tromal C-ell. Int. J. Oncol. 54, 1061–1070. doi:10.3892/ijo.2019.4685
- Zhu, B., Shang, B., Li, Y., and Zhen, Y. (2016). Inhibition of Histone Deacetylases by Trans-cinnamic Acid and its Antitumor Effect against Colon Cancer Xenografts in Athymic Mice. *Mol. Med. Rep.* 13, 4159–4166. doi:10.3892/ mmr.2016.5041
- Zhu, B. Y., Shang, B. Y., Du, Y., Li, Y., Li, L., Xu, X. D., et al. (2017). A New HDAC Inhibitor Cinnamoylphenazine Shows Antitumor Activity in Association with Intensive Macropinocytosis. *Oncotarget* 8, 14748–14758. doi:10.18632/ oncotarget.14714

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